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Visible-Light-Promoted Metal-Free Ammoxidation of C(sp3)-H bonds

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The metal-free activation of C(sp³)-H bonds to value-added products is of paramount importance in organic synthesis. Herein, we report the use of the commercially available organic dye 2,4,6-triphenylpyrylium tetrafluoroborate (TPP) for the conversion of methylarenes to the corresponding aryl nitriles via a photo process. Applying this methodology, a variety of cyanobenzenes have been synthesized in good to excellent yield under metal- and cyanide-free conditions. We demonstrate the scope of the method with over 50 examples including late-stage functionalization of drug molecules (celecoxib) and complex structures such as L-menthol, amino acids and cholesterol derivatives. Further, the presented synthetic protocol is applicable for gram-scale reactions. In addition to methylarenes, selected examples for the cyanation of aldehydes, alcohols and oximes are demonstrated as well. Detailed mechanistic investigations have been carried out using time-resolved luminescence quenching studies, control experiments and NMR- spectroscopic as well as kinetic studies, all supporting the proposed catalytic cycle.

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Visible-Light-Promoted Metal-Free Ammoxidation of C(sp³)-H bonds

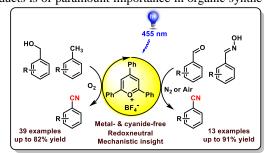
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KEYWORDS. Photoredox catalysis, nitriles, C-H functionalization, methylarenes, ammoxidation.

ABSTRACT: The metal-free activation of C(sp³)-H bonds to value-added products is of paramount importance in organic synthe-

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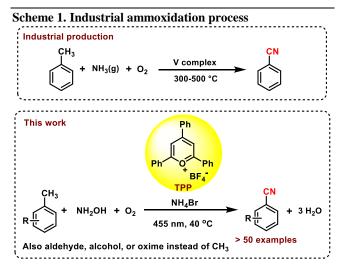


time-resolved luminescence quenching studies, control experiments and NMR- spectroscopic as well as kinetic studies, all supporting the proposed catalytic cycle.

INTRODUCTION

Transition-metal based catalysts have an indispensable role in several chemical reactions,¹ industrial production,^{1a-e, 1h} fine and bulk chemical synthesis.^{1t} However, due to their high price.² less abundance³ and toxicity.⁴ the interest in using alternative metal-free catalysts is growing within the scientific community, especially for the late-stage functionalization of inert $C(sp^3)$ -H bonds.⁵ During the last two decades, several transition-metal based catalysts^{1q, 6} have been developed for the activation of C-H bonds in sp^3 centres via thermal and photochemical pathways.^{1q, 6} One of the most important transformations is the conversion of petroleum by-products into fine chemicals,^{1a, 1b} e.g. the synthesis of benzonitrile from toluene.^{1b, 1c} Industrially, benzonitrile is produced by ammoxidation of toluene using a transition-metal catalyst (vanadium) and applying a high NH₃ and O₂ pressure at 300-500 °C.^{1b, 1c} Although the industrial ammoxidation process is an atom economic and sustainable route for the synthesis of nitriles starting from the unfunctionalized CH₃ group, but several functional groups are not tolerated due to the high reaction temperature and pressure.^{1b, 1c} The nitrile moiety is an essential functional group in various drugs and bioactive compounds,⁷ as well as an important building block for the preparation of fine and bulk chemicals, such as amines, carboxylic acids, amides and esters.8 In conventional approaches, nitriles have been synthesized using the Sandmeyer reaction,9 Rosenmundvon Braun reaction¹⁰ and halide/CN exchange.¹¹ Further, transition-metal and photoredox catalyzed ammoxidations of aldehydes have been reported.¹² A few examples for the synthesis of nitriles from sp^2 C-H bonds via photocatalysis were reported by Nicewicz¹³ and others.¹⁴ However, the abovementioned methods often suffer from several drawbacks, such as the use of transition metals leading to metal residue waste, high temperature, low selectivity or the generation of toxic cyanide. 11

The direct conversion of methylarenes to nitriles in a one-step procedure is highly desired, but one-pot protocols to prepare nitriles from toluene are underdeveloped.¹⁵ Wang et al. reported^{15a} a Palladium-catalyzed cyanation of methylarenes in 2013, using an over stoichiometric amount of *t*BuONO and NHPI at 70 °C.



Recently, Kang and co-workers^{15b} reported a similar strategy to Wang, where AlCl₃ was used as Lewis acid for the conversion of the oxime intermediates to nitriles at 80 °C. Major drawbacks of the reported methods are the harsh reaction conditions together with the use of hazardous reagents and transition metals.

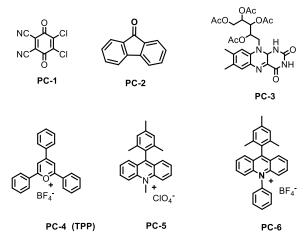
In principle, a greener way to produce nitriles is by an oxidation process using molecular oxygen or simply air. Interestingly, a few examples to synthesize nitriles from alcohols¹⁶ or amines¹⁷ employing oxygen as oxidant are known to date. However, the reported methods mainly rely on transition metal catalysts and applying harsh reaction conditions (> 100 $^{\circ}$ C).

We envisioned that an alternative metal- and cyanide-free methylarene to nitrile protocol could be developed by using a visible-light-assisted ammoxidation approach. Several challenges need to be addressed for this aim, such as the desired, yet difficult oxidation of the methylarene substrate (e.g. $E_{1/2}$ = +2.36 V vs SCE for toluene),¹⁸ especially in case of electron deficient substituents, opposed to a more facile oxidation of ammonia ($E_{1/2} = +0.63$ V vs SCE),¹⁹ and the prevention of an over oxidation of the methylarenes to the carboxylic acid. Herein we describe the use of the commercially available organic dye 2,4,6-triphenylpyrylium tetrafluoroborate (TPP)²⁰ for the synthesis of nitriles from methylarenes. In addition to this, a detailed mechanistic investigations was carried out to support our mechanistic hypothesis. All experimental results and spectroscopic analyses support the proposed catalytic cycle. To the best of our knowledge, this is the first example for a photocatalytic ammoxidation of methylarenes using abundant feedstock materials, ammonium salts and molecular oxygen without the use of metals and toxic reagents.

RESULTS AND DISCUSSION

At the start of our investigation, we tested different commercially available organic dyes for the desired transformation (Table 1).

Сн₃		Photocatalyst $\rm NH_2OH$ +HCl, $\rm NH_4Br$, 1 bar $\rm O_2$		
¢ ₽	•	ACN (0.05 M), 25 mg 4 Å MS 455 nm, 40 °C, 24 h		
1			2	
Entry	Photocatalyst (PC)	Ammonia source	Yield of 2 (%)	
1^{a}	PC-1	NH ₂ OH•HCl	26	
2 ^a	PC-2	NH ₂ OH•HCl	15	
3 ^a	PC-3	NH ₂ OH•HCl	35	
4 ^a	PC-4	NH2OH•HCl	76	
5 ^a	PC-5	NH ₂ OH•HCl	12	
6 ^a	PC-6	NH ₂ OH•HCl	14	
7 ^a	PC-4	NH ₄ (OAc) ₂	NR	
8 ^a	PC-4	Aq. NH ₃	NR	
9 ^a	PC-4	HCOONH ₄	NR	
10 ^a	PC-4	7N NH₃ in MeOH	NR	



Reaction conditions: ${}^{a}0.1$ mmol substrate, 20 mol% PC, 3 eq. ammonia source, 2.5 eq. NH₄Br, 25 mg 4 Å MS, 1 bar O₂, 2 mL acetonitrile (0.05 M), 455 nm, 40 °C, 24 h, yields were determined by GC using n-decane as standard.

The initial screening revealed DDQ and 9-fluorenone as ineffective catalysts for this reaction giving a yield of 26% and 15%, respectively (Table 1, entries 1-2). Interestingly, the use of RFTA led to a moderate yield of 35% (Table 1, entry 3), while the more sensitive photocatalyst TPP (PC-4) ($E_{1/2} = +$ 2.3 V),²¹ yielded 76% of methyl 4-cyanobenzoate (Table 1, entry 4). Surprisingly, employing acridinium photocatalysts $(E_{1/2} = + 2.28 \text{ V})^{1u}$ resulted in poor yields of 12% and 14%, respectively (Table 1, entries 5-6). Further, different ammonium salts were screened for the cyanation of methylarenes. Notably, merely hydroxylamine hydrochloride gave the desired product 2, while all other tested ammonium sources, such as ammonium acetate, aqueous ammonia, ammonium formate and methanolic ammonia failed to render the desired product (Table 1, entries 7-10). Possible reasons for this observations are that the oxidation of simple ammonia ($E_{1/2} = +$ 0.63 V vs SCE) might be faster than the required oxidation of the substrate and the potential degradation of PC-4 in the presence of a base or nucleophiles.

Kinetic investigations. After having identified a potential photocatalyst (PC-4, TPP), kinetic investigations on this system were performed, examining the effect of (a) reaction time, (b) concentration of the catalyst, (c) temperature, (d) hydroxylamine to substrate ratio, (e) ammonium bromide concentration, and (f) wavelength (Fig. 1). Regarding the reaction time (Fig. 1a), 30% of the starting material was converted after 2 h, with the major product being the oxime intermediate **3** (16%) and only small amounts of product (**2**). After 8 h, the starting material was almost completely converted. At this time, the oxime (**3**) concentration is beginning to decrease slowly, while the yield of the desired product **2** is steadily increasing.

Varying the catalyst loading (Fig. 1b), the starting material was predominant with 2 mol%. For 5 mol% catalyst, product **2**, **3** and starting material **1** were almost at an equal level (28-38%) and the maximum product yield of 76% was obtained when the photocatalyst loading was raised to 20 mol%. The reaction temperature had a significant effect on the product yield, as shown in Fig. 1(c). At 0 °C, product **2** together with **3** were observed in 42% and 38%, respectively. The oxime to

nitrile conversion was increased by elevating the reaction temperature, revealing 40 °C as the optimum. The absence or low concentration of hydroxylamine hydrochloride led to an increased amount of by-product **4** (Fig. 1d), whereas a loading of 3 eq. gave the product in a good yield. A catalytic amount of ammonium bromide was sufficient for the complete conversion of starting material. However, to reach the maximum yield an excess of ammonium bromide is required (Fig. 1e). When investigating the effect of different wavelength, we found that 365 nm and 455 nm LEDs are efficient light sources for this transformation (Fig. 1f).

In addition to this, the role of different solvents and additives were examined (Supporting Information, Table S1 and S2). Acetonitrile was identified as the ideal solvent, together with NH₄Br as the optimal additive (Supporting Information, Table S1 and S2). Crucially, control experiments revealed the necessity of light, photocatalyst, O2 and NH2OH.HCl for the formation of the desired product (Supporting Information, Table S3, entries 2-4 and 7). However, in the absence of an additive or molecular sieves the desired product was observed as well, albeit in а low vield of 25

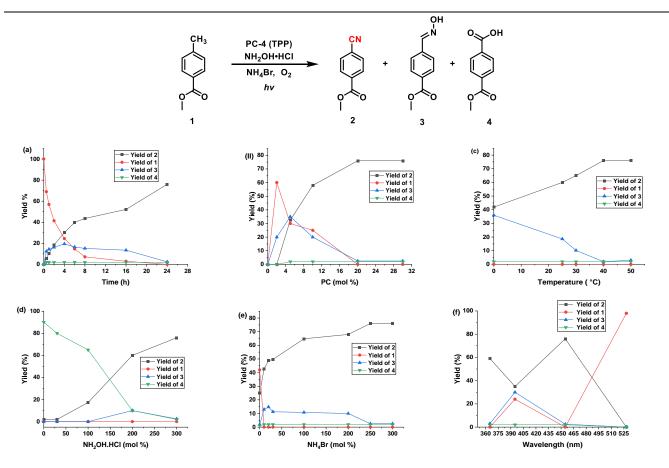


Fig. 1: Kinetic investigation on the photocatalytic ammoxidation. (a) Yield vs time, (b) yield vs concentration of TPP, (c) yield vs temperature, (d) yield vs mol% of NH₂OH•HCl, (e) yield vs mol% of NH₄Br, (f) yield vs wavelength. Reaction conditions: For Fig. 1(a): 0.1 mmol substrate, 20 mol% PC, 3 eq. NH₂OH•HCl, 2.5 eq. NH₄Br, 25 mg 4 Å MS, 1 bar O₂, 2 mL acetonitrile (0.05 M), 455 nm, 40 °C, 0-24 h. Fig. 1(b): 0.1 mmol substrate, 2-20 mol% PC, 3 eq. NH₂OH•HCl, 2.5 eq. NH₄Br, 25 mg 4 Å MS, 1 bar O₂, 2 mL acetonitrile (0.05 M), 455 nm, 40 °C, 0-24 h. Fig. 1(c): 0.1 mmol substrate, 20 mol% PC, 3 eq. NH₂OH•HCl, 2.5 eq. NH₄Br, 25 mg 4 Å MS, 1 bar O₂, 2 mL acetonitrile (0.05 M), 455 nm, 0 °C, 24 h. Fig. 1(c): 0.1 mmol substrate, 20 mol% PC, 3 eq. NH₂OH•HCl, 2.5 eq. NH₄Br, 25 mg 4 Å MS, 1 bar O₂, 2 mL acetonitrile (0.05 M), 455 nm, 0-40 °C, 24 h. Fig. 1(d): 0.1 mmol substrate, 20 mol% PC, 0.3-3 eq. NH₂OH•HCl, 2.5 eq. NH₄Br, 25 mg 4 Å MS, 1 bar O₂, 2 mL acetonitrile (0.05 M), 455 nm, 40 °C, 24 h. Fig. 1(d): 0.1 mmol substrate, 20 mol% PC, 3 eq. NH₂OH•HCl, 0.1-2.5 eq. NH₄Br, 25 mg 4 Å MS, 1 bar O₂, 2 mL acetonitrile (0.05 M), 455 nm, 40 °C, 24 h. Fig. 1(d): 0.1 mmol substrate, 20 mol% PC, 3 eq. NH₂OH•HCl, 0.1-2.5 eq. NH₄Br, 25 mg 4 Å MS, 1 bar O₂, 2 mL acetonitrile (0.05 M), 455 nm, 40 °C, 24 h. Fig. 1(e): 0.1 mmol substrate, 20 mol% PC, 3 eq. NH₂OH•HCl, 0.1-2.5 eq. NH₄Br, 25 mg 4 Å MS, 1 bar O₂, 2 mL acetonitrile (0.05 M), 455 nm, 40 °C, 24 h. Fig. 1(e): 0.1 mmol substrate, 20 mol% PC, 3 eq. NH₂OH•HCl, 0.1-2.5 eq. NH₄Br, 25 mg 4 Å MS, 1 bar O₂, 2 mL acetonitrile (0.05 M), 455 nm, 40 °C, 24 h. Fig. 1(e): 0.1 mmol substrate, 20 mol% PC, 3 eq. NH₂OH•HCl, 2.5 eq. NH₄Br, 25 mg 4 Å MS, 1 bar O₂, 2 mL acetonitrile (0.05 M), 365-528 nm, 40 °C, 24 h. Yields were determined by GC using n-decane as standard.

and 51%, respectively (Supporting Information, Table S3, entries 5-6).

Mechanistic hypothesis. The initial kinetic studies suggest that the oxime is one of the key intermediates, as its depletion is accompanied by the formation of the product. Further, the control experiment yielding no product in the absence of O_2 indicates that the oxime might be formed from the corresponding aldehyde. Based on this we envisioned following catalytic

cycle (Cycle A): the substrate $(E_{1/2} (1/1^{++}) = + 2.45 \text{ V vs SCE}$, supporting information) is oxidised by the excited photocatalyst $(E_{1/2} (PC-4^{+*/} PC-4^{+}) = + 2.3 \text{ V Vs SCE})$ via electron transfer, generating radical cation **II** and the reduced photocatalyst species (PC-4^{+}). The photocatalytic cycle is closed by the oxidation of the reduced photocatalyst with O₂, giving a superoxide anion (O₂ \cdot). Intermediate **II** loses a proton to form the more stable benzylic radical (**III**), which combines with

the superoxide anion to peroxide derivative V after protonation. Elimination of water from V leads to the corresponding aldehyde intermediate (VI). Intermediate VI can yield oxime VII upon condensation with NH₂OH•HCl, which can be converted into the desired nitrile in a second cycle (Fig. 2, Cycle A'). In this cycle, the oxime (VII) is oxidised to VIII via reductive quenching of PC-4* followed by elimination of water to give intermediate IX. The reduced PC undergoes SET with IX to render the desired product X, closing the photocatalytic cycle and completing a redox-neutral process.

The presence of NH_4Br is crucial to obtain a satisfactory yield. Thus, another catalytic cycle may be operative, yielding the same reaction product (Fig. 2, Cycle B). Bromide anions are known hydrogen atom transfer (HAT) catalysts.²³ In presence of visible-light, the excited PC can oxidise Br⁻ to generate the Br⁻ radical, which is capable to abstract the H-atom from the methylarene to yield a benzyl radical (III) and HBr. The combination of superoxide anion and III yields intermediate IV, which deprotonates HBr to regenerate the Br⁻ while generating intermediate V. After its formation, V follows the same mechanistic pathway as described above (Cycle A and A').

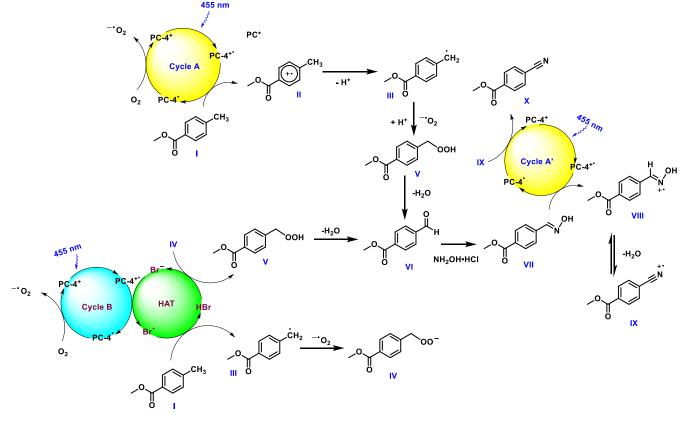


Fig. 2: Proposed catalytic cycles for the ammoxidation of methylarenes.

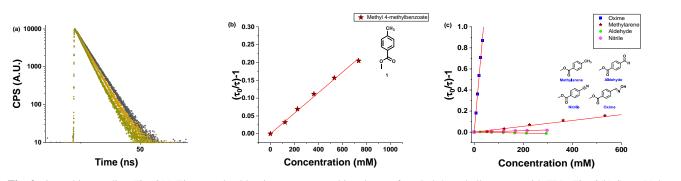
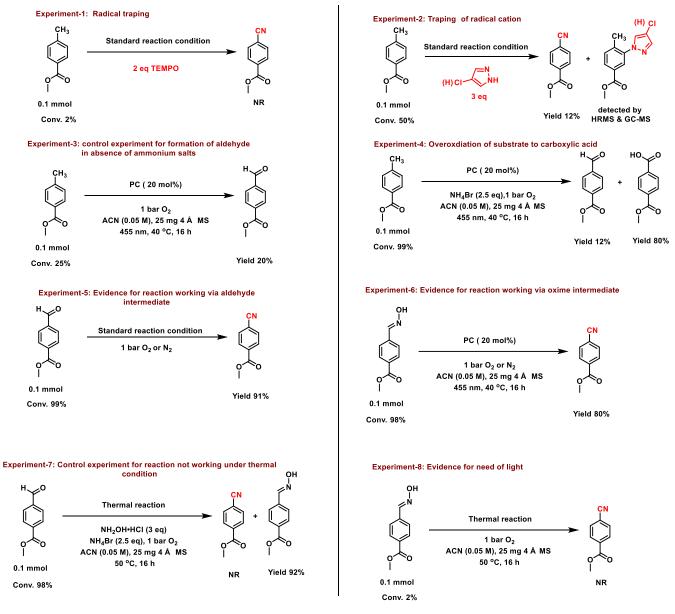


Fig. 3: Quenching studies. Fig. 3(a) Time-resolved luminescence quenching decay of methyl 4-methylbenzoate with TPP; Fig. 3(b) Stern-Volmer plot for the TPP with methyl 4-methylbenzoate; Fig. 3(c) Stern-Volmer plot for the comparison of different substances with TPP.

To support this proposed hypothesis, we performed several mechanistic experiments, starting with spectroscopic investigations. Time-resolved luminescence quenching experiments indicated that methyl 4-methylbenzoate (1) can be oxidized by

the excited TPP (PC-4) (Fig. 3), as a slight decrease of the luminescence lifetime with increasing concentration of **1** was observed (Fig. 3a). The corresponding Stern-Volmer plot

Scheme 2. Mechanistic experiments to support the proposed catalytic hypothesis



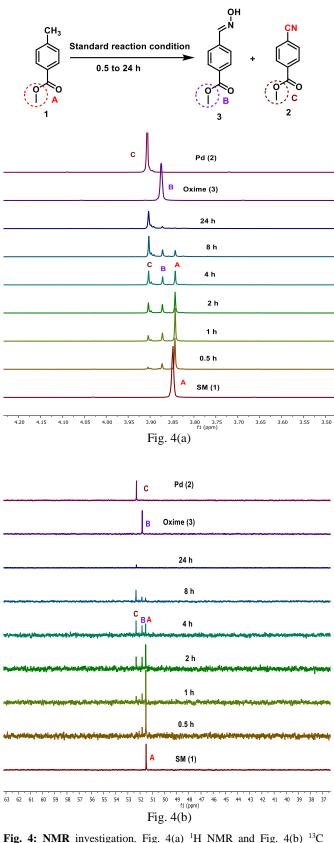
Yields were determined by GC using n-decane as standard

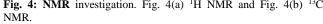
Further, the quenching of the starting material (1) was compared to the product (2) and the two crucial proposed interme-4-formylbenzoate diates methyl and methyl 4-((hydroxyimino)methyl)benzoate (3) under the same conditions. The starting material (1) was identified as a more efficient quencher than the corresponding aldehyde intermediate and nitrile product. On the other hand, the oxime intermediate (3) showed a superior quenching to methyl 4-methylbenzoate (1) (Fig. 3c), illustrating the possible interaction of the excited photocatalyst with the oxime intermediate under the applied reaction condition (Cycle A'). The quenching efficiencies of NH₂OH•HCl and NH₄Br were investigated as well, with the former exhibiting a poorer quenching than the later one (Supporting Information, Fig. S3), which reveals that Cycle B can be active as well. Comparing the luminescence lifetime quenching of NH₄Br and the methyl 4-methylbenzoate starting material (1), NH₄Br seems to decrease the lifetime more effectively (Supporting Information, Fig. S4). However, an electron-rich (4-methyl anisole) instead of an electron-deficient (1) substrate proved to be a more potent quencher as well (Supporting Information, Fig. S6), showing similar quenching ability as NH₄Br. Thus, based on the time-resolved luminescence quenching experiments, both proposed catalytic cycles A and B can be operative, with their respective importance likely being dependent on the electronic nature of the starting material.

Next, several mechanistic control experiments were performed to directly or indirectly detect reaction intermediates vital for the mechanistic process. Further, selected stable intermediates were used as starting materials for the formation of the final product under specific reaction conditions to establish the mechanistic pathways more clearly (Scheme 2). The model reaction was performed under standard reaction condition in presence of TEMPO, yielding no product, which indicates a radical pathways (Scheme 2, Experiment-1). Further, the radical cation intermediate (Fig. 2, II) could be trapped in the presence of pyrazole or 4-cholorpyrazole as nucleophiles under standard reaction condition, in a similar reaction as reported by Nicewicz,²² supporting its formation (Scheme 2, Experiment-2). Looking at the more stable intermediates, the methylarene is proposed to be oxidized to the corresponding aldehyde first (Fig. 2, VI). This was confirmed by performing the model reaction in absence of both ammonium salts (to avoid the formation of the oxime), providing the corresponding aldehyde as product in 20% GC yield (Scheme 2, experiment-3), with an incomplete starting material conversion of 25%. In contrast, a full conversion was observed when NH₄Br was added (Experiment 4, Scheme 2), showcasing its crucial role, likely due to the thus opened alternative Br-HAT-pathway^{6h, 23} (Cycle B, Fig. 2). The main product in this case was the corresponding carboxylic acid (80%) resulting from an over-oxidation of the aldehyde (12%). With no observable nitrile formation, the experiment further suggests, that NH₂OH•HCl is the main nitrogen source. It is proposed to rapidly form the oxime from the aldehyde, avoiding its oxidation to the carboxylate.

The generation of an aldehyde and oxime intermediate was further supported by using those as starting materials for the synthesis of the final nitrile product (Scheme 2, Experiment 5 and 6). In both cases, the product was formed with a yield of 91% and 80%, respectively. Additionally, the conversion of the aldehyde or oxime to the nitrile did not proceed under solely thermal condition in the absence of light (Scheme 2, Experiment 7 and 8), supporting the proposed catalytic cycle A^{*}. To further indicate the necessity of more than one photon for the formation of one product molecule, the product yield dependent on the irradiation intensity was observed as well (Supporting Information, Fig. S12).²⁴

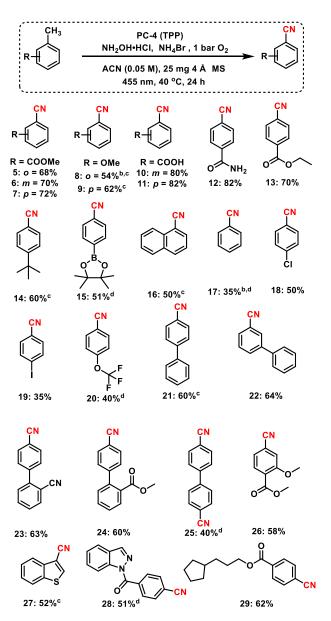
In order to follow the mechanistic pathway in more detail, NMR studies were performed analysing the reaction mixture after defined time intervals (Fig. 4). The signal corresponding to the methyl ester (-COOCH₃) of the starting material (1) shows a singlet resonance at δ 3.84 ppm in the ¹H-NMR spectrum, while the oxime (3) gives a singlet at δ 3.87, and nitrile (2) a singlet at δ 3.91, allowing for a facile distinction between the species of interest. At the beginning, the depletion of the starting material is accompanied by the increase of the oxime intermediate as well as nitrile product, with all three compounds being clearly observable at the same time after 4 h of irradiation. Further increasing the reaction time leads to the





(almost) full conversion of the starting material and the oxime with the dominant signal being the product peak.

