Supporting Information

Ligand-Controlled Direct Hydroformylation of Tri-substituted Olefins

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1. General information

Commercially available compounds were used without further purification or drying. [Rh(acac)(CO)₂] was used as received from Alfa Aesar and was stored under argon. Syngas (CO:H₂ = 1:1) was purchased from Jong-Ang Industry Gas Company, Korea. Flash column chromatography was performed using silica gel (60-Å pore size, 40-63 µm, 4-6% H₂O content, Merck). Analytical thin-layer chromatography (TLC) was performed using glass plates pre-coated with 0.25 mm silica gel impregnated with fluorescent indicator (254 nm). Thin layer chromatography plates were visualized by exposure to ultraviolet light, an aqueous solution of phosphomolybdic acid (PMA). ¹H NMR(400 MHz), ¹³C { ¹H } NMR(100 MHz) and ³¹P { ¹H } NMR(162 MHz) spectra were recorded on a Bruker Ascend 400 or a Bruker Avance III HD spectrometer. All coupling constants (*J* values) were reported in Hertz (Hz). The high-resolution mass spectra (HRMS) were obtained using SYNAPT G2 (Waters, U.K.) spectrometer at the Division of Bioconvergence analysis / Biomedical Omics Center in KBSI.

Computational details. Molecular structures were optimized using density functional theory (DFT) by employing the Minnesota 2006 functional (M06)¹ as implemented in the Jaguar 8.4 software package.² For computationally effective and accurate calculations, the Rh atom was described with a LANL2DZ effective core basis set,³ the first coordination atoms of P and N of briphos ligands were described using a standard Pople basis set 6-31G**++, and the other atoms were described using MIDIX basis set.⁴

The exact cone angle of the optimized geometry was calculated by the method defined in the previous work of Allen and co-workers⁵ using the FindConeAngle package of Mathematica 8.0 software [Mathematica 8.0; Wolfram Research, Inc.: Champaign, IL,2010.].

Percent buried volume ($%V_{bur}$) was calculated using SambVca 2.0 calculation. For $%V_{bur}$ calculation, (1) 3.50 Å was selected as the value for the sphere radius, (2) both 2.28 Å was considered as M-P bond distance, (3) crystallography hydrogen atoms were omitted, (4) scaled Bondi radii were used as recommended by Cavallo.

X-ray Crystallography. The diffraction data of Rh(CO)(acac)(L4) were collected on a Bruker SMART 1000. A suitable size and quality of crystal was coated with Paratone-N oil and mounted on a MiTeGen MicroLoop. The data were collected with graphite-monochromated Mo K α radiation ($\lambda = 0.71073$ Å) at 120 K. Cell parameters were determined and refined by SMART program.⁶ Data reduction was performed using SAINT software.⁷ An empirical absorption correction was applied using the SADABS program.⁸ The structures were solved by direct methods and all non-hydrogen atoms were subjected to anisotropic refinement by full-matrix least squares on F2 by using the SHELXTL/PC package.⁹ Unless otherwise noted, hydrogen atoms were placed at their geometrically calculated positions and refined riding on the corresponding carbon atoms with isotropic thermal parameters. Full crystallographic details can be obtained free of charge from the Cambridge Crystallographic Data Center via www.ccdc.cam.ac.uk/data_request/cif [CCDC 1897350 for Rh(CO)(acac)(L4)].

2. Preparation of briphos ligands L1-L4

OH
$$(CH_2O)_n$$
 (0.5 equiv) $(COnc. HCI)$ $($

A solution of 2,4-di-*tert*-butylphenol (10.0 g, 48.4 mmol) and paraformaldehyde (0.73 g, 24.2 mmol) in xylene (6.0 mL) was added conc. HCl (0.8 ml) and the resulting solution was stirred for 4 h at 100 °C. All volatiles were removed under reduced pressure and the red-brown residue was suspended in diethyl ether(50 mL). H₂O (50 mL) was poured into the flask and the mixture was extracted with diethyl ether (3×50 mL). The combined organic layers were dried over anhydrous MgSO₄, filtered, and concentrated under reduced pressure to give light gray solid. The solid was suspended in *n*-hexane and stirred overnight. The white solid was filtered off and washed with *n*-hexane to afford 6,6'-methylene bis(2,4-di-*tert*-butyl phenol) (**S2**).

White solid (9.2 g, 21.7 mmol, 90% yield, mp 149-150 °C).
¹H NMR (400 MHz, CDCl₃): δ ppm 1.33 (s, 18H), 1.46 (s, 18H), 3.98 (s, 2H), 5.93 (s, 2H), 7.20 (d,
$$J = 2.4$$
 Hz, 2H), 7.24 (d, $J = 2.4$ Hz, 2H). ¹³C{¹H} NMR (100 MHz, CDCl₃): δ ppm 150.0, 143.1, 135.7, 126.2, 125.3, 122.7, 34.8, 34.4, 32.7, 31.7, 30.2.

HRMS (ESI) calculated for C₂₉H₄₄O₂[M+Na]⁺: 447.3239, Found: 447.3228

A solution of 6,6'-methylene bis(2,4-di-*tert*-butyl phenol) (**S2**, 4.3 g, 10.0 mmol) in anhydrous THF (4 mL) was added a cooled suspension (0 °C) of NaH (0.6 g, 25.0 mmol) in anhydrous THF (2 mL) under N₂ atmosphere. The mixture was stirred for an additional 10 min at 0 °C. Iodomethane (1.6 mL, 25.0 mmol) was added into the flask. The mixture was allowed to ambient temperature and then refluxed for 12 h. The reaction was quenched with addition of H₂O (30 mL) and the aqueous phase was extracted with diethyl ether (3×30 mL). The combined organic layers were dried over anhydrous MgSO₄, filtered, and concentrated in vacuo to give white solid [2,2'-methylene bis(4,6-di-*tert*-butyl anisole) (**S3**)].

White solid (3.8 g, 8.4 mmol, 84% yield, mp 94-95 °C).

¹**H NMR** (400 MHz, CDCl₃): δ ppm 1.22 (s, 18H), 1.43 (s, 18H), 3.74 (s, 6H), 4.12 (s, 2H), 6.90 (d, J = 2.5 Hz, 2H), 7.21 (d, J = 2.5 Hz, 2H). ¹³C{¹**H**} **NMR** (100 MHz, CDCl₃): δ ppm 156.1, 145.3, 141.6, 133.4, 126.6, 122.0, 61.5, 35.4, 34.6, 31.6, 31.3,

31.1.

HRMS (ESI) calculated for $C_{31}H_{48}O_2[M+Na]^+$: 475.3552, Found: 475.3555

To a solution of 2,2'-methylene bis(4,6-di-*tert*-butyl anisole) (**S3**, 3.7 g, 8.2 mmol), acetic acid (16 mL), and acetic anhydride (8.8 mL) was added Na₂CrO₄ (4.0 g, 24.6 mmol) and the resulting mixture was stirred for 4.5 h at 60 °C. After cooling to room temperature, H₂O (50 mL) and ethyl acetate (50 mL) were added. The solution was washed with H₂O (3×50 mL) and the organic layer was dried over anhydrous MgSO₄. Removal of volatiles in vacuo provided the crude product as yellow solid. The solid was suspended in EtOH (30 mL) and stirred overnight. The white solid was filtered off and washed with EtOH (10 mL) to afford bis[3,5-bis(*tert*-butyl)-2-methoxyphenyl]methanone (**S4**).

White solid (2.5 g, 5.3 mmol, 65% yield, mp 182-183 °C).

¹**H NMR** (400 MHz, CDCl₃): δ ppm 1.29 (s, 18H), 1.38 (s, 18H), 3.52 (s, 6H), 7.42 (d, J = 2.6 Hz, 2H), 7.49 (d, J = 2.6 Hz, 2H). ¹³C{¹**H**} **NMR** (100 MHz, CDCl₃): δ ppm 197.8, 158.2, 144.6, 142.1, 132.4, 127.6, 127.0, 62.9, 35.5, 34.7, 31.6, 30.9. **HRMS**

(ESI) calculated for C₃₁H₄₆O₃[M+Na]⁺: 467.3525, Found: 467.3522

A cooled solution of bis[3,5-bis(*tert*-butyl)-2-methoxyphenyl]methanone (**S4**, 2.4 g, 5.1 mmol) in anhydrous CH₂Cl₂ (20 mL) was added BBr₃ (31 mmol) in CH₂Cl₂ (31 mL) dropwise over 1 h under N₂. The mixture was allowed to ambient temperature and stirred for 3 h. The reaction was quenched with addition of MeOH (100 mL). The aqueous phase was extracted with ethyl acetate (50 mL). The combined organic layer was washed with H₂O (3×50 mL), dried over anhydrous MgSO₄, filtered, and concentrated to give yellow solid product (1).

