# **Chem**Rxiv<sup>™</sup>

### Nature to Nurture- Identifying Phytochemicals from Indian Medicinal Plants as Prophylactic Medicine by Rational Screening to Be Potent Against Multiple Drug Targets of SARS-CoV-2

Pratap Kumar Parida, Dipak Paul, Debamitra Chakravorty

Submitted date: 22/05/2020 • Posted date: 22/05/2020

Licence: CC BY-NC-ND 4.0

Citation information: Parida, Pratap Kumar; Paul, Dipak; Chakravorty, Debamitra (2020): Nature to Nurture-Identifying Phytochemicals from Indian Medicinal Plants as Prophylactic Medicine by Rational Screening to Be Potent Against Multiple Drug Targets of SARS-CoV-2. ChemRxiv. Preprint. https://doi.org/10.26434/chemrxiv.12355937.v1

The COVID-19 pandemic resulted in millions of people being quarantined, impacting the world economy and health sector. There is no existing proven treatment for this disease. It may takea long time until a good candidate vaccine or a potent drug is made available in the market. Therefore, there is a need to search for alternative therapy. In the context, this work explored natural compounds from Indian medicinal plants to develop a prophylactic treatment regimen that will be instrumentalin controlling the spread of the deadly virus. In this work 1916 phytochemicals from 55 Indian medicinal plants, reported to possess anti-viral properties, were subjected to virtual screening on 8 structural and non-structural SARS-CoV-2 protein targets. Docking interactions, ADME and toxicity profiles of the 66 screened phytochemicals were correlated with 21 repurposed drugs that have been most cited in literature to be effective against SARS-CoV-2. Steroidal lactones from Withaniasomnifera and triterpenoids from Azadirachtaindica- with docking score ranging from -13 kcal/mol upto -6 kcal/mol were identified to occupy the top scoring virtually screened phytochemicals against the various targets of SARS-CoV-2. Importantly this work proposes that a concoction of these phytochemicals can act as prophylactic anti-viral medicine to control the spread of SARS-CoV-2 and also enhance natural immunity as the first line of defence towards such a deadly virus.

File list (5)

Parida et al. 21520.pdf (2.31 MiB)	view on ChemRxiv • download file
Supplemental.pdf (313.98 KiB)	view on ChemRxiv • download file
Figure1.tif (8.69 MiB)	view on ChemRxiv • download file
Figure2.tif (9.10 MiB)	view on ChemRxiv

## Nature to Nurture- Identifying phytochemicals from Indian medicinal plants as prophylactic medicine by rational screening to be potent against multiple drug targets of SARS-CoV-2

#### Authors: Pratap Kumar Parida,<sup>1#</sup>, Dipak Paul<sup>2#</sup>, DebamitraChakravorty<sup>3\*#</sup>

<sup>1</sup>Pratap Kumar Parida, PhD (Research Scientist- Computational Biology &Bioinformatics) Noor Enzymes Private Limited, 37-B, Darga Road, Kolkata - 700 017, India

<sup>2</sup>Dipak Paul, PhD(Scientist - Technical Services) Noor Enzymes Private Limited, 37-B, Darga Road, Kolkata - 700 017, India

<sup>3\*</sup>Corresponding author DebamitraChakravorty, PhD (Project Lead - Computational Biology) Novel Techsciences (OPC) Private Limited, 37-B, Darga Road, 1st Floor, Kolkata - 700 017, India E-mail: dc(at)noveltechsciences.com

<sup>#</sup>All the authors have contributed equally to the paper.

#### ABSTRACT

The COVID-19 pandemic resulted in millions of people being quarantined, impacting the world economy and health sector. There is no existing proven treatment for this disease. It may take a long time until a good candidate vaccine or a potent drug is made available in the market. Therefore, there is a need to search for alternative therapy. In the context, this work explored natural compounds from Indian medicinal plants to develop a prophylactic treatment regimen that will be instrumentalin controlling the spread of the deadly virus. In this work 1916 phytochemicals from 55 Indian medicinal plants, reported to possess antiviral properties, were subjected to virtual screening on 8 structural and non-structural SARS-CoV-2 protein targets. Docking interactions, ADME and toxicity profiles of the 66 screened phytochemicals were correlated with 21 repurposed drugs that have been most cited in literature to be effective against SARS-CoV-2. Steroidal lactones from Withaniasomnifera and triterpenoids from Azadirachtaindica- with docking score ranging from -13 kcal/mol upto -6 kcal/mol were identified to occupy the top scoring virtually screened phytochemicals against the various targets of SARS-CoV-2. Importantly this work proposes that a concoction of these phytochemicals can act as prophylactic anti-viral medicine to control the spread of SARS-CoV-2 and also enhance natural immunity as the first line of defence towards such a deadly virus.

**Key Words:**SARS-CoV-2, anti viral, India, phytochemicals, prophylaxis, concoction, repurposed drugs

#### Highlights

- Rational screening of phytochemicals that can bind to various targets of SARS-CoV-2 and proposing a concoction that can act as prophylaxis;
- Molecular docking to find interaction between phytochemicals from 55 Indian Medicinal plantstargeting eight SARS-COV-2 targets (structural and non structural proteins);
- Prediction of drug-likeness from the top hits of phytochemicals and their maximum recommended therapeutic dose;
- Comparison of the docking results with 21 repurposed drugs currently being considered to treat COVID-19.

#### Introduction

Globalization has led to increased travel and exploitation of natural environment leading to escalation in occurrence of pandemics. Despite global preparedness, challenges persistas modern medicine still lacks effective immunization and drugs due to the development of potential multi drug resistant pathogenic mutants. Among all pathogens, RNA viruses are more contagious as they havehigh rates of evolution due to natural selection on their mutation rate (Duffy, 2018; Kayla et al., 2018). This leads to their ability to emerge in novel hosts and escape vaccine-induced immunity (Duffy, 2018; Peck et al., 2018). Examples of RNA viral outbreaks are that of dengue virus, influenza virus, measles virus, severe acute respiratory syndrome (SARS) virus, West Nile virus and recently the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) (Lin et al., 2014, WHO, 2019). The latter has been declared to be pandemic on 11<sup>th</sup> March 2020 by the World Health Organization [WHO 2020]. Though vaccines and antiviral agents are the most effective against treatment of viral outbreaks, development of vaccine may take up to 12 - 18 monthsand novel drug discovery is known to take several years (Lancet, 2020). Thus prophylaxis and immediate treatment is the need of the hour. In this context natural products provide an excellent resource for novel anti-viral drug development (Lin et al., 2014). Phytochemicals from plants have been reported to possess anti-viral properties against rabies virus, Human immunodeficiency virus, Chandipura virus, Japanese Encephalitis Virus, Enterovirus, Influenza A/H1N1 and other influenza viruses, SARS coronavirus (Ganjhuet al., 2015). In the review by Ganjhuet al., 2015 it was also shown that combination of extracts from different plant species resulted in their anti-viral effect. Thus if such formulation can be developed against SARS-CoV-2 it can act as prophylaxis and aid in "flattening the curve" by attenuating viral load of the ongoing pandemic. It was also reported that combined uses of interferons and phytoconstituents facilitated anti-viral therapy (Cinatlet al., 2003). Therefore, concoction of phytoconstituents, which can be synthesized to reduce pressure on the natural sources, can also find use in combination therapy for SARS-CoV-2. The same can also be used as an immunopotentiating agent in combination with the vaccines that are being developed against SARS-CoV-2. Furthermore, semi-synthetic molecules can be developed to overcome if any inherent limitations existin using theoriginal molecule.

In the context of exploring natural compounds from plants as anti-viral agents- it can be mentioned here that India is home to plethora of indigenous medicinal plants that have been used in traditional medicine for centuries. Since the vedic period (more than 2000 years ago) medicinal plants are used as ayurvedic medicines in Indian subcontinent (Saini, 2016). Traditional Indian plants, to name a few, Azadirachtaindica, Curcuma longa, Bacopa monnieri and Zinger officinaleare known to possess anti-viral properties (Nadkaraniet al., 1976; Pathaket al., 2013). The anti-viral properties of these plants is due to the presence of phytochemicals such as polysaccharides, triterpenes, phenolic acids, alkaloids. proanthocyanidins and anthraquinones (Ganjhuet al., 2015). For example, a phenolic compound isolated from Curcumin was reported to interfere with Herpes Simplex Virus-1 replication (Kutluayet al., 2008). Chebulagic acid and Punicalagin from Curcumin have also been reported to have activity against dengue virus by inhibiting early steps of virus entry (Lin et al., 2013). Furthermore, research and digitisation have succeeded to give access to the structures of these phytochemicals (Poluret al., 2011; Pathaniaet al., 2015; Mohanrajet al., 2018). Example of one such database is IMPPAT (Mohanrajet al., 2018). This aforementioned vast literature of success presents us with a huge scope to explore the phytochemicals from these medicinal Indian plants, through in silico virtual screening, for drug development against SARS-CoV-2 targets.

Selection of drug targets from the pathogen is the first filter for the success of *in silico* drug designing. As of March 25<sup>th</sup> 2020, 104 SARS-CoV-2 structures have been deposited in the RCSB Protein Data Bank (RCSB PDB). Thorough literature survey showed that proteins involved in viral entry (Spike glycoprotein) and replication(16non structural proteins) can be targeted for drug discovery. Thus, in this work, keeping in mind that nature always nurtures us, virtual screening of a vast library of phytoconstituents from Indian medicinal plants known to have anti-viral properties, has been carried out to identify complementary or alternative therapy for SARS-CoV-2. Conclusively this work proposes a concoction of phytoconstituents

from different plant sources from India that can act as prophylactic anti-viral medicine to control the spread of SARS-CoV-2 and also enhance natural immunity as the first line of defence towards such a deadly virus.

#### **Computational methodology**

The main aim of this work was to identify natural sources and their phytochemicals from Indian medicinal plants that can be repurposed as an alternative therapy for SARS-Cov-2 or be developed as immune-potentiating agents in combination with the future vaccines or drugs. The method uses *in silico* screening techniques like rational virtual screening and drug-likeness calculations on an in- house developed 3D chemical structure database of phytochemicals from Indian medicinal plants. This was followed by their comparison (docking scores, drug-likeness and binding interactions) to the already reported repurposed drugs having potentialto treat COVID-19. The overview of the methodology has been illustrated in Figure 1.The database of the phytochemicals can be obtained on request from the authors.



**Figure 1.** The overview of the methodology employed to rationally screen phytochemicals from Indian medicinal plants as potential drug like compounds against 8 structural and non-structural targets of SARS-CoV-2. Virtual screening of 1916 phytochemicals was performed using Autodock4 and Racoon. There were 8 SARS-CoV-2 targets and 10 binding sites considered for rational screening in this work.

#### Database creation of natural compounds from Indian medicinal plants

55 Indian medicinal plants were identified from literature to have anti-viral properties(Table S1). Phytochemicals from the 55 sources were searched in literature and their three dimensional (3D) structures were obtained from PubChem which is an is an open chemistry database at the National Institutes of Health (NIH). The SMILES format

(Simplified Molecular Input Line Entry System) of the phytochemicals that lacked 3D structures in PubChem were submitted to CORINA Classic to obtain their 3D structures (Sadowski et al., 1994). We could obtain the structures of 1916 phytochemicals. To compare and validate our docking results, 3D structures of 21repurposed drugs reported to have potential inhibitory effect against SARS-CoV-2 and most cited in literature in the years 2019-2020 were also collected from PubChem and docked against the targets of SARS Cov-2 were used in this work. The number of citations were counted by literature survey and non-redundancy of the dataset was maintained. The number of citations, FDA approval status and COVID-19 FDA approval status have been presented in Table S3.