Scheme 3. Substrate scope for synthesis of functionalized cycanobenzenes

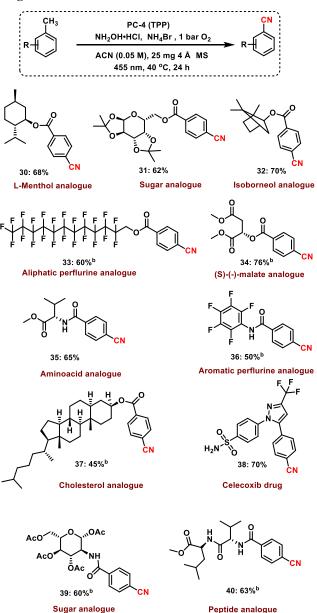


Reaction conditions: ^a0.1 mmol substrate, 20 mol% PC, 3 eq. NH₂OH•HCl, 2.5 eq. NH₄Br, 25 mg 4 Å MS, 1 bar O₂, 2 mL acetonitrile (0.05 M), 455 nm, 40 °C, 24 h, isolated yields. ^bGC yields using n-decane as standard. ^csame as 'a' using 1 eq. of PTSCl. ^dsame as 'a' 48 h instead of 24 h.

A similar tendency was observed in the ¹³C-NMR for the corresponding methyl ester carbon (Fig. 4b), all together further supporting the formation of the oxime as crucial intermediate to render the nitrile.

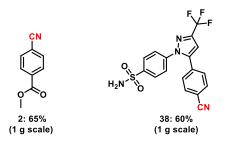
Collectively, the spectroscopy investigation, control experiments and kinetic investigation all support the proposed photocatalytic cycles. In addition, the catalyst deactivation pathway also studied under the standard reaction conditions, suggesting 2,4,6-triphenylpyridine as the degradation product, which could further be observed in the scale-up batch as minor by-product supported by HRMS (Supporting Information, Scheme-S1).

Scheme 4. Late-stage functionalization of challenging and drug-like molecules



Reaction conditions: ^a0.1 mmol substrate, 20 mol% PC, 3 eq. NH₂OH•HCl, 2.5 eq. NH₄Br, 25 mg 4 Å MS, 1 bar O₂, 2 mL acetoni-trile (0.05 M), 455 nm, 40 °C, 24 h, isolated yields. ^bsame as 'a ' 48 h instead of 24 h.

With the successful conditions in hand, we explored the $C(sp^3)$ -H functionalization of different methylarenes (Scheme 3 and 4). Substrates bearing electron-donating and -withdrawing groups, as well as heterocycles, gave the respective products in good to excellent yields (Scheme 3, 5-29). Functional groups such as carboxylic acid, ester, amide, halogens, cyanide and boronic ester were untouched during the reaction (Scheme 3, 5-13, 15, 18-20 and 23-24). Applying this methodology, a dinitrile could also be synthesized in a one-pot procedure with a moderate yield (Scheme 3, 25).

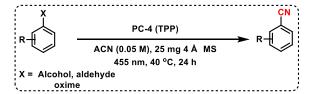


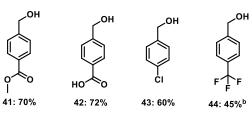
Reaction conditions: ^a1 g substrate, 20 mol% PC, 3 eq. NH₂OH•HCl, 2.5 eq. NH₄Br, 25 mg 4 Å MS for 0.1 mmol, 1 bar O_2 , acetonitrile (0.05 M), 455 nm, 40 °C, 36 h, isolated yields.

Structurally complex, bioactive- and drug- molecules could be employed for a selective cyanation as well, rendering the desired product in good to excellent yields up to 76% (Scheme 4). Delightfully, substrates bearing multiple oxygen atoms, which are usually not stable under photochemical conditions in the presence of oxygen, are viable, too.²⁵ Using the developed protocol, sugar derivatives **31** and **39** were obtained in 62% and 60% yield, respectively. Further, more challenging substrates such as cholesterol, isoborneol, amino acid and peptide derivatives gave the corresponding products in 45-70% yield. Interestingly, anti-inflammatory drug (Celecoxib) gave the desired product in 70% yield. For the example of a bulk-scale preparation, compound **2** and Celecoxib were cyanated in a 1 g scale to give the desired product in 65 and 60% yield respectively (Scheme 5).

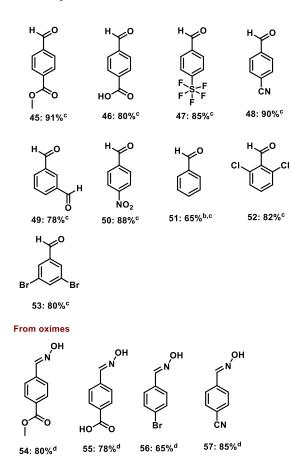
After the screening of methylarenes, we were interested to apply this methodology using alcohols, aldehydes and oximes as starting materials for the synthesis of aromatic nitriles as well (Scheme 6). Similar to methylarenes, alcohols, aldehydes and oximes gave good to excellent yields up to 91%. Notably, isophthalaldehyde (49) was cyanided twice to isophthalonitrile in 78% yield. Sterically crowded 2,6-dichlorobenzaldehyde (52) was а viable substrate, too, giving 2,6dichlorobenzonitrile (DCBN) in 82% yield, which is used as herbicide and regarded as a potential intermediate for pesticides and agrochemicals.

Scheme 6. Synthesis of nitriles from alcohols, aldehydes, and oximes





From aldehydes



Reaction conditions: ^a0.1 mmol substrate, 20 mol% PC, 3 eq. NH₂OH•HCl, 2.5 eq. NH₄Br, 25 mg 4 Å MS, 1 bar O₂, 2 mL acetonitrile (0.05 M), 455 nm, 40 °C, 24 h, isolated yields of nitriles. ^bGC yields of nitriles using n-decane as standard. ^c0.1 mmol substrate, 20 mol% PC, 3 eq. NH₂OH•HCl, 25 mg 4 Å MS, N₂, 2 mL acetonitrile (0.05 M), 455 nm, 40 °C, 24 h, isolated yields of nitriles. ^d0.1 mmol substrate, 20 mol% PC, 25 mg 4 Å MS, N₂, 2 mL acetonitrile (0.05 M), 455 nm, 40 °C, 24 h, isolated yields of nitriles. ^d0.1 mmol substrate, 20 mol% PC, 24 h, isolated yields of nitriles.

CONCLUSION

In conclusion, we present the first metal- and cyanide-free visible-light-induced photocatalytic ammoxidation of $C(sp^3)$ -H bonds using an abundant ammonia source and molecular oxygen. A detailed mechanistic investigation was carried out to support the proposed mechanistic hypothesis including various spectroscopy experiments. Applying this methodology, more than 50 aromatic and heteroaromatic substrates, as well as steroids and existing drug molecules containing methyl groups

could be converted to the nitrile in good to excellent yields. In addition to this, the method could be executed on gram scale and alcohols, aldehydes and oximes could be used as starting materials for their conversion to nitriles in up to 91% yield in the same manner.

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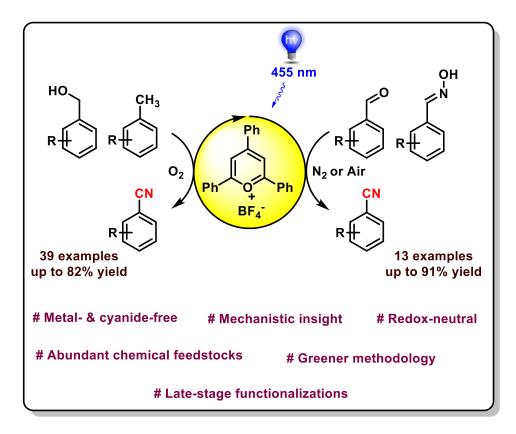
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Visible-Light-Promoted Metal-Free Ammoxidation of C(*sp*³)-H bonds

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Visible-Light-Promoted Metal-Free Ammoxidation of C(sp³)-H bonds

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Table of contents

- 1. General information
- 2. Synthetic procedure
 - 2.1 General procedure for synthesis of starting materials
 - 2.2 General procedure for synthesis of oximes
 - 2.3 General procedure for synthesis of nitriles from methylarenes or alcohols
 - 2.4 General procedure for synthesis of nitriles from aldehydes
 - 2.5 General procedure for synthesis of nitriles from oximes
 - 2.6 General procedure for gram scale reactions
- 3. Reaction optimization
- 4. Mechanistic Investigation
 - 4.1 Time-resolved Luminescence Quenching studies
 - 4.2 Trapping experiments
 - 4.3 The product yield on irradiation density
 - 4.4 NMR studies
 - 4.5 Catalyst deactivation

4.6 Cyclic Voltammetry measurement

- 5. Characterization of prepared compounds
- 6. NMR spectra
- 7. References

1. General information

All required fine chemicals were purchased from commercial suppliers (abcr, Acros, Alfa Aesar, Fluka, Fluorochem, Merck, Sigma Aldrich, TCI) and were used directly without purification unless stated otherwise. All air and moisture sensitive reactions were carried out under nitrogen atmosphere using standard Schlenk manifold technique. Extra dry anhydrous acetonitrile was purchased from Acros organics.

All NMR spectra were measured at room temperature using a Bruker Avance 300 (300 MHz for ¹H, 75 MHz for ¹³C, 282 MHz for ¹⁹F) or a Bruker Avance 400 (400 MHz for ¹H, 101 MHz for ¹³C, 376 MHz for ¹⁹F)¹ NMR spectrometer. All chemical shifts are reported in δ -scale as parts per million [ppm] (multiplicity, coupling constant *J*, number of protons) relative to the solvent residual peaks as the internal standard.² Coupling constants *J* are given in Hertz [Hz]. Abbreviations used for signal multiplicity: ¹H-NMR: b = broad, s = singlet, d = doublet, t = triplet, q = quartet, p = quintet, and m = multiplet.

GC measurements were performed on a GC 7890 from Agilent Technologies. Data acquisition and evaluation was done with Agilent ChemStation Rev.C.01.04. GC/MS measurements were performed on a 7890A GC system from Agilent Technologies with an Agilent 5975 MSD Detector. Data acquisition and evaluation was done with MSD ChemStation E.02.02.1431. A capillary column HP-5MS/30 m x 0.25 mm/0.25 µM film and helium as carrier gas (flow rate of 1 mL/min) were used. The injector temperature (split injection: 40:1 split) was 280 °C, detection temperature 300 °C (FID). GC measurements were made and investigated *via* integration of the signal obtained. The GC oven temperature program was adjusted as follows: initial temperature 40 °C was kept for 3 minutes, the temperature was increased at a rate of 15 °C/min over a period of 16 minutes until 280 °C was reached and kept for 5 minutes, the temperature was again increased at a rate of 25 °C/min over a period of 48 seconds until the final temperature (300 °C) was reached and kept for 5 minutes. *n*-Decane was used as an internal standard.

Analytical TLC was performed on silica gel coated aluminium sheets (Merck, TLC Silica gel 60 F_{254}). Compounds were visualized by exposure to UV-light (254 or 366 nm) or by dipping the plates in staining solutions (permanganate stain, bromocresol green stain, ceric ammonium molybdate stain) followed by heating. Purification by column chromatography was performed with silica gel 60 M (40-63 µm, 230-440 mesh, Merck) or with a pre-packed Biotage® Snap Ultra HP-SphereTM 25 *u*m column on a Biotage[®] IsoleraTM Spektra One device. All mixed solvent eluents are reported as v/v solutions.

High resolution mass spectrometry (HRMS) were performed at the Central Analytical Laboratory of the University of Regensburg. Mass spectra were recorded on a Finnigan MAT 95, ThermoQuest Finnigan TSQ 7000, Finnigan MAT SSQ 710 A or Agilent Q-TOF 6540 UHD instrument and a Waters Acquity UPLC system equipped with Waters PDA, sample manager, sample organiser, column oven and Waters Xevo QTOF mass spectrometer. Photoreactions in regular scale were irradiated with blue LEDs (OSRAM Oslon SSL 80 royal-blue, $\lambda = 455$ nm (± 15), average radiant flux 232 ± 23 mW, 2.9 V, 350 mA) or green LEDs (λ = 535 nm, average radiant flux, 29 ± 5 mW) and were exposed to light from the flat bottom side of the vial. The temperature of the reaction mixtures was controlled by a water-cooling circuit consisting of an aluminium cooling block connected to a thermostat (Fig. S1). An exemplary reaction in larger scale was carried out in a custom-built glass reactor which upon vigorous stirring generates a thin film of the reaction mixture between the reaction vessel and an attached cold finger. The reaction vessel was surrounded by blue LED arrays (OSRAM Oslon SSL 80 LT-2010, $\lambda = 451$ nm, 700 mA) generating a total radiant flux of 12 W (Fig. S2). Luminescence measurements were performed on a Horiba® Scientific FluoroMax-4 instrument using the above-mentioned quartz cells. Luminescence lifetime measurements were performed on a Horiba® Scientific DeltraProTM fluorescence lifetime system using a 452 nm laser diode from Horiba® Scientific DeltaDiode™ as excitation source and above-mentioned quartz cells. The instrument response function (IRF) was determined prior to measurements by using colloidal silica (LUDOX®) in water.

CV measurements were performed with the three-electrode potentiostat galvanostat PGSTAT302N from Metrohm Autolab using a glassy carbon working electrode, a platinum wire

counter electrode, a silver wire as a reference electrode and TBATFB 0.1 M as supporting electrolyte. The control of the measurement instrument, the acquisition and processing of the cyclic voltammetric data were performed with the software Metrohm Autolab NOVA 1.10.4. The measurements were carried out as follows: a 0.1 M solution of TBATFB in acetonitrile was added to the measuring cell and the solution was degassed by argon purge for 5 min. After recording the baseline the electroactive compound was added (0.01 M) and the solution was again degassed a stream of argon for 5 min. The cyclic voltammogram was recorded with one to three scans. Afterwards ferrocene (2.20 mg, 12.0 μ mol) was added to the solution which was again degassed by argon purge for 5 min and the final measurement was performed with three scans.

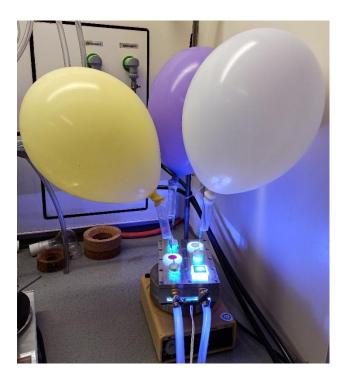


Fig. S1: Typical set-up for small scale synthesis



Fig. S2: Custom-built glass reactor for upscaling of the photocatalytic ammoxidation reaction.

2. Synthetic procedure

2.1 General procedure for the synthesis of methylarenes

Methylarenes were synthesized according to a literature procedure.³

General conditions: To a stirred solution of 4-methyl benzoyl chloride (7.5 mmol, 1.5 eq) in DCM (10 mL) and added an alcohol or amine nucleophile (5 mmol 1.0 eq), DMAP (0.5 mmol, 0.1 eq) and Et_3N (10 mmol, 2.0 eq) in DCM (10 mL). The reaction was allowed to stir overnight at room temperature. Then, the mixture was quenched upon addition of NH₄Cl (aq. 10%), and extracted with DCM (3x). The organic phase was washed with brine, concentrated and purified by silica gel flash chromatography to give the corresponding substituted methylarenes. If the HCl salt of the amine nucleophile was employed, 4 eq of Et_3N were used.

2.2 General procedure for the synthesis of oximes

Oximes were synthesized according to a literature procedure.⁴

A 100 mL round bottom flask equipped with a magnetic stirring bar was loaded with NH₂OH•HCl (1.2 eq for 1.0 g of substrate) in water (25 mL), followed by the addition of NaHCO₃ (1.75 eq for 1.0 g of substrate) at 0° C. A solution of the corresponding aldehyde (1.0 g, 1 eq) in MeOH was added to the above mixture and stirred at RT for 6h. The reaction progress was monitored by TLC (PE:EA). After completion of the reaction, MeOH was removed under reduced pressure. The precipitated solid was filtered off, washed with water and dried under high vacuum. The thus obtained product was used for following steps without further purification.

2.3 General procedure for the synthesis of nitriles from methylarenes or alcohols

A 5 mL crimp cap vial equipped with a magnetic stirring bar was loaded with TPP (20 μ mol, 8.0 mg, 20 mol%), NH₂OH•HCl (0.3 mmol, 20.7 mg, 3 eq), NH₄Br (0.25 mmol, 25.0 mg, 2.5 eq), 4 Å molecular sieves (25 mg), the corresponding methylarene or alcohol (0.1 mmol, 1 eq.) and dry ACN (2 mL, 0.05M). In doing so, all solid compounds were added before capping the vial, whereas all liquid compounds were added *via* syringe after setting the capped vial under an O₂ atmosphere (highly viscous liquids were added before capping the vial as well). The reaction mixture was stirred under 1 bar of O₂ using an O₂-filled balloon and under light irradiation using a 455 nm (± 15 nm) LED for 24 h at 40 °C using a cryostat. Four reaction batches were combined, filtered and concentrated under reduced pressure. The crude product was

purified by automated flash column chromatography using a petrolether/ethyl acetate mixture. Note: in order to get better yields, dry acetonitrile and freshly activated 4 Å molecular sieves are mandatory.

2.4 General procedure for the synthesis of nitriles from aldehydes

A 5 mL crimp cap vial equipped with a magnetic stirring bar was loaded with TPP (20 μ mol, 8.0 mg, 20 mol%), NH₂OH•HCl (0.3 mmol, 20.7 mg, 3 eq), NH₄Br (0.25 mmol, 25.0 mg, 2.5 eq), 4 Å molecular sieves (25 mg), the corresponding aldehyde (0.1 mmol, 1 eq.) and dry ACN (2 mL, 0.05M). In doing so, all solid compounds were added before capping the vial, whereas all liquid compounds were added *via* syringe after setting the capped vial under N₂ atmosphere (highly viscous liquids were added before capping the vial as well). The reaction mixture was stirred under light irradiation using a 455 nm (± 15 nm) LED for 24 h at 40 °C using a cryostat. Four reaction batches were combined, filtered and concentrated under reduced pressure. The crude product was purified by automated flash column chromatography using a petrolether/ethyl acetate mixture.

2.5 General procedure for the synthesis of nitriles from oximes

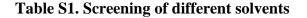
A 5 mL crimp cap vial equipped with a magnetic stirring bar was loaded with TPP (20 μ mol, 8.0 mg, 20 mol%), 4 Å molecular sieves (25 mg), the corresponding oxime (0.1 mmol, 1 eq.) and dry ACN (2 mL, 0.05M). In doing so, all solid compounds were added before capping the vial, whereas all liquid compounds were added *via* syringe after setting the capped vial under a N₂ atmosphere (highly viscous liquids were added before capping the vial as well). The reaction mixture was stirred under light irradiation using a 455 nm (± 15 nm) LED for 24 h at 40 °C using a cryostat. Four reaction batches were combined, filtered and concentrated under reduced pressure. The crude product was purified by automated flash column chromatography using a petrolether/ethyl acetate mixture.

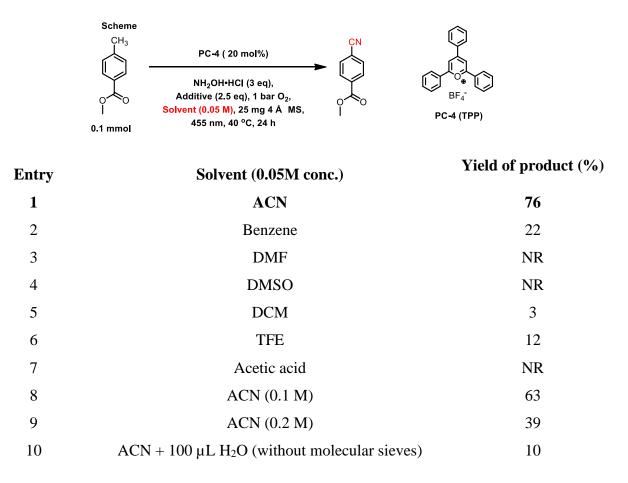
2.6 General procedure for gram scale reactions

A 200 mL glass reactor equipped with a magnetic stirring bar was loaded with TPP (20 mol% for 1 g of substrate), NH_2OH •HCl (3 eq for 1 g of substrate), NH_4Br (2.5 eq for 1 g of substrate), 4 Å molecular sieves (25 mg for 0.1 mmol of substrate), the corresponding methylarene (1 g, 1 eq.) and dry ACN (0.05 M). In doing so, all solid compounds were added first to the glass reactor, whereas all liquid compounds were added at the end under O₂ or air atmosphere. The reaction setup was connected to two balloons holding

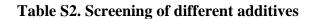
each 1 bar of oxygen (please see Fig. S2) and the mixture was stirred under light irradiation using a 455 nm $(\pm 15 \text{ nm})$ LED for 36 h at 40 °C using a cryostat. The reaction progress was monitored by GC-FID. After the completion of the reaction, the reaction mixture was filtered over a celite bed and concentrated under reduced pressure. The crude product was purified by automated flash column chromatography using a petrolether/ethyl acetate mixture.

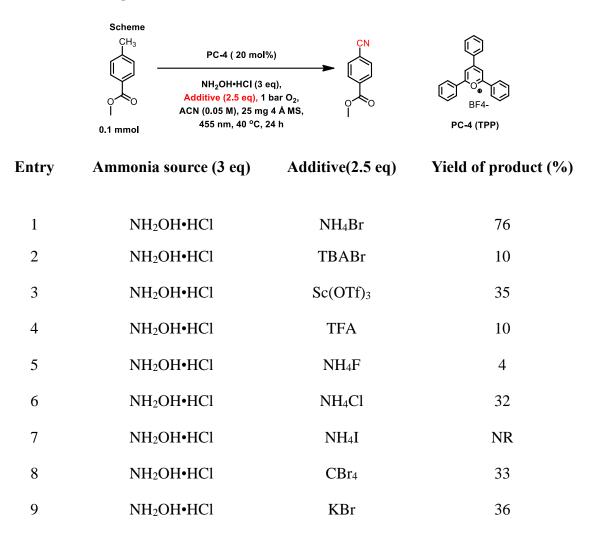
3. Reaction optimization





Reaction conditions: a0.1 mmol substrate, 20 mol% PC, 3 eq. NH₂OH•HCl, 2.5 eq. NH₄Br, 25 mg 4 Å MS, 1 bar O₂, 2 mL solvent (0.05 M), 455 nm, 40 °C, 24 h, GC yields using n-decane as standard.





Reaction conditions: ^a0.1 mmol substrate, 20 mol% PC, 3 eq. NH₂OH•HCl, 2.5 eq. additive, 25 mg 4 Å MS, 1 bar O_2 , 2 mL acetonitrile (0.05 M), 455 nm, 40 °C, 24 h, GC yields using n-decane as standard.

Table S3. Control experiments

	Scheme CH ₃ O O O O O O O O	PC 	-4 (20 mol%) OH•HCI (3 eq), (2.5 eq), 1 bar O ₂ , 5 M), 25 mg 4 Å MS, ım, 40 °C, 24 h			орания 1954- (ТРР)	
Entry	PC-4 (20 mol%)	Light (455 nm)	NH2OH•HCl (3 eq)	NH4Br (2.5 eq)	4 Å MS (25 mg)	O2 (1 bar)	Yield of product (%)
1	YES	YES	YES	YES	YES	YES	76
2	NO	YES	YES	YES	YES	YES	1
3	YES	NO	YES	YES	YES	YES	NR
4	YES	YES	NO	YES	YES	YES	2
5	YES	YES	YES	NO	YES	YES	25
6	YES	YES	YES	YES	NO	YES	51
7	YES	YES	YES	YES	YES	NO	NR

Reaction conditions: ^a0.1 mmol substrate, 20 mol% PC, 3 eq. NH₂OH•HCl, 2.5 eq. NH₄Br, 25 mg 4 Å MS, 1 bar O₂, 2 mL acetonitrile (0.05 M), 455 nm, 40 °C, 24 h, GC yields using n-decane as standard.

4. Mechanistic Investigation

4.1 Time-resolved Luminescence Quenching studies

A linear correlation between concentration of quencher [Q] and $\tau_0 \times \tau^{-1}$ indicates a dynamic luminescence quenching. The luminescence lifetime was recorded in dry ACN using a quartz cuvette (1×1 cm) with septum screw cap. The cuvette was degassed *in vacuo* and backfilled with N₂ (5×) before the stock solution of quencher and the catalyst solution were added *via* syringe. For excitation of the sample, a 452 nm laser diode was used and an optical longpass filter (cut-on wavelength 500 nm) was installed before the detection unit. The time range for the measurement was set to 100 ns. The experimental data were fitted with a monoexponential function. For liquid sample added as such for solid sample concertation solution in ACN was used. In case of NH₂OH.HCl and NH₄Br 1 mmol was taken in 1 mL ACN and stirred at 40° C for 15 minutes then passed through the filter pad and clear solution was used for quenching studies.

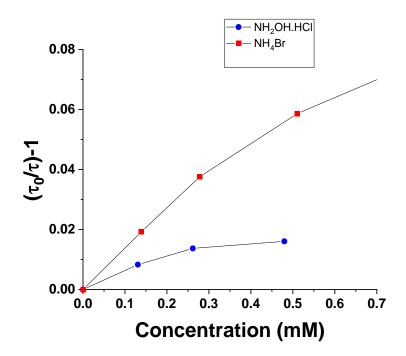


Fig. S3: Stern-Volmer plot developed with data obtained from time-resolved quenching experiments of TPP (PC-4) with NH₂OH•HCl and NH₄Br.

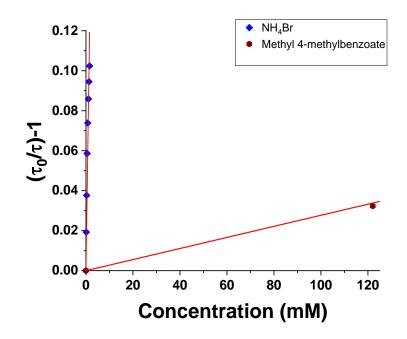


Fig. S4: Stern-Volmer plot developed with data obtained from time-resolved quenching experiments of TPP (PC-4) with methyl 4-methylbenzoate and NH₄Br.