Yellow solid (2.1 g, 4.8 mmol, 92% yield, mp 201-202 °C).

¹**H NMR** (400 MHz, CDCl₃): δ ppm 1.28 (s, 18H), 1.48 (s, 18H), 7.38 (d, J = 2.4 Hz, 2H), 7.57 (d, J = 2.4 Hz, 2H), 11.19 (d, J = 0.5 Hz, 2H). ¹³C{¹**H**} **NMR** (100 MHz, CDCl₃): δ ppm 204.9, 150.9, 140.0, 138.0, 130.7, 127.7, 119.8, 35.4, 34.5, 31.6, 29.6.

HRMS (ESI) calculated for $C_{29}H_{42}O_3[M+H]^+$: 439.3212, Found: 439.3210

A mixture of 950 mg (2.17 mmol) of 1 and (10.8 mmol) of 3,5-dimethylaniline was stirred for 12 h at 140 °C. The clean formation of imine product 2 was confirmed by ¹H NMR spectra of crude mixtures or thin-layer chromatography (TLC, ethyl acetate : n-hexane = 1:20, Rf: 0.5(UV)). After cooling to ambient temperature, the mixture, dark brown oil loaded on a pad of silica-gel (10 cm) was eluted with ethyl acetate/n-hexane (1/20) to afford yellow solid 2. The yellow solid 2 was transferred to a round bottom flask charged with THF (20.0 mL). After cooling the mixture at 0 °C, a solution of LiAlH₄ (6.51 mmol) in THF (2.8 mL) was added dropwise under N₂. The mixture was stirred for 4 h and then ethyl acetate (20 mL) was added. The crude mixture was washed with H₂O (3×30 mL), and the organic layer was dried over anhydrous MgSO₄, filtered, and concentrated under reduced pressure. The crude product dissolved in 20 mL of chloroform was mixed with 0.2 mL (2.17 mmol) of phosphorous tribromide and 0.91 mL (6.51 mmol) of Et₃N under N₂ atmosphere. The resulting solution was then stirred at ambient temperature for 4 h. The solution was filtered over a pad of silica-gel (10 cm) with ethyl acetate and concentrated under reduced pressure. The crude product dissolved in 25 mL of ethanol to induce precipitation. The precipitate was filtered and washed with ethanol to afford analytically pure briphos ligand L3.

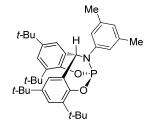
L1 (23% yield, white solid)

¹H NMR (400 MHz, CDCl₃): δ ppm 7.15 (d, J = 2.4 Hz, 2H), 7.00 (d, J = 2.4 Hz, 2H), 4.92 (d, J = 5.1 Hz, 1H), 3.07 (qt, J = 11.4, 3.6 Hz, 1H), 1.83-1.67 (m, 4H), 1.57 (d, J = 15.9 Hz, 2H), 1.46-1.41 (m, 1H), 1.38 (s, 18H), 1.28 (s, 18H), 1.21 (dt, J = 13.1, 2.9 Hz, 2H), 1.17–1.01(m, 1H). ¹³C{¹H} NMR (100 MHz, CDCl₃): δ ppm 146.3, 143.6, 137.9, 128.4, 122.8, 122.1, 59.9, 59.7, 54.3, 35.1, 34.5, 34.1, 34.0, 31.7, 29.9, 26.1, 25.7. ³¹P{¹H} NMR (162 MHz, CDCl₃): δ ppm 94.9. HRMS (ESI) calculated for C₃₅H₅₂NO₂P[M+H]⁺: 550.3816, Found: 550.3868

L2 (45% yield, white solid)

¹**H NMR** (400 MHz, CDCl₃): δ ppm 7.26 (d, J = 1.6 Hz, 2H), 7.20 (d, J = 2.4 Hz, 2H), 7.13 (dt, J = 2.2, 1.2 Hz, 1H), 7.11 (q, J = 1.0 Hz, 1H), 7.09 (d, J = 2.4 Hz, 2H), 7.00 (tt, J = 7.4, 1.0 Hz, 1H), 5.5 (d, J = 4.1 Hz, 1H), 1.39 (s, 18H), 1.29 (s, 18H). ¹³**C**{¹**H**} **NMR** (100 MHz, CDCl₃): δ ppm 145.9, 144.5, 138.2, 129.5, 127.24, 127.17, 123.3, 122.9, 122.3, 120.8, 120.7,

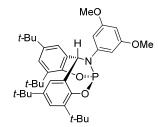
55.3, 35.1, 34.6, 31.7, 29.9. ${}^{31}P\{{}^{1}H\}$ NMR (162 MHz, CDCl₃): δ ppm 87.1. HRMS (ESI) calculated for $C_{35}H_{46}NO_{2}P[M+H]^{+}$: 544.3346, Found: 544.3330



L3 (68% yield, white solid)

¹**H NMR** (400 MHz, CDCl₃): δ ppm 7.21 (d, J = 2.5 Hz, 2H), 7.12 (d, J = 2.4 Hz, 2H), 6.76 (dq, J = 1.6, 0.8 Hz, 2H), 6.66 (tt, J = 1.5, 0.7 Hz, 1H), 5.55 (d, J = 4.1 Hz, 1H), 2.26 (q, J = 0.6 Hz, 6H), 1.42 (s, 18H), 1.31(s, 18H). ¹³**C**{¹**H**} **NMR** (100 MHz, CDCl₃): δ ppm 146.0, 145.9, 144.7, 144.4, 139.1, 138.1, 127.4, 127.3, 124.7, 123.2, 122.3, 118.6,

118.5, 56.4, 35.1, 34.6, 31.7, 29.9, 21.6. ³¹**P**{¹**H**} **NMR** (162 MHz, CDCl₃): δ ppm 87.1. **HRMS** (ESI) calculated for $C_{37}H_{50}NO_2P[M+H]^+$: 572.3659, Found: 572.3604.



L4 (45% yield, white solid)

¹**H NMR** (400 MHz, CDCl₃): δ ppm 7.21 (d, J = 2.4 Hz, 2H), 7.11 (d, J = 2.4 Hz, 2H), 6.31 (dd, J = 2.2, 1.1 Hz, 2H), 6.13 (t, J = 2.2 Hz, 1H), 5.55 (d, J = 4.1 Hz, 1H), 3.74 (s, 3H), 1.41 (s, 18H), 1.30 (s, 18H). ¹³**C**{¹**H**} **NMR** (100 MHz, CDCl₃): δ ppm 161.5, 146.7, 146.5 145.9, 145.8, 144.6, 138.24, 138.22, 127.13, 127.07, 123.3, 122.3, 100.1,

99.1, 99.0, 95.0, 56.2, 55.4, 35.1, 34.6, 31.7, 29.9. $^{31}P\{^{1}H\}$ NMR (162 MHz, CDCl₃): δ ppm 86.9. HRMS (ESI) calculated for $C_{37}H_{50}NO_{4}P[M+H]^{+}$: 604.3557, Found: 604.3532

3. ¹H NMR spectra of rhodium complexes

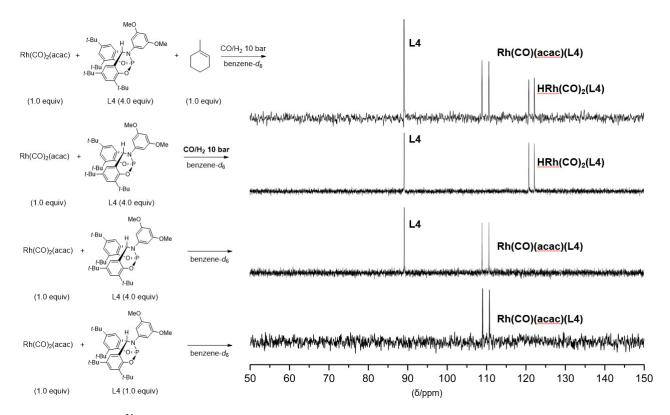


Figure S1. Stacked ³¹P{1H} NMR spectra showing ligation of L4 with Rh(CO)₂(acac).