#### Target selection from crystal structures of SARS-CoV-2

Out of the 105 structures of various proteins of SARS-CoV-2 deposited in RCSB PDB as of March, 2020, a total of 8 non redundantand high resolution structures were targeted for in silico virtual screening (Table 1). The main protease, RNA dependent RNA polymerase (NSP 12), NSP 3 (role in replication), NSP 9 (RNA-binding protein), NSP 10 (cofactor for the activation of the replicative enzyme), NSP 15 (endoribonuclease activity), NSP 16 (methyltransferase activity) and the spike glycosylated protein required for viral entry by binding to the ACE-2 receptor were targeted. The binding sites were identified from literature and HotSpot Wizard analysis (Pavelkaet al., 2009). Thekey residues for molecular docking grid generations are presented in Table 1. For example, the binding site chosen for docking in the spike protein (PDB ID: 6lzg) was with the residues which are in the ACE-2 receptor binding interface. These residues can be identified from its PDB structure. It can be noted here that in the case of the RNA dependent RNA polymerase (NSP 12), three sites were considered for docking analysis: (1) The RNA binding site, (2) NSP12-NSP7 interface and (3) NSP12-NSP8 interface. Sites 2 and 3 were considered for virtual screening because NSP 7 and NSP 8 are the cofactors of NSP 12 (Gao et al., 2020). Therefore, it was presumed that targeting these interfaces can disrupt the functioning of RNA dependent RNA polymerase and thus werechosen as allosteric drug binding sites.

Table 1: The targets of SARS Cov-2 and their binding site residues used for virtual screening

Protein Name	PDB ID	* Identified Drug Binding Site Residues
Spike Protein	6lzg	K417, G446, Y449, Y453, N487, G496, Q498, T500, N502
Main Protease (NSP 5)	6lu7	T24-T26, H41, M49, F140-C145, H163-P168, H172, D187-Q192

NSP 10-NSP16 Interface	6w4h (chain B)	N4293-Y4349
NSP 12-RNA Binding site catalytic	6m71	V557, D618, D623, G680, N691, S759-D761, D851-1864
NSP 12-NSP7 Interface	6m71	T409, K411-F415, Y420, E431, F440, F442, A443
NSP 12-NSP 8 Interface	6m71	L270-Y273, P323-K332, V338-V441, F368-F407, P505-S518, M666
NSP 16-NSP10 Interface	6w4h	K6836-M6839, V6842, T6846, K6874- P6878, Q6881-R6885, V6902-D6904, L7042-S7046
NSP 3	6w02	A21-V23, A38-A50, L126 -F132, A154-L160
NSP 9	6w4b	R100-T110
NSP 15	6w01	E234, S244-L249, P272, S289, E340, Y343, H250

\*Residue numbering follows the RCSB PDB structures.

#### Rational screening of the phytochemical library with the 8 drug targets from SARS-CoV-2

MGLTools-1.5.6 was used for the generation of grid file and the docking file parameters. The grid box parameters for each of the proteins are given in the Table S2. Molecular docking of all the compounds was performed using the Raccoon plug-in of Autodock 4. The configurations files generated from MGLTools-1.5.6 are incorporated in the Raccoon plug-in package. The docking was run with 10 Lamarkian Genetic Algorithm (LGA) runs for virtual screening with the default parameters of Autodock 4.2. Further, for each of the 8 targets, phytochemicals with lowest binding energies (kcal/mol) were identified and subjected for re-docking for 100 LGA runs. BIOVIADiscovery StuioVisualiser (Dassault Systèmes BIOVIA, 2020). Further the residue evolutionary conservation scores of the binding pocket residues were calculated using ConSurf server (Ashkenazy et al., 2016).

#### Drug-likeness Properties of the Phytochemicals and the Repurposed Drugs

Drug-likeness properties based on Lipinki Rule of 5 and Adsorption, Distribution, Metabolism, Excretion (ADME) properties were calculated using SwissADME(Daina et. al 2014, Daina et. al 2016, Daina et. al 2017)Toxicity profiles (T) were calculated by the vNN web server(Schymanet al., 2017) and compared for each top 10 scoring ligands and the 21repurposed drugshaving potential to be developed into good drug candidates against SARS CoV-2. The significant pharmacokinetic parameters for ADME/T associated properties of Gastro Intestinal absorption (GI), Blood Brain Barrier permeation (BBB), CYP inhibition, pharmacokinetic properties like Lipinski rule of 5 (Lipinski et al.,2001), PAINS (Baellet al., 2010) and Brenk (Brenket al., 2008) and Lead likeness were calculated (Teague et al., 1999). Further, comparative analysis of ADME/T properties between the phytochemicals and the 21 drugs were performed by cross correlation matrix calculated using Heatmapper (Babicket al., 2016). The maximum recommended therapeutic dose (MRTD) was also predicted using vNN-ADMETserver (Schymanet al., 2017). MRTD is the predicted upper daily safe dose (Contreraet al., 2004) for phase I clinical trials.

#### **Results and Discussion**

About half of all approved drugs between 1981 and 2010 were shown to be natural compound based (Stratton et al., 2015). It has been reported that more than 100 natural product derived drugs are undergoing clinical trials (Harvey et al., 2008). Natural compounds are unique, when compared with synthetic drugs in terms of more complex scaffolds and chiral centres and have huge diversity (Zhao et al., 2019). Examples of natural compounds with known safety profiles and potent anti-viral properties are isoflavones and artemisinin (Huang et al., 2020). Flavonoids were reported to have effect against RNA viruses (Kaul et al., 1985). Many researchers have reported the beneficial effect of natural compounds in treating HIV infections (Reiter et al., 2015; Salehiet al., 2018) and in the treatment of important chronic diseases like asthma (Moulton et al., 2011), Parkinson's (Dashtipouret al., 2007), cancer (Rajkumar et al., 2011). Recently, Khan et al., 2020 identified flavone and coumarine derivatives as promising leads targetting the main protease of SARS-CoV-2. Hydroxychloroquine was also observed to significantly decrease SARS-CoV-2 infection (Gautretet al., 2020). Li et al., 2020 reported that traditional Chinese herbal medicine treatments alone or in combination with western medicine can be successful in preventing and treating COVIS-19 (Li et al., 2020). Based on these reports it is clear that it is necessary to repurpose natural compounds especially phytochemicals as prophylaxis to manage patients with COVID-19. In this context, India is a rich resource of medicinal plants, many of which have antiviral properties (Jassimet al., 2003; Jadhav et al., 2012). Hence, the phytochemicals from these natural sources can be explored to identify bioactive drugs with no or minimal side-effects (Tahir et al., 2019).

Therefore, in this work 1916 phytochemicals from 55 Indian medicinal plants, reported to possess anti-viral properties, have been subjected to virtual screening on 8 SARS-CoV-2 targets- selected based on two broad categories: (1) Proteins required for RNA synthesis, replication and assembly and (2) proteins required for viral entry.

## Rational Screening anddrug-likeness properties of the phytochemicals and the repurposed Drugs

Rational screening made it possible to identify 66 phytochemicals that can be potent inhibitors against the 8 drug targets of SARS-COV-2. The maximum percentage oftop scoring phytochemicals that docked to the 8targets of SARS-COV-2 were steroidal lactones from *Withaniasomnifera*(37%)and triterpenoids from*Azadirachtaindica*(19%) with docking score ranging from -13 kcal/mol upto -6 kcal/mol (Fig. 2A, Table S1).It can be noted here that the steroidal lactones from *Withaniasomnifera*(against Herpes Simplex virus replication (Grover et al., 2012), H1N1 influenza virus (Cai et al., 2015) and Chikungunya Virus (Jain et al., 2018).Similarly, triterpenoidsfrom*Azadirachtaindica*(apave been reported to have anti-viral effect against Herpes Simplex virus (Ahmad et al., 2016) and coxsackievirus virus B-4 (Badamet al., 1999). Therefore, it can be mentioned here that based on these reports and our work, there arises a scope tofurther experimentally explorethese phytochemicals in order toanalyse their efficacy against treating COVID 19 patients.



**Figure 2.** A) Bar graph showing the percentage of phytochemicals with top docking score from Indian medicinal plants. The horizontal axis represents the percentage of phytochemicals. The vertical axis is the names of the Indian medicinal plants.

B) The correlation (Average linkage, Pearson's distance measurement) heat map generated by Heatmapperfrom the docking scores of the 21 drugs with the 10 binding pockets of SARS-COV-2. The numbers in the left vertical axis represent the SARS-COV-2 targets: 1: Protease; 2: Spike protein, 3: NSP 10; 4: NSP 12 catalytic pocket; 5: NSP 12-NSP7 interface; 6: NSP 12-NSP 8 interface; 7: NSP 16; 8: NSP 9; 9: NSP 15; 10: NSP 3. The horizontal axis represents the 21 drugs abbreviated as: Chloroquine phosphate (CP); Chloroquine (CL); Arbidol (AR); Remdesivir (REM); Favipiravir (FAV); Lopinavir (LOP); Ritonavir (RIT); Ribavirin (RIB); Oseltamivir (OSE); Zanamivir (ZAN); Peramivir (PER); Ganciclovir (GAN); Methylprednisolone (MEP); Dexamethasone (DEX); Baricitinib (BAR); Hydroxychloroquine (HCL); Sofosbuvir (SOF); Interferon alfa-2b (INA); Camostat mesylate (CAM); Darunavir (DAR); Galidesivir / BCX-4430 (GAL). The colors represent the z-scores of the docking scores.

C) Pairwise cross correlation map (Elucidian distance measurement) of ADME profiles of the 21 drugs abbreviated as F1...Fn and the phytochemicals abbreviated as P1...Pn. The orange box represents the region in the plot showing correlation between the drugs and the phytochemicals with ADME profiles which shows very low violations of the standard rules.

D) Comparative bar graph showing 13 toxicity profiles of the 21 drugs (Left panel) and the phytochemicals (right panel). Yes: the percentage of compounds showing toxicity and No: the percentage of compounds not showing toxicity. Horizontal axis represents the percentage of compounds and vertical axis is the various toxicity profiles which were obtained from vNN ADMET server.

Further, in this study 21 compounds, highly cited in literature to be repurposed for COVID 19 treatment, were also docked with 100 LGA runs against the 8 targets of SARS-COV-2 (Fig. 2B, Table S3A-S3B). Among the 21 compounds, hydroxychloroquine and Remdesivir were the most cited to be effective against SARS-CoV-2. The ADME/T profiles of these drugs have been presented in Table S4A-S4B.Interestingly it was observed that most of the 21 drugs bound to multiple targets of SARS-CoV-2 with docking score in the range: -11 kcal/mol to -5 kcal/mol. Additionally, two major clusters were observed in the heat map built with average linkage clustering and Pearson distance measurement methods using Heatmapper server (Fig. 2B). It can be observed from Fig. 2B that drugs targeting multiple targets with similar docking score were clustered together by the clustering algorithm. Among the 21 drugs, Baricitinib, Methylprednisolone, Arbidol, Dexamethasone, Hydroxycloroquine, Chlorquine and chloroquine phosphate bound to the 8 targets with binding energy < -5 kcal/mol. The most potent inhibitors were found to be Baricitinib (-7.62) kcal/mol) for spike, Methylprednisolone and Baricitinib (~ -8.5 kcal/mol) for the main protease (NSP 5), Methylprednisolone (-9.22 kcal/mol) for NSP 10 and NSP 12 (catalytic site) (-6.54 kcal/mol), Hydroxychloroquine (-7.73 kcal/mol) and Lopinavir (-8.33 kcal/mol) for NSP 12 -- NSP7 and NSP 8 interface, Chloroquine (-10.11 kcal/mol) for NSP 16, Camostat mesylate (-5.98 kcal/mol) for NSP 9, Dexamethasone (-7.08 kcal/mol) for NSP 15 and for NSP 3 it was Baricitinib (-10.38 kcal/mol).Further, virtual screening also showed that

NSP 16 and NSP 3 can be good drug targets for SARS CoV-2- most of the 21 drugs docked to them with low binding energy (Fig. 2B).

