OPEN LETTER

The consistency approach for the substitution of in vivo testing for the quality control of established vaccines: practical considerations and progressive vision [version 2; peer review: 1 approved, 1 approved with reservations]

Jean-François Dierick1, Marlies Halder2, Carmen Jungbaeck3, Julie Lorenz4, Jean-Marie Préaud3, Patrice Riou5, Lorenzo Tesolin6, Sylvie Uhrich5, Wim Van Molle6, Joris Vandeputte3

1GSK Vaccines, 89 Rue de l'Institut, Rixensart, 1330, Belgium
2Joint Research Centre, European Commission, Via E. Fermi 2749, Ispra (VA), 21027, Italy
3IABS-EU, 24 Rue Baldassini, Lyon, 69007, France
4Zoetis Inc., 333 Portage St, Kalamazoo, MI, 49007, USA
5Sanofi Vaccine, 1541 avenue marcel Mérieux, Marcy l'Etoile, 69280, France
6Quality of Vaccines and Blood Products, Sciensano, 14 rue J. Wytsman, Brussels, 1050, Belgium

Open Peer Review

Approval Status

1
2

version 2
(revision)
13 Dec 2022

version 1
05 Oct 2022

1. Elliot Lilley, NC3Rs, London, UK
2. Kirsty Reid, European Federation of Pharmaceutical Industry and Association, Brussels, Belgium
Laura Viviani, Humane Society International, Geneva, Switzerland

Any reports and responses or comments on the article can be found at the end of the article.

Abstract

The aim of this letter is to share the discussions and proposals made by the VAC2VAC consortium on how to support the deployment of the “Consistency Approach” for quality control of established vaccines and thus facilitate the substitution of in vivo testing. This work answers specific questions about “what does a control strategy according to the consistency testing look like” and “how to submit a control strategy defined according to the consistency testing”. Some topics were answered in a very straightforward manner. This was the case when the deployment of the consistency approach and the corresponding changes in vaccines control strategy was supported by the generic application of procedures already described in regulatory guidelines/requirements and related to the establishment or change in the control strategy of vaccines. The application of other procedures required more specific attention and some were deeply debated before reaching a proposal. The key outcomes of this work are that robust science must be used to develop a substitution strategy and generate supportive data packages. And this good science can best occur with good scientific collaboration between the different parties involved. Therefore, early interaction between manufacturers and competent authorities before and during dossier submission is critical to success. The consistency approach, when approved and in place, will ensure vaccine products of assured quality
reach the patient in a more efficient manner than when relying on in vivo testing. Adapting the mindset was one of the major hurdles to a progressive vision but there is now consensus between manufacturers and competent authorities to foster the elimination of in vivo testing for routine vaccine release testing.

**Keywords**
consistency testing, vaccines, in vitro, in vivo, validation, analytical method

This article is included in the Microbiology gateway.

**Corresponding author:** Jean-Francois Dierick (jean-francois.m.dierick@gsk.com)

**Author roles:** Dierick JF: Conceptualization, Writing – Original Draft Preparation, Writing – Review & Editing; Halder M: Conceptualization, Writing – Original Draft Preparation, Writing – Review & Editing; Jungbaeck C: Conceptualization, Writing – Review & Editing; Lorenz J: Conceptualization, Writing – Review & Editing; Préaud JM: Conceptualization, Writing – Review & Editing; Riou P: Conceptualization, Writing – Review & Editing; Tesolin L: Conceptualization, Writing – Original Draft Preparation, Writing – Review & Editing; Ulrich S: Conceptualization, Writing – Review & Editing; Van Molle W: Conceptualization, Writing – Review & Editing; Vandeputte J: Conceptualization, Writing – Review & Editing

**Competing interests:** No competing interests were disclosed for Marlies Halder, Carmen Jungbaeck, Julie Lorenz, Jean-Marie Préaud, Lorenzo Tesolin, Wim Van Molle and Joris Vandeputte. Jean-Francois Dierick is an employee of the GSK group of companies. Patrice Riou and Sylvie Ulrich are Sanofi employees and may hold shares and/or stock options in the company.

**Grant information:** This project was financially supported by the European Union's Horizon 2020 research and innovation program and the Innovative Medicines Initiative 2 Joint Undertaking under grant agreement No [115924] (Vaccine lot to Vaccine lot comparison by consistency testing [VAC2VAC]). Financial support was also provided by the EFPIA.

**Copyright:** © 2022 Dierick JF et al. This is an open access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

**How to cite this article:** Dierick JF, Halder M, Jungbaeck C et al. The consistency approach for the substitution of in vivo testing for the quality control of established vaccines: practical considerations and progressive vision [version 2; peer review: 1 approved, 1 approved with reservations] Open Research Europe 2022, 2:116 https://doi.org/10.12688/openreseurope.15077.2

**First published:** 05 Oct 2022, 2:116 https://doi.org/10.12688/openreseurope.15077.1
Plain language summary

VAC2VAC is a collaborative research project funded by IMI2, which aims to develop approaches for the testing of vaccines (human or animal) using non-animal methods. In that perspective, the authors launched a reflection on how to support the deployment of the “Consistency Approach” which had been defined in order to facilitate and accelerate the substitution of current in vivo methods with in vitro alternatives for quality control of vaccines.