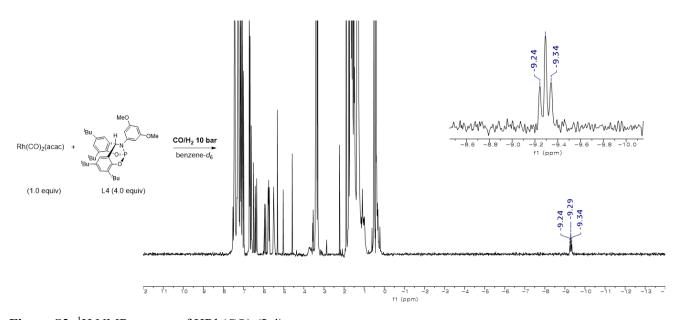


Figure S2. ¹H NMR spectra of HRh(CO)₂(L4)

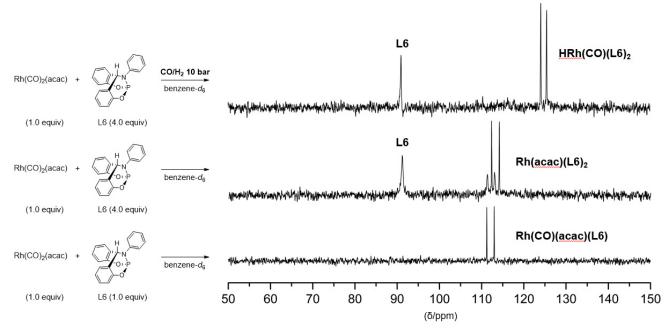


Figure S3. Stacked ³¹P{1H} NMR spectra showing ligation of L6 with Rh(CO)₂(acac).

4. General procedure for hydroformylation

In a glovebox filled with argon, a 5 mL vial equipped with a magnetic bar was added Rh(acac)(CO)₂ (0.01 mmol in 0.5 mL toluene) and ligand (0.02 mmol). After stirring for 30 min, 1-methyl-1-cyclohexene (1.0 mmol) and additional toluene was charged to bring the total volume of 2.0 mL. The vial was transferred into an autoclave and taken out of the glovebox then 20 bar of syngas (CO:H₂=1:1) was charged. The reaction mixture was stirred at 100 °C for 12 h. After the reaction, autoclave reactor was cooled to ambient temperature and the pressure was carefully released in a well-ventilated fume hood. The conversion of 1-methyl-1-cyclohexene was determined by ¹H NMR spectra of the crude reaction mixture. The ratio of isomers (5a to 5d) was determined by GC-MS analysis of the crude reaction mixture. The conversion and selectivity of other substrates were determined by GC-MS spectra with mesitylene as internal standard. Isolated yields were determined after purifying the crude product by silica-gel column chromatography (ethyl acetate : *n*-hexane = 1:10). For volatile aldehyde products, reduction with NaBH₄ was performed and the resulting alcohols were characterized.

- Optimization of Rh/L4-catalyzed hydroformylation

Entry	Catalyst	L/Rh	Pressure	Temperature	Conversion	5a/5b
1	0.1 mol%	2	20 bar	100 °C	30%	2.5
2	0.5 mol%	2	20 bar	100 °C	83%	3.3
3	1.0 mol%	2	20 bar	100 °C	83%	4.0
4	4.0 mol%	2	20 bar	100 °C	>99%	4.0
5	1.0 mol%	2	20 bar	80 °C	17%	2.3
6	1.0 mol%	2	20 bar	120 °C	48%	3.8
7	1.0 mol%	10	20 bar	100 °C	54%	3.8
8	1.0 mol%	5	20 bar	100 °C	70%	4.0
9	1.0 mol%	1	20 bar	100 °C	33%	4.0
10	1.0 mol%	2	10 bar	100 °C	68%	4.0
11	1.0 mol%	2	40 bar	100 °C	63%	3.9
12	1.0 mol%	2	CO 20 bar H ₂ 10 bar	100 °C	10%	2.3
13	1.0 mol%	2	CO 10 bar H ₂ 20 bar	100 °C	7%	4.7

- Product characterization

2,3-Dimethylbutan-1-ol (6): colorless oil. ¹H NMR (400 MHz, CDCl₃):
$$\delta$$
 ppm 3.59 (dd, J = 10.6, 5.9 Hz, 1H), 3.44 (dd, J = 10.6, 7.0 Hz, 1H), 1.74-1.66 (m, 1H), 1.53-1.47 (m, 1H), 0.91 (d, J = 6.9 Hz, 3H), 0.86 (d, J = 6.9 Hz, 3H), 0.83 (d, J = 6.8 Hz,

3H). ¹³C{¹H} NMR (100 MHz, CDCl₃): δ ppm 66.8, 41.6, 29.0, 20.8, 18.2, 12.7. HRMS (ESI) calculated for $C_6H_{14}O[M-H]^+$: 101.0966, Found: 101.0968

2-Isopropylpentan-1-ol (7): colorless oil. ¹H NMR (400 MHz, CDCl₃):
$$\delta$$
 ppm 3.50 (dd, J = 10.5, 5.8 Hz, 1H), 3.41 (dd, J = 10.5, 6.6 Hz, 1H), 1.67-1.42 (m, 2H), 1.35-1.26 (m, 1H), 1.26-1.04 (m, 3H), 0.96-0.86 (m, 9H). ¹³C{¹H} NMR (100 MHz,

CDCl₃): δ ppm 68.6, 36.4, 36.2, 31.0, 28.4, 22.9, 22.6, 16.7. **HRMS** (ESI) calculated for $C_8H_{18}O[M-H]^+$: 129.1279, Found: 129.1275

2-(Hydroxymethyl)-3-methylbutanal (8): colorless oil. ¹**H NMR** (400 MHz, CDCl₃): δ ppm 9.81 OH (d, J = 1.3 Hz, 1H), 3.96 (dd, J = 11.2, 8.1 Hz, 1H), 3.75 (dd, J = 11.2, 3.6 Hz, 1H), 2.40 (m, 1H), 2.17 (m, 1H), 1.03 (d, J = 6.7 Hz, 3H), 1.00 (d, J = 7.1 Hz, 3H). ¹³**C**{¹**H**} **NMR** (100 MHz, CDCl₃): δ ppm 206.3, 60.0, 59.7, 26.5, 20.7, 20.2. **HRMS** (ESI) calculated for C₆H₁₂O₂[M+H]⁺: 117.0916, Found: 117.0912.

2-Formyl-3-methylbutyl acetate (9): pale-yellow oil. ¹H NMR (400 MHz, CDCl₃): δ ppm 9.71 (d, J = 2.6 Hz, 1H), 4.38 (dd, J = 11.4, 8.3 1H), 4.27 (dd, J = 11.4, 4.5 Hz, 1H), 2.47 (dddd, J = 8.3, 6.4, 4.5, 2.6 Hz, 1H), 2.13 (dq, J = 13.6, 6.8 Hz, 1H), 2.03 (s, 3H), 1.02 (d, J = 3.5 Hz, 3H), 0.93 (d, J = 3.6 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃): δ ppm 202.7, 170.8, 61.0, 56.8, 26.3, 20.7, 20.3, 19.7. HRMS (ESI) calculated for $C_8H_{14}O_3[M+Na]^+$: 181.0841, Found: 181.0842.

3-Methyl-2-phenylbutanal (10): pale-yellow oil. ¹H NMR (400 MHz, CDCl₃): δ ppm 9.67 (d, J = 3.3 Hz, 1H), 7.35-7.25 (m, 3H), 7.18-7.16 (m, 2H), 3,17 (dd, J = 9.5, 3.3 Hz, 1H), 2.40 (dp, J = 9.5, 6.7 Hz, 1H), 1.03 (d, J = 6.6 Hz, 3H), 0.75 (d, J = 6.7 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃): δ ppm 201.1, 135.5, 129.3, 128.9, 127.5, 66.8, 28.8, 21.2, 20.0. HRMS (ESI) calculated for C₁₁H₁₄O[M+H]⁺: 163.1123, Found: 163.1128.

2-Cyclohexylpropan-1-ol (11): colorless oil. 1 H NMR (400 MHz, CDCl₃): δ ppm 3.59 (dd, J = 10.5, 5.7 Hz, 1H), 3.43 (dd, J = 10.5, 6.9 Hz, 1H), 1.71 (m, 2H), 1.62 (m, 3H), 1.47 (tt, J = 6.9 Hz, 5.4 Hz, 1H), 1.38-1.27 (m, 1H), 1.25-1.05 (m, 4H), 1.01-0.91 (m, 1H), 0.86 (d, J = 6.9 Hz, 3H). 13 C{ 1 H} NMR (100 MHz, CDCl₃): δ ppm 66.4, 41.0, 39.5, 31.1, 28.9, 26.9, 26.8, 26.7, 13.5. HRMS (ESI) calculated for C₉H₁₈O[M-H]⁺: 141.1279, Found: 141.1281.