Further, clustering of the binding sites can be attributed to the docking scores of the drugs (Fig. 2B). The sites which docked multiple drugs with similar docking scores were clustered together. This led us to analyze the similarity among the binding pocket residues of the targets that clustered together. For example, NSP 16 and NSP 3, NSP 12-NSP 8 interface and the main protease were separately clustered in two groups- the former bound to the same drugs with similar docking scores which were better thanthe latter. NSP 15 and Spike protein binding to these drugs with lesser negative docking scores were also clustered together.NSP12 D1 (catalytic site) and NSP 9 showed the worst docking scores with these 21 drugs and thus were orphans closer to the NSP 15 and Spike protein group in the clustering heat map. Whereas NSP 12-NSP7 interface and NSP 3 showing better binding with these drugs fell in between the NSP 16 and NSP 3, NSP 12-NSP 8 interface and the main protease groups (Fig. 2B, S Table S3B). It can also be comprehended from the binding site analysis that binding pockets which are more hydrophobic bound to multiple drugs with better binding scores and were thus clustered together. Very similar observations were made when the 100 top hits (66 unique) phytochemicals from virtual screening were re-docked to the 10 binding sites from 8 SARS-CoV-2 targets.

Further, the re-docking results after 100 LGA runs of the virtually screened phytochemicals have been presented in Table S4along with their ADME/T properties(Table S6). The compounds which can be good leads in terms of medicinal chemistry- showing 0 Lipinski, Veber and Egan rules violations, high Gastro Intestinal (GI) absorption, 0 PAINS (pan assay interference compounds) alert and 1-2 Brenk (to bear poor pharmacokinetics properties) and very low number of lead likeness violations (250<= molecular weight<=350, XLOGP <=3.5, rotable bonds <= 7) have been presented in Table 2.

Phytochemical	Docking Score (Kcal/Mol)	MRTD Dose (md/day)	Source					
Main Protease (PDB; 6lu7)								
		8.7						
27-Deoxy-14-hydroxywithaferin A	-10.8		Withaniasomnifera					
Nimolicinol	-10.09	84	Azadirachtaindica					
17-Hydroxywithaferin	-10.08	342	Withaniasomnifera					
WithanolideR	-9.63	46	Withaniasomnifera					
	Spike (PDB 6	LZG)						
27-Hydroxywithanone	-8.47	49	Withaniasomnifera					

Table 2: Good drug leads in terms of medicinal chemistry obtained after re-docking of phytochemicals from Indian medicinal plants

Deoxywithastramonolide	-8.27	54	Withaniasomnifera
27-Deoxywithaferin A	-7.82	303	Withaniasomnifera
2,3-Dihydrowithaferin A	-7.45	319	Withaniasomnifera
	NSP 10 (P	'DB 6w4h)	
27-Hydroxywithanolide B	-8.32	54	Withaniasomnifera
Anaferine	-6.22	196	Withaniasomnifera
NS	P12 D1 (RNA Bind	ling Site) (PDB 6M7	71)
12-Deoxywithastramonolide	-7.58	54	Withaniasomnifera
NSP1	2 D2 (NSP12-NSP	7 Interface) (PDB 6	M71)
Withastramonolide	-9.72	60	Withaniasomnifera
Withanolide B	-9.2	51	Withaniasomnifera
12-Deoxywithastramonolide	-9.05	54	Withaniasomnifera
Withanolide R	-8.94	46	Withaniasomnifera
Withaferin A	-7.9	342	Withaniasomnifera
NSP	12 D3 (NSP-NSP8	Interface) (PDB 6M	171)
Withaferin A	-9.94	342	Withaniasomnifera
	NSP16(P	DB 6w4h)	
Solvanol	-10.98	473	Solanum nigrum
(-)-Anaferine	-10.91	196	Withaniasomnifera
Limonin	-10.73	172	Nigella sativa
	NSP9 (PI	DB 6w4b)	
27-Hydroxywithanolide B	-8.28	54	Withaniasomnifera
12-Deoxywithastramonolide	-8.19	54	Withaniasomnifera
Azadiradionolide	-7.3	220	Azadirachtaindica
	NSP15 (P	,	
Somniferine	-7.21	45	Withaniasomnifera
Vindolinine	-6.87	102	Catharanthus roseus
	NSP3 (Pl	DB 6w02)	
2,3-Dehydrosomnifericin	-12.3	127	Withaniasomnifera
WithanolideB	-11.44	51	Withaniasomnifera
24,25-dihydrowithanolide D	-10.24	54	Withaniasomnifera
27-Deoxy-14-hydroxywithaferin A	-9.49	8.7	Withaniasomnifera

Other than these phytochemicals reported in Table 2, many phytochemicals with good docking scores could not satisfy the standard rules of drug likeness. It can be noted from Table S4 that Serpentinine, though violates drug-likeness rules, is the most potent phytochemical among the 1916 compound with the lowest docking score against the SARS COV-2 main protease (-13 kcal/mol). Serpentinine also docked with NSP 16 (-12 kcal/mol). Similarly, Terchebulin docked against three binding sites of NSP 12 (~ -11 kcal/mol), 7-Deacetyl-7-benzoylepoxyazadiradione with NSP 9 (-8.74 kcal/mol), Ashwagandhanolide docked with NSP 15 (-10.17 kcal/mol) and 2,3-Dehydrosomnifericin docked with NSP 3 (-12.3 kcal/mol). The justification for these drugs to be tested experimentallycomes the observation of the cross correlation matrix constructed on the ADME properties of Lipinski,

Ghose, Veber, Egan, Muegge, Lead likeness rule violations, PAINS and Brenk alerts for the 66 phytochemicals along with the 21 drugs (Fig. 2C). It was observed that the ADME profiles of most of the phytochemicals have positive correlation with the 21 drugs. Further, it can be observed from the bar graph in Fig. 2D that the phytochemicals show very less toxicity which is again comparable to the 21 drugs. Another justification comes from the reports where drug-likeness rules have been shown to be biased measurement for natural compounds as they were derived using small molecules (Loureiro et al., 2019). It was also reported that it is still a challenge to find alternative drug-likeness rules for natural compounds (Katiyar et al. 2012). Further, there are many examples of successful natural drugs which also do not obey the usual drug-likeness rules (Loureiro et al., 2019). For example, the drug paclitaxel (chemotherapy medication) does not comply with Lipinski rule of five. Conclusively it can be said that these compounds need to be studied further *in vitro* to confirm their experimental drug likeness.

An interesting observationmade from virtual screening results, was that the phytochemicals like the 21 repurposed drugs were able to bind to multiple targets with good docking scores. From Table S4 it can be observed that Terchebulin (*Terminalia chebula*) docked to 7 binding sites except NSP 16. Similar multi-potency was also observed for compounds like Serpentinine (*Rauvolfia serpentine*), Nimocin (Azadirachtaindica) and Ashwagandhanolide (*Withaniasomnifera*). It can be mentioned here that natural products are known to bind to multiple targets (Efferthet al., 2011). Moreover, according to previous research, it has been reported that binding to more than one target at the same time can lead to therapeutic efficiency decreasing probability of resistance to one target (Kadiogluet al., 2020).

A novel observation that can be reported in this paper isbased on the predicted MRTD dose (maximum recommended therapeutic dose) for the phytochemicals and the 21 drugs-calculated employing the vNN server (Table 2, Supplementary Table 3B and Supplementary Table 6). It is worth mentioning that most of the phytochemicals have low MRTD dose (maximum: 691 mg/day for a 60 Kg body weigh individual) as compared to the repurposed drugs (maximum: 10779 mg/day for a 60 Kg body weigh individual). Thus it can be said that these phytochemicals possess high pseudo-potency. These predicted MRTD dose can be instrumental to estimate the starting dose in phase I clinical trials, reducing the number of animals used in preliminary toxicology studies (Schymanet al., 2017). It was also reported that low MRTD drugs often have high pseudo-potency (Liu et al., 2016). On the other hand,

drugs of low pseudo-potency need high dose to reach the desired therapeutic effects (Liu et al., 2016).

## Analysis of the intermolecular interactions of the docked phytochemicals and the repurposed drugswith the drug targets of SARS-CoV-2

The binding interactions of the lead like phytochemicals in comparison to the 21 drugs have been illustrated in Figure 3. It can be observed that residues which were reported to be structurally and functionally important in the 8 SARS-COV-2 binding sites were visualized to interact with the phytochemicals through various *van der Waal's* interactions (hydrogen bonds, hydrophobic contacts, aromatic ring  $\pi$ - $\pi$  interactions, etc.). It can also be observed that interacting residues in these binding sites were similar for the repurposed drugs and the phytochemicals.For example, in the case of the main protease (3CLpro) of SARS-CoV-2, phytochemical moieties formed strong hydrogen bonds with the catalytic dyad residues (Cys-145 and His-41) as well as significant interactions were formed with the receptor-binding residues Gly143, His163, Met165, Glu166, Pro168 and Gln189.

It is known that the spike glycoprotein aids in virus entry by priming to angiotensinconverting enzyme 2 (ACE-2) as a receptor (McKee et al., 2020). Residues reported to be instrumental in viral entry by interacting with the ACE2 receptor were reported from the crystal structure (6lzg) are S494, N501, Q493, L455 (Shang et al. 2020). These residues were observed to interact with the phytochemicals and the repurposed drugs. It was interesting to observe that the span of residue coverage by the phytochemicals were greater than the repurposed drugs(Figure 3). It was also reported that L453 and N501 were responsible for stabilizing virus binding "hotspots" in the ACE2 receptors (Shang et al. 2020). These residues were observed to form strong hydrogen bonds <3Å in length with the phytochemicals and the repurposed drugs.

Further, the phytochemicals and the repurposed drugs were able to form strong hydrogen bonds with the residues at the interface of NSP10-NSP16, NSP12-NSP7 and NSP12-NSP 8. Therefore, disruption of interaction among these proteins can inhibit the viral replication machinery. Further, NSP 10 in SARS-CoV with two zinc fingers is known to stimulate methyl transferase activity of NSP 16 (Bouvet et al. 2014). Thus similar role is expected in SARS-CoV-2. Residues involved in zinc coordination (C4327, C4330, C4343, H4336) identified in the crystal structure (6w4h) were observed to interact with the docked molecules. Thus binding of inhibitors at this region can inhibit its interaction with NSP 16.Nsp12 is an RNA-dependent RNA polymerase (RdRp) involved in viral replication. Nsp8

synthesizesthe primer required for Nsp12-RdRp RNA synthesis. Further, the Nsp7 and Nsp8 enhances the docking of Nsp12 (Wu et al. 2020). Phytochemicals and the repurposed drugs docked poorly with the catalytic pocket with Val555 as the catalytic residue. However, theywere observed to interact with these interface residues and thus can interrupt RNA synthesis by RdRp by inhibiting NSP7 and NSP 8 docking with NSP 12. NSP 3 is the ATP ribose phosphatase of SARS-CoV-2 is required for virus replication. Binding of phytochemicals and repurposed drugs were observed to form hydrogen bonds and hydrophobic interactions with the Adenosine-5-Diphosphoribose binding site in the crystal structure (6wo2). NSP 9 is most likely involved in viral replication as in SARS CoV (Friemanet al., 2012). Residues important for its dimerization can be targeted for drug development as this effects viral proliferation in SARS-CoV (Friemanet al., 2012). Phytochemicals were observed to interact with this interface residue with strong non-covalent bonds. NSP15 is an uridylate specific endoribonuclease. His235, His250, Lys290, Thr341, Tyr343, and Ser294 were reported as the catalytic residues (highly conserved) in its active site pocket (Kim et al. 2020). The phytochemicals were observed to interact with these residues better than the repurposed drugs. Thus it can be comprehended that these phytochemicals along with repurposed drugs (in conjugation or alone) can aid in COVID 19 treatment by blocking the association of these key non-structural proteins required for replication of the SARS-CoV-2 virus.

