S1

## **Supporting Information**

## RNA Aptamer-Based Electrochemical Biosensor for Selective and Label-Free Analysis of Dopamine

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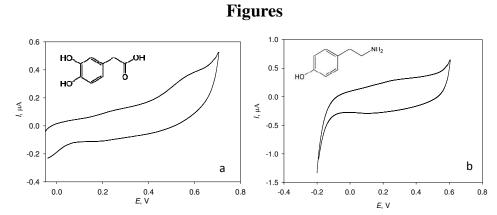
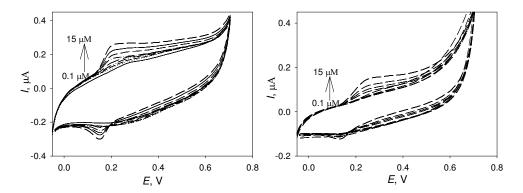


Figure S1. Representative CVs of 30 μM (a) DOPAC and (b) tyramine at Au electrodes, potential scan rate 0.1 V s<sup>-1</sup>.



**Figure S2.** Representative CVs of dopamine at (a) bare Au, (b) cysteamine-modified Au electrodes, dopamine concentrations: 0, 0.1, 0.5, 1, 2, 5, 7, 10, and  $15 \,\mu\text{M}$ , pH 7.4, the potential scan rate is  $50 \,\text{mV s}^{-1}$ .

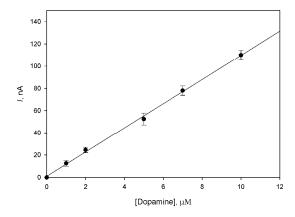
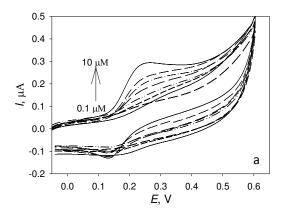
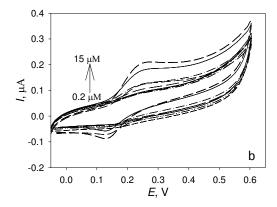
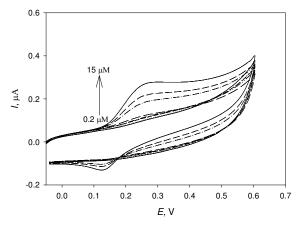


Figure S3. Calibration curve for dopamine detection at the bare Au electrodes, based on the dependence of the CV peak currents of dopamine oxidation on concentration of dopamine. Dopamine detection limit was 1  $\mu$ M dopamine.

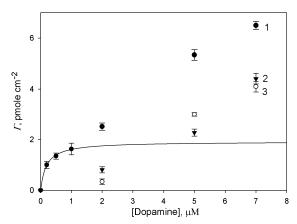




**Figure. S4.** Representative CVs of (a) the Dop\_RNA aptamer/Cys/Au electrode in the presence of 0, 0.1, 0.2, 0.5, 1, 2, 5, 7,  $10\mu M$  dopamine, and (b) the Mut\_aptamer/Cys/Au electrode in the presence of 0, 0.2, 0.5, 1, 2, 5, 7, 10,  $15\mu M$  dopamine. Potential scan rate is 50 mVs<sup>-1</sup>.



**Figure. S5**. Representative CVs of dopamine at cysteamine-modified Au electrodes, additionally modified with 20 nts DNA sequence of the arbitrary composition (not aptamer). Dopamine concentrations: 0, 0.2, 0.5, 1, 2, 5, 7, 10, and 15  $\mu$ M, PBS, pH 7.4, potential scan rate 50 mV s<sup>-1</sup>.



**Figure S6.** Presumable surface coverage of the RNA aptamer estimated by integration of the dopamine peak in CV measured at different concentrations of dopamine at differently modified electrodes: (1) Dopa\_RNA aptamer/cysteamine/Au, (2) arbitrary DNA/cysteamine/Au (20nts) and (3) Dopa\_Mut\_aptamer/cysteamine/Au.

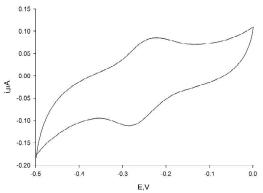
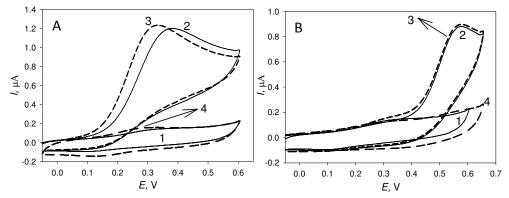
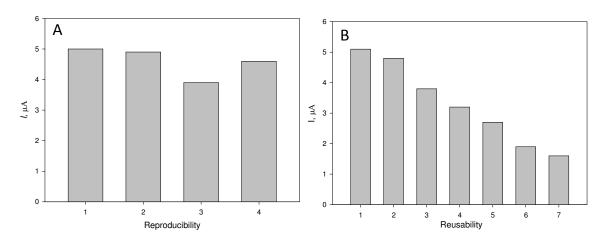


Figure S7. Representative CV recorded with the MB-labeled aptamer-modified electrode, scan rate 50 mV s<sup>-1</sup>.



**Figure S8.** (A) Representative CV recorded with the aptamer-modified electrodes in solutions of (1)  $5\mu M$  dopamine, (2) 0.3 mM AA, (3)  $5\mu M$  dopamine and 0.3 mM AA, and (4)  $5\mu M$  dopamine and 0.3 mM AA with the electrode covered by the Nafion membrane. (B) Similar data for UA.



**Figure S9.** Reproducibility of the CV oxidation signal from the Dopa\_RNA aptamer-modified electrodes in the presence of 5  $\mu$ M dopamine for (A) four individual electrodes and (B) a single electrode in the course of continuous measurements, the electrode was regenerated by successive washing with water.

Table S1. DNA sequences and produced RNAs in the present work

Name	Sequences (5'3')
Dopa_RNA aptamer <sup>a</sup>	GUCUCUG <b>UG</b> CGCCAGAGACAGUGGGGCAGAUAUGGGC CA <b>GCACA</b> GAAUGAGGCCC
EcoRI_T7_DopaMut_Apt_F <sup>b,c</sup>	TATAGAATTC <u>TAATACGACTCACTATAG</u> GTCTCTGTGTGCaa CAGAGAACACTGGGGCAGATAT
DopaMut_Apt_R	GGGCCggATTCTGTGCgGGCCCATATCTGCCCCAGTGTTCTC
Dopa_RNA Mut_aptamer a,b	GGUCUCUGUGCaaCAGAGAACACUGGGGCAGAUAUGGG CCcGCACAGAAUccGGCCC

<sup>&</sup>lt;sup>a</sup> Domains responsible for dopamine binding are indicated in bold letters; <sup>b</sup>Primers used: T7 promoter sites are underlined; <sup>c</sup> Mutations are indicated by lower case letters.