The key outcomes of this work are that robust science must be used to develop a substitution strategy and produce supportive data packages. And, this good science will be more efficient when supported by scientific collaboration between the vaccine manufacturers and competent authorities before and during dossier submission are key elements to success.

Disclaimer

The views expressed in this article are those of the authors. Publication in Open Research Europe does not imply endorsement of the European Commission. The views expressed by the regulators who have contributed to this article do not per se represent the views of the organizations they belong to.

Introduction

VAC2VAC is a wide-ranging collaborative research project funded by IMI2 (2016–2022), which aims to develop and validate quality testing approaches (physicochemical, immunochemical methods, cell-based and multi-parametric) for routine batch quality, safety and efficacy testing of human and veterinary vaccines using non-animal methods. More information about the VAC2VAC program can be found on the Cordis database from The European Commission: Vaccine lot to Vaccine lot comparison by consistency testing. | VAC2VAC Project | Fact Sheet | H2020 | CORDIS | European Commission (europa.eu), and https://www.imi.europa.eu/projects-results/project-factsheets/vac2vac. As part of the project, an initiative was launched to canvas opinion and understand best options to support the deployment of the “Consistency Approach” for quality control of established vaccines, where current approaches often rely on in vivo methods.

Discussions started between manufacturers and competent authorities, involved in the VAC2VAC project, on what does a control strategy according to consistency testing entail and how to submit a control strategy defined according to the consistency testing. Then the consortium organized two workshops (on June 22, 2020 and June 24, 2021) inviting VAC2VAC experts and experts not involved in the VAC2VAC consortium and who represent important stakeholders into the discussions (academics, National Control Laboratories, regulators, governmental and trans-governmental organizations). The goal was to address all topics (simple or complex) that could be related to the change of control strategy and facilitate a common view from regulators and industries in anticipation of future submission of a method variation dossier or a marketing authorization application. Consensus was reached, and proposals were made on several topics. The aim of this document is to share these proposals to a larger audience and stimulate discussion and engagement toward the consistency approach for the substitution of in vivo testing for quality control of established vaccines.

This paper summarizes the discussions and proposals made by the VAC2VAC consortium composed of experts, regulators and scientists from different institutions, manufacturers and competent authorities on how to support the deployment of the “Consistency Approach” for quality control of established vaccines. Table 1 summarizes the current regulatory framework taken into consideration during these discussions.

The consistency approach can replace the current in vivo testing, known to be laborious and highly variable for many established vaccines, with in vitro assays now seen as the Golden standard (see Stalpers et al., 2021). Robust science and early interaction between manufacturers and competent authorities before and during dossier submission are certainly key elements to success. This consistency approach, when approved and in place, will allow new and established products of assured quality to reach the patient in a more efficient manner.

Science, newly developed methods and relevant guidelines are available to support the move to a Consistency Testing-based control strategy (see Akkermans et al., 2020; Schutte et al., 2017 and Viviani et al., 2022). Adapting the mindset was one of the major hurdles to a progressive vision but there is now a consensus between manufacturers and competent authorities to foster the elimination of in vivo assays for routine vaccinerelease testing.

The consistency approach

According to de Mattia et al. (2011): “The consistency approach implies the use of a set of parameters to constitute a product profile (e.g., antigen content, antigen integrity, etc.) that can replace current release tests. The product profile is established to the satisfaction of the regulators at the time of licensing, and is monitored throughout production under a strict quality system. The product profile ensures that each batch or lot released is similar to a manufacturer-specific vaccine
### Table 1.

#### RELEVANT EU-PROVISIONS

<table>
<thead>
<tr>
<th>(Veterinary) International Conference on Harmonisation (V)ICH</th>
</tr>
</thead>
<tbody>
<tr>
<td>ICH Q6B &quot;Specifications: Test Procedures and Acceptance Criteria for Biotechnological/Biological Products&quot; for setting specifications limits.</td>
</tr>
<tr>
<td>ICH Q2 (R1) &quot;Validation of analytical procedures&quot;</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>European Union</th>
</tr>
</thead>
</table>