Further, evolutionary conservation scores were also calculated for these residues forming favorable interactions with the phytochemicals- through the Consurf server(Figure 3). This was performed to analyze whether these interacting residues are functionally important to the virus and if they are prone to mutations. It is well known according to the neutral theory of molecular evolution that functionally important residues vary less than less important ones (Saldanoet al., 2016). It is also known now that SARS-CoV-2 is fast evolving due to its high rate of mutations (Pachettiet al., 2020). The concern was that if the phytochemicals were interacting more with non-conserved residues than occurrence of mutation of these residues in different strains of the virus and using observed that most of the strong interactions were formed between highly conserved residues and the phytochemicals (Figure 3). These high affinity interactions with the conserved residues were more profound with the phytochemicals than the 21 repurposed drugs. Thus it can be predicted that the phytochemicals can be developed into better drug candidates can inhibit SARS-CoV-2 as positive results were obtained. A concoction of these phytochemicals

can target the different SARS-CoV-2 proteins. This natural concoction can thus act as prophylaxis for treating SARS-CoV-2 infections along with the repurposed drugs.



Figure 3. Interaction of amino acid residues in the binding pocket of 8 SARS-COV-2 targets (10 binding pockets). I) Cartoon representation of the 10 binding sites with the re-docked and superimposed phytochemicals(top scoring with best drug likeness properties).Reference Table 2 for the docked phytochemicals. II) Color map of the residues in the targets interacting with the top

scoring phytochemicals and the top scoring repurposed drugs. The evolutionary conservation scores calculated by the ConSurf server have also been illustrated: 1: Low conservation to 9: high conservation (Ashkenazy et al., 2016).

#### Conclusions

This is the first systematic work of exploring 1916 phytochemicals from 55 Indian medicinal plants having anti-viral properties by in silico rational screening approaches to develop prophylactic treatment against SARS-Cov-2. 21 repurposed drugs were also subjected to molecular docking and compared to the phytochemicals. Top scoring phytochemicals screened were steroidal lactones from Withaniasomnifera (37%) and triterpenoids from Azadirachtaindica (19%) with docking score ranging from -13 kcal/mol upto -6 kcal/mol. Comparison of ADME/T profiles of the screened phytochemicals with the repurposed drugs showed positive correlation. Further, high affinity interactions of the binding site residues of these targets were dominated by hydrogen bonds and hydrophobic interactions. Strong inter-molecular interactions were also observed to be profound with the evolutionary conserved residues- with the phytochemicals than the 21 repurposed drugs. Thus, it can be said that the phytochemicals will be effective against different mutated strains of the virus. We have also predicted the MRTD dose of these screened phytochemicals along with the repurposed drugs that can be used as a safe dose for the phase I clinical trials. It is worth mentioning that most of the phytochemicals have low MRTD dose and thus can be developed into good drug leads. Conclusively it can be proposed here that a concoction of these phytochemicals can target the different SARS-CoV-2 proteins at the same time and this natural concoction can act as prophylactic medication for treating SARS-CoV-2 infections. It is also proposed here that these phytochemicals being natural in origin can lead to develop immunity which is the first line of defense against such deadly virus. Further, this natural concoction can aid in treatment if given in conjugation with other drug therapy or used as an immunopotentiating agent with the future developed vaccines against SARS-CoV-2.

#### Acknowledgements

We would like to acknowledge Noor Enzymes Private Limited for providing the infrastructure and computational facilities for successfully completing this work.

#### **Disclosure statement**

No potential conflict of interest was reported by the authors.

#### Author contributions statement

All authors conceived, conducted and analyzed the results of experiments equally.

Supplemental online material

Supplemental data for this article is available online

#### References

Ahmad, A., Javed, M. R., Rao, A. Q., &Husnain, T. (2016). Designing and screening of universal drug from neem (*Azadirachtaindica*) and standard drug chemicals against influenza virus nucleoprotein. *BMC Complementary and Alternative Medicine*, 16(1), 519. https://doi.org/10.1186/s12906-016-1469-2

Ashkenazy, H., Abadi, S., Martz, E., Chay, O., Mayrose, I., Pupko, T., & Ben-Tal, N. (2016). ConSurf 2016: an improved methodology to estimate and visualize evolutionary conservation in macromolecules. *Nucleic Acids Research*, 44(W1), W344-W350. <u>https://doi.org/10.1093/nar/gkw408</u>

Babicki, S., Arndt, D., Marcu, A., Liang, Y., Grant, J. R., Maciejewski, A., & Wishart, D.

- S. (2016). Heatmapper: web-enabled heat mapping for all. *Nucleic Acids Research*, 44, W147-W153. <u>https://doi.org/10.1093/nar/gkw419</u>
- Badam, L., Joshi, S. P., &Bedekar, S. S. (1999). 'In vitro' antiviral activity of neem (Azadirachtaindica. A. Juss) leaf extract against group B coxsackieviruses. The Journal of Communicable Diseases, 31(2), 79-90.
- Baell, J. B., & Holloway, G. A. (2010). New substructure filters for removal of pan assay interference compounds (PAINS) from screening libraries and for their exclusion in bioassays. *Journal of Medicinal Chemistry*, 53(7), 2719-2740. https://doi.org/10.1021/jm901137j
- Bharitkar, Y. P., Bathini, S., Ojha, D., Ghosh, S., Mukherjee, H., Kuotsu, K., Kuotsu, K., Chattopadhyay, D., & Mondal, N. B. (2014). Antibacterial and antiviral evaluation of sulfonoquinovosyldiacylglyceride: a glycolipid isolated from *Azadirachtaindica* leaves. *Letters* in *Applied Microbiology*, 58(2), 184-189. <u>https://doi.org/10.1111/lam.12174</u>

Bouvet, M., Lugari, A., Posthuma, C. C., Zevenhoven, J. C., Bernard, S., Betzi, S., Imbert, I., Canard, B., Guillemot, J. C., Lécine, P., &Pfefferle, S. (2014). Coronavirus Nsp10, a critical co-factor for activation of multiple replicative enzymes. *Journal of Biological Chemistry*, 289(37), 25783-25796. <u>https://doi.org/10.1074/jbc.M114.577353</u>

- Brenk, R., Schipani, A., James, D., Krasowski, A., Gilbert, I. H., Frearson, J., & Wyatt, P.
- G. (2008). Lessons learnt from assembling screening libraries for drug discovery for neglected diseases. *ChemMedChem: Chemistry Enabling Drug Discovery*, 3(3), 435-444. <u>https://doi.org/10.1002/cmdc.200700139</u>

Cai, Z., Zhang, G., Tang, B., Liu, Y., Fu, X., & Zhang, X. (2015). Promising antiinfluenza properties of active constituent of Withaniasomnifera ayurvedic herb in neuraminidase of H1N1 influenza: computational study. *Cell Biochemistry and Biophysics*, 72(3), 727-739. <u>https://doi.org/10.1007/s12013-015-0524-9</u>

- Cinatl, J., Morgenstern, B., Bauer, G., Chandra, P., Rabenau, H., &Doerr, H. W. (2003). Treatment of SARS with human interferons. *The Lancet*, *362*(9380), 293-294. <u>https://doi.org/10.1016/S0140-6736(03)13973-6</u>
- Contrera, J. F., Matthews, E. J., Kruhlak, N. L., & Benz, R. D. (2004). Estimating the safe starting dose in phase I clinical trials and no observed effect level based on QSAR modeling of the human maximum recommended daily dose. *Regulatory Toxicology* and Pharmacology, 40(3), 185-206. <u>https://doi.org/10.1016/j.yrtph.2004.08.004</u>
- Daina, A., &Zoete, V. (2016). A BOILED-Egg to predict gastrointestinal absorption and brain penetration of small molecules. *ChemMedChem*, 11(11), 1117-1121. <u>https://doi.org/10.1002/cmdc.201600182</u>

- Daina, A., Michielin, O., &Zoete, V. (2014). iLOGP: a simple, robust, and efficient description of n-octanol/water partition coefficient for drug design using the GB/SA approach. *Journal of Chemical Information and Modeling*, 54(12), 3284-3301. https://doi.org/10.1021/ci500467k
- Daina, A., Michielin, O., &Zoete, V. (2017). SwissADME: a free web tool to evaluate pharmacokinetics, drug-likeness and medicinal chemistry friendliness of small molecules. *Scientific Reports*, 7, 42717. <u>https://doi.org/10.1038/srep42717</u>
- Dashtipour, K., Chung, J. S., Wu, A. D., & Lew, M. F. (2007). Amantadine and Anticholinergics. In: Pahwa, RLyons, K.E.. (eds), *Handbook of Parkinson's Disease*, 293-308.
- Dassault SystèmesBIOVIA,. (2020). BIOVIA Discovery Studio Visualiser, v20.1..0.19295, San Diego, CA, USA.
- Duffy, S. (2018). Why are RNA virus mutation rates so damn high? *PLoS Biology*, *16*(8), e3000003. <u>https://doi.org/10.1371/journal.pbio.3000003</u>

Efferth, T., & Koch, E. (2011). Complex interactions between phytochemicals. The multitarget therapeutic concept of phytotherapy. *Current Drug Targets*, *12*(1), 122-132. https://doi.org/10.2174/138945011793591626

- Forli, S., Huey, R., Pique, M. E., Sanner, M. F., Goodsell, D. S., & Olson, A. J. (2016). Computational protein–ligand docking and virtual drug screening with the AutoDock suite. *Nature Protocols*, 11(5), 905-919. <u>https://doi.org/10.1038/nprot.2016.051</u>
- Frieman, M., Yount, B., Agnihothram, S., Page, C., Donaldson, E., Roberts, A., Vogel,
- L., Woodruff, B., Scorpio, D., Subbarao, K., & Baric, R. S. (2012). Molecular determinants of severe acute respiratory syndrome coronavirus pathogenesis and virulence in young and aged mouse models of human disease. *Journal of Virology*, 86(2), 884-897. <u>https://doi.org/10.1128/JVI.05957-11</u>

Ganjhu, R. K., Mudgal, P. P., Maity, H., Dowarha, D., Devadiga, S., Nag, S., &Arunkumar, G. (2015). Herbal plants and plant preparations as remedial approach for viral diseases. *VirusDisease*, *26*(4), 225-236. <u>https://doi.org/10.1007/s13337-015-0276-6</u>

Gao, Y., Yan, L., Huang, Y., Liu, F., Zhao, Y., Cao, L., Wang, T., Sun, Q., Ming, Z., Zhang, L., Ge, J., Zheng, L., Zhang, Y., Wang, H., Zhu, Y., Zhu, C., Hu, T., Hua, T., Zhang, B., Yang, X., Li, J., Yang, H., Liu, Z., Xu, W., Guddat, L. W., Wang, Q., Lou, Z., & Rao, Z., (2020). Structure of RNA-dependent RNA polymerase from 2019nCoV, a major antiviral drug target. *BioRxiv*. https://doi.org/10.1101/2020.03.16.993386

Gautret, P., Lagier, J. C., Parola, P., Meddeb, L., Mailhe, M., Doudier, B., Courjon, J., Giordanengo, V., Vieira, V. E., Dupont, H. T., &Honoré, S. (2020).
Hydroxychloroquine and azithromycin as a treatment of COVID-19: results of an open-label non-randomized clinical trial. *International Journal of Antimicrobial Agents*, 105949. <u>https://doi.org/10.1016/j.ijantimicag.2020.105949</u>

Grover, A., Agrawal, V., Shandilya, A., Bisaria, V. S., & Sundar, D. (2011). Nonnucleosidic inhibition of Herpes simplex virus DNA polymerase: mechanistic insights into the anti-herpetic mode of action of herbal drug withaferin A. *BMC Bioinformatics*. *12*, S22. <u>https://doi.org/10.1186/1471-2105-12-S13-S22</u>

Harvey, A. L. (2008). Natural products in drug discovery. *Drug Discovery Today*, *13*(19-20), 894-901. <u>https://doi.org/10.1016/j.drudis.2008.07.004</u>

Huang, J., Song, W., Huang, H., & Sun, Q. (2020). Pharmacological Therapeutics Targeting RNA-Dependent RNA Polymerase, Proteinase and Spike Protein: From Mechanistic Studies to Clinical Trials for COVID-19. *Journal of Clinical Medicine*, 9(4), 1131. <u>https://doi.org/10.3390/jcm9041131</u>