CDCl₃): δ ppm 3.71 (dd, J = 10.7, 2.7 Hz, 1H), 3.52 (dd, J = 10.7, 5.8 Hz, 1H), 1.82-1.73 (m, 2H), 1.69-1.64 (m, 2H), 1.33 (s, 1H), 1.28-1.20 (m, 3H), 1.11-1.07 (m, 2H), 0.92 (d, J = 6.2 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃): δ ppm 66.2, 46.7, 35.8, 33.8, 29.7, 26.5, 26.4, 20.3. HRMS (ESI) calculated for C₈H₁₆O[M+Na]⁺: 151.1099, Found: 151.1088

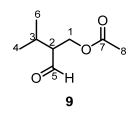
trans-2-(Methylcyclopentyl)methanol (12): colorless oil. ¹H NMR (400 MHz, CDCl₃): δ ppm 3.65 (dd, J = 10.5, 5.0 Hz, 1H), 3.47 (dd, J = 10.5, 7.0 Hz, 1H), 1.87-1.76 (m, 2H), 1.65-1.49 (m, 4H), 1.39-1.32 (m, 1H), 1.22-1.15 (m, 1H), 1.01 (d, J = 6.4 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃): δ ppm 66.7, 49.9, 37.1, 35.2, 29.6, 24.1, 20.2. HRMS (ESI) calculated for C₇H₁₄O[M-H]⁺: 113.0966, Found: 113.0964.

O H (1*R*,2*R*,6*S*)-2-Hydroxy-6-methylcyclohexane-1-carbaldehyde (13): pale-yellow oil. ¹H NMR (400 MHz, CDCl₃): δ ppm 9.76 (d, J = 3.6 Hz, 1H), 3.87-3.81 (td, J = 10.3, 4.5 Hz, 1H), 2.04 (dd, J = 11.9, 3.1 Hz, 1H), 1.93 (td, J = 10.4, 10.0, 3.6 Hz, 1H), 1.81-1.63 (m, 3H), 1.40-1.24 (m, 3H), 1.01 (d, J = 6.4 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃): δ ppm 205.6, 69.6, 64.6, 34.0, 33.7, 31.8, 23.3, 19.7. HRMS (ESI) calculated for $C_8H_{14}O_2[M+H]^+$: 143.1072, Found: 143.1082.

OH ((1*S*,2*S*,3*S*,5*R*)-2,6,6-Trimethyl bicyclo[3.1.1]heptan-3-yl)methanol (14): colorless oil. ${}^{1}H$ NMR (400 MHz, CDCl₃): δ ppm 3.58 (dd, J = 10.2, 5.7 Hz, 1H), 3.45 (dd, J = 10.2, 7.5 Hz, 1H), 2.31-2.25 (dtd, J = 9.6, 6.2, 2.1 Hz, 1H), 2.16-2.09 (dddd, J = 13.5, 10.1, 3.4, 2.1 Hz, 1H), 1.94-1.86 (m, 2H), 1.78-1.75 (m, 1H), 1.72-1.68 (m, 1H), 1.55 (ddd, J = 13.5, 5.8, 2.7 Hz, 1H), 1.19 (s, 3H), 1.07 (d, J = 7.2 Hz, 3H), 1.01 (s, 3H), 0.73 (d, J = 9.6 Hz, 1H). ${}^{13}C\{{}^{1}H\}$ NMR (100 MHz, CDCl₃): δ ppm 70.1. 47.9. 41.5. 39.5. 39.1. 39.0. 33.6. 31.3. 28.1, 23.0, 22.2. HRMS (ESI) calculated for $C_{11}H_{20}O[M-H]^{+}$: 167.1436, Found: 167.1432

5. Determination of aldehyde structures

For substrate No. 9



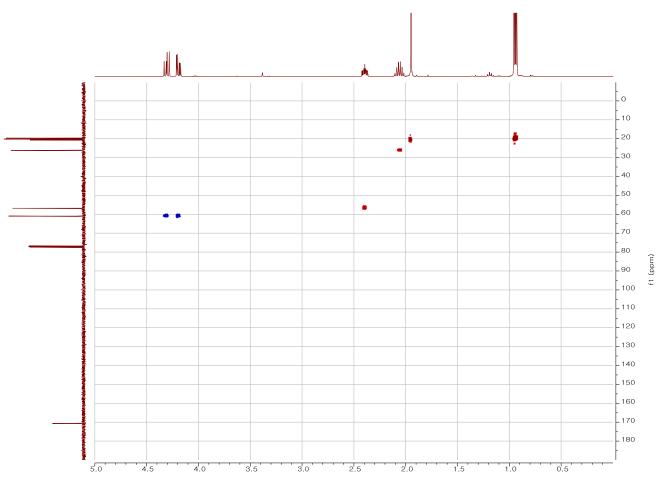


Figure S4. HSQC spectrum of 9 in CDCl₃.

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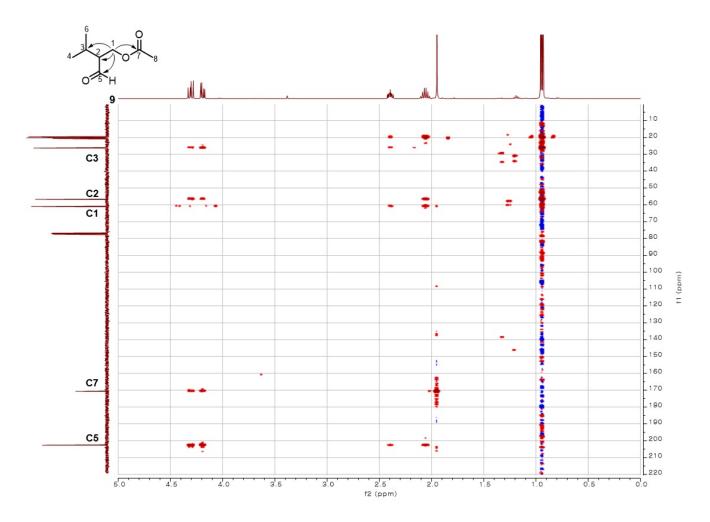


Figure S5. HMBC spectrum of 9 in CDCl₃

For substrate No. 13

HO
$$H_a$$
 H_b H_c $J_{ab} = 10.4 \text{ Hz}$ $J_{bc} = 10.0 \text{ Hz}$ $J_{bd} = 3.6 \text{ Hz}$ H_a H_c H_a H_c H_c H_a H_c H_c

For substrate No. 14

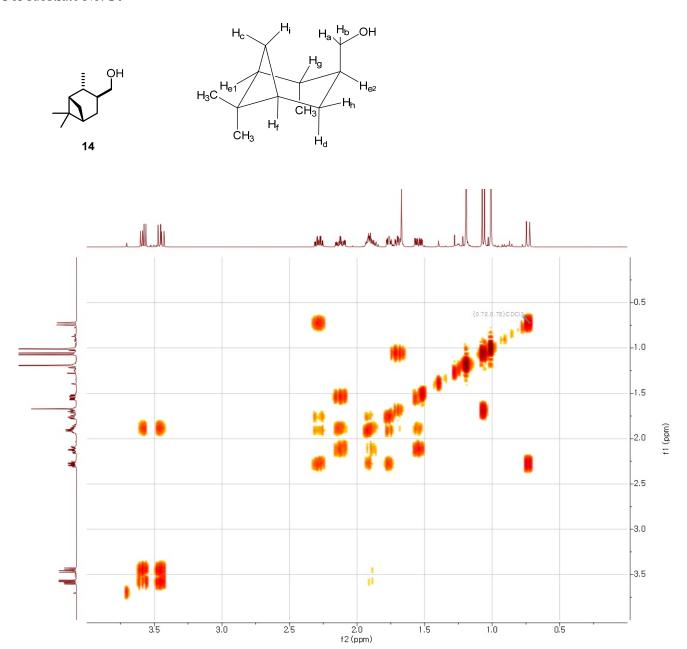


Figure S6. COSY spectrum of 14 in CDCl₃.

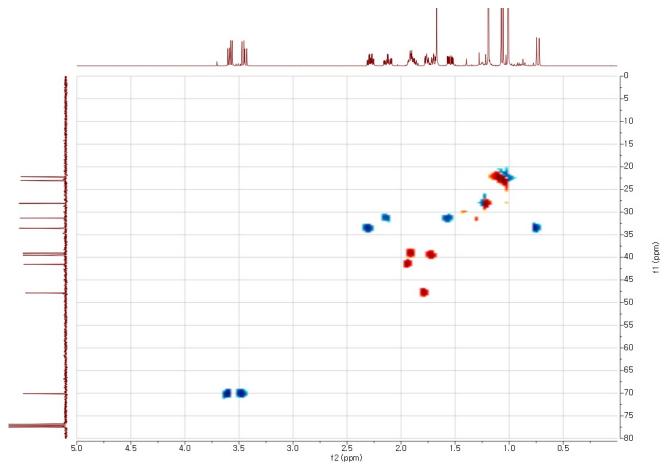


Figure S7. HSQC spectrum of 14 in CDCl₃.