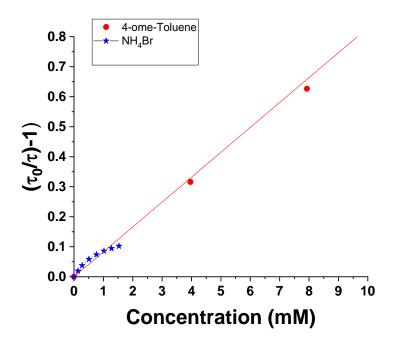


Fig. S5: Stern-Volmer plot developed with data obtained from time-resolved quenching experiments of TPP (PC-4) with 4-methoxy toluene and NH₄Br.

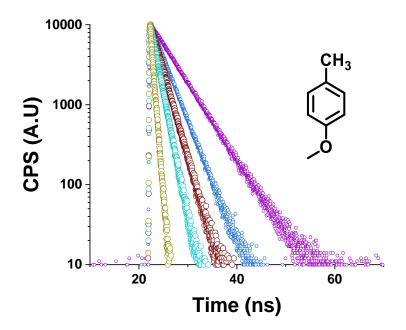


Fig. S6: Luminescence decay of 4-methoxy toluene with TPP (PC-4). The lifetime was determined by employing an exponential fit function.

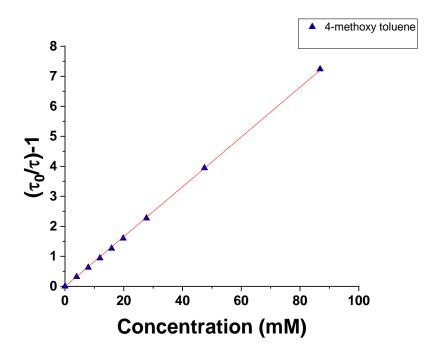
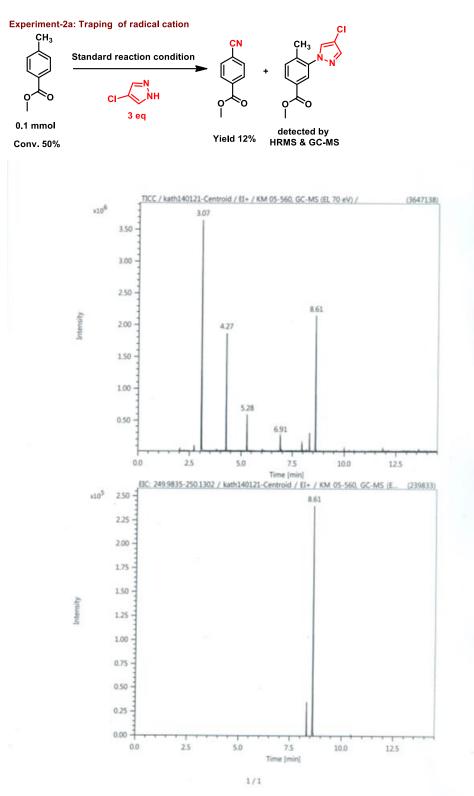
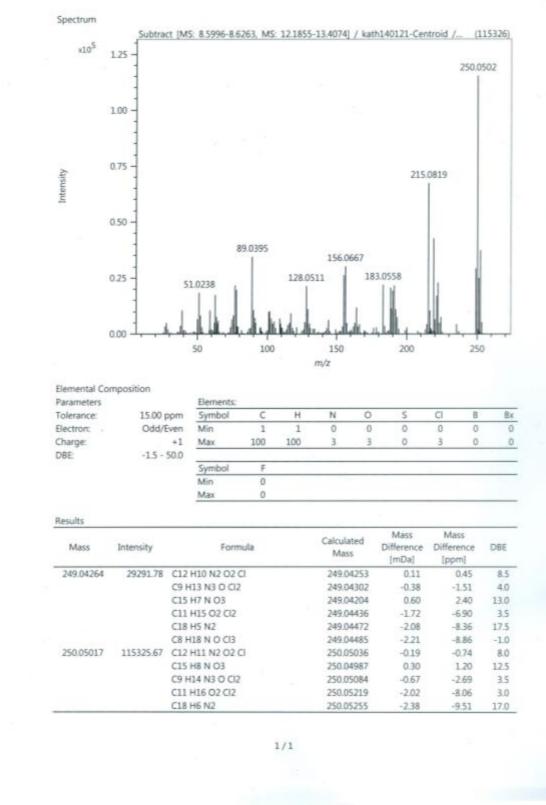
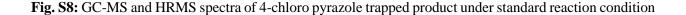


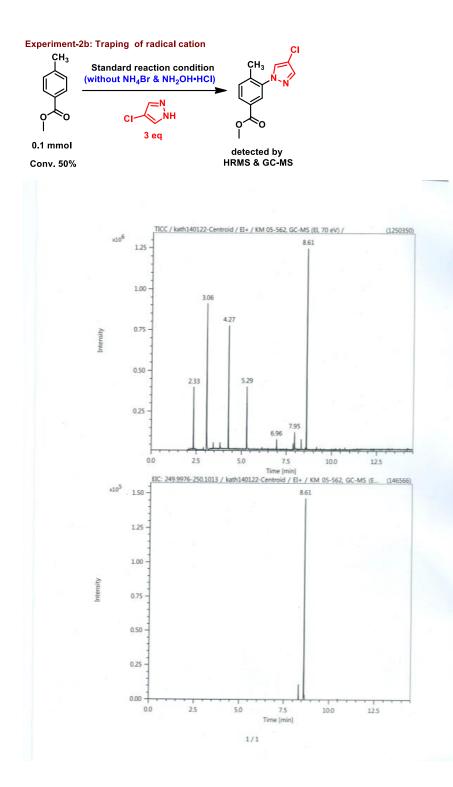
Fig. S7: Stern-Volmer plot developed with data obtained from time-resolved quenching experiments of TPP (PC-4) with 4-methoxy toluene.

4.2 Trapping experiments⁵









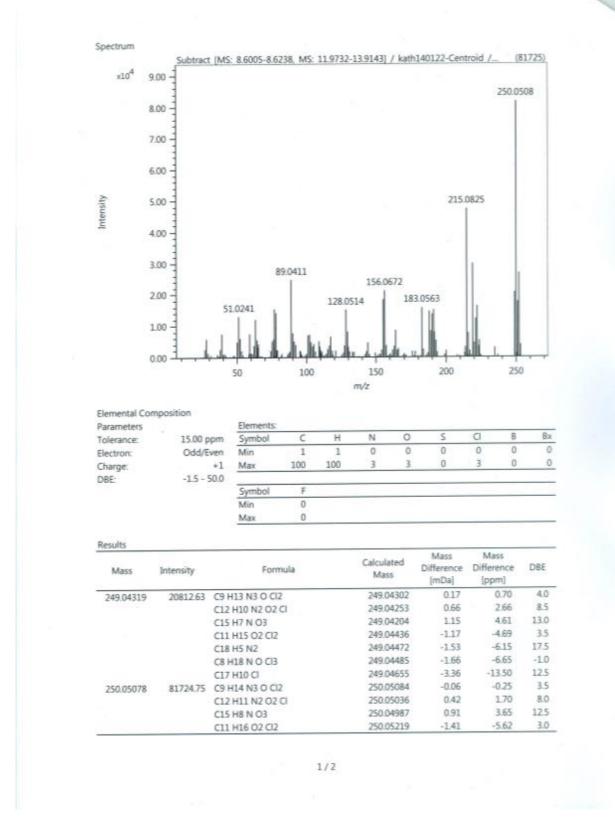
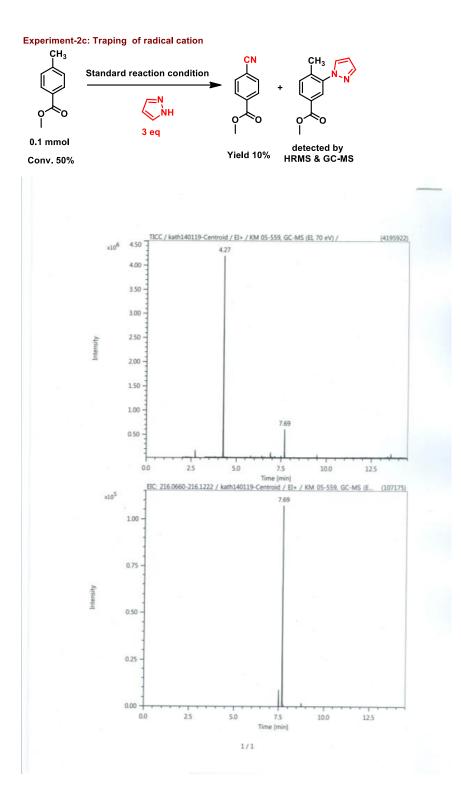
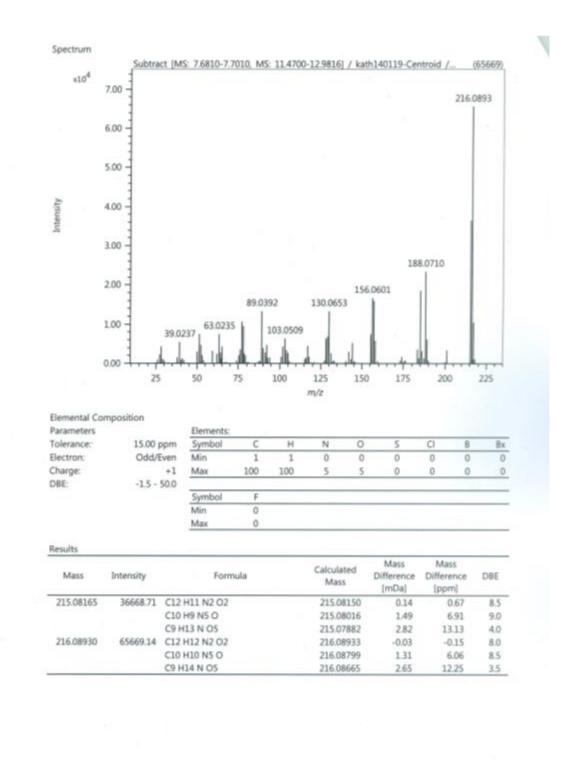


Fig. S9: GC-MS and HRMS spectra of 4-chloro pyrazole trapped product without NH₄Br and NH₂OH•HCl

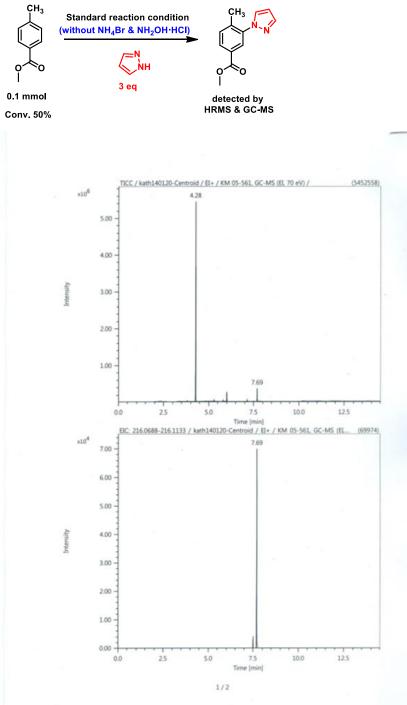


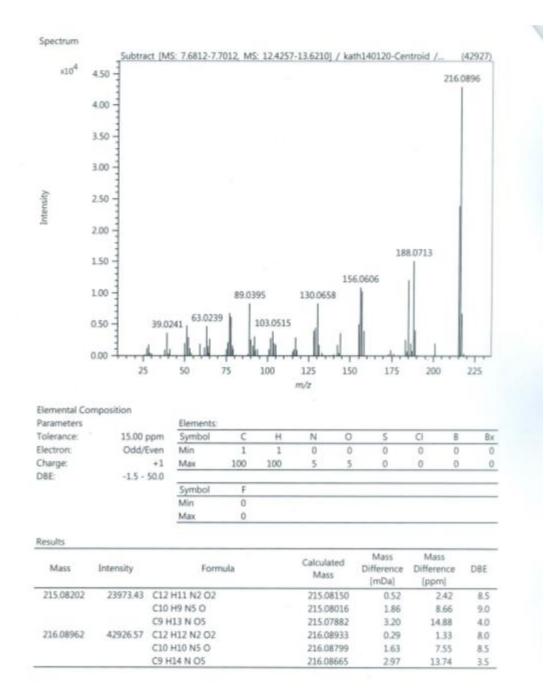


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Fig. S10: GC-MS and HRMS spectra of pyrazole trapped product under standard reaction condition

Experiment-2d: Traping of radical cation





1/1

Fig. S11: GC-MS and HRMS spectra of 4-chloro pyrazole trapped product without NH_4Br and NH_2OH •HCl

4.3 The product yield on irradiation density

The irradiation power of blue LED was adjusted in Photoreactor TAK 120 by turning the voltage or current knob. PowerMax USB-PS19Q Power Sensor was also used to verify the emitted light power.

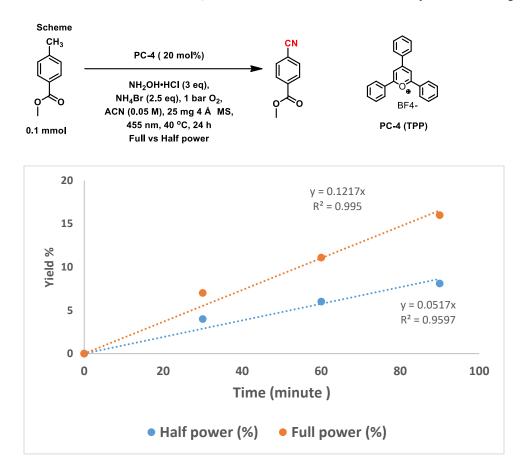


Fig. S12: Two, according to the standard reaction conditions (Section 2.3) identically prepared samples, containing methyl 4-methylbenzoate (1 equiv.), TPP (20 mol%), NH₄Br (2.5 eq.), NH₂OH•HCl (3 eq), 4 Å molecular sieves (25 mg) and 2 mL ACN, were irradiated with the same blue LED using different intensities. To exclude other influencing factors, such as, adverse effects of prolonged reaction time, the reactions were irradiated only for 90 minutes. After 30, 60 and 90 minutes, the irradiation was stopped for both reactions. Yields to the given time were determined by GC analysis using n-decane as internal standard. The slope showed a 2.3:1 (full power: half power) relation, which supports a mechanism involving more than one photon.

4.4 NMR studies

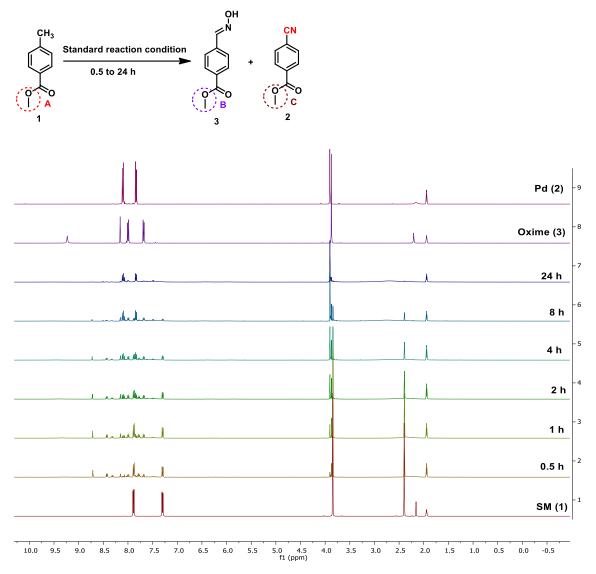


Fig. S13: ¹H NMR spectra for different time interval using CD₃CN as a solvent.

Reaction conditions: ^a0.1 mmol substrate, 20 mol% PC, 3 eq. NH₂OH•HCl, 2.5 eq. NH₄Br, 25 mg 4 Å MS, 1 bar O₂, 1 mL CD₃CN (0.1 M), 455 nm, 40 °C, 0.5-24 h. After each time interval, the reaction mixture was filtered through filter pad and the NMR of the solution was measured as such.

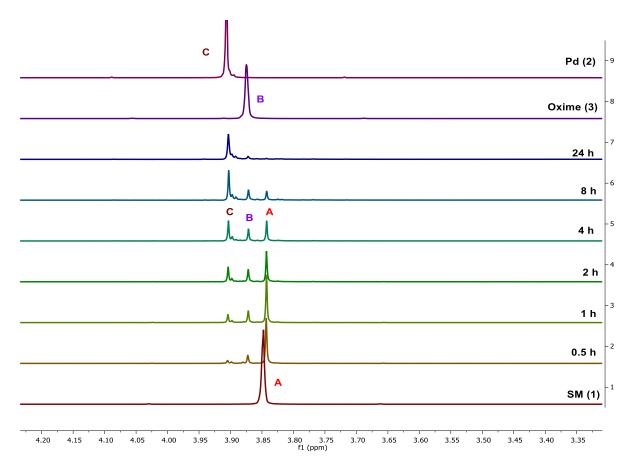


Fig. S13a: ¹H NMR spectra for different time interval using CD₃CN as a solvent.

Reaction conditions: ${}^{a}0.1$ mmol substrate, 20 mol% PC, 3 eq. NH₂OH•HCl, 2.5 eq. NH₄Br, 25 mg 4 Å MS, 1 bar O₂, 1 mL CD₃CN (0.1 M), 455 nm, 40 °C, 0.5-24 h. After each interval reaction mixture was filtered through filtered pad and measured as such a for NMR analysis.

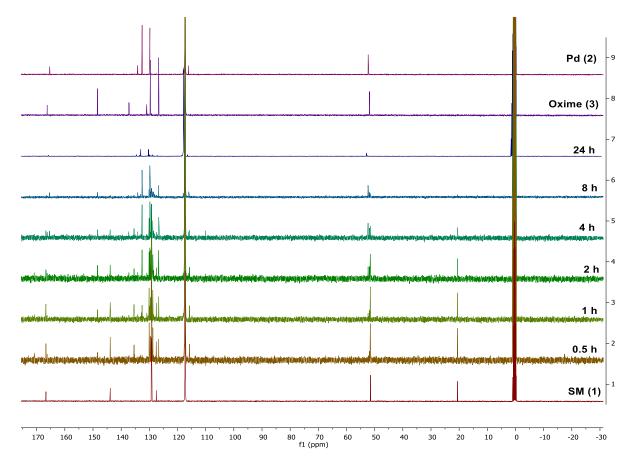


Fig. S14: ¹³C NMR spectra for different time interval using CD₃CN as a solvent.

Reaction conditions: ^a0.1 mmol substrate, 20 mol% PC, 3 eq. NH₂OH•HCl, 2.5 eq. NH₄Br, 25 mg 4 Å MS, 1 bar O₂, 1 mL CD₃CN (0.1 M), 455 nm, 40 °C, 0.5-24 h. After each interval reaction mixture was filtered through filtered pad and measured as such a for NMR analysis.

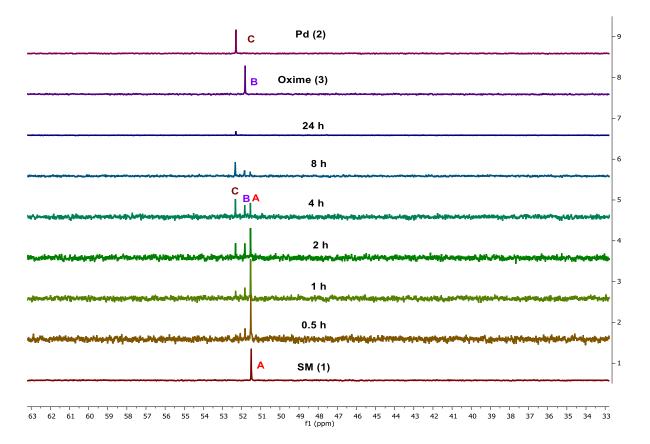
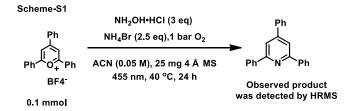
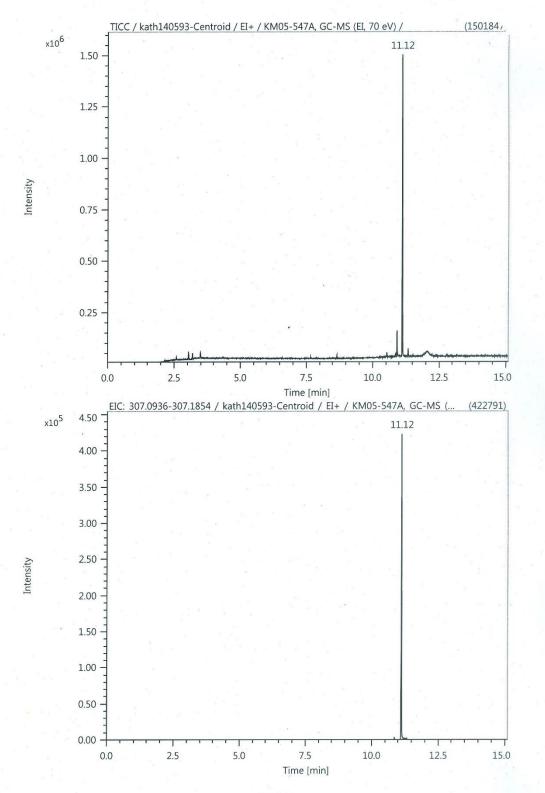


Fig. S14a: ¹³C NMR spectra for different time interval using CD₃CN as a solvent.

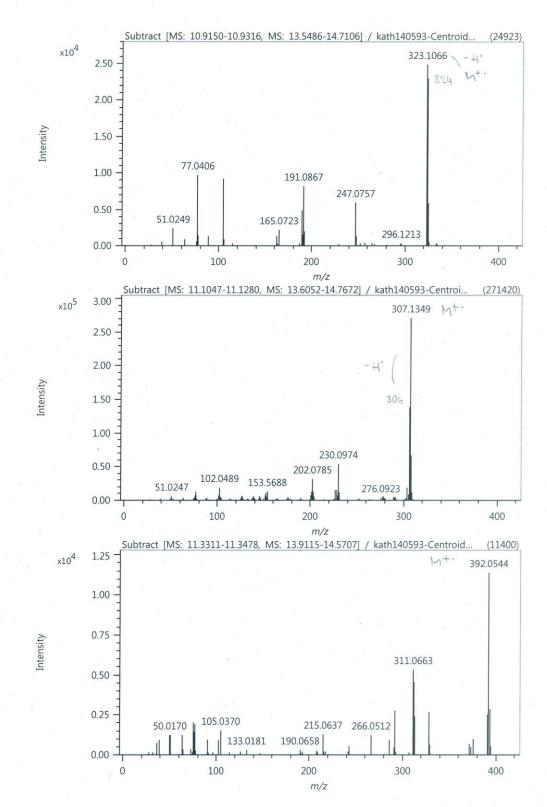
Reaction conditions: ^a0.1 mmol substrate, 20 mol% PC, 3 eq. NH₂OH•HCl, 2.5 eq. NH₄Br, 25 mg 4 Å MS, 1 bar O₂, 1 mL CD₃CN (0.1 M), 455 nm, 40 °C, 0.5-24 h. After each interval reaction mixture was filtered through filtered pad and measured as such a for NMR analysis.

4.5 Catalyst deactivation



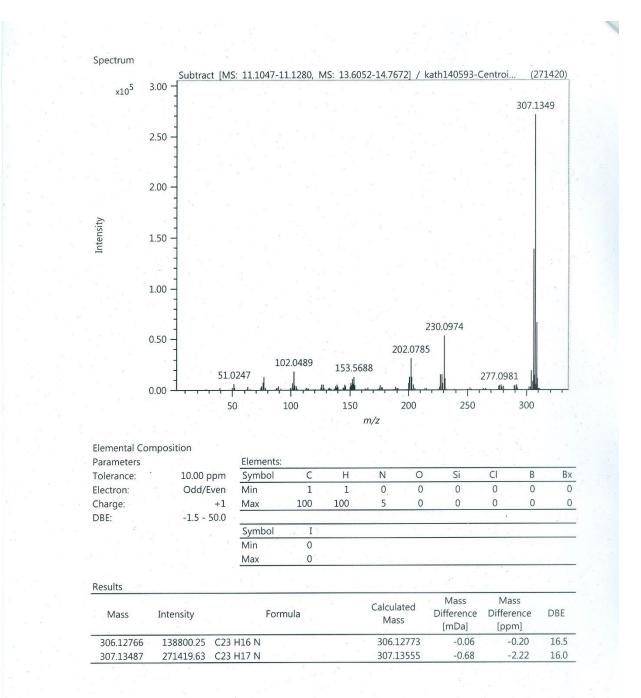


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2/2



** Search Report Page 1 of 1 **

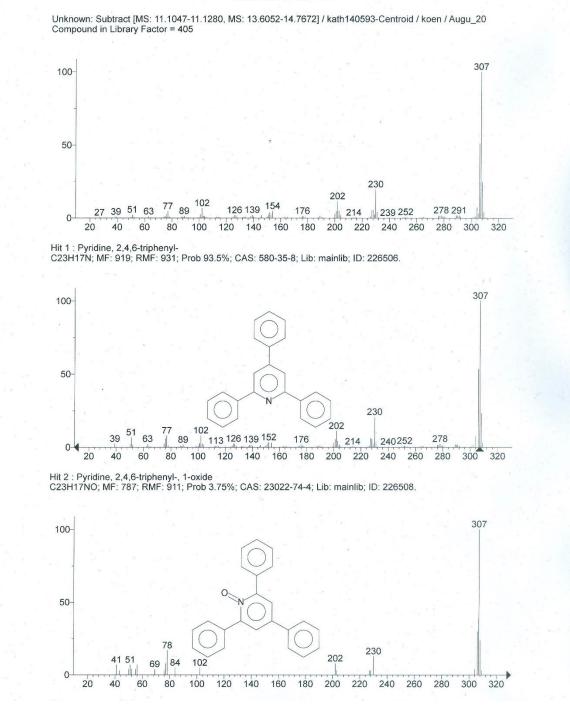


Fig. S15: HRMS and GC-MS spectra for catalyst deactivation

Reaction conditions: ^a0.1 mmol PC, 3 eq. NH₂OH•HCl, 2.5 eq. NH₄Br, 25 mg 4 Å MS, 1 bar O₂, 1 mL acetonitrile (0.05 M), 455 nm, 40 °C, 24 h.