Jadhav, P., Kapoor, N., Thomas, B., Lal, H., &Kshirsagar, N. (2012). Antiviral potential

- of selected Indian medicinal (ayurvedic) plants against herpes simplex virus 1 and 2. North American Journal of Medical Sciences, 4(12), 641-647. https://doi.org/10.4103/1947-2714.104316
- Jain, J., Narayanan, V., Chaturvedi, S., Pai, S., & Sunil, S. (2018). In vivo evaluation of Withaniasomnifera-based Indian traditional formulation (AmukkaraChoornam), against chikungunya virus-induced morbidity and arthralgia. *Journal of Evidence-Based Integrative Medicine*, 23, 1-7. <u>https://doi.org/10.1177/2156587218757661</u>
- Jassim, S. A. A., &Naji, M. A. (2003). Novel antiviral agents: a medicinal plant perspective. *Journal of Applied Microbiology*, 95(3), 412-427. https://doi.org/10.1046/j.1365-2672.2003.02026.x

Kadioglu, O., Saeed, M., Johannes Greten, H., &Efferth, T. (2020). Identification of compounds against three targets of SARS CoV-2 coronavirus by combined virtual screening and supervised machine learning. [Preprint]. *Bulletin of the World Health Organization*. https://doi.org/10.2471/BLT.20.255943

Katiyar, C., Gupta, A., Kanjilal, S., &Katiyar, S. (2012). Drug discovery from plant sources: An integrated approach. *Ayu*, 33(1), 10-19. <u>https://doi.org/10.4103/0974-8520.100295</u>

- Kaul, T. N., Middleton Jr, E., &Ogra, P. L. (1985). Antiviral effect of flavonoids on human viruses. *Journal of Medical Virology*, 15(1), 71-79. https://doi.org/10.1002/jmv.1890150110
- Khan, Y. D., &Roomi, M. S. (2020). Promising compounds for treatment of COVID-19. VAWKUM Transactions on Computer Sciences, 17(1), 1-8.
- Kim, Y., Jedrzejczak, R., Maltseva, N. I., Wilamowski, M., Endres, M., Godzik, A., Michalska, K., &Joachimiak, A. (2020). Crystal structure of Nsp15 endoribonuclease NendoU from SARS-CoV-2. *Protein Science*. https://doi.org/10.1002/pro.3873

Kutluay, S. B., Doroghazi, J., Roemer, M. E., &Triezenberg, S. J. (2008). Curcumin inhibits herpes simplex virus immediate-early gene expression by a mechanism independent of p300/CBP histone acetyltransferase activity. *Virology*, 373(2), 239-24. <u>https://doi.org/10.1016/j.virol.2007.11.028</u>

Li, Y., Liu, X., Guo, L., Li, J., Zhong, D., Zhang, Y., Clarke, M., &Jin, R. (2020). Traditional Chinese herbal medicine for treating novel coronavirus (COVID-19) pneumonia: protocol for a systematic review and meta-analysis. *Systematic Reviews*, 9, 75. <u>https://doi.org/10.1186/s13643-020-01343-4</u>

Lin, L. T., Chen, T. Y., Lin, S. C., Chung, C. Y., Lin, T. C., Wang, G. H., Anderson, R., Lin, C. C., & Richardson, C. D. (2013). Broad-spectrum antiviral activity of

- chebulagic<br/>entry. BMCacid and punicalagin against viruses that use glycosaminoglycans for<br/>Microbiology, 13, 187. <a href="https://doi.org/10.1186/1471-2180-13-187">https://doi.org/10.1186/1471-2180-13-187</a>
- Lin, L. T., Hsu, W. C., & Lin, C. C. (2014). Antiviral natural products and herbal medicines. *Journal of Traditional and Complementary Medicine*, 4(1), 24-35. https://doi.org/10.4103/2225-4110.124335
- Lipinski, C. A., Lombardo, F., Dominy, B. W., & Feeney, P. J. (1997). Experimental and computational approaches to estimate solubility and permeability in drug discovery and development settings. *Advanced Drug Delivery Reviews*, 23(1-3), 3-25. <u>https://doi.org/10.1016/S0169-409X(96)00423-1</u>

Liu, T., Oprea, T., Ursu, O., Hasselgren, C., & Altman, R. B. (2016). Estimation of maximum recommended therapeutic dose using predicted promiscuity and potency. *Clinical and Translational Science*, 9(6), 311-320. https://doi.org/10.1111/cts.12422 Loureiro, D. R., Soares, J. X., Costa, J. C., Magalhães, Á. F., Azevedo, C. M., Pinto, M. M., & Afonso, C. M. (2019). Structures, activities and drug-likeness of antiinfective xanthone derivatives isolated from the marine environment: a

review. *Molecules*, 24(2), 243. https://doi.org/10.3390/molecules24020243

McKee, D. L., Sternberg, A., Stange, U., Laufer, S., &Naujokat, C. (2020). Candidate drugs against SARS-CoV-2 and COVID-19. *Pharmacological Research*, *157*, 104859. https://doi.org/10.1016/j.phrs.2020.104859

- Mohanraj, K., Karthikeyan, B. S., Vivek-Ananth, R. P., Chand, R. B., Aparna, S. R., Mangalapandi, P., &Samal, A. (2018). IMPPAT: A curated database of Indian medicinal plants, phytochemistry and therapeutics. *Scientific Reports*, 8, 4329. https://doi.org/10.1038/s41598-018-22631-z
- Morris, G. M., Huey, R., Lindstrom, W., Sanner, M. F., Belew, R. K., Goodsell, D. S., & Olson, A. J. (2009). AutoDock4 and AutoDockTools4: Automated docking with selective receptor flexibility. *Journal of Computational Chemistry*, 30(16), 2785-2791. <u>https://doi.org/10.1002/jcc.21256</u>
- Moulton, B. C., & Fryer, A. D. (2011). Muscarinic receptor antagonists, from folklore to pharmacology; finding drugs that actually work in asthma and COPD. *British Journal of Pharmacology*, *163*(1), 44-52. <u>https://doi.org/10.1111/j.1476-5381.2010.01190.x</u>
- Nadkarni, K. M., & Nadkarni, A. K. (1976). *Indian Materia Medica*, Vol 1, 3rd ed., M/S Popular PrakasanPvt. Ltd., Mumbai, India.
- Pachetti, M., Marini, B., Benedetti, F., Giudici, F., Mauro, E., Storici, P., Masciovecchio,
- C., Angeletti, S., Ciccozzi, M., Gallo, R. C., & Zella, D. (2020). Emerging SARS-CoV-2 mutation hot spots include a novel RNA-dependent-RNA polymerase variant. *Journal of translational medicine*, *18*(1), 1-9. <u>https://doi.org/10.1186/s12967-020-02344-6</u>
- Pachetti, M., Marini, B., Benedetti, F., Giudici, F., Mauro, E., Storici, P., Masciovecchio,
- C., Angeletti, S., Ciccozzi, M., Gallo, R.C. and Zella, D., 2020. Emerging SARS-CoV-2 mutation hot spots include a novel RNA-dependent-RNA polymerase variant. *Journal of translational medicine*, *18*, 179. <u>https://doi.org/10.1186/s12967-020-02344-6</u>

Pathak, K., & Zaman, K. (2013). Antiviral activity of some Indian medicinal herbs. *Indian Journal of Pharmacy Practice*, 6(2), 13-15.

- Pathania, S., Ramakrishnan, S. M., Randhawa, V., &Bagler, G. (2015). SerpentinaDB: a database of plant-derived molecules of Rauvolfiaserpentina. *BMC Complementary and Alternative Medicine*, 15, 262. <u>https://doi.org/10.1186/s12906-015-0683-7</u>
- Pavelka, A., Chovancova, E., &Damborsky, J. (2009). HotSpot Wizard: a web server for identification of hot spots in protein engineering. *Nucleic Acids Research*, 37(suppl\_2), W376-W383. <u>https://doi.org/10.1093/nar/gkp410</u>
- Peck, K. M., &Lauring, A. S. (2018). Complexities of viral mutation rates. Journal of Virology, 92(14), e01031-17. <u>https://doi.org/10.1128/JVI.01031-17</u>

Polur, H., Joshi, T., Workman, C. T., Lavekar, G., & Kouskoumvekaki, I. (2011). Back to

the roots: prediction of biologically active natural products from Ayurveda traditional medicine. *Molecular Informatics*, *30*(2-3), 181-187. <u>https://doi.org/10.1002/minf.201000163</u>

Rajkumar, V., Guha, G., & Kumar, R. A. (2011). Antioxidant and anti-neoplastic activities of *Picrorhizakurroa* extracts. *Food and Chemical Toxicology*, 49(2), 363-369. <u>https://doi.org/10.1016/j.fct.2010.11.009</u>

- Reiter, R. J., Tan, D. X., Zhou, Z., Cruz, M. H. C., Fuentes-Broto, L., &Galano, A. (2015). Phytomelatonin: assisting plants to survive and thrive. *Molecules*, 20(4), 7396-7437. <u>https://doi.org/10.3390/molecules20047396</u>
- Sadowski, J., Gasteiger, J., &Klebe, G. (1994). Comparison of automatic threedimensional model builders using 639 X-ray structures. *Journal of Chemical*

Information and Computer https://doi.org/10.1021/ci00020a039

Saini, A. (2016). Physicians of ancient India. Journal of Family Medicine and Primary Care, 5(2), 254-258. https://doi.org/10.4103/2249-4863.192322

- Saldaño, T. E., Monzon, A. M., Parisi, G., & Fernandez-Alberti, S. (2016). Evolutionary conserved positions define protein conformational diversity. *PLoS Computational Biology*, 12(3), e1004775. <u>https://doi.org/10.1371/journal.pcbi.1004775</u>
- Salehi, B., Kumar, N. V. A., Şener, B., Sharifi-Rad, M., Kılıç, M., Mahady, G. B., Vlaisavljevic, S., Iriti, M., Kobarfard, F., Setzer, W. N., & Avatollahi, S. A. (2018). Medicinal plants used in the treatment of human immunodeficiency virus. International Molecular Journal of Sciences, 19(5), 1459. https://doi.org/10.3390/ijms19051459
- Schyman, P., Liu, R., Desai, V., &Wallqvist, A. (2017). vNN web server for ADMET predictions. *Frontiers* in *Pharmacology*, 8, 889. https://doi.org/10.3389/fphar.2017.00889
- Shang, J., Ye, G., Shi, K., Wan, Y., Luo, C., Aihara, H., Geng, Q., Auerbach, A., & Li, F. (2020). Structural basis of receptor recognition by SARS-CoV-2. *Nature*, 581, 221-224. <u>https://doi.org/10.1038/s41586-020-2179-y</u>
- Stratton, C. F., Newman, D. J., & Tan, D. S. (2015). Cheminformatic comparison of approved drugs from natural product versus synthetic origins. *Bioorganic & Medicinal Chemistry Letters*, 25(21), 4802-4807. <u>https://doi.org/10.1016/j.bmcl.2015.07.014</u>

Tahir ul Qamar, M., Maryam, A., Muneer, I., Xing, F., Ashfaq, U. A., Khan, F. A.,
Anwar, F., Geesi, M. H., Khalid, R. R., Rauf, S. A., & Siddiqi, A. R. (2019).
Computational screening of medicinal plant phytochemicals to discover potent panserotype inhibitors against dengue virus. *Scientific Reports*, 9, 1443. https://doi.org/10.1038/s41598-018-38450-1

Teague, S. J., Davis, A. M., Leeson, P. D., &Oprea, T. (1999). The design of leadlike combinatorial libraries. *AngewandteChemie International Edition*, 38(24), 3743-3748. <u>https://doi.org/10.1002/(SICI)1521-3773(19991216)38:24%3C3743::AID-ANIE3743%3E3.0.CO;2-U</u>

The Lancet. (2020). COVID-19: fighting panic with information. *Lancet*, 395(10224), 537. <u>https://doi.org/10.1016/S0140-6736(20)30379-2</u>

World Health Organization. (2020). Coronavirus disease 2019 (COVID-19): situation report, 35.<u>https://www.who.int/docs/default-source/coronaviruse/situation-</u>

reports/20200224-sitrep-35-covid-19.pdf?sfvrsn=1ac4218d\_2(Published online February 24<sup>th</sup>, 2020; Date of access: May 14<sup>th</sup>, 2020).