<table>
<thead>
<tr>
<th>European Medicines Agency (EMA)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Guideline on the principles of regulatory acceptance of 3Rs (replacement, reduction, refinement) testing approaches (EMA/CHMP/CVMP/JEG-3Rs/450091/2012)</td>
</tr>
<tr>
<td>CMDh Recommendation for classification of unforeseen variations according to Article 5 of Commission Regulation (EC) 1234/2008 <a href="https://www.hma.eu/fileadmin/dateien/Human_Medicines/CMD_h_/procedural_guidance/Variations/Art_5_Recommendations/CMDh_172_2010_09_2019_Tracking_Table_Article_5.xls">https://www.hma.eu/fileadmin/dateien/Human_Medicines/CMD_h_/procedural_guidance/Variations/Art_5_Recommendations/CMDh_172_2010_09_2019_Tracking_Table_Article_5.xls</a></td>
</tr>
<tr>
<td>CMDv Recommendation for classification of unforeseen variations according to Article 5 of Commission Regulation (EC) 1234/2008 <a href="https://www.hma.eu/fileadmin/dateien/Human_Medicines/CMD_h_/procedural_guidance/Variations/Art_5_Recommendations/CMDh_172_2010_09_2019_Tracking_Table_Article_5.xls">https://www.hma.eu/fileadmin/dateien/Human_Medicines/CMD_h_/procedural_guidance/Variations/Art_5_Recommendations/CMDh_172_2010_09_2019_Tracking_Table_Article_5.xls</a></td>
</tr>
<tr>
<td>Reflection paper providing an overview of the current regulatory testing requirements for veterinary medicinal products and opportunities for implementation of the 3Rs (EMA/CHMP/CVMP/3Rs/164002/2016)</td>
</tr>
<tr>
<td>Guidance for individual laboratories for transfer of quality control methods validated in collaborative trials with a view to implementing 3Rs (EMA/CHMP/CVMP/3Rs/94436/2014)</td>
</tr>
<tr>
<td>Recommendation to MAHs, highlighting the need to ensure compliance with 3Rs methods described in the Ph. Eur. – applicable to all medicinal products regardless of type. (EMA/CHMP/CVMP/JEG-3Rs/252137/2012)</td>
</tr>
<tr>
<td>Recommendation to MAHs, highlighting recent measures in the veterinary field to promote 3Rs measures described in the Ph. Eur. – applicable to veterinary vaccines from 01/01/2017.(EMA/CHMP/CVMP/3Rs/336802/2017)</td>
</tr>
<tr>
<td>Statement of the CVMP position on the ethical use of animals in the testing, development and manufacture of veterinary medicines (EMA/CVMP/3Rs/506841/2017)</td>
</tr>
<tr>
<td>EU-Regulation 2019/6 Draft Annex 2</td>
</tr>
<tr>
<td>SECTION IIIb REQUIREMENTS FOR IMMUNOLOGICAL VETERINARY MEDICINAL PRODUCTS</td>
</tr>
<tr>
<td>IIIb.2D. Control tests during the manufacturing process</td>
</tr>
<tr>
<td>IIIb.2E. Control tests on the finished product</td>
</tr>
</tbody>
</table>

#### RELEVANT US-PROVISIONS

<table>
<thead>
<tr>
<th>USDA Veterinary Services Memorandum No. 800.112, Guidelines for Validation of In Vitro Potency Assays, April 10, 2015.</th>
</tr>
</thead>
</table>
of proven clinical efficacy and safety, with respect to all characteristics agreed upon at licensing between manufacturer and regulator.”

The VAC2VAC consortium proposes the following update of definition from de Mattia et al. (2011) to reflect current developments: “The consistency approach implies the use of a (set of) parameter(s) to constitute an integrated control strategy preferably relying on non-animal testing (e.g. antigen content, antigen integrity, etc.) and taking into account production controls (e.g. defined ranges, harvest criteria etc.). Taken together it can be seen as an approach which can replace the current approach for many established vaccines which involves in vivo testing. The new integrated control strategy has to be established to the satisfaction of the regulators and is monitored throughout production under a strict quality system. The integrated control strategy ensures that each batch or lot released is similar to a manufacturer-specific vaccine of proven clinical efficacy and safety, with respect to all characteristics agreed upon at licensing between manufacturer and regulator.

If available, comparison to the batches used in the clinical studies / challenge studies / field trials to support the marketing authorization application may be an option. For older vaccines, parallel testing of batches may be the better way. The number of batches to be tested in parallel should be reasonable, depending on the number of batches normally manufactured in one year (blockbuster versus orphan products / MUMS).

Proposals on the use of the consistency approach

The following paragraphs describe the proposals emerging from the work from the VAC2VAC consortium on the use of the consistency approach for the substitution of in vivo testing for the quality control of established vaccines.

The simple straightforward topics

Some very simple straightforward conclusions will be shared in this first part in order to express clearly what does not need to change in order to apply the consistency approach.

First, it is acknowledged that a Consistency Testing-based control strategy can be defined using currently existing concepts and terms (Critical Quality Attribute, In Process Control, release assays, characterization assays, monitoring assays, …).

For instance, the Critical Quality Attribute (CQA) “potency/safety” might be replaced by several CQAs that are relevant for the biological activity/safety of a given vaccine. Process Control might be considered as part of the Consistency Testing-based control strategy if relevant and necessary in combination with Critical Quality Attributes for the Drug Product (final product).

“Relevant In Process Controls” refer to control tests that have been shown to ensure that a given component of the Critical Quality Attribute “potency/safety” (example: conformation) is maintained when the Critical Process Parameter (CPP) is met throughout the production step.

