



Instruction Sheets

HYGIENE

Hygiene Surface Monitoring Proficiency Testing Scheme

lgcstandards.com/AXIO

Detailed information about samples, matrices, ranges, units, etc. can be found in the [HYGIENE Scheme Description](#)

Issue No: 22

Issued: 18/11/25

Sample ID	Protocol	Page
General Information		3
PT-HY-01	H1V1P-A	4
PT-HY-02	H1V1P-B	5
PT-HY-03	H1V1P-C	6
PT-HY-05	H2T	7
PT-HY-06	H1V1P-C	6
PT-HY-07	H1V1P-C	6
PT-HY-08	H2S	8
PT-HY-09	H2F	9
PT-HY-10	H1T1D-A	10
PT-HY-11	H1T1D-B	11
PT-HY-12	H1V1P-C	6
PT-HY-13	H1SP	12
PT-HY-14	H1SP	12
PT-HY-15	H1SW	13

GENERAL INFORMATION

Storage after receipt

On receipt of the test material, record the date and store as appropriate.

Storage temperature	Samples			
Refrigerated (2 to 8°C)	PT-HY-01	PT-HY-06	PT-HY-10	PT-HY-14
	PT-HY-02	PT-HY-07	PT-HY-11	
	PT-HY-03	PT-HY-08	PT-HY-12	
	PT-HY-05	PT-HY-09	PT-HY-13	

Please note:

- Conditions for storage after receipt of the sample may differ from the conditions under which the PT sample was transported
- Stability data have shown that uncooled transport of samples intended to be stored at 2-8°C does not affect the stability of the test materials for the time of the PT round

The test material(s) should be analysed in accordance with the deadlines shown on the website: <https://portal.lgcstandards.com>

Choice of method or procedure

Participants are expected to use the test method, calibration, or measurement procedure of their choice. This method should be consistent with the participant's normal procedures, for example, duplicate analysis should only be performed if that is part of the routine analytical process. Some samples may require specific preparation or analysis, if so, this is indicated in this document.

Participants may submit results for some, or all the parameters requested.

Precautions

- Microbiological test materials contain viable micro-organisms and are supplied on the understanding that the purchaser has suitably competent and qualified personnel to handle them safely. Test materials must only be opened in a laboratory by qualified personnel.
- Refer to the Safety Data Sheet for information on the safe handling and disposal of the test materials

Reporting Results





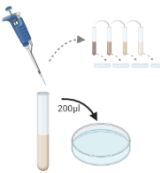




- All results should be submitted using PORTAL
- Please go to <https://portal.lgcstandards.com>
- Login using your Lab ID, username, and password.
- A PORTAL user guide can be downloaded from the help section.

If you need any help at all, please do not hesitate to contact our support team using the details below or your local LGC representative.

Tel: +44(0)161 762 2500



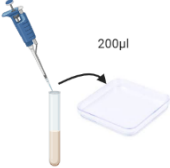

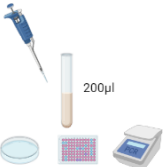



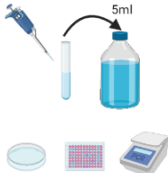
Email: axiopt@lgcgroup.com

Protocol H1V1P-A	Sample codes: PT-HY-01	Description: Square plastic plate + vial	


Step 1 Resuscitate the tablet by adding it to 5ml of suitable microbiological diluent (such as maximum recovery diluent)		Step 2 Leave for 15 to 45 minutes. Mix the sample thoroughly immediately before the next step		Step 3 Remove the top lid of the square plastic plate and discard. Pipette 200µl onto the surface of the plate	
Step 4 Using a sterile spreader, spread the inoculum over the surface of the plate and allow to dry for 15-30mins		Step 5 Analyse 200µl of the inoculum (serial dilutions may be required) and report results in 'direct count' analyte as cfu/200 µl		Step 6 Prepare your chosen volume of diluent	
Step 7 Use a sterile swab to swab the entire surface of the plate		Step 8 Place the swab in the diluent to transfer the microorganisms from the swab and leave 30-60 min at room temperature		Step 9 Enumerate using your usual method. Participants reporting results as per cm ² should treat the plate as 100cm ²	






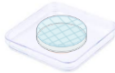
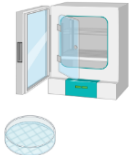
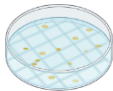
[Back to sample list](#)

Protocol H1V1P-B	Sample codes:	Description:	
	PT-HY-02	Square plastic plate + vial	