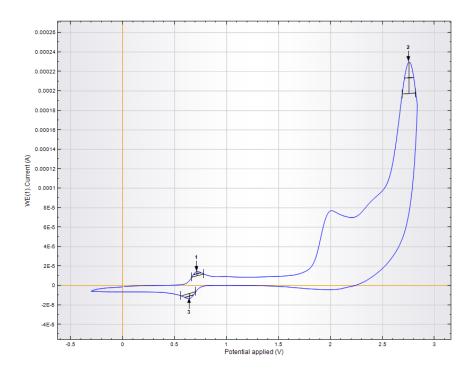
0.0003 0.0002 0.00026 0.00024 0.0002 0.0002 0.00018 (F) 0.00016 0.00014 0.00012 0.00012 0.0001 8E-5 6E-5 4E-5 2E-5 -2E-5 -4E-8 -0.5 0.5 Potential applied (V)

4.5 Cyclic Voltammetry measurement





Fig. S16: Cyclic voltammogram of methyl 4-methylbenzoate (1) in acetonitrile under argon. The peak at +2.74 V shows the oxidation of 1. The measurement was performed with a scan rate of 50 mV/s and with TBATFB (0.1M) as supporting electrolyte.



Index Peak position

- 1 0.71503
- 2 2.7544
- 3 0.64453

Fig. S17: Cyclic voltammogram of methyl 4-methylbenzoate (1) in the presence of ferrocene as internal standard in acetonitrile under argon. The peak at +2.75 V shows the oxidation of 1, whereas the reversible peak at +0.64 and +0.72 V corresponds to ferrocene. The measurement was performed with a scan rate of 50 mV/s and with TBATFB (0.1M) as supporting electrolyte.

With these values, the oxidation potential of **1** was calculated as follows:

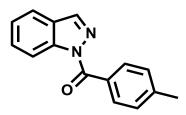
$$E_{1/2}(\mathbf{1}^{\bullet+}/\mathbf{1}) = \left(2.754 \, V - \frac{0.715 \, V + 0.645 \, V}{2}\right) \, vs. \, Fc^+/Fc = 2.07 \, V \, vs. Fc^+/Fc$$

This value was converted from the ferrocene reference to the SCE⁶:

$$E_{1/2}(1^{+}/1) = (2.07 V + 0.38 V) vs. SCE = 2.45 V vs. SCE$$

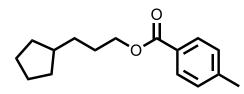
5. Characterization of prepared compounds

(1H-indazol-1-yl)(p-tolyl)methanone⁷



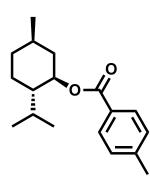
¹**H** NMR (400 MHz, Chloroform-*d*) δ 8.50 (dd, J = 8.4, 0.9 Hz, 1H), 8.12 (d, J = 0.8 Hz, 1H), 7.92 (d, J = 8.2 Hz, 2H), 7.69 (dd, J = 7.9, 1.0 Hz, 1H), 7.54 (ddd, J = 8.3, 7.1, 1.2 Hz, 1H), 7.32 (ddd, J = 8.0, 7.1, 0.9 Hz, 1H), 7.25 (d, J = 7.6 Hz, 2H), 2.38 (s, 3H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 168.35, 143.01, 140.21, 140.08, 131.14, 130.45, 129.43, 128.74, 126.11, 124.72, 120.91, 115.93, 21.70. HRMS (EI) (m/z): [M⁺] (C₁₅H₁₂N₂O⁺) calc. 236.0944; observed 236.0947. Off-white solid.

3-cyclopentylpropyl 4-methylbenzoate



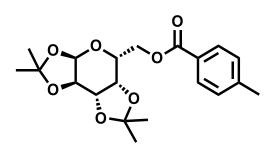
¹**H** NMR (400 MHz, Chloroform-*d*) δ 7.95 (d, *J* = 8.2 Hz, 2H), 7.25 (d, *J* = 8.0 Hz, 2H), 4.31 (t, *J* = 6.7 Hz, 2H), 2.42 (s, 3H), 1.87 – 1.72 (m, 5H), 1.68 – 1.40 (m, 6H), 1.18 – 1.02 (m, 2H). ¹³**C** NMR (101 MHz, Chloroform-*d*) δ 166.77 , 143.39 , 129.56 , 129.02 , 127.83 , 65.16 , 39.84 , 32.69 , 32.43 , 28.02 , 25.18 , 21.65 . HRMS (APCI) (m/z): [M+H⁺] ([C₁₆H₂₂O₂+H]⁺) calc. 247.1693; observed 247.1698. Yellow oil.

(1R,2S,5R)-2-isopropyl-5-methylcyclohexyl 4-methylbenzoate⁸



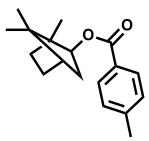
¹**H NMR** (**400 MHz, Chloroform-***d***)** δ 7.95 (d, J = 8.2 Hz, 2H), 7.25 (d, J = 7.9 Hz, 2H), 4.93 (td, J = 10.9, 4.4 Hz, 1H), 2.42 (s, 3H), 2.20 – 2.09 (m, 1H), 2.04 – 1.88 (m, 1H), 1.83 – 1.70 (m, 2H), 1.63 – 1.46 (m, 2H), 1.24 – 1.02 (m, 2H), 0.98 – 0.90 (m, 7H), 0.81 (d, J = 7.0 Hz, 3H). ¹³C NMR (**101 MHz, Chloroform-***d***)** δ 166.16 , 143.28 , 129.58 , 128.99 , 128.15 , 74.59 , 47.31 , 41.02 , 34.37 , 31.46 , 26.52 , 23.69 , 22.06 , 21.63 , 20.77 , 16.56 . **HRMS (APCI)** (m/z): [M+H⁺] ([C₁₈H₂₆O₂+H]⁺) calc. 275.2006; observed 275.2011. **Colorless oil.**

((3aR,5R,5aS,8aS,8bR)-2,2,7,7-tetramethyltetrahydro-3aH-bis([1,3]dioxolo)[4,5-b:4',5'd]pyran-5-yl)methyl 4-methylbenzoate⁹



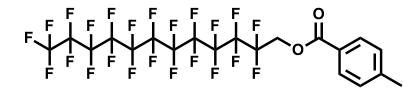
¹H NMR (400 MHz, Chloroform-*d*) δ 7.97 (d, *J* = 8.2 Hz, 2H), 7.25 (d, *J* = 8.0 Hz, 2H), 5.59 (d, *J* = 4.9 Hz, 1H), 4.68 (dd, *J* = 7.9, 2.5 Hz, 1H), 4.55 (dd, *J* = 11.5, 4.9 Hz, 1H), 4.47 – 4.33 (m, 3H), 4.25 – 4.16 (m, 1H), 2.43 (s, 3H), 1.54 (s, 3H), 1.50 (s, 3H), 1.38 (s, 3H), 1.36 (s, 3H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 166.49, 143.62, 129.74, 129.04, 127.33, 109.67, 108.80, 96.33, 71.17, 70.74, 70.56, 66.20, 63.69, 26.03, 25.99, 25.00, 24.50, 21.66. HRMS (ESI) (m/z): [M+H⁺] ([C₂₀H₂₆O₇+H]⁺) calc. 379.1751; observed 379.1753. Colorless gum.

1,7,7-trimethylbicyclo[2.2.1]heptan-2-yl 4-methylbenzoate



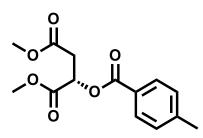
¹**H** NMR (400 MHz, Chloroform-*d*) δ 7.92 (d, *J* = 8.2 Hz, 2H), 7.25 (d, *J* = 7.8 Hz, 2H), 4.92 (dd, *J* = 7.1, 4.5 Hz, 1H), 2.42 (s, 3H), 1.99 – 1.69 (m, 4H), 1.68 – 1.57 (m, 1H), 1.30 – 1.18 (m, 2H), 1.14 (s, 3H), 0.95 (s, 3H), 0.91 (s, 3H). ¹³**C** NMR (101 MHz, Chloroform-*d*) δ 166.13 , 143.30 , 129.49 , 129.05 , 128.18 , 81.33 , 49.01 , 47.03 , 45.14 , 38.95 , 33.78 , 27.10 , 21.64 , 20.16 , 20.09 , 11.60 . HRMS (APCI) (m/z): [M+H⁺] ([C₁₈H₂₄O₂+H]⁺) calc. 273.1849; observed 273.1852. Colorless oil.

2,2,3,3,4,4,5,5,6,6,7,7,8,8,9,9,10,10,11,11,12,12,12-tricosafluorododecyl 4-methylbenzoate



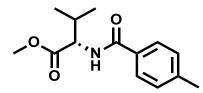
¹**H** NMR (400 MHz, Chloroform-*d*) δ 7.95 (d, *J* = 8.2 Hz, 2H), 7.27 (d, *J* = 8.3 Hz, 2H), 4.92 – 4.68 (m, 2H), 2.43 (s, 3H). ¹³**C** NMR (101 MHz, Chloroform-*d*) δ 164.96 , 144.84 , 130.03 , 129.35 , 125.57 , 59.96 , 21.74 . ¹⁹**F** NMR (282 MHz, Chloroform-*d*) δ -81.31 (t, *J* = 10.0 Hz, 3F), -119.58 – -120.10 (m, 2F), -121.95 – -122.72 (m, 12F), -123.02 – -123.47 (m, 2F), -123.49 – -124.17 (m, 2F), -126.40 – -127.02 (m, 2F). HRMS (APCI) (m/z): [M+NH₄⁺] ([C₂₀H₉F₂₃O₂+NH₄]⁺) calc. 736.0574; observed 736.0589. White solid.

(S)-dimethyl 2-((4-methylbenzoyl)oxy)succinate



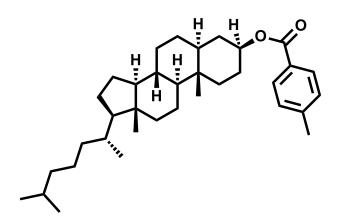
¹**H** NMR (400 MHz, Chloroform-*d*) δ 7.95 (d, *J* = 8.2 Hz, 2H), 7.25 (d, *J* = 8.0 Hz, 2H), 5.72 (dd, *J* = 6.9, 5.4 Hz, 1H), 3.79 (s, 3H), 3.73 (s, 3H), 3.04 (dd, *J* = 6.2, 1.8 Hz, 2H), 2.42 (s, 3H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 169.62, 169.50, 165.51, 144.30, 129.99, 129.15, 126.31 , 68.60, 52.73, 52.20, 36.21, 21.72. HRMS (APCI) (m/z): [M+H⁺] ([C₁₄H₁₆O₆+H]⁺) calc. 281.1020; observed 281.1023. Yellow oil.

(S)-methyl 3-methyl-2-(4-methylbenzamido)butanoate¹⁰



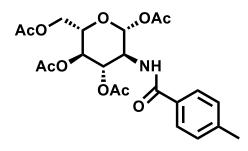
¹**H NMR** (**400 MHz**, **Chloroform**-*d*) δ 7.71 (d, *J* = 8.2 Hz, 2H), 7.25 (d, *J* = 8.0 Hz, 2H), 6.59 (d, *J* = 8.6 Hz, 1H), 4.79 (dd, *J* = 8.7, 4.9 Hz, 1H), 3.78 (s, 3H), 2.41 (s, 3H), 2.35 – 2.22 (m, 1H), 1.00 (dd, *J* = 9.2, 6.9 Hz, 6H). ¹³**C NMR** (**101 MHz**, **Chloroform**-*d*) δ 172.75 , 167.18 , 142.19 , 131.32 , 129.26 , 127.05 , 57.34 , 52.22 , 31.68 , 21.47 , 19.00 , 17.98 . **HRMS** (**ESI**) (m/z): [M+H⁺] ([C₁₄H₁₉NO₃+H]⁺) calc. 250.1438; observed 250.1440. White solid.

(3S,5S,8R,9S,10S,13R,14S,17R)-10,13-dimethyl-17-((R)-6-methylheptan-2yl)hexadecahydro-1H-cyclopenta[a]phenanthren-3-yl 4-methylbenzoate¹¹



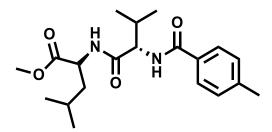
¹H NMR (400 MHz, Chloroform-*d*) δ 7.95 (d, J = 8.2 Hz, 2H), 7.25 (d, J = 8.0 Hz, 2H), 5.29 – 4.47 (m, 1H), 2.43 (s, 3H), 2.07 – 0.84 (m, 43H), 0.78 – 0.61 (m, 3H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 166.21 , 143.22 , 129.54 , 128.94 , 128.24 , 74.15 , 56.44 , 56.29 , 54.26 , 44.73 , 42.61 , 40.01 , 39.53 , 36.83 , 36.18 , 35.82 , 35.54 , 35.52 , 34.17 , 32.03 , 28.67 , 28.02 , 27.62 , 26.23 , 24.23 , 23.85 , 22.83 , 22.57 , 21.64 , 21.24 , 18.69 , 12.31 , 12.09 . HRMS (APCI) (m/z): [M+H⁺] ([C₃₅H₅₄O₂+H]⁺) calc. 507.4197; observed 507.4210. White solid.

(2R,3S,4S,5R,6S)-6-(acetoxymethyl)-3-(4-methylbenzamido)tetrahydro-2H-pyran-2,4,5triyl triacetate¹²



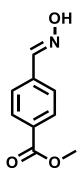
¹**H NMR** (**400 MHz, Chloroform-***d***)** δ 7.58 (d, J = 8.2 Hz, 2H), 7.19 (d, J = 8.0 Hz, 2H), 6.33 (d, J = 9.5 Hz, 1H), 5.79 (d, J = 8.8 Hz, 1H), 5.37 – 5.15 (m, 2H), 4.57 (dt, J = 10.4, 9.1 Hz, 1H), 4.29 (dd, J = 12.5, 4.7 Hz, 1H), 4.16 (dd, J = 12.5, 2.2 Hz, 1H), 3.86 (ddd, J = 9.8, 4.7, 2.2 Hz, 1H), 2.37 (s, 3H), 2.10 (s, 3H), 2.06 (s, 6H), 1.98 (s, 3H). ¹³C NMR (101 MHz, Chloroform-*d***)** δ 171.54, 170.69, 169.62, 169.25, 167.21, 142.55, 130.69, 129.39, 126.93, 92.88, 73.15, 72.73 , 67.77 , 61.76 , 53.22 , 21.45 , 20.86 , 20.75 , 20.63 , 20.60 . HRMS (ESI) (m/z): [M+H⁺] ([C₂₂H₂₇NO₁₀+H]⁺) calc. 466.1708; observed 466.1706. White solid.

(R)-methyl 4-methyl-2-((S)-3-methyl-2-(4-methylbenzamido)butanamido)pentanoate



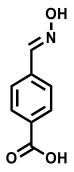
¹**H NMR** (**400 MHz**, **Chloroform**-*d*) δ 7.70 (d, J = 8.2 Hz, 2H), 7.22 (d, J = 7.4 Hz, 2H), 6.88 (d, J = 8.7 Hz, 1H), 6.81 (d, J = 7.8 Hz, 1H), 4.68 – 4.51 (m, 2H), 3.73 (s, 3H), 2.38 (s, 3H), 2.29 – 2.12 (m, 1H), 1.69 – 1.42 (m, 3H), 1.03 (dd, J = 8.2, 6.8 Hz, 6H), 0.84 (dd, J = 6.2, 4.9 Hz, 6H). ¹³**C NMR** (**101 MHz**, **Chloroform**-*d*) δ 173.11, 171.34, 167.31, 142.20, 131.23, 129.23, 127.12 , 58.49 , 52.24 , 50.92 , 41.07 , 31.75 , 24.80 , 22.65 , 21.81 , 21.47 , 19.15 , 18.37 . **HRMS** (**ESI**) (m/z): [M+H⁺] ([C₂₀H₃₀N₂O₄+H]⁺) calc. 363.2278; observed 363.2285. **White solid.**

(E)-methyl 4-((hydroxyimino)methyl)benzoate⁴



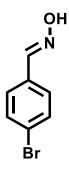
¹H NMR (400 MHz, Chloroform-*d*) δ 7.86 (s, 1H), 7.27 (d, *J* = 8.7 Hz, 2H), 6.66 (d, *J* = 8.7 Hz, 2H), 3.58 (s, 3H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 161.10 , 149.92 , 128.56 , 124.56 , 114.27 , 113.96 , 55.36 . White solid. (Exchangeable OH-proton was not picked-up)

(E)-4-((hydroxyimino)methyl)benzoic acid¹³



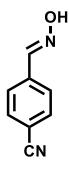
¹**H** NMR (400 MHz, DMSO-*d*₆) δ 13.03 (br s, 1H), 11.53 (s, 1H), 8.21 (s, 1H), 7.94 (d, *J* = 8.3 Hz, 2H), 7.69 (d, *J* = 8.3 Hz, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 167.38, 147.99, 137.65, 131.56, 130.15, 126.88. White solid.

(E)-4-bromobenzaldehyde oxime¹⁴



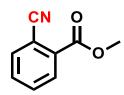
¹H NMR (400 MHz, DMSO-*d*₆) δ 11.35 (s, 1H), 8.12 (s, 1H), 7.59 (d, *J* = 8.6 Hz, 2H), 7.53 (d, *J* = 8.6 Hz, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 147.68 , 132.82 , 132.16 , 128.75 , 122.86 . White solid.

(E)-4-((hydroxyimino)methyl)benzonitrile¹⁵



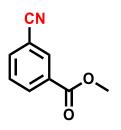
¹H NMR (400 MHz, DMSO-*d*₆) δ 11.72 (s, 1H), 8.23 (s, 1H), 7.85 (d, *J* = 8.4 Hz, 2H), 7.76 (d, *J* = 8.4 Hz, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 147.57 , 138.08 , 133.14 , 127.47 , 119.16 , 111.83 . White solid.

methyl 2-cyanobenzoate¹⁶



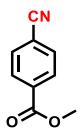
¹**H NMR (400 MHz, Chloroform-***d***)** δ 8.18 – 8.09 (m, 1H), 7.83 – 7.77 (m, 1H), 7.70 – 7.63 (m, 2H), 3.99 (s, 3H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 164.51 , 134.82 , 132.72 , 132.50 , 132.42 , 131.19 , 117.53 , 112.95 , 52.87 . Yellow solid.

methyl 3-cyanobenzoate¹⁷



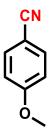
¹H NMR (400 MHz, Chloroform-*d*) δ 8.32 (td, *J* = 1.7, 0.6 Hz, 1H), 8.26 (dt, *J* = 8.0, 1.5 Hz, 1H), 7.83 (dt, *J* = 7.8, 1.4 Hz, 1H), 7.58 (td, *J* = 7.9, 0.7 Hz, 1H), 3.95 (s, 3H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 165.09, 135.98, 133.65, 133.27, 131.44, 129.46, 117.88, 112.99, 52.72. White solid.

methyl 4-cyanobenzoate¹⁸



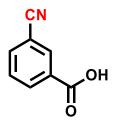
¹**H NMR (400 MHz, Chloroform-***d***)** δ 8.13 (d, *J* = 8.5 Hz, 2H), 7.74 (d, *J* = 8.4 Hz, 2H), 3.95 (s, 3H). ¹³**C NMR (101 MHz, Chloroform-***d***)** δ 165.43 , 133.94 , 132.24 , 130.11 , 117.97 , 116.42 , 52.74 . White solid.

4-methoxybenzonitrile¹⁹



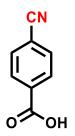
¹**H NMR (400 MHz, Chloroform-***d***)** δ 7.58 (d, *J* = 8.9 Hz, 2H), 6.95 (d, *J* = 8.8 Hz, 2H), 3.86 (s, 3H). ¹³**C NMR (101 MHz, Chloroform-***d***)** δ 162.86 , 134.00 , 119.24 , 114.77 , 104.00 , 55.57 . White solid.

3-cyanobenzoic acid²⁰



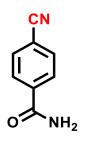
¹**H** NMR (400 MHz, DMSO-*d*₆) δ 8.27 (d, *J* = 1.7 Hz, 1H), 8.22 (dt, *J* = 7.9, 1.5 Hz, 1H), 8.08 (dt, *J* = 7.8, 1.5 Hz, 1H), 7.78 – 7.66 (m, 1H). ¹³**C** NMR (101 MHz, DMSO-*d*₆) δ 166.15, 136.72 , 134.22 , 133.22 , 132.51 , 130.55 , 118.53 , 112.38 . White solid. (Exchangeable OH-proton was not picked-up)

4-cyanobenzoic acid²¹



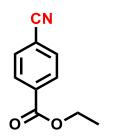
¹H NMR (400 MHz, DMSO-*d*₆) δ 13.55 (br s, 1H), 8.08 (d, *J* = 8.5 Hz, 2H), 7.98 (d, *J* = 8.4 Hz, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 166.48 , 135.32 , 133.12 , 130.37 , 118.64 , 115.51 . Off-white solid.

4-cyanobenzamide²²



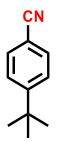
¹**H NMR (400 MHz, DMSO-***d*₆) δ 8.19 (br s, 1H), 8.01 (d, *J* = 8.7 Hz, 2H), 7.93 (d, *J* = 8.4 Hz, 2H), 7.66 (br s, 1H). ¹³**C NMR (101 MHz, DMSO-***d*₆) δ 166.90, 138.75, 132.84, 128.71, 118.83, 114.10. White solid.

ethyl 4-cyanobenzoate²³



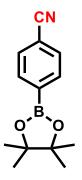
¹**H NMR** (**400 MHz**, **Chloroform**-*d*) δ 8.14 (d, J = 8.7 Hz, 2H), 7.74 (d, J = 8.7 Hz, 2H), 4.42 (q, J = 7.1 Hz, 2H), 1.41 (t, J = 7.1 Hz, 3H). ¹³**C NMR** (**101 MHz**, **Chloroform**-*d*) δ 164.96, 134.32, 132.19, 130.08, 118.03, 116.30, 61.84, 14.26. White solid.

4-(tert-butyl)benzonitrile²³



¹H NMR (400 MHz, Chloroform-d) δ 7.58 (d, J = 8.7 Hz, 2H), 7.48 (d, J = 8.5 Hz, 2H), 1.33 (s, 9H).
¹³C NMR (101 MHz, Chloroform-d) δ 156.66, 131.98, 126.19, 119.17, 109.33, 35.29, 30.97.

4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)benzonitrile²⁴



¹**H NMR (400 MHz, Chloroform-***d***)** δ 7.88 (d, *J* = 8.2 Hz, 2H), 7.64 (d, *J* = 8.3 Hz, 2H), 1.35 (s, 12H). ¹³**C NMR (101 MHz, Chloroform-***d***)** δ 135.11, 135.10, 131.15, 118.88, 114.56, 84.52, 24.89. White solid.

<u>1-naphthonitrile²⁵</u>



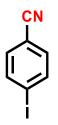
¹**H NMR** (**400 MHz**, **Chloroform**-*d*) δ 8.28 – 8.19 (m, 1H), 8.08 (dd, J = 8.4, 1.1 Hz, 1H), 7.91 (ddd, J = 7.1, 6.3, 1.1 Hz, 2H), 7.69 (ddd, J = 8.3, 6.9, 1.4 Hz, 1H), 7.62 (ddd, J = 8.2, 6.9, 1.3 Hz, 1H), 7.52 (dd, J = 8.3, 7.2 Hz, 1H). ¹³**C NMR** (**101 MHz**, **Chloroform**-*d*) δ 133.31 , 132.95 , 132.65 , 132.38 , 128.69 , 128.62 , 127.58 , 125.17 , 124.95 , 117.85 , 110.22 . **Off-white solid.**

4-chlorobenzonitrile²⁵



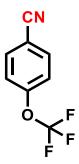
¹H NMR (400 MHz, Chloroform-*d*) δ 7.60 (d, *J* = 8.6 Hz, 2H), 7.46 (d, *J* = 8.6 Hz, 2H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 139.57, 133.40, 129.72, 117.98, 110.82. White solid.

4-iodobenzonitrile²⁶



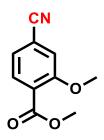
¹H NMR (400 MHz, Chloroform-*d*) δ 7.85 (d, *J* = 8.4 Hz, 2H), 7.37 (d, *J* = 8.4 Hz, 2H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 138.54, 133.18, 118.23, 111.78, 100.32. Pale brown solid.

4-(trifluoromethoxy)benzonitrile²³



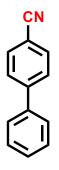
¹H NMR (400 MHz, Chloroform-*d*) δ 7.72 (d, *J* = 8.9 Hz, 2H), 7.32 (d, *J* = 8.0 Hz, 2H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 152.21 (d, *J* = 1.9 Hz), 134.19 ,120.18 (q, *J* = 259.7 Hz), 121.23 , 117.65 , 110.85 . ¹⁹F NMR (376 MHz, Chloroform-*d*) δ -58.30 . Yellow oil.

methyl 4-cyano-2-methoxybenzoate²⁷



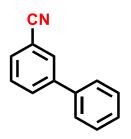
¹H NMR (400 MHz, Chloroform-*d*) δ 7.68 – 7.58 (m, 3H), 3.98 (s, 3H), 3.93 (s, 3H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 165.50 , 161.12 , 135.49 , 133.74 , 121.65 , 115.61 , 112.03 , 105.84 , 56.36 , 52.78 . HRMS (EI) (m/z): [M⁺] (C₁₀H₉NO₃⁺) calc. 191.0576; observed 191.0578. Yellow solid.

[1,1'-biphenyl]-4-carbonitrile²⁶



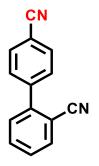
¹H NMR (400 MHz, Chloroform-*d*) δ 7.76 – 7.66 (m, 4H), 7.62 – 7.56 (m, 2H), 7.53 – 7.39 (m, 3H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 145.70 , 139.20 , 132.62 , 129.14 , 128.68 , 127.76 , 127.25 , 118.96 , 110.95 . White solid.

[1,1'-biphenyl]-3-carbonitrile²⁶



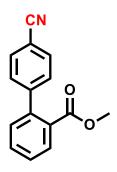
¹H NMR (400 MHz, Chloroform-*d*) δ 7.90 – 7.84 (m, 1H), 7.82 (dt, *J* = 7.8, 1.6 Hz, 1H), 7.64 (dt, *J* = 7.7, 1.4 Hz, 1H), 7.59 – 7.52 (m, 3H), 7.52 – 7.38 (m, 3H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 142.48, 138.91, 131.51, 130.74, 130.71, 129.62, 129.15, 128.41, 127.11, 118.87, 112.99. Yellow gum.