Wu, C., Liu, Y., Yang, Y., Zhang, P., Zhong, W., Wang, Y., Wang, Q., Xu, Y., Li, M.,
Li, X., & Zheng, M. (2020). Analysis of therapeutic targets for SARS-CoV-2 and
discovery of potential drugs by computational methods. *Acta PharmaceuticaSinica B*. https://doi.org/10.1016/j.apsb.2020.02.008

Zhao, C., Ye, Z., Ma, Z. X., Wildman, S. A., Blaszczyk, S. A., Hu, L., Guizei, I. A., & Tang, W. (2019). A general strategy for diversifying complex natural products to polycyclic scaffolds with medium-sized rings. *Nature Communications*, 10(1), 1-10. <u>https://doi.org/10.1038/s41467-019-11976-2</u>

#### **Supplemental Materials**

Nature to Nurture- Identifying phytochemicals from Indian medicinal plants as prophylactic medicine by rational screening to be potent against multiple drug targets of SARS-CoV-2

Authors: Pratap Kumar Parida,<sup>1#</sup>, Dipak Paul <sup>2#</sup>, Debamitra Chakravorty <sup>3\*#</sup>

<sup>1</sup>Pratap Kumar Parida, PhD (Research Scientist - Computational Biology & Bioinformatics) Noor Enzymes Private Limited, 37-B, Darga Road, Kolkata - 700 017, India

<sup>2</sup>Dipak Paul, PhD (Scientist - Technical Services) Noor Enzymes Private Limited, 37-B, Darga Road, Kolkata - 700 017, India

<sup>3\*</sup>Corresponding author Debamitra Chakravorty, PhD (Project Lead - Computational Biology) Novel Techsciences (OPC) Private Limited, 37-B, Darga Road, 1st Floor, Kolkata - 700 017, India E-mail: dc@noveltechsciences.com

<sup>#</sup>All the authors have contributed equally to the paper.

Sources	Top 10 hits of phytochemicals
Withania somnifera	37
Azadirachta indica	19
Catharanthus roseus	9
Terminalia arjuna	7
Aloe barbadensis	5
Nigella sativa	5
Solanum nigrum	5
Rauvolfia serpentina	4
Anodendron paniculatum	2
Aconitum heterophyllum	1
Aegle marmelos	1
Alstonia venenata	1
Bacopa monnieri	1
Centella asiatica	1
Phyllanthus amarus	1
Swertia chirayita	1
Allium sativum	0
Aloe vera	0
Amaranthus tricolor	0
Andrographis paniculata	0
Annona reticulate	0

Table S1: Sources of phytochemicals from Indian Medicinal plants reported for their anti-viral properties

Aquilaria agallocha	0
Artemisia annua	0
Asparagus racemosus	0
Cassia angustifolia	0
Cinnamomum verum	0
Crocus sativus	0
Curcuma longa	0
Cymbopogon flexuosus	0
Cymbopogon martinii	0
Eucalyptus citriodora	0
Gymnema sylvestre	0
Hedychium spicatum	0
Hibiscus rosa-sinensis	0
Inula racemosa	0
Litsea glutinosa	0
Mucuna pruriens	0
Oroxylum indicum	0
Pelargonium graveolens	0
Phyllanthus emblica	0
Picrorhiza kurroa	0
Piper betle	0
Piper longum	0
Plantago ovata	0
Plumbago zeylanica	0
Pogostemon cablin	0
Pterocarpus marsupium	0
Santalum album	0

Saraca asoca	0	
Smilax china	0	
Terminalia bellirica	0	
Terminalia chebula	0	
Tinospora cordifolia	0	
Valeriana jatamansi	0	
Vetiveria zizanioides	0	

#### Table S2: Grid parameters of targets

Protein Name	XYZ Coordinates of Grid Centre (Å)	Grid Spacing (Å)
Main Protease (PDB; 6lu7)	-19.417, 12.599, 63.803	0.425
Spike (PDB 6LZG)	-35.544, 34.286, 6.97	0.403
NSP 10 (PDB 6w4h)	67.879, 21.337, 12.249	0.375
NSP12_d1 (RNA Binding Site) (PDB 6M71)	122.503, 108.491, 131.873	0.792
NSP12_d2 (NSP12-NSP7 Interface) (PDB 6M71)	117.599, 107.015, 130.936	0.375
NSP12_d2 (NSP12-NSP8 Interface) (PDB 6M71)	120.051, 141.341, 138.381	0.375
NSP16 (PDB 6w4h)	90.908, 18.518, 23.35	0.509
NSP9 (PDB 6w4b)	45.366, -17.534, 18.126	0.375
NSP15 (PDB 6w01)	-64.205, 69.315, 34.572	0.375
NSP3 (PDB 6w02)	2.622, -5.785, -21.408	0.375

Compound name	Molecular weight	Chemical formula	*References	FDA Approval status	COVID-19 FDA approval status	No. of citations (2019- 2020)
Chloroquine phosphate	515.9 g/mol	C18H32CIN3O8P2	Gao et al., 2020	Approved	Emergency Use Authorization (EUA)	382
Chloroquine	319.9 g/mol	C18H26CIN3	Dong et al., 2020; Wang et al., 2020	Approved	Emergency Use Authorization (EUA)	856
Arbidol	477.4 g/mol	C22H25BrN2O3S	Dong et al., 2020	Not approved	Not approved	374
Remdesivir	602.6 g/mol	C27H35N6O8P	Dong et al., 2020; Wang et al., 2020	Approved (emergency use)	Emergency Use Authorization (EUA)	981
Favipiravir	157.1 g/mol	C5H4FN3O2	Dong et al., 2020	Not approved yet (FDA Phase II and Phase III clinical trials)	Not Yet Approved	442
Lopinavir	628.8 g/mol	C37H48N4O5	Lu, 2020	Approved	Not Yet Approved	849
Ritonavir	720.9 g/mol	C37H48N6O5S2	Lu, 2020	Approved	Not Yet Approved	897
Ribavirin	244.2 g/mol	C8H12N4O5	Lu, 2020	Approved	Not Yet Approved	552
Oseltamivir	312.4 g/mol	C16H28N2O4	Lu, 2020	Approved	Not Yet Approved	486
Zanamivir	332.31 g/mol	C12H20N4O7	Lu, 2020	Approved	Not Yet Approved	47
Peramivir	328.41 g/mol	C15H28N4O4	Lu, 2020	Approved	Not Yet Approved	43
Ganciclovir	255.23 g/mol	C9H13N5O4	Chen et al., 2020	Approved	Not Yet Approved	114
Methylprednisolone	374.5 g/mol	C22H30O5	Chen et al., 2020	Approved	Not Yet Approved	450
Dexamethasone	392.5 g/mol	C22H29FO5	Chen et al., 2020	Approved	Not Yet Approved	191
Baricitinib	371.4 g/mol	C16H17N7O2S	Stebbing et al., 2020	Approved	Not Yet Approved	207
Hydroxychloroquin e	335.9 g/mol	C18H26CIN3O	Colson et al., 2020	Approved	Emergency Use Authorization (EUA)	1,280
Sofosbuvir	529.5 g/mol	C22H29FN3O9P	Elfiky, 2020	Approved	Not Yet Approved	107

#### Table S3 A: Drugs used in this study for comparison based on reported drugs in literature being tested for efficacy against COVID 19

Interferon alfa-2b	746.5 g/mol	C16H17Cl3I2N3NaO5	Xu et al., 2020	Approved	Not Yet Approved	40
		S				
Camostat mesylate	494.5 g/mol	C21H26N4O8S	Hoffmann et al., 2020	Not approved	Not Yet Approved	154
Darunavir	547.7 g/mol	C27H37N3O7S	Liu et al., 2020	Approved	Not Yet Approved	290
Galidesivir / BCX- 4430	265.27 g/mol	C11H15N5O3	Liu et al., 2020	Approved	Not Yet Approved	80

Drugs	Protease (PDB: 6lu7)	Spike (PDB 6LZG)	NSP10 (PDB 6w4h)	NSP12 (RNA- Binding site) (PDB 6m71)	NSP12- NSP7 Interface (PDB 6M71)	NSP12- NSP8 Interfac e (PDB 6M71)	NSP16 (PDB: 6w4h)	NSP9 (PDB 6w4b)	NSP15 (PDB 6w01)	NSP3 (PDB 6w02)
Chloroquine phosphate	-7.37	-5.55	-7.38	-5.15	-6.7	-7.21	-9.54	-4.79	-5.8	-8.07
Chloroquine	-7	-5.27	-7.13	-4.77	-6.76	-7.21	-10.11	-4.3	-6.23	-8.35
Arbidol	-7.1	-5.18	-7.53	-5.75	-6.25	-6.44	-8.98	-5.21	-5.98	-7.8
Remdesivir	-6.06	-5.32	-3.74	-3.66	-4.26	-6.1	-6.09	-3.4	-3.42	-9.79
Favipiravir	-4.05	-4.94	-4.04	-3.76	-4.94	-4.6	-5.25	-3.43	-4.08	-4.77
Lopinavir	-6.5	-3.77	-4.46	-2.31	-6.79	-8.33	-9.41	-5.21	-4.62	-9.36
Ritonavir	-6.43	-3.22	-3.91	-2.77	-3.68	-5.68	-8.1	-3.25	-4.69	-7.2
Ribavirin	-5.14	-4.44	-5	-2.97	-3.83	-4.7	-5.99	-3.44	-4.39	-6.17
Oseltamivir	-6.42	-5.21	-6.51	-4.65	-5.93	-6.32	-8.04	-3.77	-6.04	-6.22
Zanamivir	-3.18	-2.66	-5.11	-1.14	-4.39	-2.95	-3.02	-2.19	-4.11	-3.19
Peramivir	-5.32	-4.47	-6.46	-2.97	-5.96	-7.08	-5.34	-3.83	-4.46	-6.99
Ganciclovir	-3.72	-4.12	-4.57	-2.35	-3.45	-3.77	-4.07	-3.04	-3.62	-7.2
Methylprednisolone	-8.57	-5.77	-8.92	-6.54	-7.18	-7.93	-7.61	-5.88	-5.61	-7.98
Dexamethasone	-7.08	-5.32	-7.23	-5.4	-6.29	-7.17	-8.21	-5.82	-7.08	-9.56
Baricitinib	-8.46	-7.62	-9.22	-5.87	-6.54	-7.88	-9.08	-6	-6.76	-10.38
Hydroxychloroquine	-7.02	-4.6	-7.2	-5.23	-7.73	-7.12	-8.58	-5.63	-4.86	-7.63
Sofosbuvir	-5.76	-2.84	-4.85	-3.94	-4.57	-5.37	-6.84	-5.18	-5.09	-9.09
Interferon alfa-2b	-6.35	-5.58	-7.39	-4	-5.89	-6.57	-6.29	-5.13	-4.67	-8.04
Camostat mesylate	-7.19	-5.49	-7.85	-4.18	-6.2	-7.17	-8.14	-5.98	-6.07	-10.2
Darunavir	-8.03	-5.5	-5.2	-3	-5.74	-7.4	-7.86	-5.66	-5.33	-7.02
Galidesivir / BCX-4430	-5.76	-5.56	-6.32	-4.92	-5.97	-6.29	-7.01	-3.74	-5.46	-6.26

Table S3 B: The docking scores of 21 FDA approved drugs found effective against SARS-COV 2