In all these cases, currently existing concepts and terms are sufficient to define and clearly explain a control strategy that is based on the Consistency Approach.

Some common processes and practices are also fully applicable without any specific attention: Analytical Method Validation and Certificates of Analysis. The current provisions on validation should be applied as required under the current control strategy. Results derived with new methods must be included in the Certificate of Analysis when applying (or not) the Consistency Approach. Critical Process Parameters used in the context of Consistency Testing should not be included in the Certificate of Analysis (i.e., no change compared to the current situation). In Process Control assays might be used for release on a Consistency Testing-based control strategy (for instance, when they cannot be performed at the final stage due to product/matrix changes). In that case, In Process Control results may appear on the Certificate of Analysis.

In these discussions, we considered whether new method(s) developed in the context of the deployment of a Consistency Testing-based control strategy could become compendial? Any new method, e.g., for potency testing, can become compendial (e.g., become part of a monograph from the European Pharmacopeia (Ph.Eur.), initiated by a request for revision). It is understood that new technologies/methods should ideally be platform technologies and/or product group specific and/or product or manufacturer specific. The rationale for a method to become compendial is for harmonization purposes. An international reference can be developed, and a monograph can be published using common units and specifications. It is important to note that having a method recognized as compendial will promote its usage. Therefore, the introduction of methods that are alternatives to in vivo methods in the compendia is an important lever towards the reduction of animal use in vaccine quality control.

Some other common processes and practices require specific attention and are presented hereunder.

Product characterization package

When shifting to a Consistency Testing-based control strategy, the characterization package that will be deployed to assess process changes should be defined and endorsed by a competent authority. It may contain new in vitro characterization assays but no in vivo assay(s). Indeed, when approved by competent authorities as part of the registration dossier, new methods are considered to be in force. Substituted in vivo methods should no longer be used and therefore should not be requested in case of a process change.

Acceptance criteria

Acceptance criteria (or other types of limits) will be set for the new assay(s) that will substitute in vivo potency/safety tests.
The VAC2VAC proposal highlights that there is no need to change the way acceptance criteria are defined for the new assay(s) that will substitute in vivo potency/safety tests. When properly defined, these criteria are reflecting the variability of the (new) method(s) combined with normal variability of the manufacturing process. It is important to insist on the fact that the introduction of a new analytical method neither changes the variability of the manufacturing process nor the quality of the product.

It is also important to acknowledge that, in the current control strategy, the Critical Quality Attribute (CQA) “potency” is addressed by an in vivo test. These tests have an inherently high variability with a range difficult to quantify. Setting specifications for the new in vitro methods may therefore be challenging when seeking correlation with the specification established for the in vivo assay. This correlation might even be impossible to establish in case the units used in the in vivo and the in vitro assay are different. Moreover, head-to-head comparison to clinical batches used to demonstrate efficacy/safety may not be possible for already licensed vaccines, since these batches will no longer be available or may be expired.

Taking these elements into consideration, the acceptance criteria of proposed in vitro tests will, in most cases, be set on collection of data from released batches – in order to maintain consistency of batch quality – rather than through seeking correlation to acceptance criteria established for the in vivo test. Data collection for setting the acceptance criteria must be performed in representative conditions of the future batch release (e.g., if the product is demonstrated to be stable, batches of different ages can be tested retrospectively with the in vitro assay. If the product is known to change over time, batches must be tested at the time of release with the in vitro assay in order to acquire representative data).

This approach should be supported by data demonstrating the ability of the proposed assay(s) to control key quality attributes of the vaccine and maintain the link between the quality of the batches to be released using the proposed Consistency Testing-based control strategy with those batches released as “compliant” when tested with the in vivo - based control strategy. Those latter “compliant” batches having been consistently demonstrated to be of the same quality as the clinical batches which have been demonstrated safe and efficacious through clinical studies.

The release and stability specifications and acceptance criteria for the new method(s) have to be set on a sufficient number of batches, also taking into account variability of the new method(s). Only limited information will be available on the variability between batches for the new assay at the time of implementation because only a few batches will have been tested with the new assay. When only limited batch results for the new method are available the acceptance criteria may initially be set somewhat wider to account for potential inter-batch variability. A predicted revision of the release specification acceptance criteria after sufficient batches have been tested could be part of the implementation strategy.

This proposal is supported by the following elements. The quality of vaccines is determined by their design, development, in-process controls, release controls, and process validation, and by specifications applied to them throughout development and manufacture. Specifications, i.e., those tests, validated procedures, and acceptance criteria play a major role in assuring the quality of the product and intermediates at release and during shelf life. Acceptance criteria need to be set in line with the various guidance e.g., text from the European Pharmacopeia issued by the European Directorate for the Quality of Medicines & HealthCare (EDQM) and other legal documents of relevance issued by other competent authorities outside the EU wherever this approach will be accepted.

Recently produced batches can be considered comparable to the original clinical batches, because they are also demonstrated by testing against bridged reference material equivalent to original clinical batches (e.g., Diphtheria and Tetanus) and/or are manufactured by a process demonstrated to be comparable, in cases where manufacturing changes have been introduced.