[1,1'-biphenyl]-2,4'-dicarbonitrile²⁸



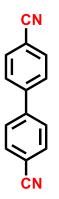
¹H NMR (400 MHz, Chloroform-*d*) δ 7.59 – 7.52 (m, 3H), 7.50 – 7.40 (m, 3H), 7.34 – 7.24 (m, 2H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 143.33 , 142.53 , 133.99 , 133.20 , 132.54 , 129.93 , 129.59 , 128.80 , 118.40 , 118.03 , 112.66 , 111.25 . HRMS (EI) (m/z): [M⁺] (C₁₄H₈N₂⁺) calc. 204.0682; observed 204.0682. Off-white solid.

methyl 4'-cyano-[1,1'-biphenyl]-2-carboxylate²⁹



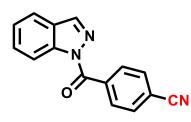
¹**H NMR** (400 MHz, Chloroform-*d*) δ 7.78 (dd, J = 7.7, 1.4 Hz, 1H), 7.54 (d, J = 8.3 Hz, 2H), 7.47 – 7.38 (m, 1H), 7.38 – 7.29 (m, 1H), 7.25 (d, J = 8.3 Hz, 2H), 7.16 (dd, J = 7.7, 1.3 Hz, 1H), 3.52 (s, 3H). ¹³**C NMR** (101 MHz, Chloroform-*d*) δ 167.93 , 146.39 , 141.09 , 131.79 , 131.77 , 130.52 , 130.45 , 130.07 , 129.19 , 128.29 , 118.89 , 111.10 , 52.11 . HRMS (EI) (m/z): [M⁺] (C₁₅H₁₁NO₂⁺) calc. 237.0784; observed 237.0786. Colorless gum.

[1,1'-biphenyl]-4,4'-dicarbonitrile²³



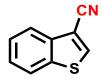
¹H NMR (400 MHz, Chloroform-*d*) δ 7.78 (d, *J* = 8.6 Hz, 4H), 7.69 (d, *J* = 8.6 Hz, 4H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 146.76, 132.91, 127.95, 118.41, 112.48. HRMS (EI) (m/z): [M⁺] (C₁₄H₈N₂⁺) calc. 204.0682; observed 204.0684. White solid.

4-(1H-indazole-1-carbonyl)benzonitrile³⁰



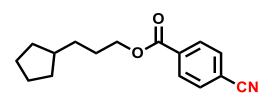
¹**H NMR** (**400 MHz**, **Chloroform-***d***)** δ 8.57 (dt, *J* = 8.4, 0.9 Hz, 1H), 8.22 (s, 1H), 8.16 (d, *J* = 8.4 Hz, 2H), 7.87 – 7.76 (m, 3H), 7.66 (ddd, *J* = 8.4, 7.2, 1.1 Hz, 1H), 7.46 (ddd, *J* = 8.0, 7.1, 0.9 Hz, 1H). ¹³**C NMR** (**101 MHz**, **Chloroform-***d***)** δ 166.57, 141.22, 139.96, 137.36, 131.74, 131.38, 130.03, 126.28, 125.45, 121.21, 118.06, 115.93, 115.53. **HRMS** (**EI**) (m/z): [M⁺] (C₁₅H₉N₃O⁺) calc. 247.0740; observed 247.0749. **White solid.**

benzo[b]thiophene-3-carbonitrile²³



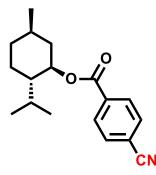
¹H NMR (400 MHz, Chloroform-*d*) δ 7.89 (s, 1H), 7.81 – 7.74 (m, 1H), 7.72 – 7.65 (m, 1H), 7.38 – 7.20 (m, 2H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 138.52 , 137.54 , 137.30 , 126.21 , 126.01 , 122.86 , 122.58 , 114.34 , 107.17 . HRMS (EI) (m/z): [M⁺] (C₉H₅NS⁺) calc. 159.0137; observed 159.0133. Brown solid.

3-cyclopentylpropyl 4-cyanobenzoate



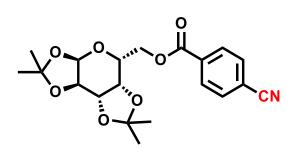
¹**H** NMR (400 MHz, Chloroform-*d*) δ 8.13 (d, *J* = 8.4 Hz, 2H), 7.74 (d, *J* = 8.4 Hz, 2H), 4.34 (t, *J* = 6.7 Hz, 2H), 1.86 – 1.70 (m, 5H), 1.66 – 1.36 (m, 6H), 1.17 – 1.03 (m, 2H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 164.99, 134.34, 132.19, 130.05, 118.02, 116.27, 66.18, 39.78, 32.67, 32.34, 27.89, 25.16. HRMS (APCI) (m/z): [M+NH₄⁺] (C₁₆H₁₉NO₂+NH₄⁺) calc. 275.1754; observed 275.1760. Colorless oil.

(1R,2S,5R)-2-isopropyl-5-methylcyclohexyl 4-cyanobenzoate³¹



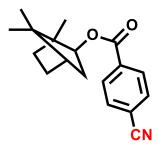
¹H NMR (400 MHz, Chloroform-*d*) δ 8.13 (d, *J* = 8.4 Hz, 2H), 7.73 (d, *J* = 8.4 Hz, 2H), 4.95 (td, *J* = 10.9, 4.4 Hz, 1H), 2.18 – 2.03 (m, 1H), 1.96 – 1.84 (m, 1H), 1.78 – 1.67 (m, 2H), 1.63 – 1.49 (m, 2H), 1.19 – 1.02 (m, 2H), 0.98 – 0.87 (m, 7H), 0.78 (d, *J* = 7.0 Hz, 3H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 164.42, 134.65, 132.16, 130.06, 118.05, 116.16, 75.94, 47.20, 40.85, 34.20 , 31.45 , 26.57 , 23.59 , 22.00 , 20.73 , 16.49 . HRMS (APCI) (m/z): [M+NH₄⁺] ([C₁₈H₂₃NO₂+NH₄]⁺) calc. 303.2067; observed 303.2068. Yellow gum.

((3aR,5R,5aS,8aS,8bR)-2,2,7,7-tetramethyltetrahydro-3aH-bis([1,3]dioxolo)[4,5-b:4',5'd]pyran-5-yl)methyl 4-cyanobenzoate³²

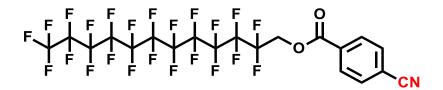


¹**H NMR** (**400 MHz, Chloroform-***d***)** δ 8.13 (d, *J* = 8.5 Hz, 2H), 7.72 (d, *J* = 8.5 Hz, 2H), 5.55 (d, *J* = 4.9 Hz, 1H), 4.64 (dd, *J* = 7.9, 2.5 Hz, 1H), 4.53 (dd, *J* = 11.6, 4.5 Hz, 1H), 4.45 (dd, *J* = 11.6, 7.8 Hz, 1H), 4.34 (dd, *J* = 5.0, 2.5 Hz, 1H), 4.30 (dd, *J* = 7.9, 1.9 Hz, 1H), 4.22 – 4.11 (m, 1H), 1.47 (d, *J* = 13.4 Hz, 6H), 1.33 (d, *J* = 7.6 Hz, 6H). ¹³**C NMR** (**101 MHz, Chloroform-***d***)** δ 164.84 , 133.89 , 132.23 , 130.20 , 117.98 , 116.46 , 109.82 , 108.85 , 96.32 , 71.10 , 70.76 , 70.46 , 66.06 , 64.74 , 26.02 , 25.98 , 24.95 , 24.50 . **HRMS** (**ESI**) (m/z): [M+H⁺] ([C₂₀H₂₃NO₇+H]⁺) calc. 390.1547; observed 390.1559. **Yellow gum.**

1,7,7-trimethylbicyclo[2.2.1]heptan-2-yl 4-cyanobenzoate

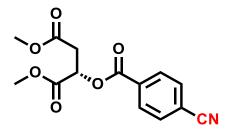


¹**H** NMR (400 MHz, Chloroform-*d*) δ 8.09 (d, J = 8.4 Hz, 2H), 7.73 (d, J = 8.5 Hz, 2H), 4.93 (dd, J = 7.4, 4.2 Hz, 1H), 1.97 – 1.85 (m, 2H), 1.85 – 1.70 (m, 2H), 1.67 – 1.55 (m, 1H), 1.28 – 1.12 (m, 2H), 1.10 (s, 3H), 0.91 (s, 3H), 0.89 (s, 3H). ¹³**C** NMR (101 MHz, Chloroform-*d*) δ 164.37, 134.68, 132.24, 129.94, 118.03, 116.18, 82.60, 49.13, 47.06, 45.07, 38.82, 33.69, 27.01, 20.08, 20.06, 11.59. HRMS (APCI) (m/z): [M+NH₄⁺] ([C₁₈H₂₁NO₂+NH₄]⁺) calc. 301.1911; observed 301.1918. White solid.



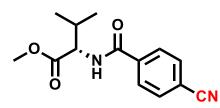
¹**H** NMR (400 MHz, Chloroform-*d*) δ 8.17 (d, *J* = 8.5 Hz, 2H), 7.79 (d, *J* = 8.4 Hz, 2H), 4.85 (t, *J* = 13.1 Hz, 2H).¹³**C** NMR (101 MHz, Chloroform-*d*) δ 163.35, 132.45, 132.03, 130.44, 117.61 , 117.45 , 60.56 (t, *J* = 27.8 Hz). ¹⁹**F** NMR (282 MHz, Chloroform-*d*) δ -81.35 (t, *J* = 10.0 Hz, 3F), -119.41 - -120.36 (m, 2F), -121.97 - -122.76 (m, 12F), -123.19 - -123.37 (m, 2F), -123.58 - -123.80 (m, 2F), -126.26 - -126.97 (m, 2F). HRMS (APCI) (m/z): [M+NH₄⁺] ([C₂₀H₆F₂₃NO₂+NH₄]⁺) calc. 747.0370; observed 747.0391. White solid.

(S)-dimethyl 2-((4-cyanobenzoyl)oxy)succinate



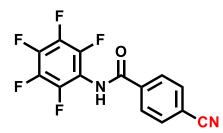
¹**H** NMR (400 MHz, Chloroform-*d*) δ 8.13 (d, J = 8.6 Hz, 2H), 7.74 (d, J = 8.7 Hz, 2H), 5.72 (dd, J = 6.8, 5.3 Hz, 1H), 3.78 (s, 3H), 3.71 (s, 3H), 3.07 – 3.00 (m, 2H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 169.39 , 168.82 , 163.90 , 132.89 , 132.29 , 130.42 , 117.83 , 116.89 , 69.27 , 52.93 , 52.31 , 35.92 . HRMS (APCI) (m/z): [M+NH₄⁺] ([C₁₄H₁₃NO₆+NH₄]⁺) calc. 309.1081; observed 309.1089. Yellow gum.

(S)-methyl 2-(4-cyanobenzamido)-3-methylbutanoate



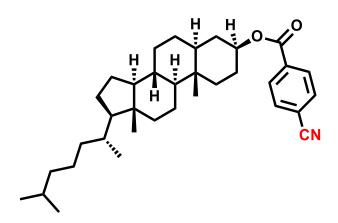
¹**H NMR** (**400 MHz, Chloroform-***d***)** δ 7.89 (d, J = 8.4 Hz, 2H), 7.73 (d, J = 8.6 Hz, 2H), 6.74 (d, J = 8.6 Hz, 1H), 4.75 (dd, J = 8.6, 4.9 Hz, 1H), 3.78 (s, 3H), 2.44 – 2.08 (m, 1H), 0.99 (dd, J = 7.9, 6.9 Hz, 6H). ¹³**C NMR** (**101 MHz, Chloroform-***d***)** δ 172.43 , 165.60 , 138.02 , 132.49 , 127.83 , 117.96 , 115.33 , 57.71 , 52.46 , 31.59 , 19.00 , 18.00 . HRMS (ESI) (m/z): [M+H⁺] ([C₁₄H₁₆N₂O₃+H]⁺) calc. 261.1234; observed 261.1238. **Off-white solid.**

4-cyano-N-(perfluorophenyl)benzamide³³



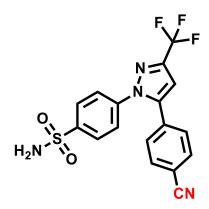
¹H NMR (400 MHz, DMSO-*d*₆) δ 10.85 (s, 1H), 8.15 (d, *J* = 8.4 Hz, 2H), 8.07 (d, *J* = 8.4 Hz, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 164.71 , 153.11 – 151.07 (m), 145.22 – 143.67 (m), 139.64 – 138.22 (m), 136.69 , 134.29 – 132.51 (m), 130.10 – 128.57 (m), 118.57 , 115.30 , 113.35 – 112.62 (m). ¹⁹F NMR (376 MHz, DMSO-*d*₆) δ -144.66 (d, *J* = 21.0 Hz, 2F), -156.39 (t, *J* = 22.9 Hz, 1F), -159.09 – -168.13 (m, 2F). HRMS (EI) (m/z): [M⁺] (C₁₄H₅N₂F₅O⁺) calc. 312.0316; observed 312.0308. Colorless gum.

(3S,5S,8R,9S,10S,13R,14S,17R)-10,13-dimethyl-17-((R)-6-methylheptan-2yl)hexadecahydro-1H-cyclopenta[a]phenanthren-3-yl 4-cyanobenzoate



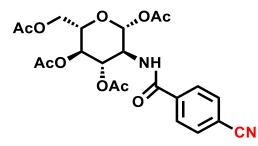
¹H NMR (400 MHz, Chloroform-*d*) δ 8.12 (d, J = 8.4 Hz, 2H), 7.72 (d, J = 8.4 Hz, 2H), 5.04 – 4.86 (m, 1H), 2.03 – 0.81 (m, 42H), 0.75 – 0.55 (m, 4H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 164.43, 134.78, 132.12, 130.06, 118.08, 116.14, 75.47, 56.43, 56.30, 54.24, 44.71, 42.62, 40.00, 39.54, 36.77, 36.20, 35.83, 35.53, 35.51, 34.05, 32.00, 28.65, 28.27, 28.04, 27.53, 24.24, 23.87, 22.85, 22.59, 21.26, 18.71, 12.31, 12.11. HRMS (APCI) (m/z): [M+NH₄⁺] ([C₃₅H₅₁NO₂+NH₄]⁺) calc. 535.4258; observed 535.4255. White solid.

4-(5-(4-cyanophenyl)-3-(trifluoromethyl)-1H-pyrazol-1-yl)benzenesulfonamide³⁴



¹H NMR (**300** MHz, Chloroform-*d*) δ 7.96 (d, J = 8.8 Hz, 2H), 7.69 (d, J = 8.7 Hz, 2H), 7.46 (d, J = 8.7 Hz, 2H), 7.37 (d, J = 8.6 Hz, 2H), 6.87 (s, 1H), 4.95 (s, 2H). ¹³C NMR (**101** MHz, Chloroform-*d*) δ 144.56 (q, J = 39.0 Hz), 143.00 , 142.20 , 141.85 , 132.94 , 132.87 , 129.41 , 127.90 , 125.64 , 120.74 (q, J = 269.6 Hz), 117.82 , 113.47 , 107.51 . ¹⁹F NMR (**376** MHz, Chloroform-*d*) δ -63.03 . HRMS (ESI) (m/z): [M+H⁺] ([C₁₇H₁₁F₃N₄O₂S+H]⁺) calc. 393.0628; observed 393.0636. Off-white solid.

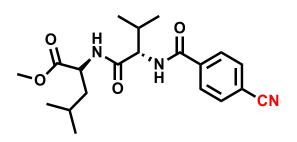
<u>(2R,3S,4S,5R,6S)-6-(acetoxymethyl)-3-(4-cyanobenzamido)tetrahydro-2H-pyran-2,4,5-triyl</u> triacetate



¹**H NMR** (**400 MHz**, **Chloroform-***d*) δ 7.79 (d, J = 8.6 Hz, 2H), 7.71 (d, J = 8.6 Hz, 2H), 6.54 (d, J = 9.4 Hz, 1H), 5.81 (d, J = 8.7 Hz, 1H), 5.36 – 5.15 (m, 2H), 4.53 (dt, J = 10.4, 9.1 Hz, 1H), 4.29 (dd, J = 12.5, 4.7 Hz, 1H), 4.20 – 4.06 (m, 1H), 3.91 – 3.82 (m, 1H), 2.10 (s, 3H), 2.07 (s, 3H), 2.06 (s, 3H), 2.00 (s, 3H). ¹³**C NMR** (**101 MHz**, **Chloroform-***d*) δ 171.63 , 170.69 , 169.58 , 169.25 , 165.57 , 137.34 , 132.62 , 127.66 , 117.78 , 115.64 , 92.72 , 73.14 , 72.67 , 67.63 , 61.69

, 53.79 , 20.86 , 20.74 , 20.65 , 20.60. HRMS (ESI) (m/z): $[M+H^+]$ ($[C_{22}H_{24}N_2O_{10}+H]^+$) calc. 477.1504; observed 477.1505. Brown solid.

(R)-methyl 2-((S)-2-(4-cyanobenzamido)-3-methylbutanamido)-4-methylpentanoate



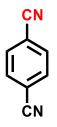
¹**H NMR** (**400 MHz**, **Chloroform**-*d*) δ 7.82 (d, J = 8.4 Hz, 2H), 7.60 (d, J = 8.4 Hz, 2H), 7.27 (d, J = 8.7 Hz, 1H), 6.86 (d, J = 7.8 Hz, 1H), 4.57 – 4.41 (m, 2H), 3.63 (s, 3H), 2.16 – 2.03 (m, 1H), 1.58 – 1.35 (m, 3H), 0.91 (dd, J = 9.7, 6.7 Hz, 6H), 0.74 (d, J = 6.1 Hz, 6H). ¹³**C NMR** (**101 MHz**, **Chloroform**-*d*) δ 173.00 , 171.20 , 165.65 , 137.85 , 132.38 , 127.96 , 117.94 , 115.29 , 58.94 , 52.32 , 51.02 , 41.00 , 31.74 , 24.81 , 22.64 , 21.81 , 19.08 , 18.48 . **HRMS** (**ESI**) (m/z): [M+H⁺] ([C₂₀H₂₇N₃O₄+H]⁺) calc. 374.2074; observed 374.2078. **Brown solid**

4-(pentafluorothio)benzonitrile³⁵



¹H NMR (400 MHz, Chloroform-*d*) δ 7.89 (d, J = 8.8 Hz, 2H), 7.79 (d, J = 9.6 Hz, 2H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 157.62 – 155.08 (m), 132.72 , 128.44 – 125.04 (m), 116.90 , 115.87 . White solid

terephthalonitrile²⁵



¹H NMR (400 MHz, Chloroform-*d*) δ 7.80 (s, 4H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 132.80, 117.01, 116.73. White solid

isophthalonitrile²⁵



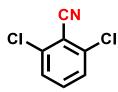
¹**H NMR (400 MHz, Chloroform-***d***)** δ 7.99 – 7.93 (m, 1H), 7.91 (d, *J* = 1.6 Hz, 1H), 7.89 (d, *J* = 1.6 Hz, 1H), 7.70 – 7.61 (m, 1H). ¹³**C NMR (101 MHz, Chloroform-***d***)** δ 136.00, 135.43, 130.35 , 116.60, 114.19. White solid.

3,5-dibromobenzonitrile³⁶

CN Br

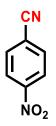
¹H NMR (400 MHz, Chloroform-*d*) δ 7.91 (t, *J* = 1.8 Hz, 1H), 7.74 (d, *J* = 1.7 Hz, 2H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 138.84, 133.45, 123.60, 115.92, 115.47. White solid.

2,6-dichlorobenzonitrile³⁷



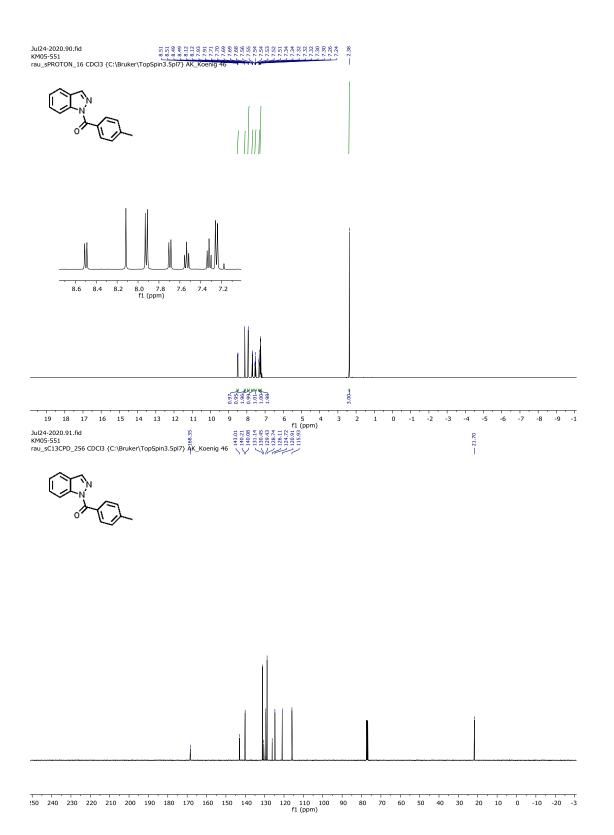
¹H NMR (400 MHz, Chloroform-*d*) δ 7.96 – 6.74 (m, 3H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 138.51, 133.86, 128.17, 114.44, 113.35. White solid.

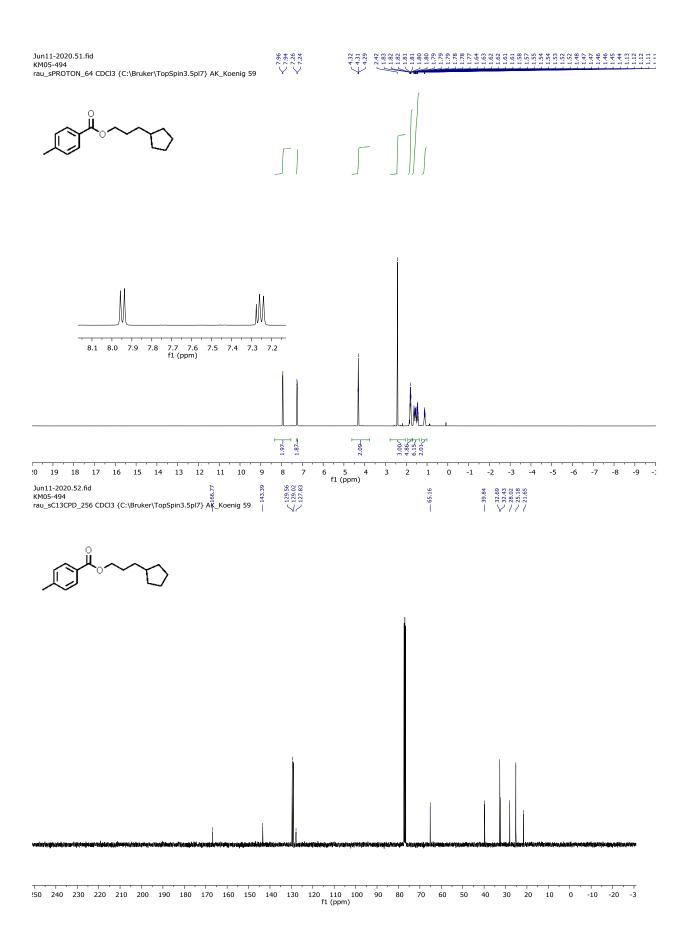
4-nitrobenzonitrile²⁵



¹**H NMR (400 MHz, Chloroform-***d***)** δ 8.36 (d, *J* = 8.8 Hz, 2H), 7.89 (d, *J* = 8.8 Hz, 2H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 150.04, 133.48, 124.30, 118.35, 116.79. **Yellow solid.**

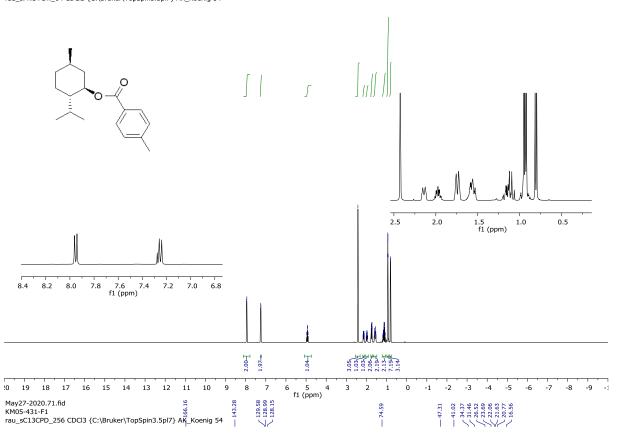
6. NMR Spectra

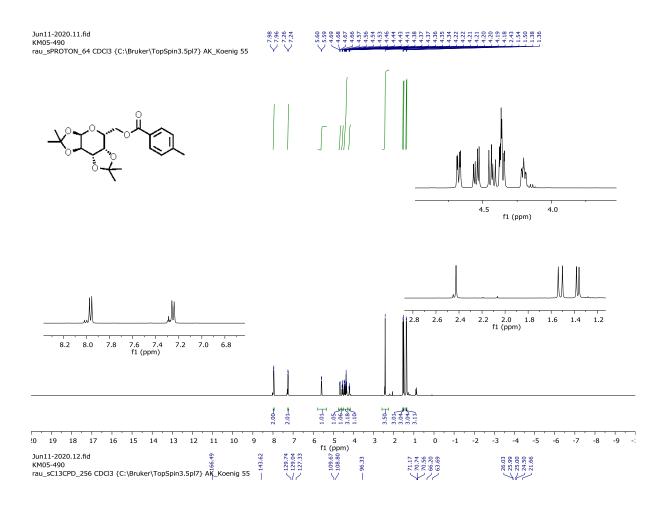




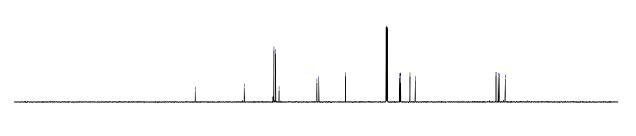
May27-2020.70.fid KM05-431-F1 rau_sPROTON_64 CDCl3 {C:\Bruker\TopSpin3.5pl7} AK_Koenig 54