Molecule	Dockin	Lipinski	Ghose	Veber	Egan	Muegge	Bioavailabili	PAIN	Bren	Leadlikene
	g score	#violatio	#violatio	violatio	violatio	violatio	ty Score	S	k	SS
	kcal/m	ns	ns	ns	ns	ns		alerts	alert	#violations
	ol		Protea	)SA					S	
	0.02	1	1	1		1	0.55		0	2
7-Benzoyl-17-hydroxynimbocinol	-9.03	1	4	0	0	1	0.55	0	0	2
Serpentinine	-13	1	3	0	0	3	0.56	1	3	2
Limocinin	-8.83	2	4	0	1	1	0.17	0	1	2
27-Deoxy-14-hydroxywithaferin A	-10.8	0	1	0	0	0	0.55	0	1	1
Terchebulin	-10.86	3	3	1	1	5	0.17	1	4	1
17-Hydroxywithaferin	-10.08	0	1	0	0	0	0.55	0	1	2
Nimolicinol	-10.09	0	1	0	0	0	0.55	0	1	2
Nimocin	-9.92	1	4	0	1	1	0.55	0	1	2
7-Deacetyl-7-benzoylgedunin	-10.75	1	3	0	0	1	0.55	0	2	2
Withanolide	-9.63	0	1	0	0	0	0.55	0	1	1
	•		Spik	e	1					•
Serpentinine	-6.04	1	3	0	0	3	0.56	1	3	2
Terchebulin	-8.73	3	3	1	1	5	0.17	1	4	1
27-Deoxywithaferin A	-7.82	0	1	0	0	0	0.55	0	1	2
27-Hydroxywithanone	-8.47	0	2	0	0	0	0.55	0	1	1
Hydroxywithanolide B	-8.28	1	2	0	1	0	0.55	0	1	1
Deoxywithastramonolide	-8.27	0	1	0	0	0	0.55	0	1	2
2,3-Dihydrowithaferin A	-7.45	0	1	0	0	0	0.55	0	1	2
	•		NSP	10	1					•
MELIANONE	-11.13	1	3	0	1	1	0.55	0	2	2
27-Hydroxywithanolide B	-8.09	0	1	0	0	0	0.55	0	1	2
Anaferine	-6.22	0	0	0	0	0	0.55	0	0	1

Table S4: Top hits of Phytochemicals from Indian medicinal plants after re-docking (100 LGA runs) with their ADME properties drom SwissADME.

Amyrin acetate	-10.97	1	3	0	1	1	0.55	0	1	2
Nimocin	-10.21	1	4	0	1	1	0.55	0	1	2
Lophenol	-9.48	1	2	0	1	2	0.55	0	1	2
Meldenin	-10.46	0	1	0	0	0	0.55	0	1	2
Epoxyazadiradione	-8.29	0	0	0	0	0	0.55	0	1	2
			NSI	P 12 D1						
Ashwagandhanolide	-6.83	3	4	1	2	6	0.17	0	1	3
Pelankine	-8.49	1	3	0	0	2	0.55	0	3	2
beta-Amyrin	-8.24	1	3	0	1	2	0.55	0	1	2
Terchebulin	-10.16	3	3	1	1	5	0.17	1	4	1
Solasodine	-8.19	1	1	0	0	1	0.55	0	1	2
Somniferine	-8.15	1	3	0	0	2	0.55	0	0	1
12-Deoxywithastramonolide	-7.58	0	1	0	0	0	0.55	0	1	2
Spirostan-3-ol	-7.28	1	2	0	0	1	0.55	0	0	2
Campestanol	-8.18	1	2	0	1	2	0.55	0	0	2
		_	NSI	P 12 D2				I		
Terchebulin	-10.16	3	3	1	1	5	0.17	1	4	1
1,3,6,8-Tetranitro-4,5-Dihydroxy-2- Hydroxymethylanthraquinones	-10.35	1	0	1	1	2	0.55	1	2	1
Withastramonolide	-9.72	0	2	0	0	0	0.55	0	1	1
Somniferine	-8.15	1	3	0	0	2	0.55	0	0	1
Withanolide B	-9.2	0	1	0	0	0	0.55	0	1	2
Withanolide R	-7.63	0	1	0	0	0	0.55	0	1	2
12-Deoxywithastramonolide	-9.05	0	1	0	0	0	0.55	0	1	2
Withaferin A	-7.9	0	1	0	0	0	0.55	0	1	2
Lutein	-8.94	2	4	0	1	1	0.17	0	2	3
	1		NS	P12 D3		I	I	I	1	1

Terchebulin	-12.46	3	3	1	1	5	0.17	1	4	1
Ashwagandhanolide	-12.09	3	4	1	2	6	0.17	0	1	3
Pelankine	-11.06	1	3	0	0	2	0.55	0	3	2
Serpentinine	-10.7	1	3	0	0	3	0.56	1	3	2
7-Deacetyl-7-benzoylgedunin	-10.89	1	3	0	0	1	0.55	0	2	2
Somniferine	-10.7	1	3	0	0	2	0.55	0	0	1
Withaferin A	-9.94	0	1	0	0	0	0.55	0	1	2
3-Methoxy-5,7-dimethyl-2-naphthyl 6-O- [(10eta,13alpha)-20-oxopimaran-20-yl]-L- glucopyranoside	-10.24	1	4	0	1	2	0.55	0	0	3
Dammarane	-10.82	1	3	0	1	2	0.55	0	0	2
			NSP	16						
Serpentinine	-12.1	1	3	0	0	3	0.56	1	3	2
Solasodine	-11.81	1	1	0	0	1	0.55	0	1	2
Ashwagandhanolide	-10.84	3	4	1	2	6	0.17	0	1	3
Solvanol	-10.98	0	1	0	0	0	0.55	0	1	2
(-)-Anaferine	-10.91	0	0	0	0	0	0.55	0	0	1
Phyllanthosterol	-10.84	1	3	0	1	2	0.55	0	1	2
Cycloartenol	-10.89	1	3	0	1	2	0.55	0	1	2
Limonin	-10.73	0	0	0	0	0	0.55	0	2	1
Isonimolicinolide	-10.73	1	3	0	1	0	0.55	0	1	1
veratridine	-11.3	3	3	1	1	5	0.17	0	0	1
			NSI	29						
7-Benzoyl-17-hydroxynimbocinol	-8.73	1	4	0	0	1	0.55	0	0	2
Nimocin	-8.82	1	4	0	1	1	0.55	0	1	2
Terchebulin	-8.71	3	3	1	1	5	0.17	1	4	1
Serpentinine	-7.89	1	3	0	0	3	0.56	1	3	2
12-Deoxywithastramonolide	-8.19	0	1	0	0	0	0.55	0	1	2

27-Hydroxywithanolide B	-8.28	0	1	0	0	0	0.55	0	1	2
Bacosine	-8.22	1	3	0	1	1	0.56	0	1	2
7-Deacetyl-7-benzoylepoxyazadiradione	-8.74	1	4	0	1	1	0.55	0	1	2
Azadiradionolide	-7.3	0	0	0	0	0	0.55	0	1	2
Taraxasterol acetate	-8.16	1	3	0	1	1	0.55	0	1	2
			N	SP 15						
Ashwagandhanolide	-10.17	3	4	1	2	6	0.17	0	1	3
Anodendroside A	-8.93	2	3	1	1	3	0.17	0	2	1
Anodendroside F	-8.94	2	3	1	1	3	0.17	0	2	1
Somnifericin	-7.21	0	3	0	0	0	0.55	0	0	1
Terchebulin	-8.72	3	3	1	1	5	0.17	1	4	1
Rescinnamine	-8.59	2	3	1	0	1	0.17	1	2	3
7-Deacetyl-7-benzoylgedunin	-8.38	1	3	0	0	1	0.55	0	2	2
Vindolinine	-6.87	0	0	0	0	0	0.55	0	1	0
campest-4-en-3-one	-7.92	1	2	0	1	2	0.55	0	0	2
		_	N	SP 3						
Delta5-Avenasterol	-11.35	1	3	0	1	2	0.55	0	1	2
Physagulin-D	-8.87	2	3	1	1	3	0.17	0	1	1
Withanolide P	-11.44	0	1	0	0	0	0.55	0	1	2
2,3-Dehydrosomnifericin	-12.3	0	3	0	0	0	0.55	0	0	1
Gramisterol	-11.22	1	3	0	1	2	0.55	0	1	2
Lophenol	-11.4	1	2	0	1	2	0.55	0	1	2
AC1L1UKB	-11.82	1	2	0	1	2	0.55	0	1	2
24,25-dihydrowithanolide D	-10.24	0	1	0	0	0	0.55	0	1	2
27-Deoxy-14-hydroxywithaferin A	-9.49	0	1	0	0	0	0.55	0	1	1
Nimocin	-11.78	1	4	0	1	1	0.55	0	1	2

Molecule	Lipinski #violatio ns	Ghose #violations	Veber #violations	Egan #violations	Muegge #violations	Bioavailabilit y Score	PAINS #alerts	Brenk #alerts	Leadlikenes s #violations	Synthetic Accessibilit v
Favipiravir	3	1	1	1	2	0.17	0	1	2	3.6
Chloroquine	0	0	0	0	0	0.55	0	0	2	2.76
Arbidol	0	0	0	0	0	0.55	1	0	3	3.57
Remdesivir	2	3	2	1	3	0.17	0	1	2	6.33
Favipiravir	0	4	0	0	1	0.55	0	0	1	2.08
Lopinavir	1	3	1	0	3	0.55	0	0	3	5.67
Ritonavir	2	4	2	1	4	0.17	0	0	3	6.45
Ribavirin	0	1	1	1	0	0.55	0	0	1	3.89
Oseltamivir	0	0	0	0	0	0.55	0	0	1	4.44
Zanamivir	2	1	1	1	3	0.17	0	2	0	4.96
Peramivir	0	0	1	1	1	0.55	0	2	1	4.31
Ganciclovir	0	1	0	1	0	0.55	0	0	0	2.61
Methylprednisolone	0	0	0	0	0	0.55	0	0	1	5.42
Dexamethasone	0	0	0	0	0	0.55	0	0	1	5.47
Baricitinib	0	0	0	0	0	0.55	0	0	1	3.07
Hydroxychloroquine	0	0	0	0	0	0.55	0	0	2	2.82
Sofosbuvir	2	1	2	1	2	0.17	0	1	2	6.02
Interferon alfa-2b	1	3	0	1	2	0.55	0	3	3	3.1
Camostat mesylate	1	1	1	1	1	0.55	0	4	2	3.46
Darunavir	1	3	2	1	0	0.55	0	1	2	5.67
Galidesivir	1	1	1	1	2	0.55	0	0	0	3.48

Table S5 A: ADMET profiles of the 21 drugs calculated from Siss ADME and Vnn-ADMET

Query	Cyto- toxicity	HLM	Cyp1A 2 Inhibit or	Cyp3A 4 Inhibit or	Cyp2D 6 Inhibit or	Cyp2C 9 Inhibit or	Cyp2C 19 Inhibit or	BBB	P-gp Inhibit or	P-gp Substra te	hERG Blocker	MMP	AMES	MRTD (mg/da y)
Chloroquine phosphate	No	Yes	No	No	No	No	No	Yes	No	Yes	No	No	Yes	430
Chloroquine	No	Yes	No	No	No	No	No	Yes	No	Yes	No	No	Yes	430
Arbidol	No	Yes	No	No	No	No	No	Yes	No	Yes	No	No	No	201
Remdesivir	Yes	Yes	No	No	No	No	No	No	Yes	Yes	No	No	No	229
Favipiravir	No	Yes	No	No	No	No	No	No	No	No	No	No	No	170
Lopinavir	No	Yes	No	No	No	No	No	No	Yes	Yes	No	Yes	No	327
Ritonavir	Yes	Yes	No	Yes	No	No	No	No	Yes	Yes	No	No	No	367
Ribavirin	No	Yes	No	No	No	No	No	No	No	No	No	No	No	10779
Oseltamivir	No	Yes	No	No	No	No	No	Yes	No	Yes	No	No	No	165
Zanamivir	No	Yes	No	No	No	No	No	No	No	No	No	No	No	19
Peramivir	No	Yes	No	No	No	No	No	No	No	Yes	No	No	No	19
Ganciclovir	No	Yes	No	No	No	No	No	No	No	No	No	No	Yes	183
Methylprednis olone	No	Yes	No	No	No	No	No	No	No	Yes	No	No	No	75
Dexamethason e	No	Yes	No	No	No	No	No	No	No	Yes	No	No	No	8.4
Baricitinib	No	Yes	No	No	No	No	No	No	Yes	No	Yes	No	Yes	136
Hydroxychloro quine	No	Yes	No	No	No	No	No	Yes	No	Yes	No	No	Yes	478
Sofosbuvir	Yes	Yes	No	No	No	No	No	Yes	Yes	No	No	No	No	277
Interferon alfa- 2b	No	Yes	No	Yes	No	No	No	No	No	No	No	Yes	No	217
Camostat mesylate	No	Yes	No	No	No	No	No	Yes	No	No	No	No	No	31
Darunavir	No	Yes	No	No	No	No	No	No	Yes	Yes	No	No	No	246
Galidesivir	No	Yes	No	No	No	No	No	No	No	No	No	No	No	164