**Stability studies**

In vivo potency tests are often used in stability studies. Given the potential configuration of the Consistency Testing-based control strategy it is important to consider how stability studies should be managed after the substitution or the removal of in vivo potency.

Stability studies per se, do not need to be managed in a different way. After removal of the in vivo assay, results generated with the new method(s) being indicative for stability can be accepted, provided that they are within approved specifications throughout product shelf-life.

The strategy for stability testing needs to be adapted to the new control strategy and according to the respective stability-indicating capacity of the new assays as well as the process steps where they are performed. For instance, in a scenario where an in vivo potency assay is replaced by an assay measuring the antigen content and another assay assessing the structural integrity of the antigen, the assay for content will not be introduced in the stability plans because it is likely not stability indicating. The structural assay, being demonstrated to be stability indicating could be introduced in stability plans if the stability of the antigen requires evaluation according to current product stability knowledge.

The definition of the acceptance criteria should take into account stability data. If the results generated with the in vivo substitution method(s) show a decrease/increase over time, an end-of-shelf-life acceptance criterion can be defined for the vaccine provided that it is not in contradiction to current regulatory requirements and that it is in line with principles of e.g., ICH Q6B “Specifications: Test Procedures and Acceptance Criteria for Biotechnological/Biological Products”.

...
In vivo assays used in stability studies generally have a different discriminative power than in vitro assays regarding the various effects that may impact the product along stability. Therefore, the strategy for stability testing needs to be adapted to the new control strategy.

Change of the shelf-life criteria during storage can partly be assessed by accelerated stability studies but can only be confirmed in real-time stability studies. Final stability criteria have to be based on batches at the end of shelf-life. Again, a foreseen revision of end of shelf-life acceptance criteria may be part of the implementation strategy.

The role of Official Medicines Control Laboratories (OMCLs)

Technically all assays can be performed by Official Medicines Control Laboratories (OMCLs) regardless of the production step at which they are performed and this does not change with a control strategy based on the Consistency Approach.

Providing samples either of the final lot or from upstream process steps is required in the regulation of official batch release procedures. An upstream sample might need to be sent to the control laboratories before the final lot submission to gain time for release testing and to achieve testing within shelf-life.

In Europe, after approval of the data package submitted to EMA, the list of tests to be performed is defined by the EDQM - Official Control Authority Batch Release (OCABR) drafting groups and approved by consultation of the OMCL network.

In the European batch release framework (as laid down in Council Directive 2001/83/EC, Article 114 for medicinal products for human use and in Regulation 2019/6/EU Art 128 for medicinal products for veterinary use), an Official Medicines Control Laboratory (OMCL) tests a biological product before it can be marketed. The list of tests to be performed is prepared by the OCABR drafting group of experts addressing product quality and approved by consultation of the relevant EDQM OMCL network. Tests are chosen from those approved in the data package submitted to EMA. For example, the potency test is part of product quality evaluation package and, therefore, any tests replacing the potency test should be performed by the OMCLs. By extension, National Control Laboratories outside Europe could also perform the new assay(s) for the product release in their market or rely on the European batch release certificate.

Additional key elements to success

The keys to success are in the scientific approach combined with early interactions between manufacturers and competent authorities.

Even beyond the development of fit-for-purpose, alternative in vitro assays, the topic that triggered most discussions in the VAC2VAC consortium is how to build data packages that demonstrate that the change towards the new proposed Consistency Testing-based control strategy can be achieved.

The answer to that challenge is composed of three elements: including scientific relevance of the proposed in vitro assays, the use of non-compliant batches in the assessment of the control strategy and early interactions between manufacturers and authorities to discuss the proposed change(s). This approach reflects the fact that applying good science is the best way to move forward on such complex and sometimes sensitive topics. And this good science will be made more efficient when supported by scientific collaboration between the different parties involved (and not simply scientific communication).

The science: the use of non-compliant batches

Demonstration of agreement between in vitro and in vivo methods may not be scientifically justified and should not always be expected (Ph.Eur. 5.2.14 Substitution of in vitro method(s) by in vitro method(s) for the quality control of vaccines). Batches that differ from compliant batches (e.g., partially degraded, lower content, impurities) tested by in vivo tests in terms of quality should be used to demonstrate that the in vitro assay is fit-for-purpose.

The use of non-compliant batches is important for the demonstration of the ability of the new assay(s) to discriminate between compliant and non-compliant batches when changing from an existing in vivo - based - control strategy to a proposed Consistency Testing-based control strategy.

However, the direct comparison of results obtained on these non-compliant batches using an in vivo method and the proposed in vitro method(s) is neither considered as the best nor as a sufficient approach. Indeed, Ph.Eur. 5.2.14 states the following “…a demonstration of agreement between the 2 methods is generally not scientifically justified and should not always be expected. Even where pass/fail results from the 2 test procedures are in agreement, the correlation between 2 quantitative methods across the assay range may still be low. Regardless, the in vitro method(s) or testing strategy must provide at least the same confidence that the key quality attributes, which are necessary to ensure the consistency of a product’s safety and effectiveness, are adequately controlled.”