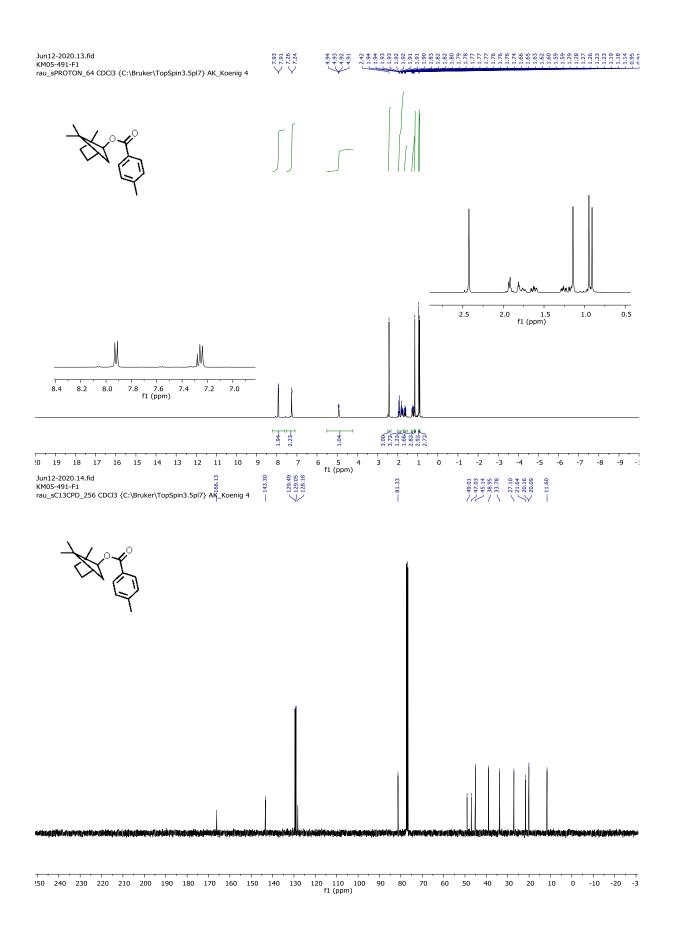


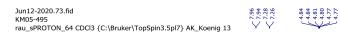


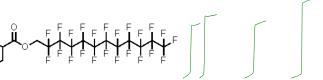
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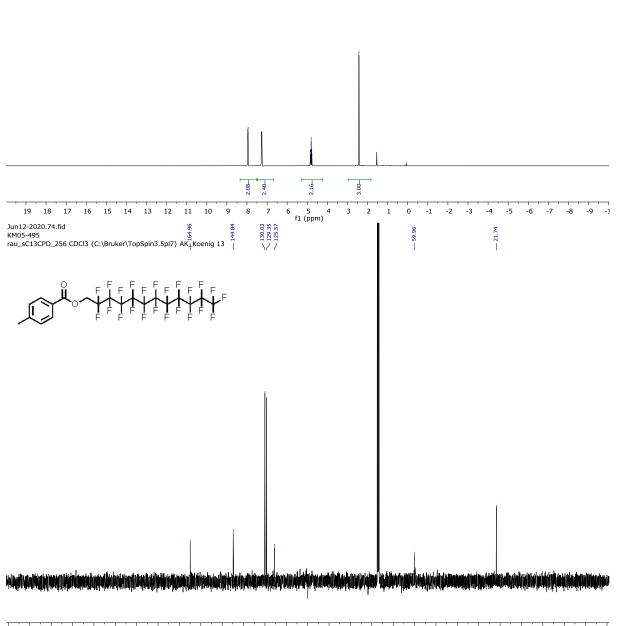


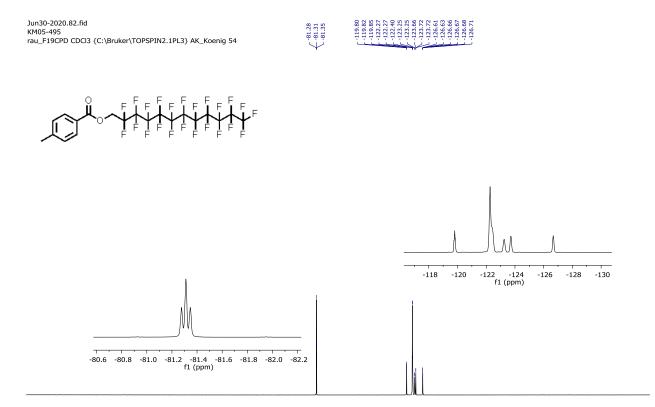
250 240 230 220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 -20 -3 f1 (ppm)



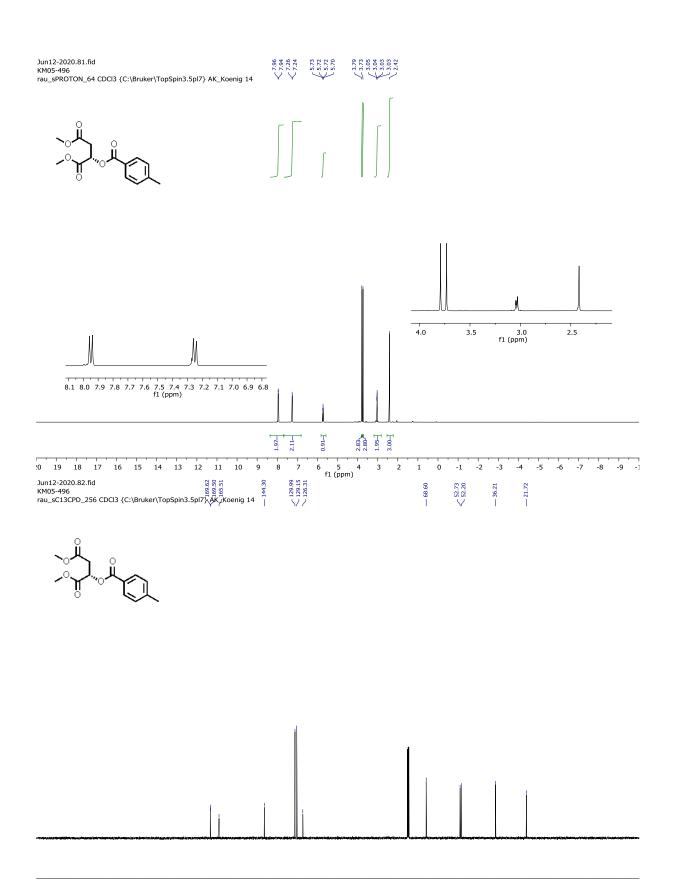


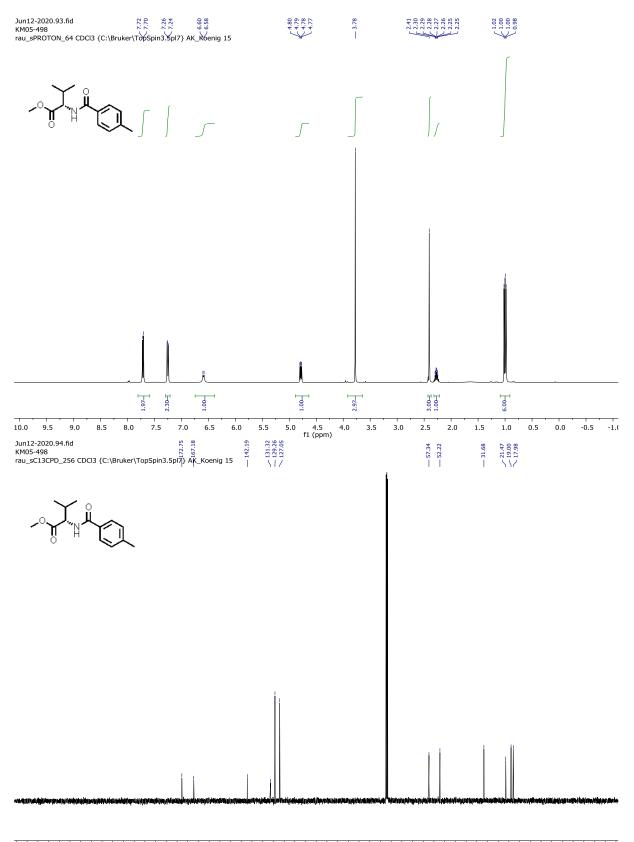


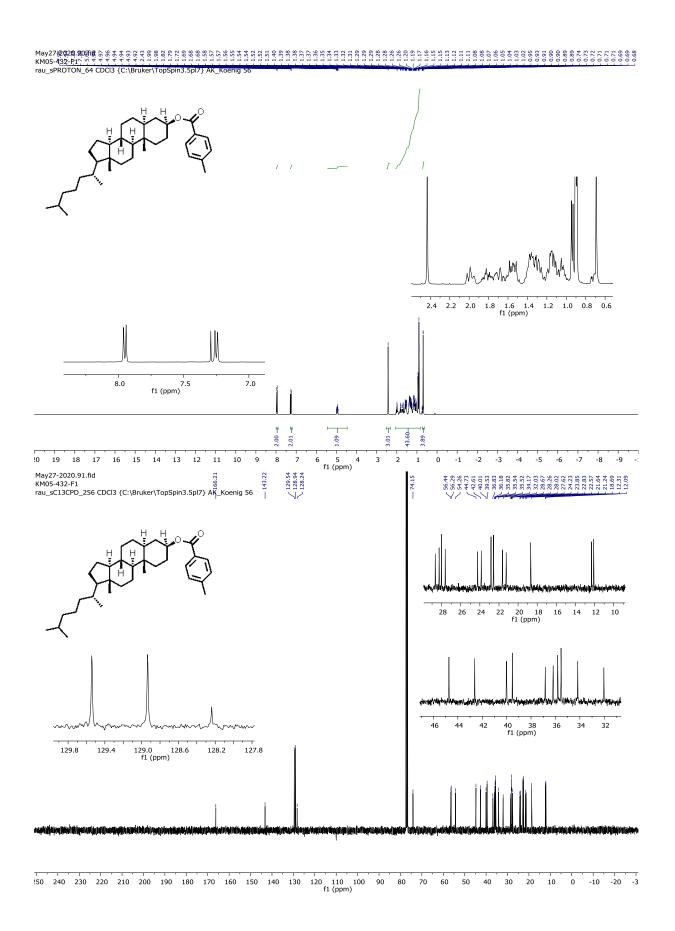




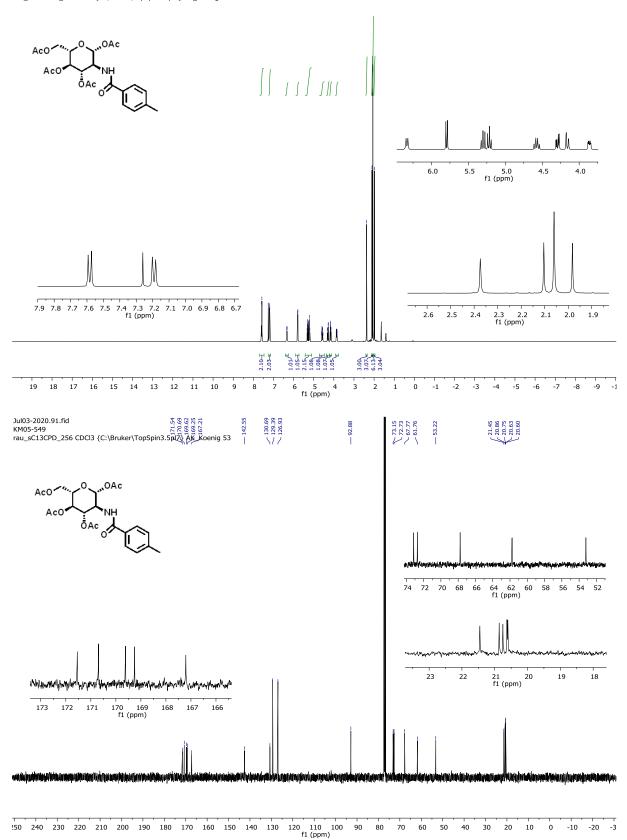
40 30 20 10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 -220 f1 (ppm)

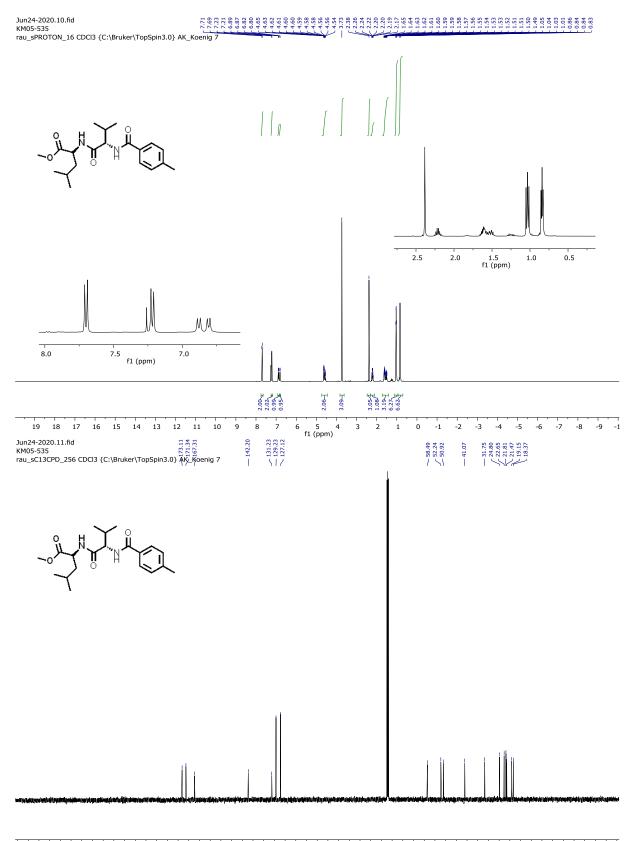




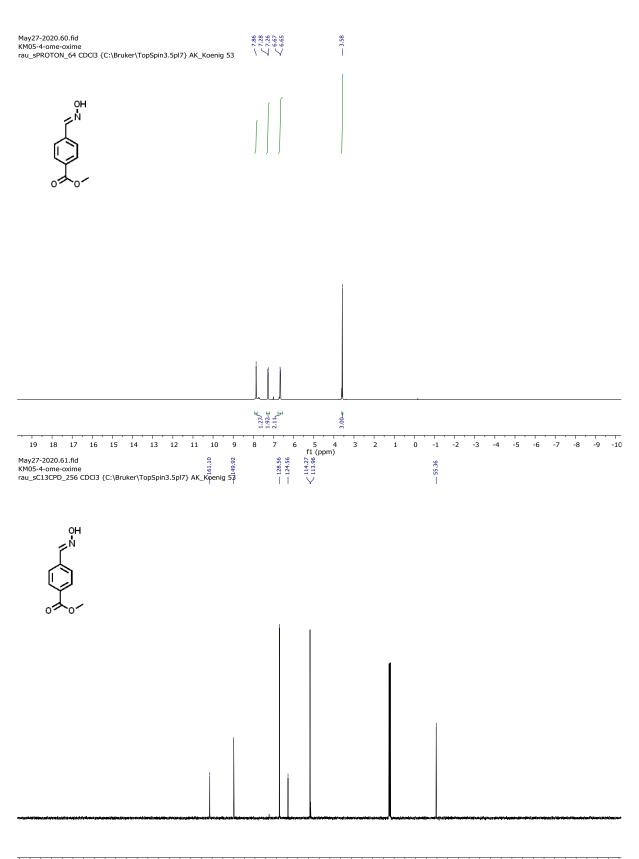


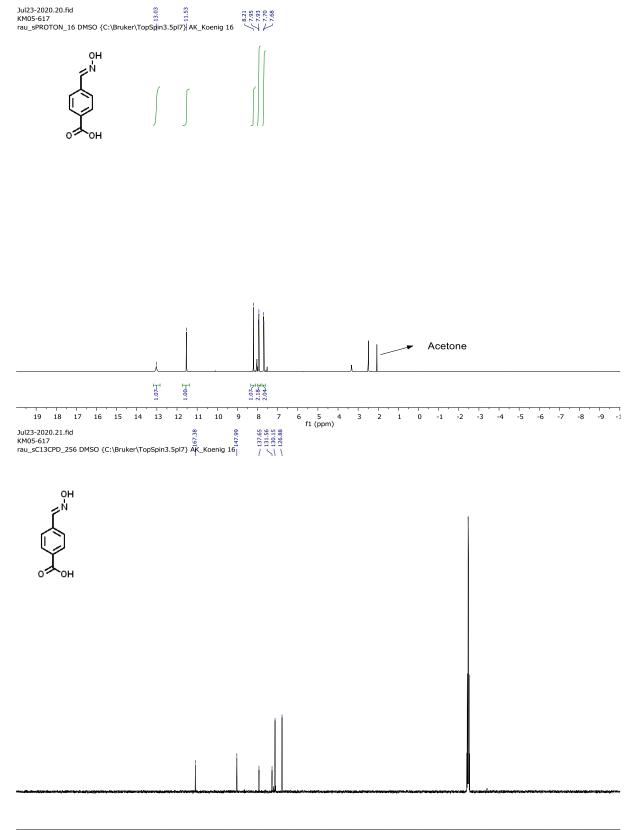
Jul03-2020.90.fid KM05-549 rau_sPROTON_64 CDCl3 {C:\Bruker\TopSpin3.5pl7} AK_Koenig 53 7.55
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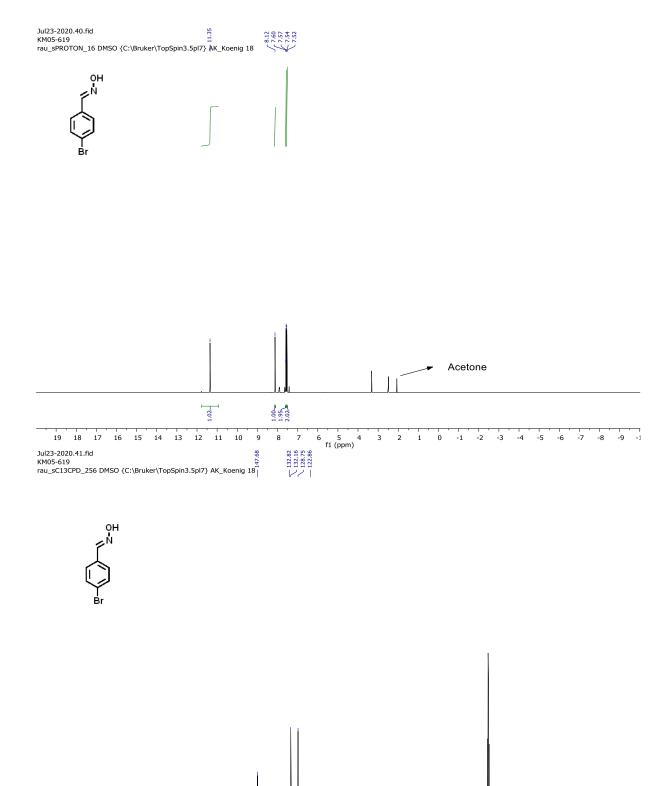


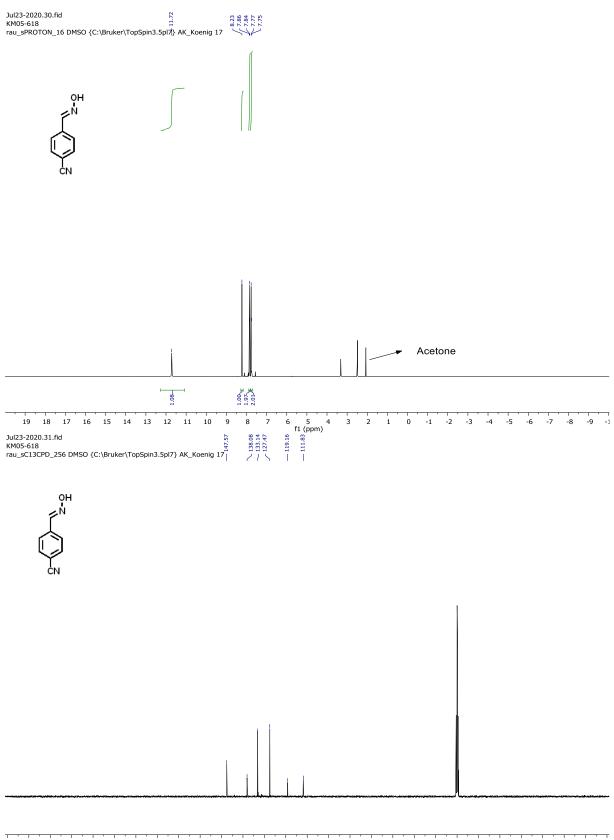


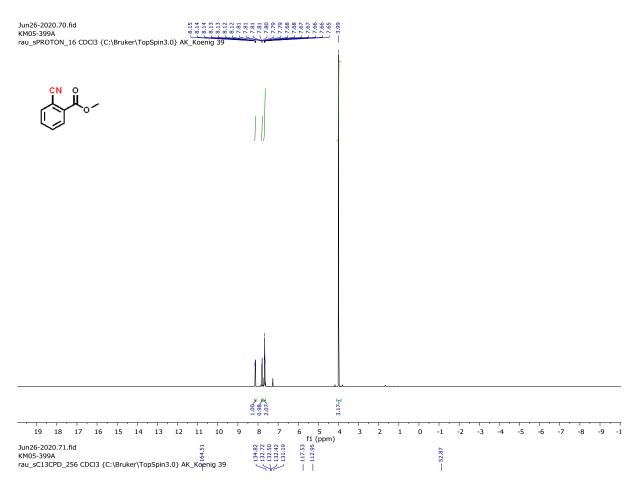
^{250 240 230 220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 -20 -3} f1 (ppm)