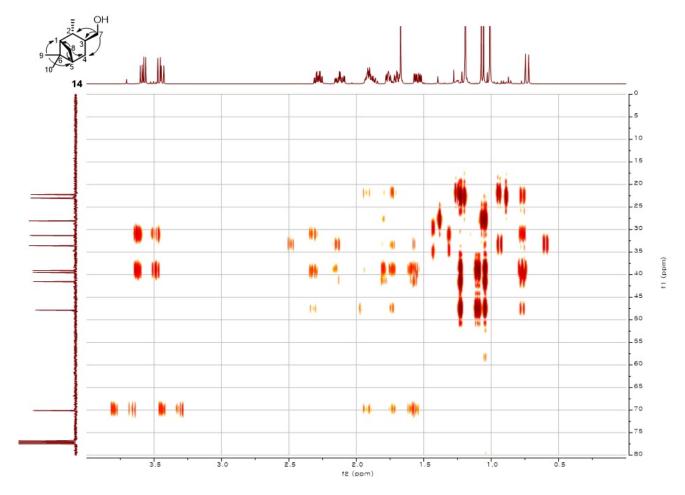
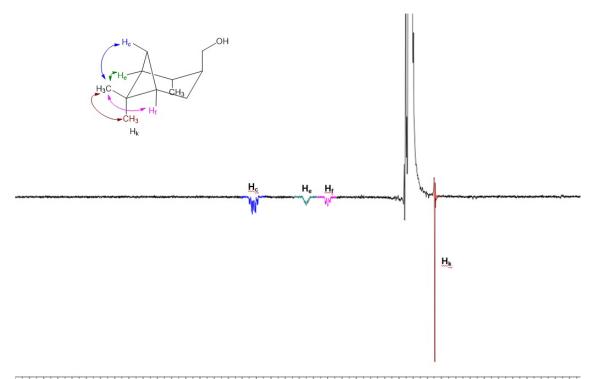
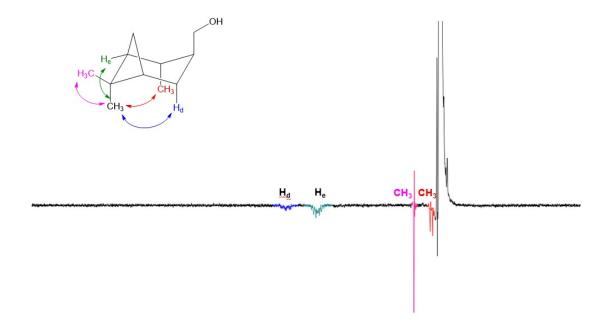


Figure S8. HMBC spectrum of 14 in CDCl₃.



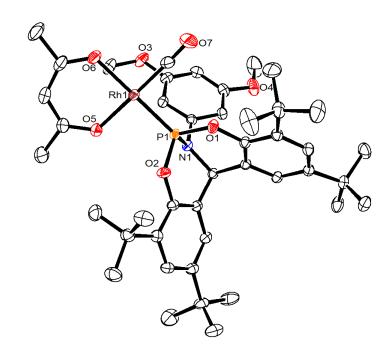
0 3/9 3/8 3/7 3/6 3/5 3/4 3/3 3/2 3/1 3/0 2/9 2/8 2/7 2/6 2/5 2/4 2/3 2/2 2/1 2/0 1/9 1/8 1/7 1/6 1/5 1/4 1/3 1/2 1/1 1/0 0/9 0/8 0/7 0/6 0/5 0/4 0/3 0/2 0/1 ff (gpm)



0 3.9 3.8 3.7 3.6 3.5 3.4 3.3 3.2 3.1 3.0 2.9 2.8 2.7 2.6 2.5 2.4 2.3 2.2 2.1 2.0 1.9 1.8 1.7 1.6 1.5 1.4 1.3 1.2 1.1 1.0 0.9 0.8 0.7 0.6 0.5 0.4 0.3 0.2 0.1 ft (ppm)

Figure S9. NOE spectrum of 14 in CDCl₃.

6. Crystal data



ORTEP representation (50% probability) of the crystal structure of Rh(CO)(acac)(L4)

Identification of	code	0521B-1	a

Empirical formula C43 H57 N O7 P Rh

Formula weight 833.77

Temperature 120(2) K

Wavelength 0.71073 Å

Crystal system Monoclinic

Space group P 21/c

Unit cell dimensions a = 16.1403(6) Å $\alpha = 90^{\circ}$.

b = 12.6980(5) Å $\beta = 92.570(2)^{\circ}.$

c = 20.9045(7) Å $\gamma = 90^{\circ}$.

Volume 4280.1(3) Å³

Z 4

Density (calculated) 1.294 Mg/m³
Absorption coefficient 0.484 mm⁻¹

F(000) 1752

Crystal size $0.230 \times 0.130 \times 0.110 \text{ mm}^3$

Theta range for data collection 2.238 to 33.802°.

Index ranges -25 <= h <= 25, -19 <= k <= 19, -32 <= l <= 32

Reflections collected 148484

Independent reflections 17093 [R(int) = 0.0519]

Completeness to theta = 25.242° 99.8 %

Absorption correction Semi-empirical from equivalents
Refinement method Full-matrix least-squares on F²

Data / restraints / parameters 17093 / 0 / 494

Goodness-of-fit on F² 1.044

Final R indices [I>2sigma(I)] R1 = 0.0312, wR2 = 0.0776R indices (all data) R1 = 0.0444, wR2 = 0.0847

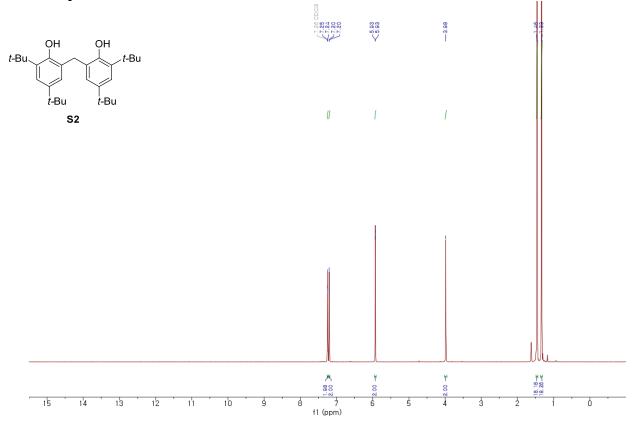
Extinction coefficient n/a

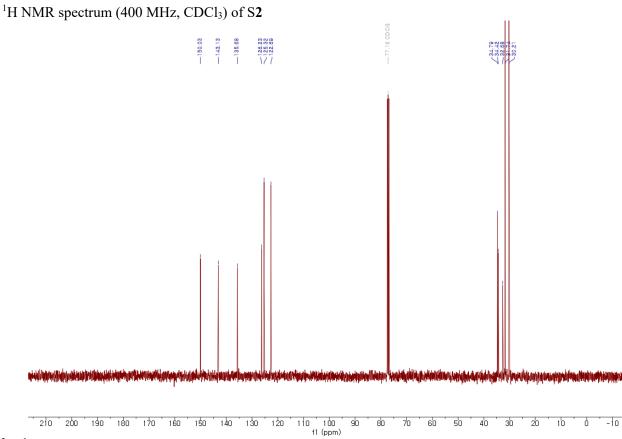
Largest diff. peak and hole 0.552 and -0.400 e.Å-3

7. Reference

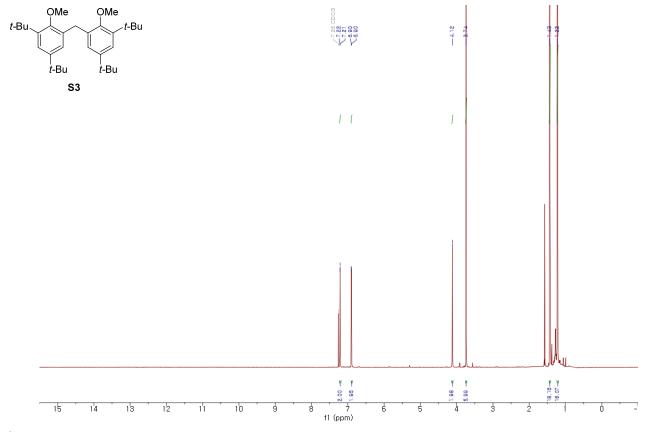
- 1. Y. Zhao and D. G. Truhlar, Theor. Chem. Acc. 2008, 120, 215–241.
- 2. A.D. Bochevarov, E. Harder, T.F. Hughes, J.R. Greenwood, D.A. Braden, D.M. Philipp, D. Rinaldo, M.D. Halls, J. Zhang, R.A. Friesner
- 3. J. P. Hay, W. R. Wadt, J. Chem. Phys. 1985, 82, 299.
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- 5. J. A. Bilbrey, A. Kazez, J. Locklin, W. D. Allen, J. Comput. Chem. 2013, 34, 1189–1197.
- 6. SMART (version 5.0), data collection software; Bruker AXS, Inc., Madison, WI, 1998.
- 7. SAINT (version 5.0), data integration software; Bruker AXS, Inc., Madison, WI, 1998.
- 8. G. M. Sheldrick, SADABS: program for absorption correction with the Bruker SMART system; Universität Göttingen, Germany, 1996.
- 9. G. M. Sheldrick, SHELXTL (version 6.1), Bruker AXS, Inc., Madison, WI, 2000.