The reasons supporting that statement are:

- In vivo methods present a high level of variability that do not allow proper, statistically-relevant comparison of methods based on a reasonable amount of data (and animals).

- Historically, in vivo methods were generally not designed to address a well-defined Critical Quality Attribute but mostly rely on the belief that the product will trigger the same overall response in test animals as in target species.

- Modern analytical technologies allow for more relevant and comprehensive data sets about product characteristics/knowledge than in vivo assays.

Therefore, the demonstration that changing from an existing in vivo-based-control strategy to a proposed Consistency Testing-based control strategy must rely on data/evidence
that the proposed Consistency Testing-based control strategy can ensure the control of well-defined Critical Quality Attribute(s) that are contributing directly or indirectly to the quality attributes being measured by the in vivo assay(s) to be substituted. One must demonstrate that the level of control of the proposed Consistency Testing-based control strategy is appropriate in order to consistently provide a product of the desired quality.

Early scientific interactions between manufacturers and authorities
Manufacturers and regulator(s) must agree on an overall strategy and the type of data to be provided in a future variation/licensing application. A scientific discussion with competent authorities is of utmost importance and should start as early as possible. It is necessary to define different data/justification packages recommended for proposing in vivo potency substitution for different assays/individual products. It is recommended that specific data/justification packages are defined for proposing in vivo potency/safety test substitution of the various assays/individual products.

The various guidance documents issued by the competent authorities on how to apply for scientific advice should be followed.

The manufacturer is advised to have an early scientific discussion with competent authorities.

These discussions can address multiple levels of complexity:

- Rational explaining the scientific relevance and the added value of the proposed control strategy
- Outline of the proposed control strategy (new assays to be introduced, usage of these assays, e.g., stability)
- Proposed plan to generate the data package to support the change in control strategy

The data/justification package needs to be approved by competent agencies on a case-by-case basis. The approach should be in line with Ph.Eur. Chapter 5.2.14.: “The test methods used for routine quality control of vaccines are intended to monitor production consistency and to ensure comparability of the quality attributes between commercial batches and those batches originally found to be safe and efficacious in clinical studies, or for veterinary vaccines, in the target species.”

At any stage of the method development and validation, the scientific discussion with the competent authorities helps the manufacturer to develop the appropriate strategies, the tests including specification limits and the studies, so that major objections regarding the design of the in vivo substitution tests are not likely to be raised during the evaluation of the variation/licensing data package. The discussions can be very useful in light of the complexity of this type of change in the control strategy.

The data/justification packages should be supported by research and development studies, historical results generated on manufactured batches, stability studies, and non-conforming batch studies, in accordance with the advice given by regulatory agencies during the staggered approach and in line with the different guidelines in force. They can differ from one case to another due to the following: potency vs safety test, the number of in vitro test(s) to substitute an in vivo test, drug substance vs drug product stage, etc. Those data packages should therefore be considered on a case-by-case basis.

The discussions between manufacturers and regulators are thus critically important in light of the complexity of this type of change in the control strategy.

Conclusions
This paper summarizes the discussions and proposals made by the VAC2VAC consortium composed of experts, regulators and scientists from different institutions, manufacturers and competent authorities on how to support the deployment of the “Consistency Approach” for quality control of established vaccines. The consistency approach can replace the current in vivo testing, known to be laborious and highly variable for many established vaccines, with in vitro assays now seen as the Golden standard. Robust science and early interaction between manufacturers and competent authorities before and during dossier submission are certainly key elements to success. This consistency approach, when approved and in place, will allow new and established products of assured quality to reach the patient in a more efficient manner.

Science, newly developed methods and relevant guidelines are available to support the move to a Consistency Testing-based control strategy. Adapting the mindset was one of the major hurdles to a progressive vision but there is now a consensus between manufacturers and competent authorities to move firmly towards eliminating in vivo testing.

Definitions
Release assay. Assay which is used to assess the quality of a given batch of the final product before the release on the market, or a production intermediate prior to its entry into further manufacturing steps. Its result must be within approved pre-defined acceptance criteria.

Characterization assay. Physicochemical, immunochemical or biological assay which is used to accumulate knowledge on the product (final product or production intermediate) and that will contribute to the overall product understanding. A characterization assay is deployed specifically in the context of evaluating the impact of process changes on the product.

Critical Quality Attributes (CQA). A physical, chemical, biological or microbiological property or characteristic that is related to the product’s safety or efficacy and that should be within an appropriate limit, range, or distribution to ensure the desired product quality.
**Critical Process Parameter (CPP).** A process parameter whose variability has an impact on a critical quality attribute and therefore should be monitored or controlled to ensure the process yields a product of desired quality.

**Acceptance Criteria.** Numerical limits, ranges, or other suitable measures defining acceptance of analytical results and which the drug substance or drug product, intermediates or materials at defined stages of manufacturing process should meet.