 Table S5 B: Toxicity profiles of the 21 drugs calculated from Siss ADME and Vnn-ADMET

Phytochemicals	Cyto- toxici ty	HL M	Cyp1A 2 Inhibit	Cyp3A 4 Inhibit	Cyp2D 6 Inhibit	Cyp2C 9 Inhibit	Cyp2C 19 Inhibit	BB B	P-gp Inhibit or	P-gp Substr ate	hER G Block	MM P	AM ES	MRT D (mg/d
Serpentinine	No	Yes	or No	or Yes	or Yes	or No	or No	No	No	No	er No	No	No	ay) 0.99
Aegeline	No	Yes	Yes	No	No	No	No	Yes	No	No	No	No	No	43
Trypan Blue free acid	No	Yes	No	No	No	No	No	No	No	No	No	Yes	Yes	300
Terchebulin	No	Yes	No	Yes	No	No	No	No	No	No	No	No	No	501
27-Deoxywithaferin A	Yes	Yes	No	No	No	No	No	No	Yes	Yes	No	No	No	303
27-Hydroxywithanone	Yes	Yes	No	No	No	No	No	No	No	Yes	No	No	No	49
Melianin A	No	Yes	No	No	No	No	No	Yes	Yes	Yes	Yes	No	No	100
Hydroxywithanolide B	Yes	Yes	No	No	No	No	No	No	No	No	No	No	No	33
Deoxywithastramonolide	Yes	Yes	No	No	No	No	No	No	No	Yes	No	No	No	54
2,3-Dihydrowithaferin A	Yes	Yes	No	No	No	No	No	No	Yes	Yes	No	No	No	319
7-Benzoyl-17-hydroxynimbocinol	No	Yes	No	No	No	No	No	Yes	Yes	Yes	Yes	No	No	148
Limocinin	No	Yes	No	No	No	No	No	No	Yes	Yes	Yes	No	No	134
27-Deoxy-14-hydroxywithaferin A	Yes	Yes	No	No	No	No	No	No	No	No	No	No	No	8.7
17-Hydroxywithaferin	Yes	Yes	No	No	No	No	No	No	Yes	Yes	No	No	No	342
Nimolicinol	No	Yes	No	No	No	No	No	No	Yes	Yes	No	No	No	84
Nimocin	No	Yes	No	No	No	No	No	Yes	Yes	Yes	Yes	No	No	162
7-Deacetyl-7-benzoylgedunin	No	Yes	No	Yes	No	No	No	Yes	Yes	Yes	Yes	No	No	182
Withanolide	Yes	Yes	No	No	No	No	No	No	Yes	No	No	No	No	48
Solasodine	No	Yes	No	No	No	No	No	No	No	Yes	No	No	No	657
Ashwagandhanolide	Yes	Yes	No	No	No	No	No	No	Yes	Yes	No	No	No	189
Solvanol	No	Yes	No	No	No	No	No	No	No	Yes	No	No	No	473
(-)-Anaferine	No	Yes	No	No	No	No	No	Yes	No	No	Yes	No	No	196
Phyllanthosterol	No	Yes	No	No	No	No	No	Yes	No	Yes	No	No	No	452

 Table S6: Toxicity profile of the phytochemicals form Indian Medicinal plants

Cycloartenol	No	No	No	No	No	No	No	No	Yes	No	No	No	No	633
Limonin	No	Yes	No	Yes	No	No	No	No	No	No	No	No	No	172
Isonimolicinolide	No	Yes	No	No	No	No	No	No	Yes	Yes	No	No	No	66
veratridine	No	Yes	No	No	No	No	No	No	Yes	Yes	Yes	No	No	1.9
Ashwagandhanolide	Yes	Yes	No	No	No	No	No	No	Yes	Yes	No	No	No	189
Anodendroside A	No	Yes	No	No	No	No	No	No	No	Yes	No	Yes	No	0.07
Anodendroside F	No	Yes	No	No	No	No	No	No	No	Yes	No	Yes	No	0.07
Somnifericin	Yes	Yes	No	No	No	No	No	No	Yes	Yes	No	No	No	158
Rescinnamine	No	Yes	No	No	No	No	No	Yes	Yes	Yes	No	No	No	1.1
Vindolinine	No	Yes	No	No	No	No	No	No	No	Yes	No	No	No	102
campest-4-en-3-one	No	Yes	No	No	No	No	No	No	Yes	No	No	No	No	58
Pelankine	No	Yes	No	No	No	No	No	No	Yes	Yes	No	No	No	182
Somniferine	No	Yes	No	No	No	No	No	No	No	No	No	No	No	45
Withaferin A	Yes	Yes	No	No	No	No	No	No	Yes	Yes	No	No	No	342
3-Methoxy-5,7-dimethyl-2-naphthyl 6-O-[(10eta,13alpha)-20-oxopimaran- 20-yl]-L-glucopyranoside	No	Yes	No	No	No	No	No	No	No	No	No	No	No	157
Dammarane	No	No	No	No	No	No	No	No	Yes	Yes	No	Yes	Yes	59
1,3,6,8-Tetranitro-4,5-Dihydroxy-2- Hydroxymethylanthraquinones	No	Yes	No	Yes	No	Yes	No	No	No	No	No	Yes	Yes	329
Withastramonolide	Yes	Yes	No	No	No	No	No	No	No	No	No	No	Yes	60
Withanolide B	Yes	Yes	No	No	No	No	No	No	No	Yes	No	No	No	51
Withanolide R	Yes	Yes	No	No	No	No	No	No	No	Yes	No	No	No	46
12-Deoxywithastramonolide	Yes	Yes	No	No	No	No	No	No	No	Yes	No	No	No	54
Lutein	No	Yes	No	No	No	No	No	No	No	No	No	No	Yes	35
beta-Amyrin	No	No	No	No	No	No	No	No	No	No	No	No	No	691
Spirostan-3-ol	No	Yes	No	No	No	No	No	No	Yes	Yes	No	No	No	136
Campestanol	No	Yes	No	No	No	No	No	No	Yes	Yes	No	No	No	180

MELIANONE	Yes	Yes	No	No	No	No	No	No	No	No	No	No	No	310
27-Hydroxywithanolide B	Yes	Yes	No	No	No	No	No	No	No	Yes	No	No	No	54
Anaferine	No	Yes	No	No	No	No	No	Yes	No	No	Yes	No	No	196
Amyrin acetate	No	Yes	No	No	No	No	No	No	Yes	No	No	No	No	259
Hygrine	No	Yes	No	No	No	No	No	Yes	Yes	No	Yes	No	No	3.3
Lophenol	No	Yes	No	No	No	No	No	No	No	Yes	No	No	No	341
Meldenin	No	Yes	No	No	No	No	No	No	Yes	No	No	No	No	105
Epoxyazadiradione	No	Yes	No	Yes	No	No	No	Yes	Yes	No	No	No	No	131
Nimocin	No	Yes	No	No	No	No	No	Yes	Yes	Yes	Yes	No	No	162
Terchebulin	No	Yes	No	Yes	No	No	No	No	No	No	No	No	No	501
Bacosine	No	No	No	No	No	No	No	No	Yes	No	No	No	No	496
7-Deacetyl-7- benzoylepoxyazadiradione	No	Yes	No	Yes	No	No	No	Yes	Yes	Yes	Yes	No	No	182
Azadiradionolide	No	Yes	No	No	No	No	No	Yes	Yes	Yes	No	No	No	220
Taraxasterol acetate	No	Yes	No	No	No	No	No	No	Yes	No	No	No	No	102
Delta5-Avenasterol	No	Yes	No	No	No	No	No	No	No	Yes	No	No	No	191
Physagulin-D	Yes	Yes	No	No	No	No	No	No	No	Yes	No	No	No	2.6
2,3-Dehydrosomnifericin	Yes	Yes	No	No	No	No	No	No	Yes	Yes	No	No	No	127
Gramisterol	No	Yes	No	No	No	No	No	No	Yes	Yes	No	No	No	546
AC1L1UKB	No	Yes	No	No	No	No	No	No	No	Yes	No	No	No	313
24,25-dihydrowithanolide D	Yes	Yes	No	No	No	No	No	No	Yes	No	No	No	No	54



Figure1.tif (8.69 MiB)

view on ChemRxiv • download file



Figure2.tif (9.10 MiB)

view on ChemRxiv • download file

A) Spike with docked superimposed phytochemicals	н	
2	A) Spike Phytochemicals Repurposed Drugs Conservation Scores	R403 E406 Q409 K417/418 V453 L455 F456 E484 F400 Q403 S404 V405 Q406 Q408 T500 N501 V505
B) Main Protease with docked superimposed phytochem	nicals	
	B) Protease Phytochemicals Repurposed Drugs Conservation Scores	H41 M49 G143 C145 H163 M165 E166 P168 G189 T160 G182
C) NSP 10 with docked superimposed phytochemicals	C) NSP 10 Phytochemicals Repurposed Drugs Conservation Scores	A4277 K4281 H308 C4327 L4328 Y4329 H4336 P4337 F4342 C4343 D4344 L4345 T4364 L4365 N4367 T4368 V4369
D) NSP 12 D1 with docked superimposed phytochemica	ls	
	D) NSP 12 D1 Phytochemicals Repurposed Drugs Conservation Scores	0413 F415 L437 F440 F441 F442 F843
E) NSP 12 D2 with docked superimposed phytochemic	cals	
	E) NSP12 D2 Phytochemicals Repurposed Drugs Conservation Scores	D164 V166 V455 I553 W617 V619 P620 K621 C622 D623 I8624 C662 D760 D761 F793 5795 K798 W600 H810 E811 S814
F) NSP 12 D3 with docked superimposed phytochemic	cals	
	© NSP12D3 Phytochemicals Repurposed Drugs Conservation Scores	L271 Y273 P323 F326 P328 P329 P329 Y330 A383 V398 F396 8664 V675
G) NSP 16 with docked superimposed phytochemicals		
	G) NSP 16 Phytochemicals Repurposed Drugs Conservation Scores	D5897 L6898 Q6911 D6912 C6913 A6914 M5929 F6947 Y8950
H) NSP 3 with docked superimposed phytochemicals		
) NSP 9 with docked superimposed phytochemicals	HD NGP 3 Phytochemicals Repurposed Drugs Conservation Ocores	123 A38 V49 A50 P125 L128 A129 1131 F132 A154 V155 F156
	D NSP 9 Phytochemicals Repurposed Drugs Conservation Scores	C74 R75 F76 V77 L89 L104 L107 A108 V111 L113 Q114
J) NSP 15 with docked superimposed phytochemicals		
SE Er	J) NSP 15 Phytochemicals Repurposed Drugs Conservation Scores	11235 F241 8242 5244 0245 0247 0248 11250 K290 W333 T341 Y343
		Evolutionary conservation scores from Consurf
		1 2 3 4 5 4 7 8 7
		Interacting residues with Phytochemicals Interacting residues with repurposed drugs

Figure3.tif (3.09 MiB)

I.

view on ChemRxiv • download file