8. NMR Spectra

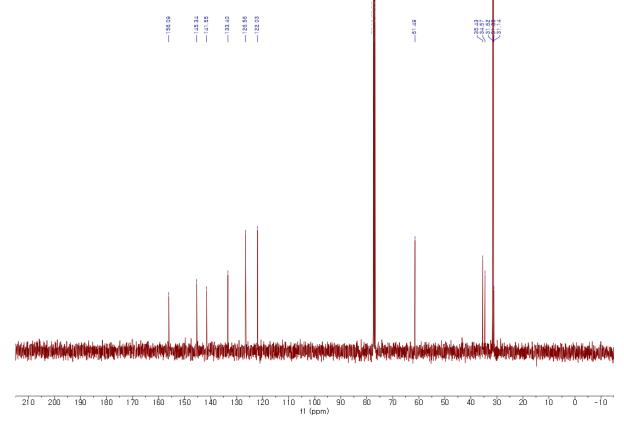




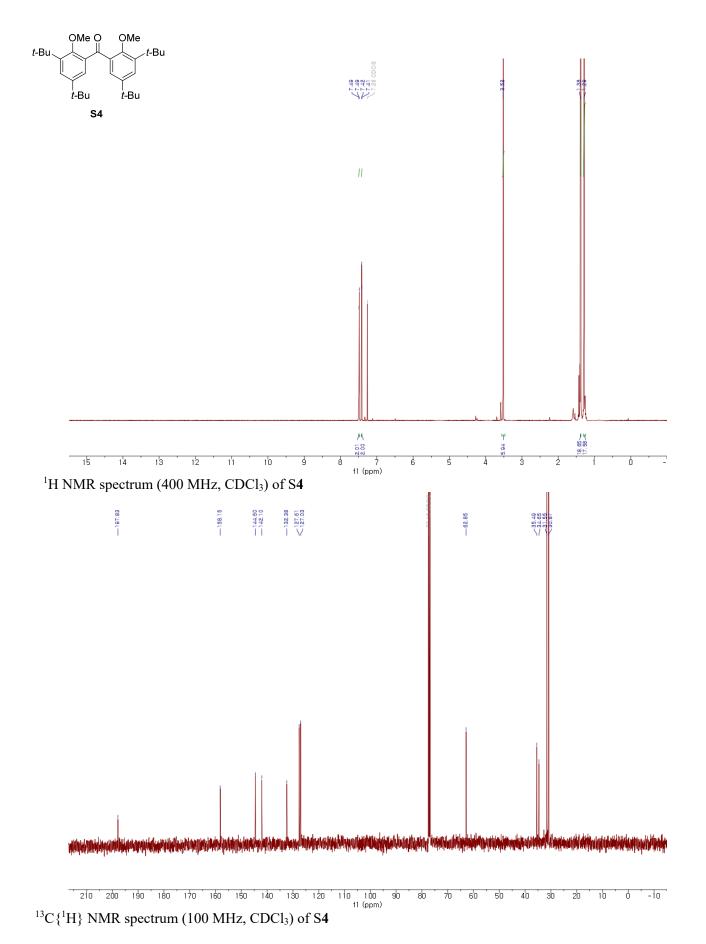
 $^{13}C\{^1H\}$ NMR spectrum (100 MHz, CDCl₃) of S2



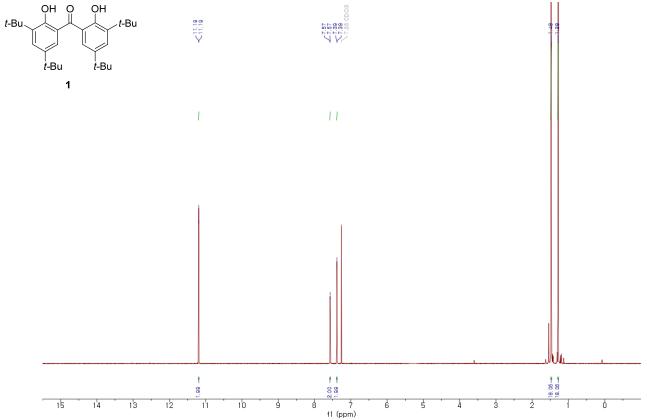




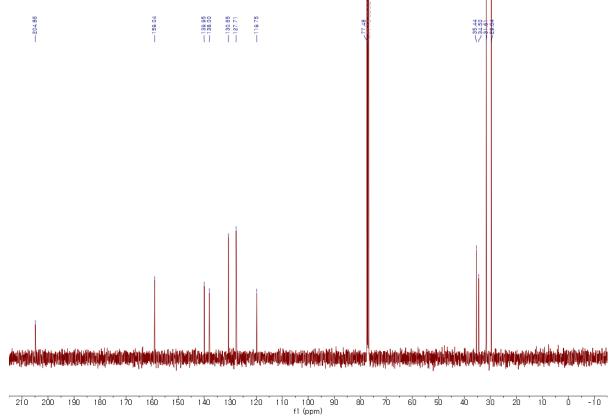
 $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (100 MHz, CDCl₃) of S3