**Ethics and consent**
Ethical approval and consent were not required.

---

### Data availability

**Underlying data**
No data are associated with this article.

### Acknowledgments
The authors would like to thank the members of the VAC2VAC consortium as well as the experts and scientists who participated in the two workshops held on June 22, 2020 and June 24, 2021 for their valuable contributions to this progressive vision.

---

### References


It was a privilege to review a letter on such an important topic. We have some general comments for further consideration:

- Throughout the text in the abstract, summary, additional key elements, conclusion - there is a mix of use of words manufactures vs industry, regulatory bodies vs competent authorities. Just ensure consistency and wording used is that which best describes the stakeholder referenced.

- Is there a need to elaborate briefly on what is IMI?

- In the first paragraph of the Introduction section, I recommend including the link to the IMI webpage on the topic also: https://www.imi.europa.eu/projects-results/project-factsheets/vac2vac

- In the last paragraph of the Introduction could you consider referencing the following publications as they provide, not only the state of the art of the developing of in vitro opportunities to vaccines batch release testing, but also share the discussion, engagement and consensus level of various type of stakeholders on the topic. The manuscript mentions the shift of mindset that happened in the last years, and those publications reported such shift so they strengthen the points made in the letter.

3. Modern science for better quality control of medicinal products “Towards global

Kirsty Reid
European Federation of Pharmaceutical Industry and Association, Brussels, Belgium

Laura Viviani
Humane Society International, Geneva, Switzerland
harmonization of 3Rs in biologicals**: The report of an EPAA workshop: https://www.sciencedirect.com/science/article/pii/S1045105617300647

- In the text there is a need for a final check of spacings, duplication of words (e.g. p5 ‘that are’ is repeated in paragraph 6).

- P5 - for consideration in the text. An example below is covid specific but the concept of regulatory alignment, harmonization and resilience is well described. The same concept is key in the case of the substitution of in vivo methods. Lack of regulatory alignment is one of the major hurdles. Similarly, the consistency approach concept might incur in resistance in its acceptance and implementation.


- Suggestion for the repeated use of the phrase 'Consistency Testing hyphen based hyphen control strategy'. Here it seems that “based” is a separate word, while, it means the Consistency Testing-based control strategy. Consider use of this suggested structure. Furthermore for the same phrase, sometimes the words all start with an upper case, others only certain words are upper case. Decide on one approach and keep it consistent.

- Abbreviations: First time European Directorate for the quality of Medicines and HealthCare is used, include the abbreviation (EDQM) next to it (p6). OCABR is mentioned, however never included in full in text - add Official Control Authority Batch Release to p7 before the abbreviation.

- P6 in the first paragraph under the heading ‘stability studies’ there is the reference to ‘removal of the in vivo’ consider also including the word ‘substitution or removal’.

- On p7 Paragraph starting ‘However, the comparison of results....’ is very confusing. It refers to 2 methods, then 2 procedures, then 2 quantitative methods. Please try to clarify for lay person.

- On p7 paragraph starting 'Modern analytical technologies..' please consider that the in vivo method might fail the requirement of the ICH Q2(R2) criteria for validation

- p8 when a paragraph is quoted from the PhEur, this should be in italics.

- Final sentence of final paragraph of the Conclusion, the word ‘aggressive’ is used. This is an odd choice of words. Reconsider using a different term here.

- In addition to the suggested publications above, I would recommend strengthening the letter by considering other publications which have come through the VAC2VAC projects: https://europevaccine.wixsite.com/vac2vac-eu/publications
References

Is the rationale for the Open Letter provided in sufficient detail? (Please consider whether existing challenges in the field are outlined clearly and whether the purpose of the letter is explained)
Yes

Does the article adequately reference differing views and opinions?
Yes

Are all factual statements correct, and are statements and arguments made adequately supported by citations?
Yes

Is the Open Letter written in accessible language? (Please consider whether all subject-specific terms, concepts and abbreviations are explained)
Yes

Where applicable, are recommendations and next steps explained clearly for others to follow? (Please consider whether others in the research community would be able to implement guidelines or recommendations and/or constructively engage in the debate)
Yes

**Competing Interests:** Laura Viviani has co-authored a number of publications in the present article under review. She is a biologics expert and represents an NGO which is a stakeholder group actively engaging with other stakeholders in this field on vaccines and non-animal methods. This is a small sector of experts and therefore there are possibilities of overlap however this has not influenced her expert judgement on the accuracy and purpose of the letter.

**Reviewer Expertise:** Animal research, animal welfare, alternatives, IMI, IHI, biology

**We confirm that we have read this submission and believe that we have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.**
Thank you very much for your constructive comments. We have adapted our publication accordingly and believe this leads to an improved version. We’d like to provide you our responses to the following comments:

○ Is there a need to elaborate briefly on what is IMI?

We prefer to keep our text focused on the main purpose of the publication. Several links included in the manuscript are directing towards websites that are referring to IMI.