Step 1 Resuscitate the tablet by adding it to 5ml of suitable microbiological diluent (such as maximum recovery diluent)		Step 2 Leave for 15 to 45 minutes. Mix the sample thoroughly immediately before the next step		Step 3 Remove the top lid of the square plastic plate and discard. Pipette 200µl onto the surface of the plate	
Step 4 Using a sterile spreader, spread the inoculum over the surface of the plate and allow to dry for 15-30 min		Step 5 Analyse 200µl of the inoculum and report results in 'direct testing' analyte as 'detected/not detected' in 200 µl		Step 6 Prepare 10ml of diluent; this should be non-selective such as maximum recovery diluent	
Step 7 Use a sterile swab to swab the entire surface of the plate		Step 8 Place the swab in the diluent to transfer the microorganisms from the swab and leave 30-60 min at room temperature		Step 9 Transfer 5ml of the primary diluent to the enrichment broth required for each pathogen. Test using your usual method	

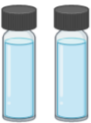


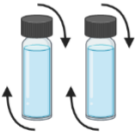


[Back to sample list](#)

Protocol H1V1P-C	Sample codes:			Description: Square plastic plate + vial	
	PT-HY-03 PT-HY-12	PT-HY-06	PT-HY-07		

Step 1 Resuscitate the tablet by adding it to 5ml of suitable microbiological diluent (such as maximum recovery diluent)		Step 2 Leave for 15 to 45 minutes. Mix the sample thoroughly immediately before the next step		Step 3 Remove the top lid of the square plastic plate and discard. Pipette 50µl onto the surface of the plate	
Step 4 Using a sterile spreader, spread the inoculum, in the center of the plate, over an area of 55mm (Ø of a contact plate)		Step 5 Allow to dry for 15-30 min. The sample is now ready for analysis		Step 6 Apply your contact plate once only in the centre of the plate, ensuring that the entire contact plate is in contact with the surface	
Step 7 Incubate the contact plate		Step 8 Enumerate the number of colonies and report results per contact plate or per cm ²			

[Back to sample list](#)


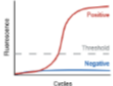
Protocol H2T	Sample codes: PT-HY-05	Description: 2 x tube containing tablet	
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Step 1 Prepare two 10ml aliquots of sterile water in appropriate containers	 2 x 10ml	Step 2 Add the lyophilised tablet labelled 'Sample A' to one of the containers, and 'Sample B' to the other		Step 3 Leave both samples at room temperature for 15 to 45 minutes	 15-45 min resuscitation
Step 4 Immediately before testing invert the samples 5 times		Step 5 Test both samples using your routine method for ATP		Step 6 In PORTAL, select the sample with the highest level of ATP. Also enter the type of equipment used in the comments section	

[Back to sample list](#)

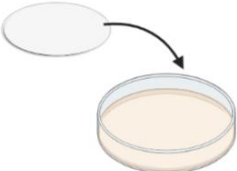
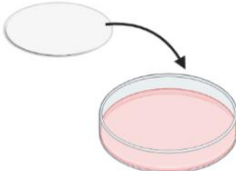
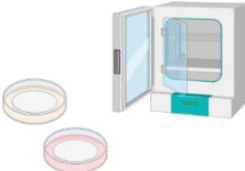
Protocol H2S	Sample codes:	Description:	 OR 
	PT-HY-08	2 x swab rinsate (liquid) OR 2 x dry swab	

Step 1
Process the test materials using your routine testing method following the manufacturer's recommended instructions



[Back to sample list](#)

Protocol H2F	Sample codes: PT-HY-09	Description: 2 x 0.45µm membrane filter in a sterile petri dish	
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Step 1 Place one of the pre-spiked membrane filter on your routine non-selective agar used for total viable count		Step 2 Place the second pre-spiked membrane filter on your routine selective agar used for yeast and mould	
Step 3 Incubate and enumerate according to your routine methods			





[Back to sample list](#)

Protocol H1T1D-A	Sample codes: PT-HY-10	Description: 1 x tube containing tablet and 10ml diluent	
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Step 1 Add lyophilised tablet to 10ml diluent		Step 2 Leave at room temperature for 15-45 minutes	 This represents neat swab rinsate sample
Step 3 Mix thoroughly before processing according to your routine testing methods			




[Back to sample list](#)

Protocol H1T1D-B	Sample codes: PT-HY-11	Description: 1 x tube containing tablet and 10ml diluent	
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Step 1 Add lyophilised tablet to 10ml diluent		Step 2 Leave at room temperature for 15-45 minutes	 This represents neat swab rinsate sample
Step 3 Mix thoroughly before adding 2.5ml to your enrichment broths		Step 4 Test according to your routine methods	




[Back to sample list](#)

Protocol H1SP	Sample codes:		Description: 1 x sponge (ready to test)	
	PT-HY-13	PT-HY-14		

Step 1 Prepare appropriate volume of your routine enrichment media (typically 100ml)		Step 2 Add to the sample and follow your routine enrichment protocol	
Step 3 Test according to the method used in your laboratory			

[Back to sample list](#)

Protocol H1SW	Sample codes: PT-HY-15	Description: 1 x swab (ready to test)	
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Step 1 Prepare appropriate volume of your routine enrichment media (typically 10ml)		Step 2 Add to the sample and follow your routine testing protocol	
Step 3 Test according to the method used in your laboratory			

[Back to sample list](#)