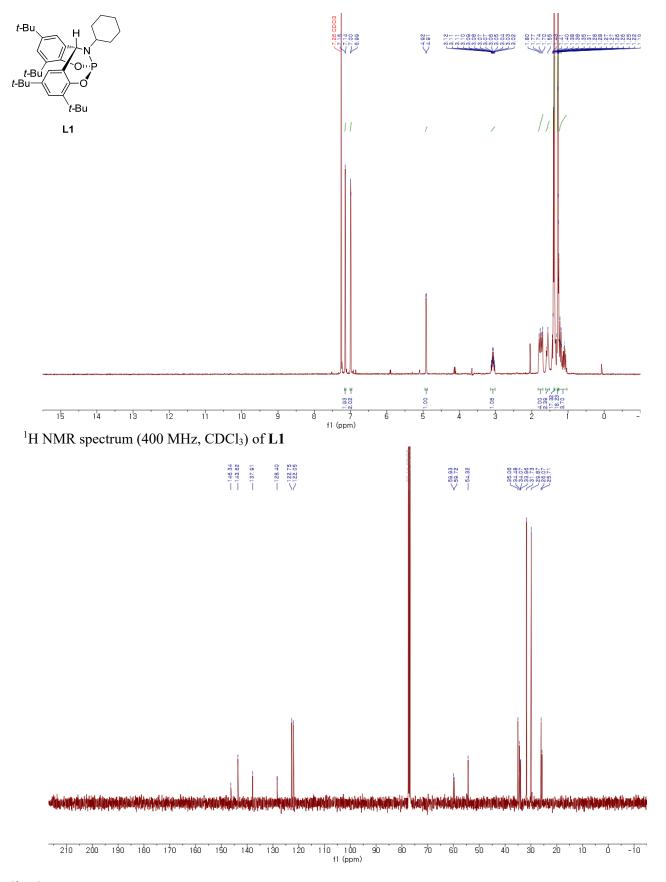
S22



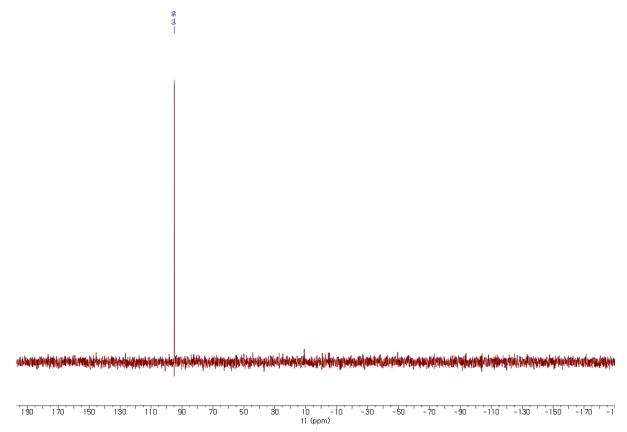
¹H NMR spectrum (400 MHz, CDCl₃) of 1



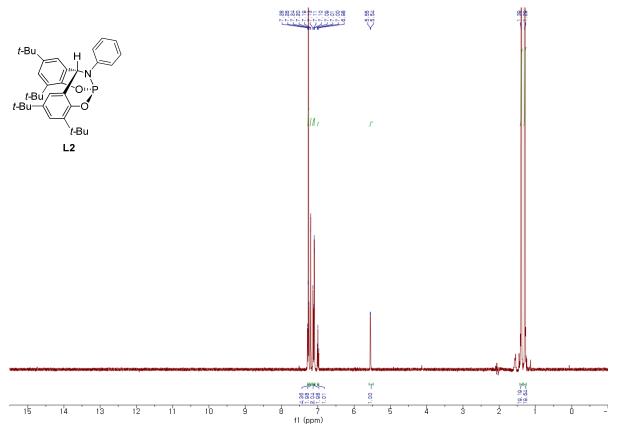
 $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (100 MHz, CDCl₃) of 1



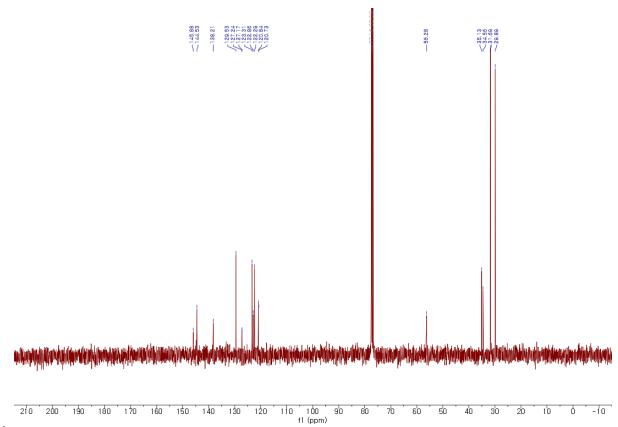
 $^{13}C\{^1H\}$ NMR spectrum (100 MHz, CDCl3) of $\boldsymbol{L1}$



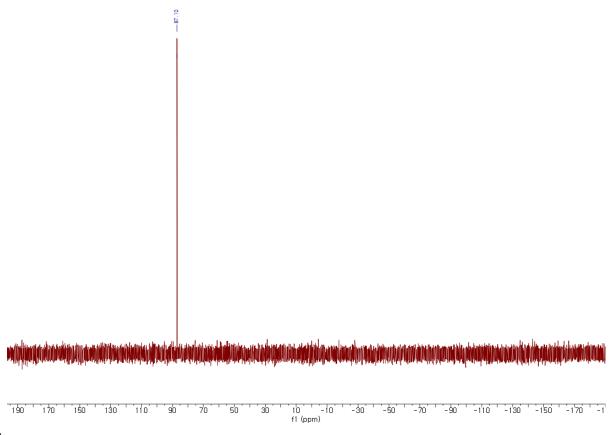
 $^{31}P\{^{1}H\}$ NMR spectrum (162 MHz, CDCl3) of $\boldsymbol{L1}$



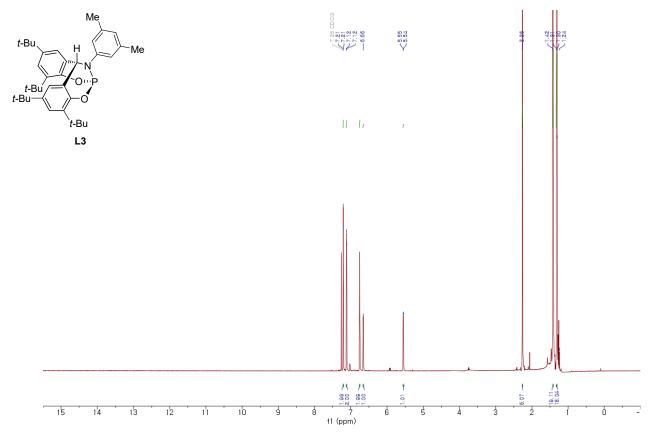
¹H NMR spectrum (400 MHz, CDCl₃) of **L2**



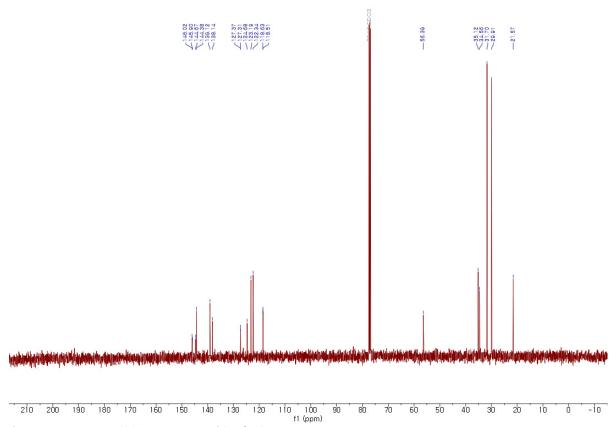
 $^{13}C\{^1H\}$ NMR spectrum (100 MHz, CDCl₃) of $\boldsymbol{L2}$



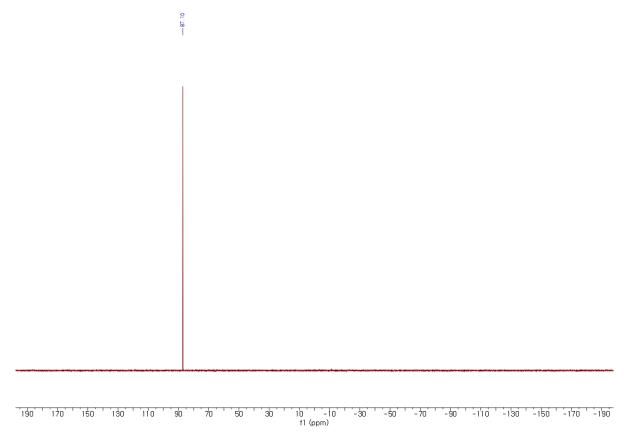
 $^{31}P\{^{1}H\}$ NMR spectrum (162 MHz, CDCl3) of $\boldsymbol{L2}$



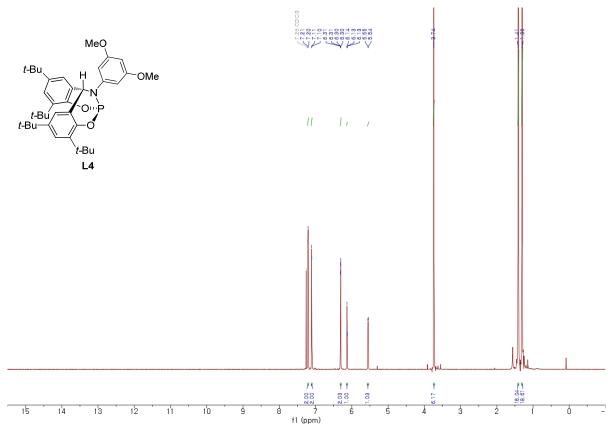
 ^{1}H NMR spectrum (400 MHz, CDCl₃) of **L3**



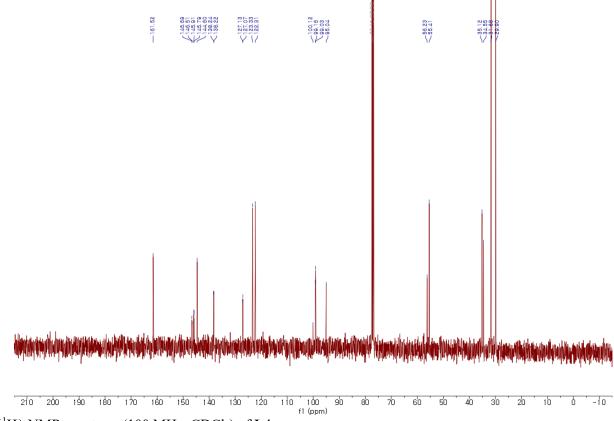
 $^{13}C\{^1H\}$ NMR spectrum (100 MHz, CDCl3) of $\boldsymbol{L3}$