○ P5 - for consideration in the text. An example below is covid specific but the concept of regulatory alignment, harmonization and resilience is well described. The same concept is key in the case of the substitution of in vivo methods. Lack of regulatory alignment is one of the major hurdles. Similarly, the consistency approach concept might incur in resistance in its acceptance and implementation.


The authors consider that this publication is not necessary for the reader to understand the manuscript and prefer not to add this reference that is related to the very specific case of Covid vaccines and is not focused on 3Rs objectives.

○ On p7 Paragraph starting 'However, the comparison of results....' is very confusing. It refers to 2 methods, then 2 procedures, then 2 quantitative methods. Please try to clarify for lay person

We agree with your comment, however, the confusion is coming from the original text from Ph.Eur. 5.2.14 that uses both “method” and “procedure”. We have therefore proposed not to change the text.

○ On p7 paragraph starting 'Modern analytical technologies...' please consider that the in vivo method might fail the requirement of the ICH Q2(R2) criteria for validation

We are not confident in writing that “the in vivo method might fail the requirement of the ICH Q2(R2) criteria for validation” since this could be misinterpreted as the if the current control strategy was not appropriate. And this is not the case. We understand your comment and believe its essence is covered by the bullet point above the one discussed: “In vivo methods present a high level of variability that do not allow proper, statistically-relevant comparison of methods based on a reasonable amount of data (and animals).”

○ In addition to the suggested publications above, I would recommend strengthening the letter by considering other publications which have come through the VAC2VAC projects: https://europevaccine.wixsite.com/vac2vac-eu/publications

The VAC2VAC consortium has indeed produced several very interesting publications in the context of the replacement of in vivo assays like, for instance the development of alternative in vitro assays. However, these publications are not illustrating or supporting the deployment of the consistency approach per se. We therefore propose not to add these references that risk to blur the specific message of this manuscript We sincerely hope that the revised version together with our responses to your comments will meet your expectations.
Competing Interests: No competing interests were disclosed.

Reviewer Report 14 October 2022

https://doi.org/10.21956/openreseurope.16305.r30253

© 2022 Lilley E. This is an open access peer review report distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Elliot Lilley
NC3Rs, London, UK

Overall, this is a very important letter and worthy of publication. The authors have summarised the output from two workshops, organised by the VAC2VAC consortium, to discuss how manufacturers, control laboratories and regulators can work together to promote the use of the “Consistency Approach” to avoid the use of animals in quality control, batch and lot-release testing of vaccines. The letter sets out clear recommendations and makes a valuable contribution to the promotion of the 3Rs in routine vaccine testing.

Sometimes the language is a little over-complicated in places and there are a number of typographical and grammatical errors.

For example, in the introduction the following sentence:
‘In that perspective, the initiative launched reflection on how to support the deployment of the “Consistency Approach” for quality control of established vaccines, where current quality control approaches are often relying on in vivo methods.’ does not read well.

I would suggest the following: ‘As part of the project, an initiative was launched to canvas opinion on how to support the deployment of the “Consistency Approach” for quality control of established vaccines, where current approaches often rely on in vivo methods.’

I would suggest that the manuscript is revised to improve the written English. This would greatly help to clarify the important message that the authors wish to communicate.

Is the rationale for the Open Letter provided in sufficient detail? (Please consider whether existing challenges in the field are outlined clearly and whether the purpose of the letter is explained)
Yes

Does the article adequately reference differing views and opinions?
Yes

Are all factual statements correct, and are statements and arguments made adequately
supported by citations?
Yes

Is the Open Letter written in accessible language? (Please consider whether all subject-specific terms, concepts and abbreviations are explained)
Partly

Where applicable, are recommendations and next steps explained clearly for others to follow? (Please consider whether others in the research community would be able to implement guidelines or recommendations and/or constructively engage in the debate)
Yes

**Competing Interests:** I was chair of the mid term review panel for the VAC2VAC project on behalf of the European Commission. A number of the authors of this letter are current (CJ & J-MP) or past (MH & JV) members of a working group that I manage as part of a project to review WHO guidelines to improve implementation of the 3Rs principles. I confirm I can review this article objectively.

**Reviewer Expertise:** I am Programme Manager for the 3Rs in vaccine batch release and quality control testing at the UK National Centre for the 3Rs (NC3Rs). After a 15-year career in the pharmaceutical industry, I spent 9 years with the RSPCA research animals department where I worked to promote refinement. I joined the NC3Rs in 2020 to work on a project to apply the 3Rs to batch testing of biologics, working closely with the WHO and a working group of international experts. I am a Fellow of the British Pharmacological Society and animal ethics editor of the British Journal of Pharmacology.

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however I have significant reservations, as outlined above.

Author Response 22 Nov 2022

Jean-Francois Dierick, GSK Vaccines, 89 Rue de l’Institut, Rixensart, Belgium

Thank you for your constructive comments. The entire text has been reviewed by a native speaker and has been improved. We hope these improvement will meet your expectations.

**Competing Interests:** No competing interests were disclosed.