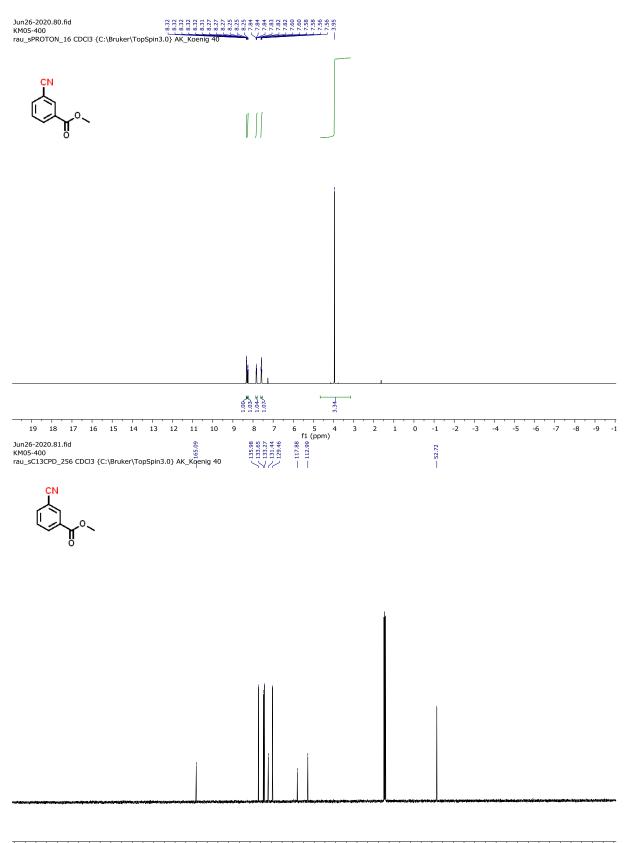


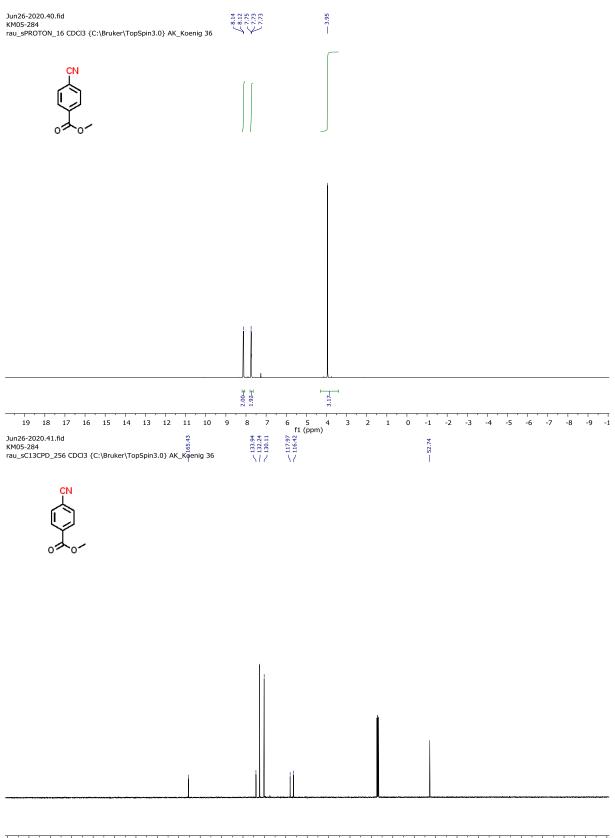


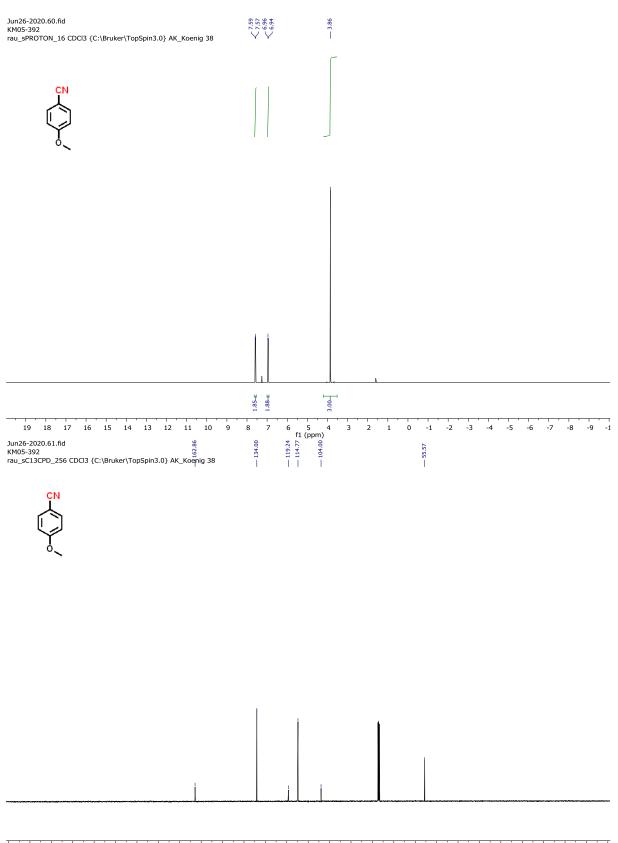


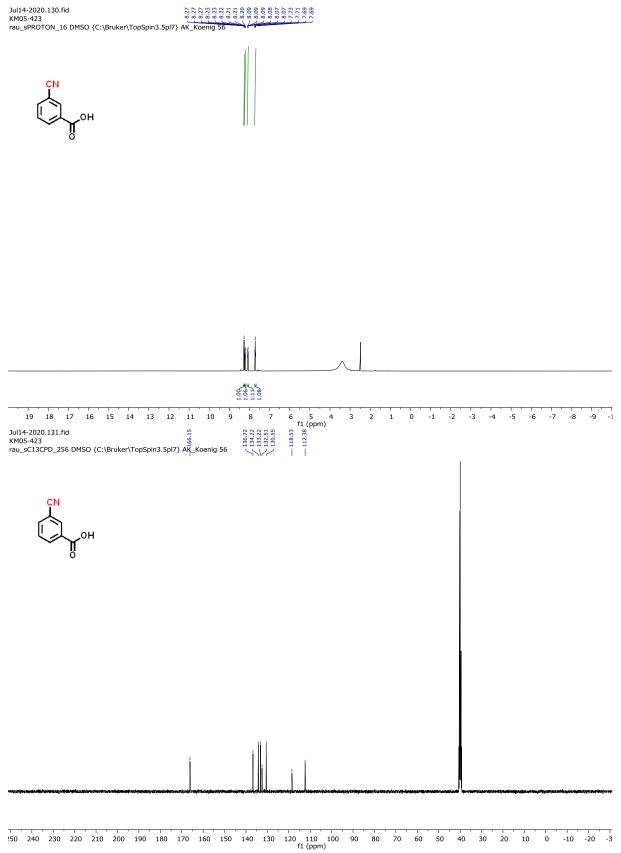


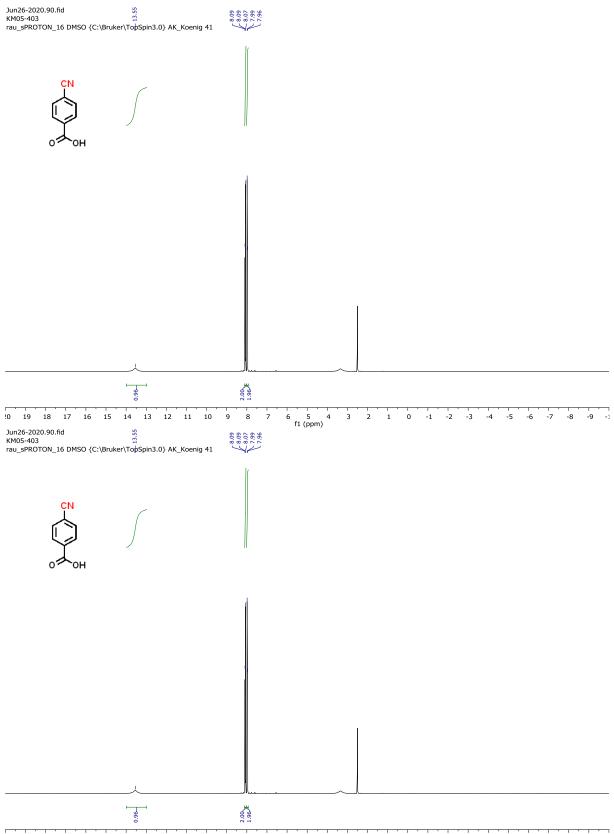
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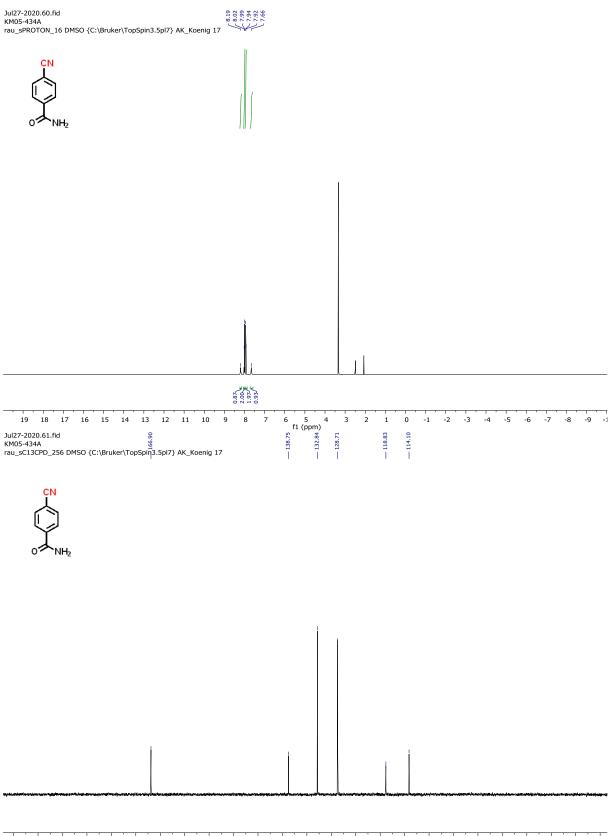








10 19 18 17 16 15 14 13 12 11 10 9 8 7 6 5 4 3 2 1 0 -1 -2 -3 -4 -5 -6 -7 -8 -9 -: f1 (ppm)

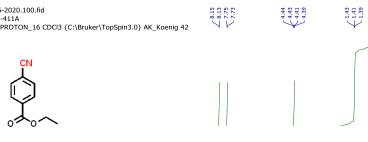


195 190 185 180 175 170 165 160 155 150 145 140 135 130 125 120 115 110 105 100 95 90 85 80 75 f1 (ppm)

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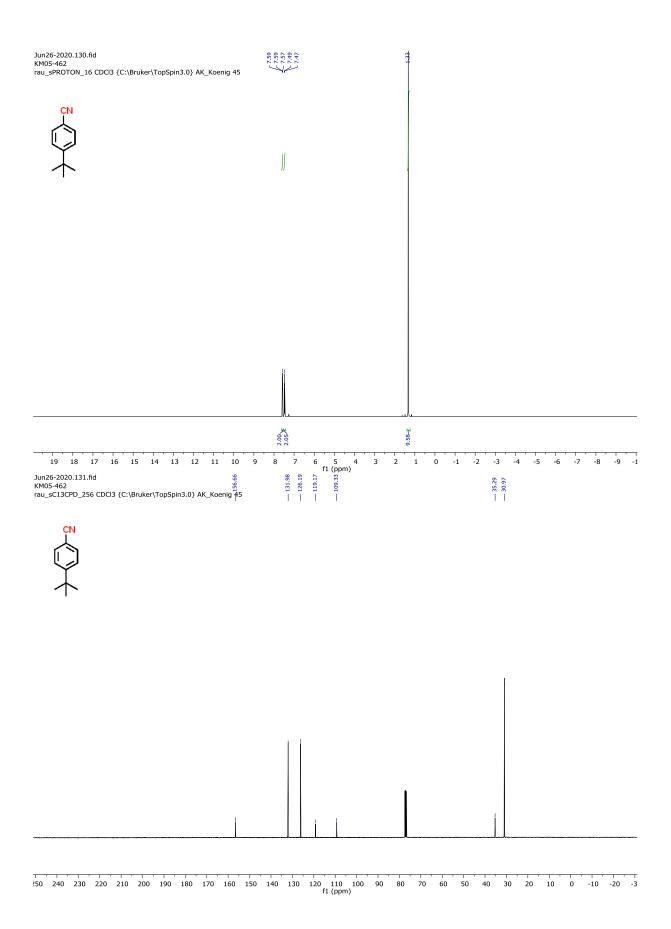
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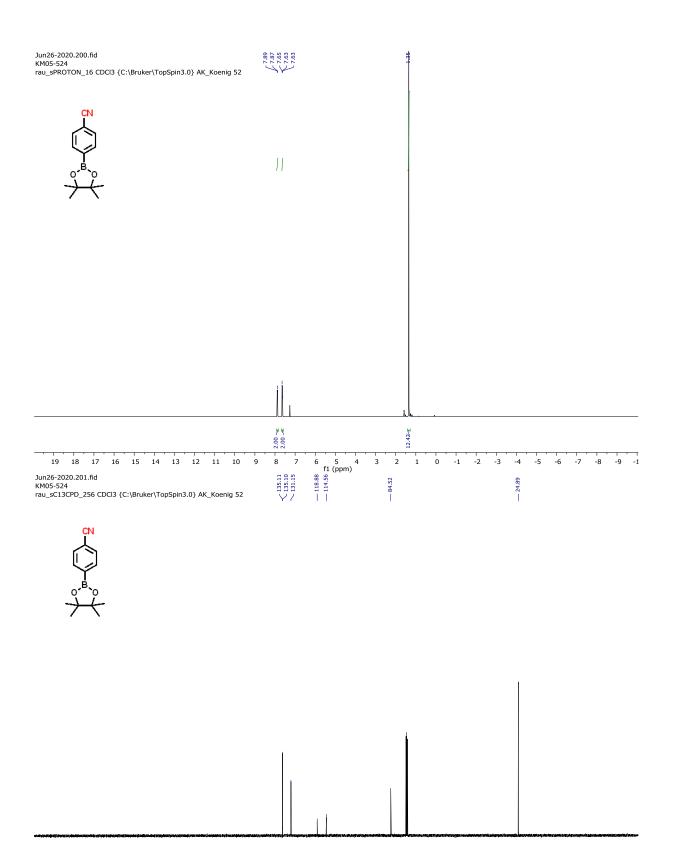
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19 18 17 16 15 14 13 12 11 10	9 8 7 6 5 4 3	2 1 0 -1 -2 -	3 -4 -5 -6 -7 -8 -9 -1
Jun26-2020.101.fid KM05-411A rau_sC13CPD_256 CDCl3 {C:\Bruker\TopSpin3.0} AK_Kqenig 42	f1 (ppm) 617021 617021 617021 1110 617021 1110		— 14.26

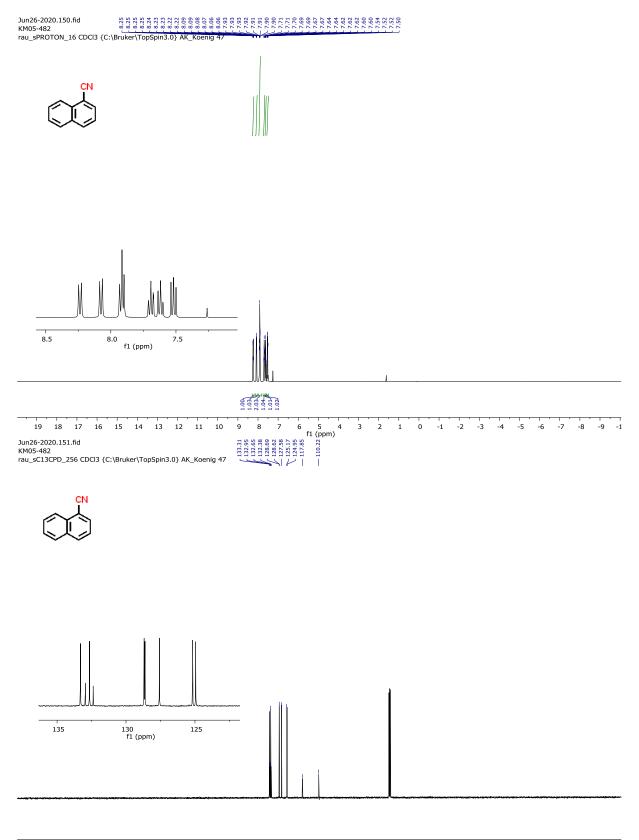


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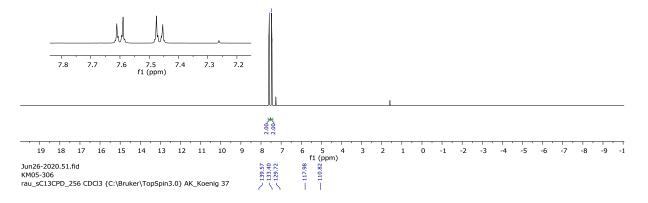




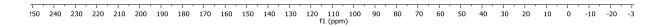






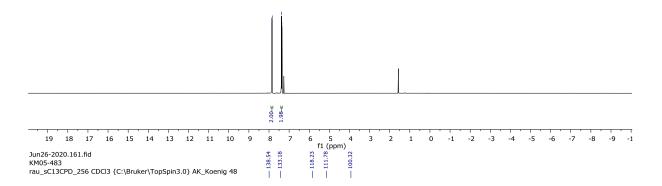




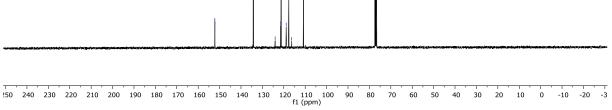


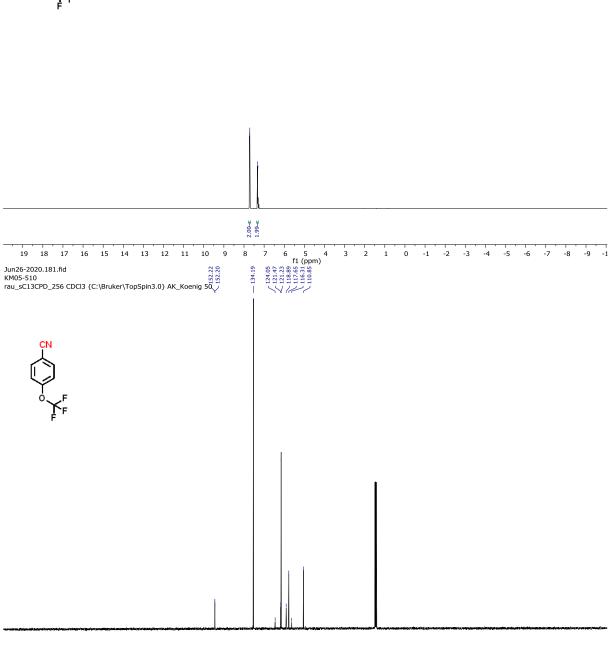
Jun26-2020.160.fid KM05-483 rau_sPROTON_16 CDCl3 {C:\Bruker\TopSpin3.0} AK_Koenig 48











Jun26-2020.180.fid KM059548 (400 MHz, Chloroforat)-ຈັ 7.72 (d,J = 8.9 Hz, 1H), 7.32 (d,= 8.0 Hz, 0H); rau_sPROTON_16 CDCl3 {C:\Bruker\TopSpin3.0} AK_Koenig 50

Jun26-2020.182.fid KM05-510 rau_sF19CPD CDCl3 {C:\Bruker\TopSpin3.0} AK_Koenig 50

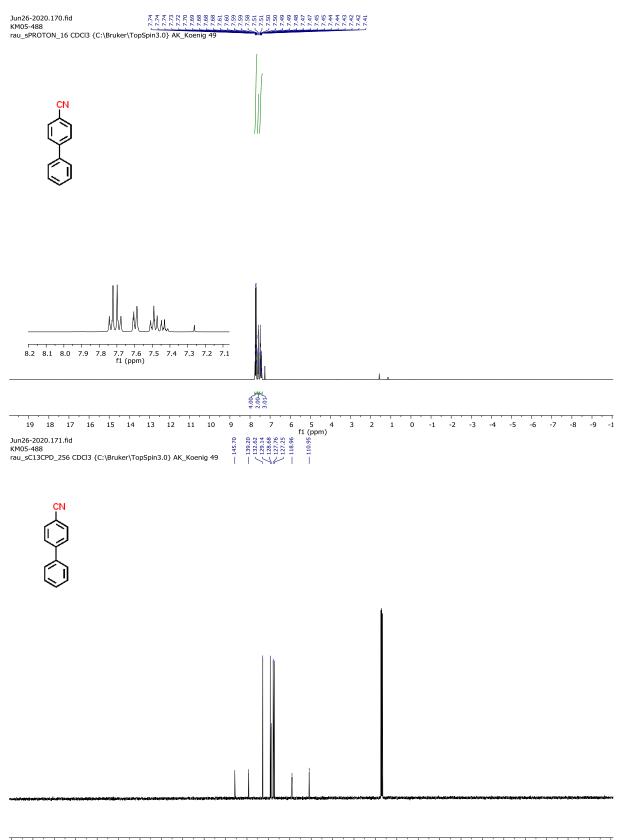


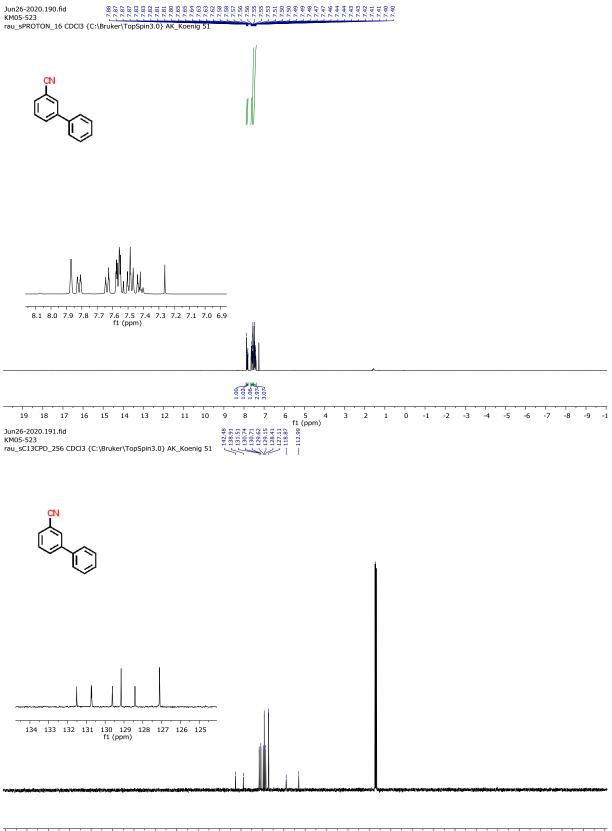
40 30 20 10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 -220 f1 (ppm)

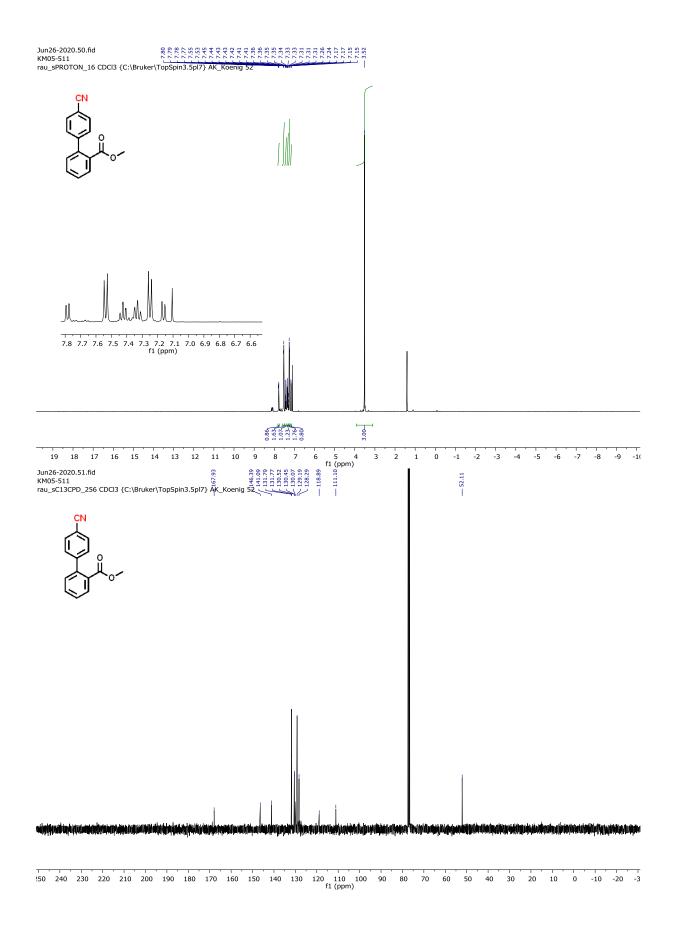
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 $\overbrace{3.93}^{3.98}$

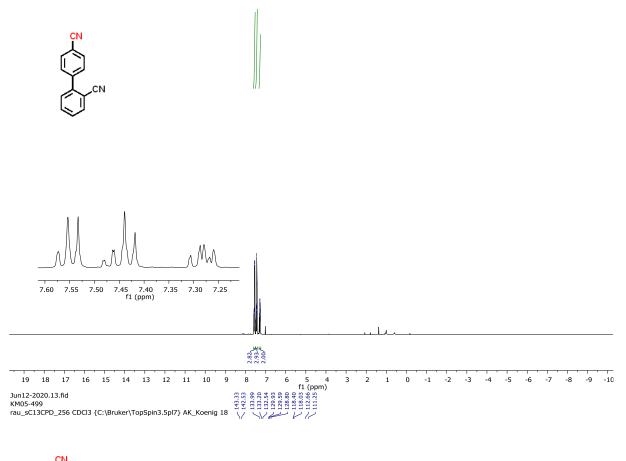
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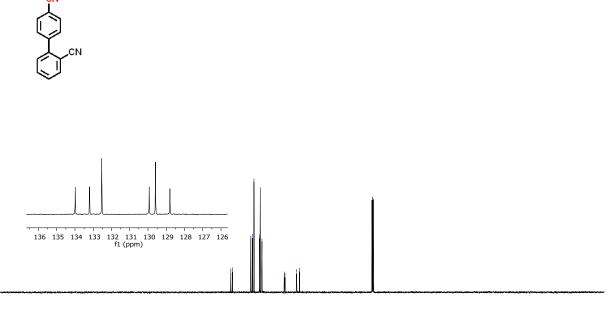


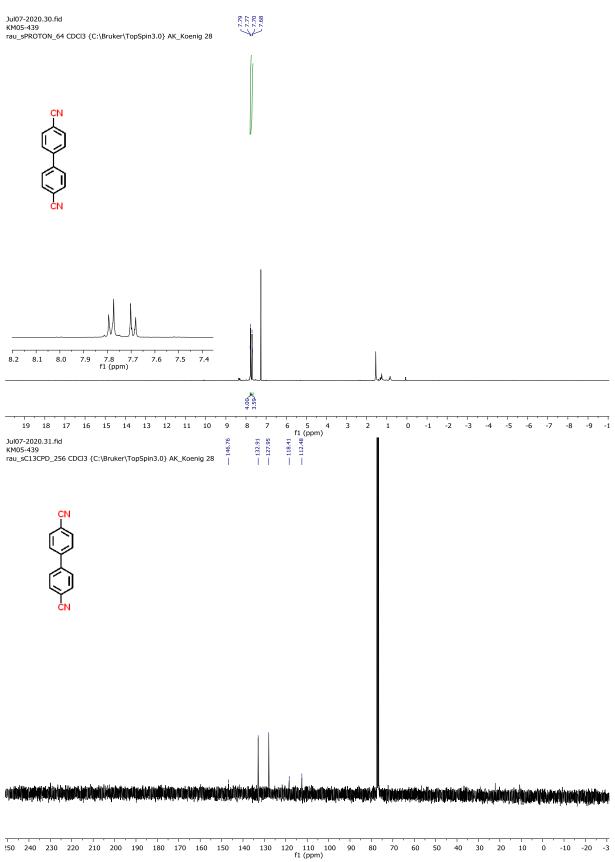


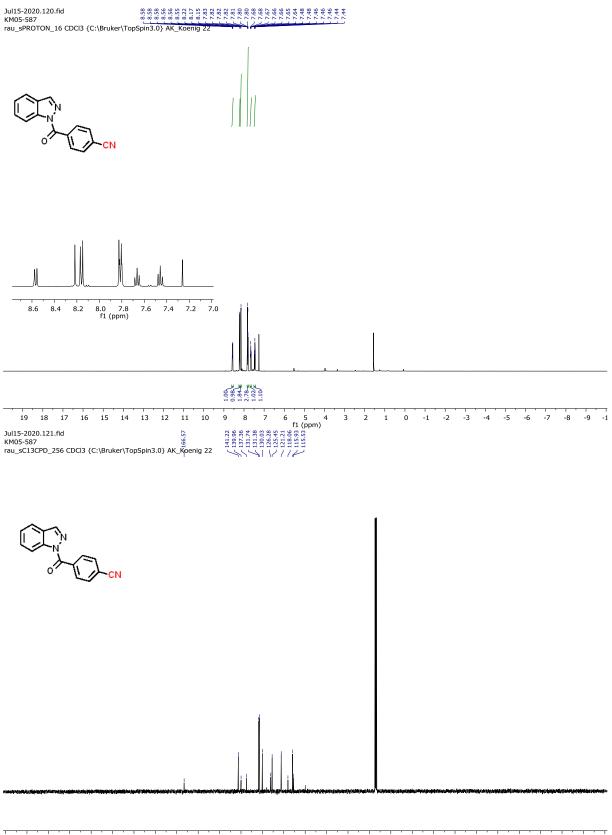


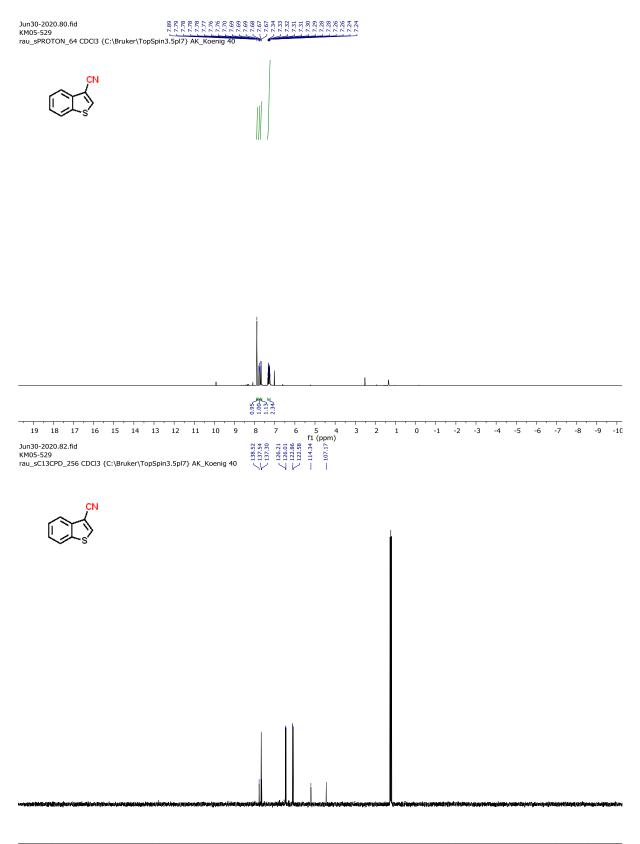


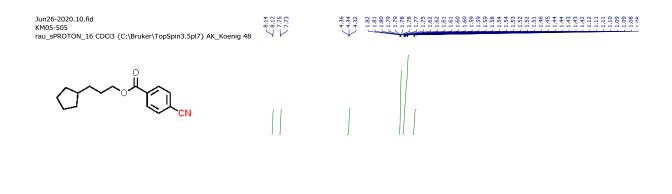


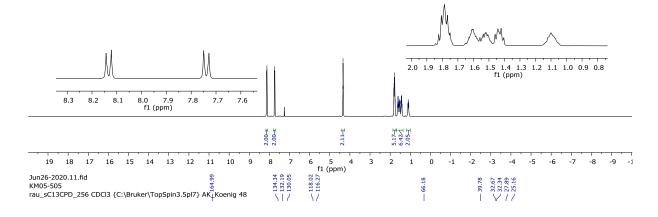


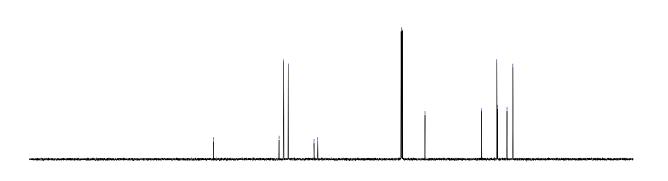


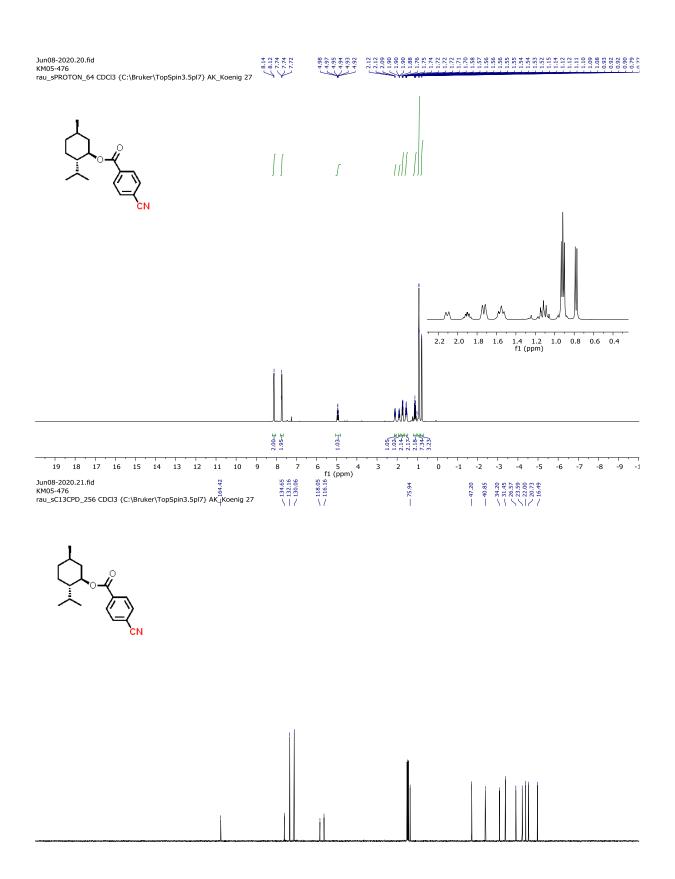


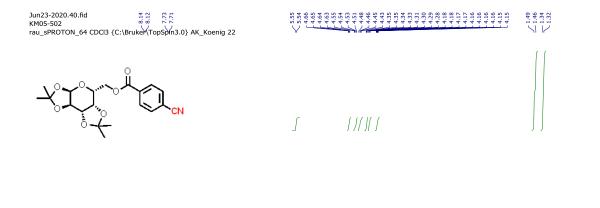


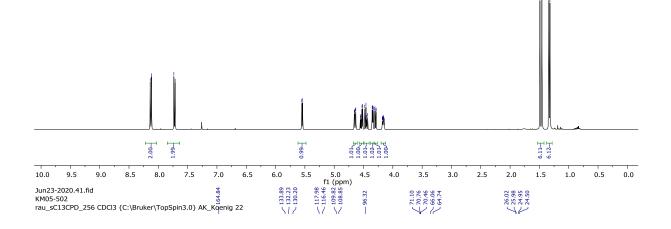




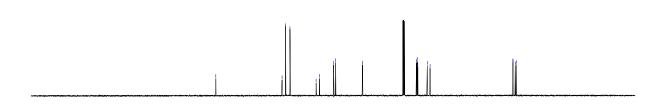


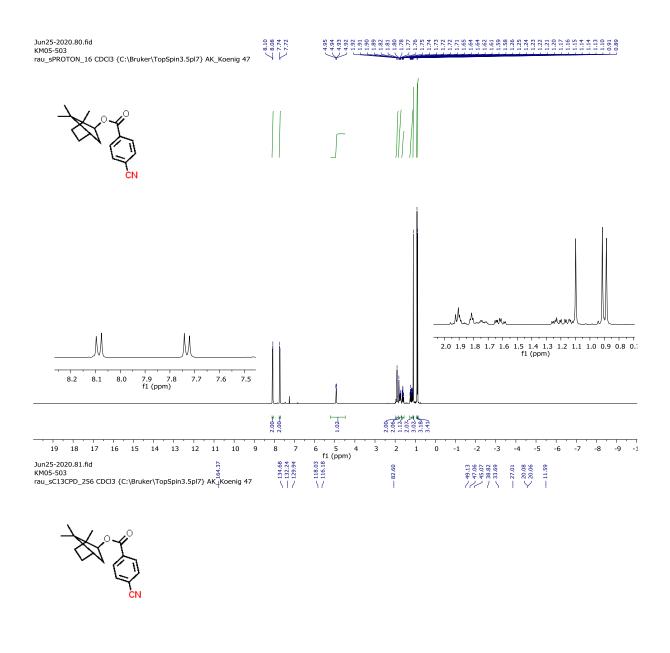


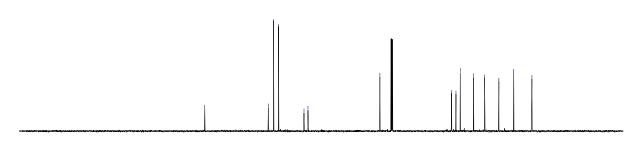


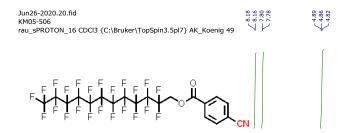


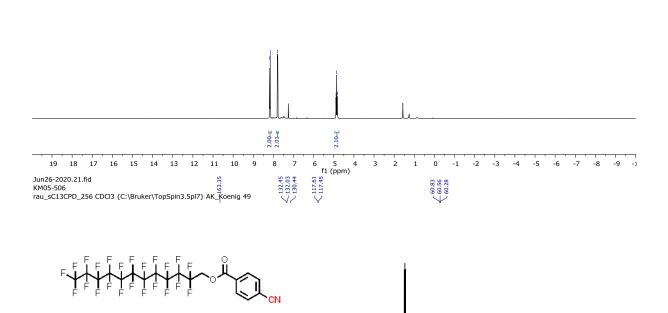
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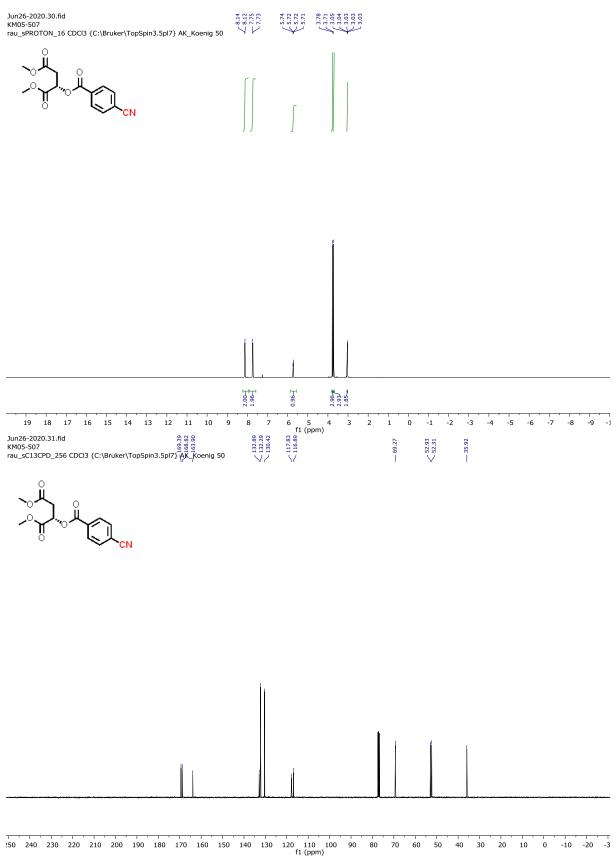


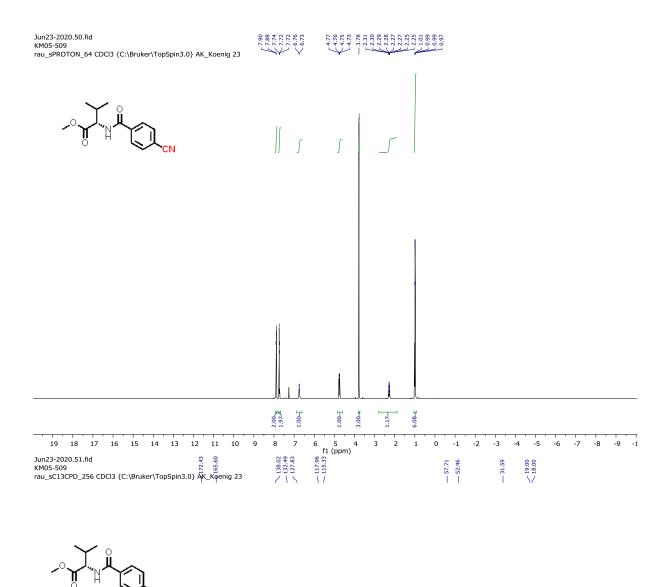


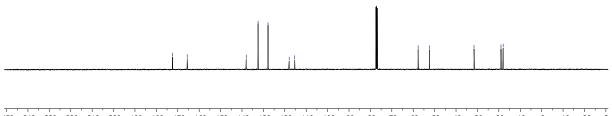


-119.79 -119.83 -119.85 -119.85 -119.85 -122.29 -122.29 -122.29 -122.29 -123.28 -123.28 -123.28 -123.68 -123.68 -123.66 -126.66 -126.66 -126.66 -126.69 -126.67 -126.74 -126.74 -126.77 -126.74 -126.74 -126.74 -126.74 -126.75 -126.69 -126.75 -126.69 -126.75 -126.69 -126.75 -126.66 -126.6 $\underbrace{\left\{\begin{smallmatrix} -81.31 \\ -81.35 \\ -81.38 \end{smallmatrix}\right\}}_{-81.38}$

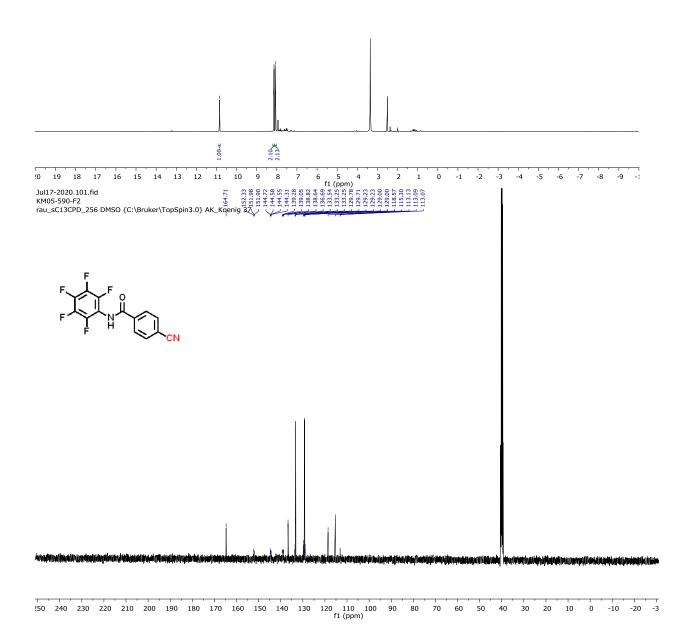
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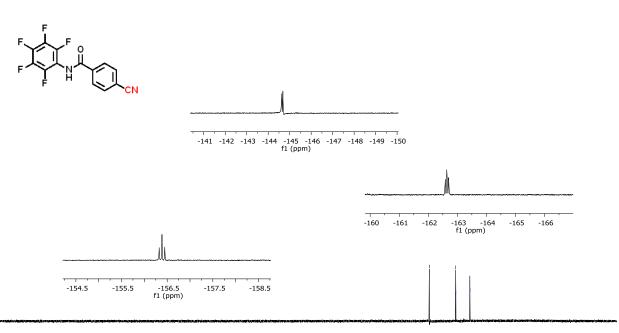




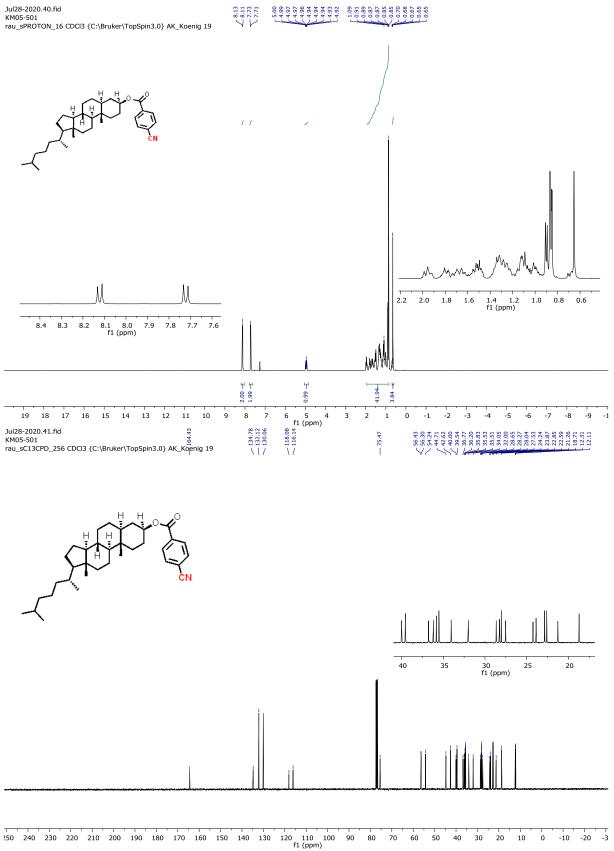


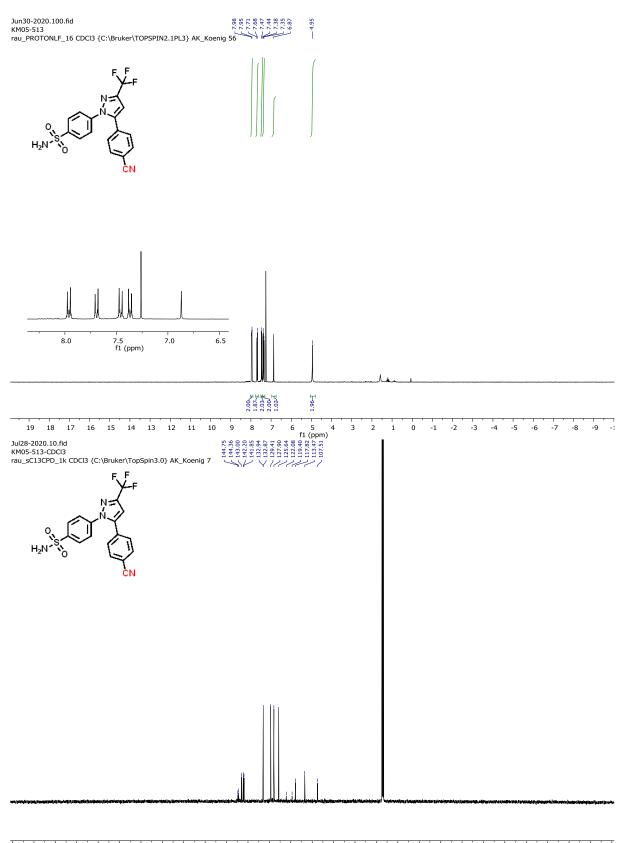


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40 30 20 10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 -220 f1 (ppm)

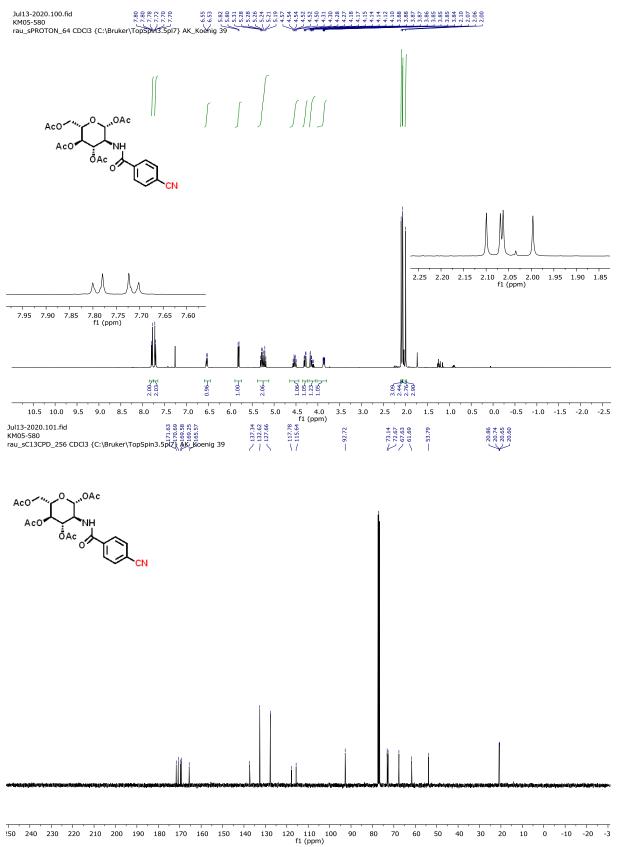


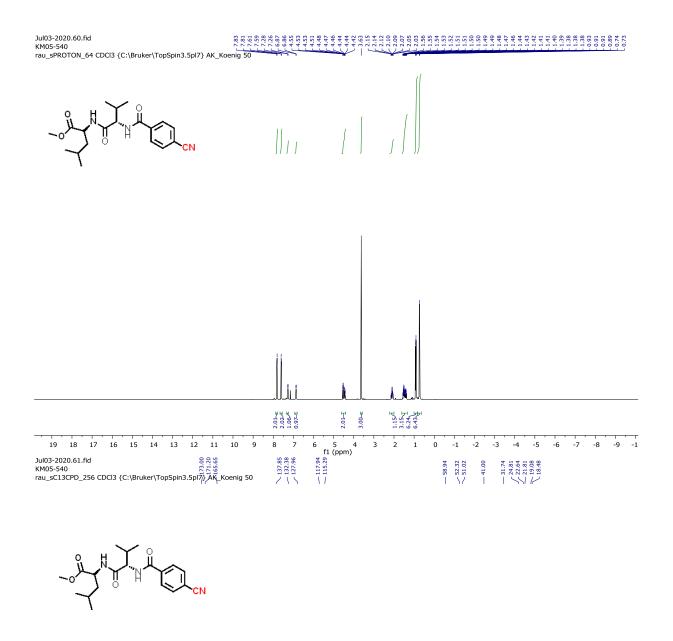


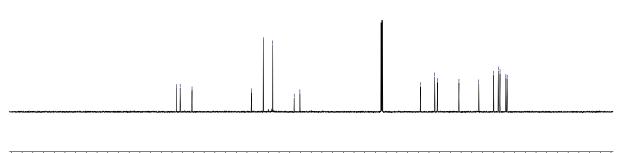
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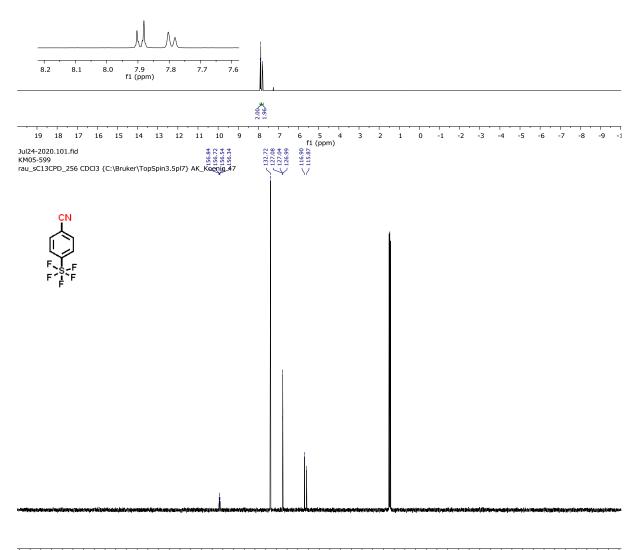


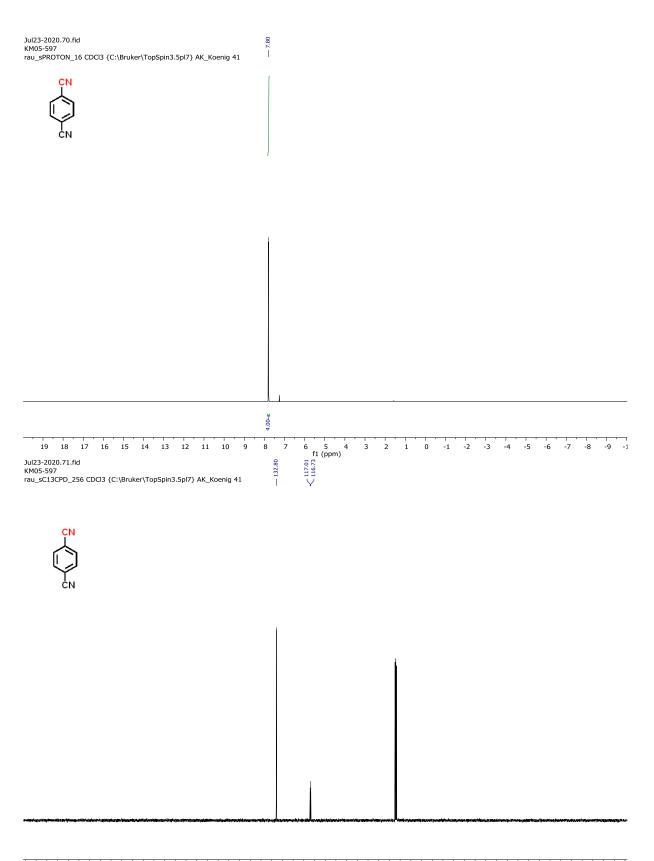






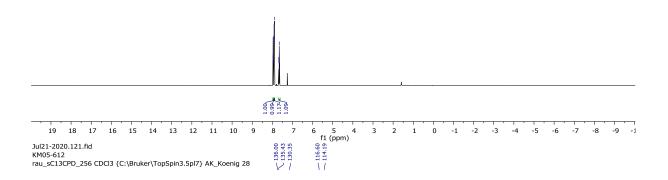






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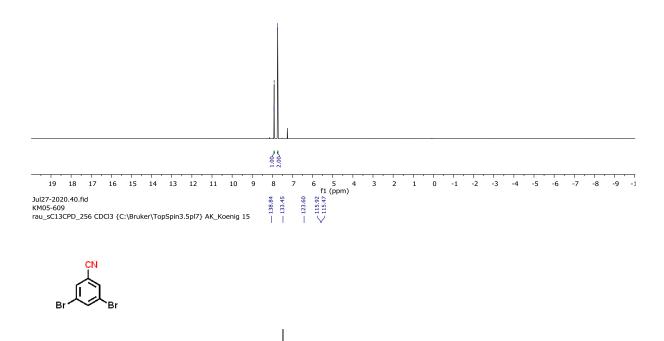


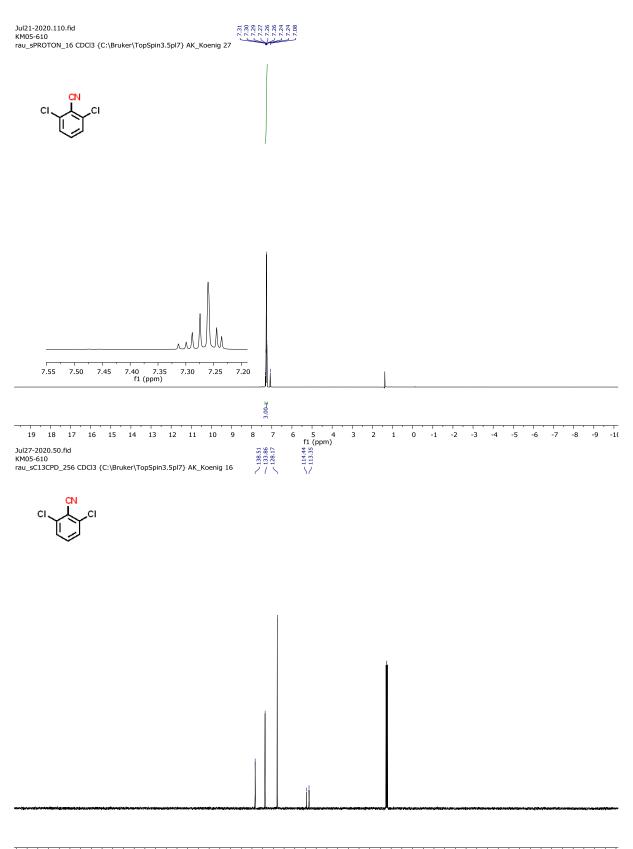




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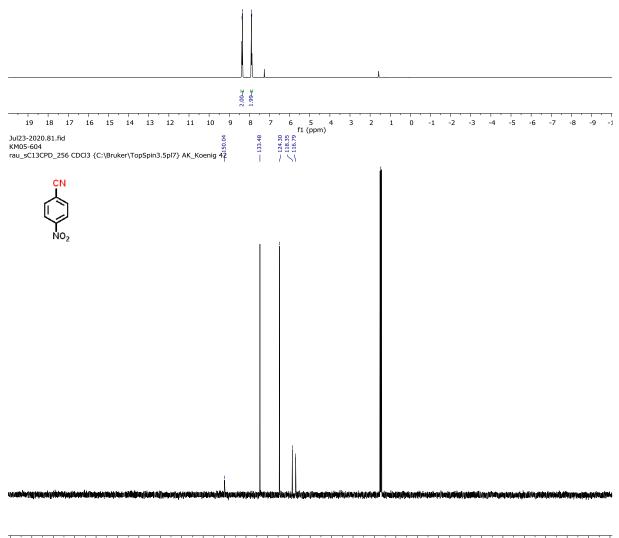












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