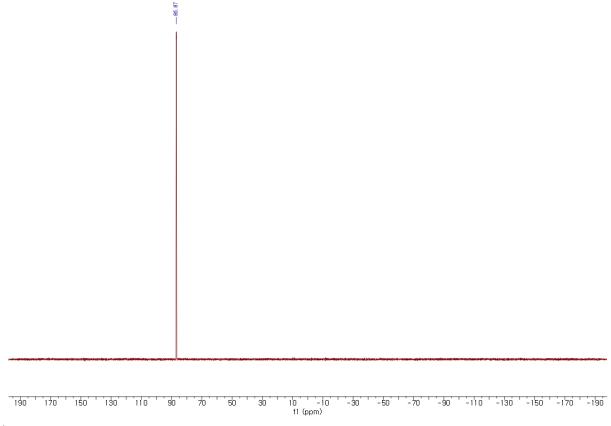
³¹P{¹H} NMR spectrum (162 MHz, CDCl₃) of **L3**



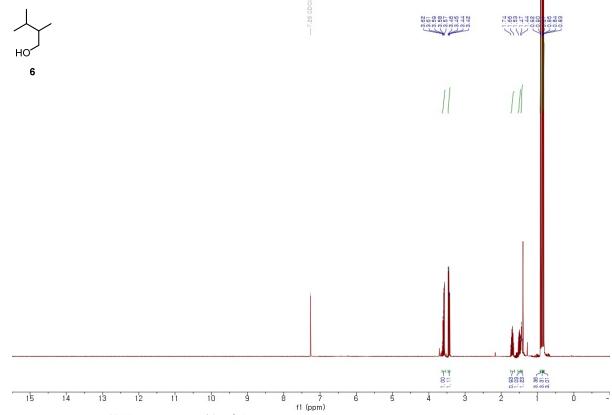
¹H NMR spectrum (400 MHz, CDCl₃) of L4

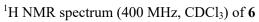


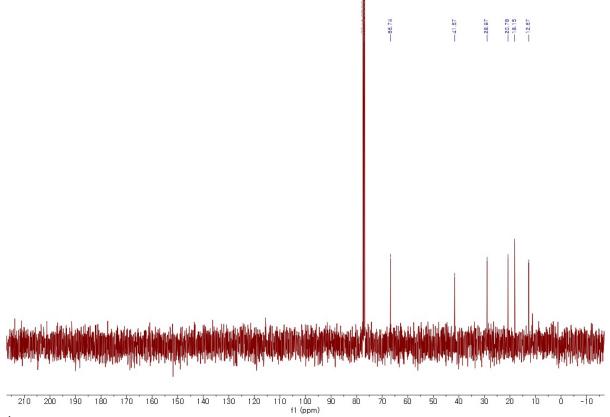
 $^{13}C\{^1H\}$ NMR spectrum (100 MHz, CDCl3) of $\boldsymbol{L4}$



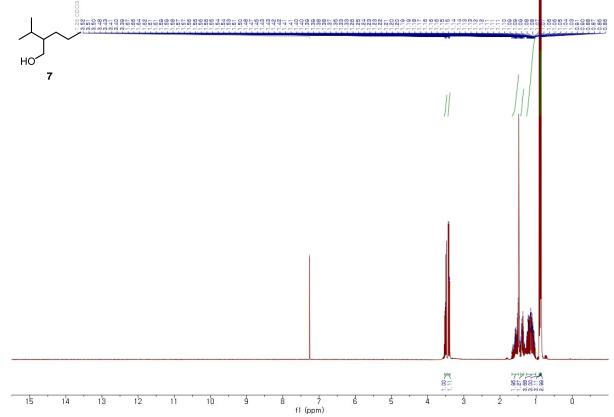
 $^{31}P\{^{1}H\}$ NMR spectrum (162 MHz, CDCl₃) of L4



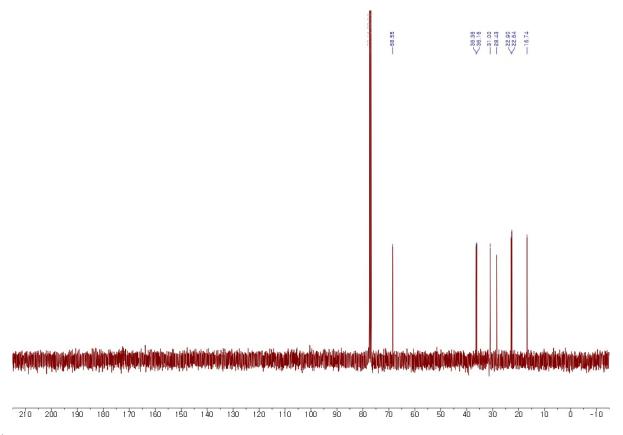




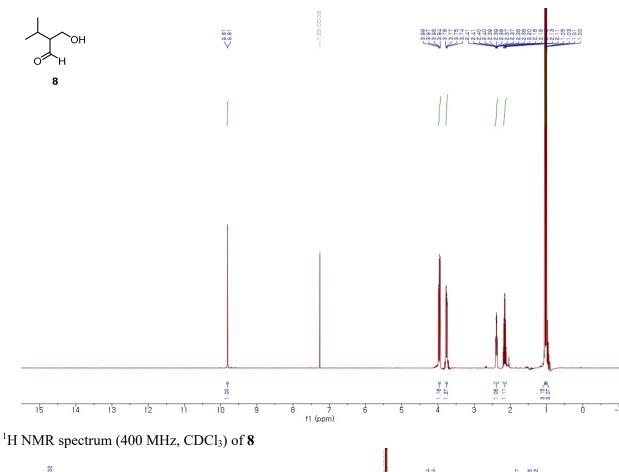
 $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (100 MHz, CDCl₃) of $\boldsymbol{6}$

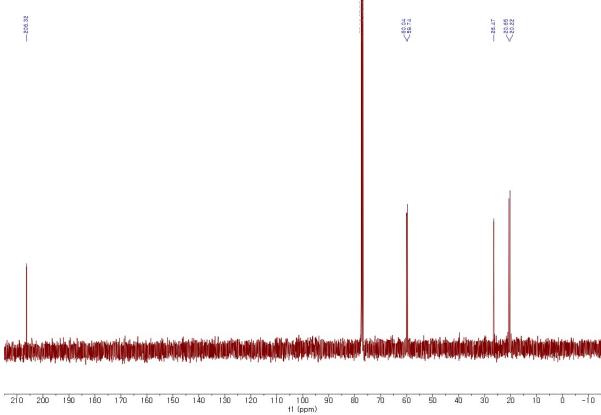


 $^{1}\text{H NMR}$ spectrum (400 MHz, CDCl₃) of 7

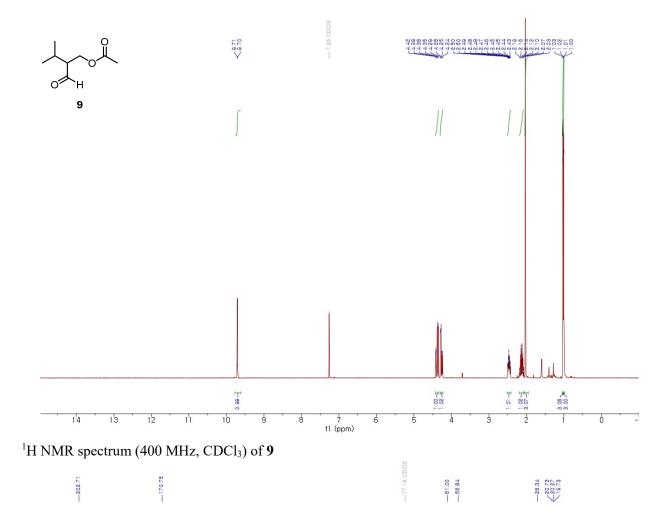


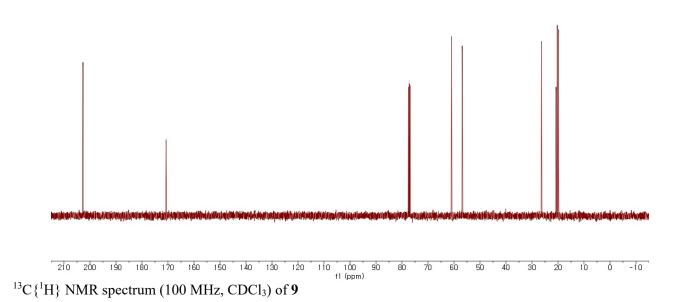
 13 C $\{^{1}$ H $\}$ NMR spectrum (100 MHz, CDCl₃) of 7

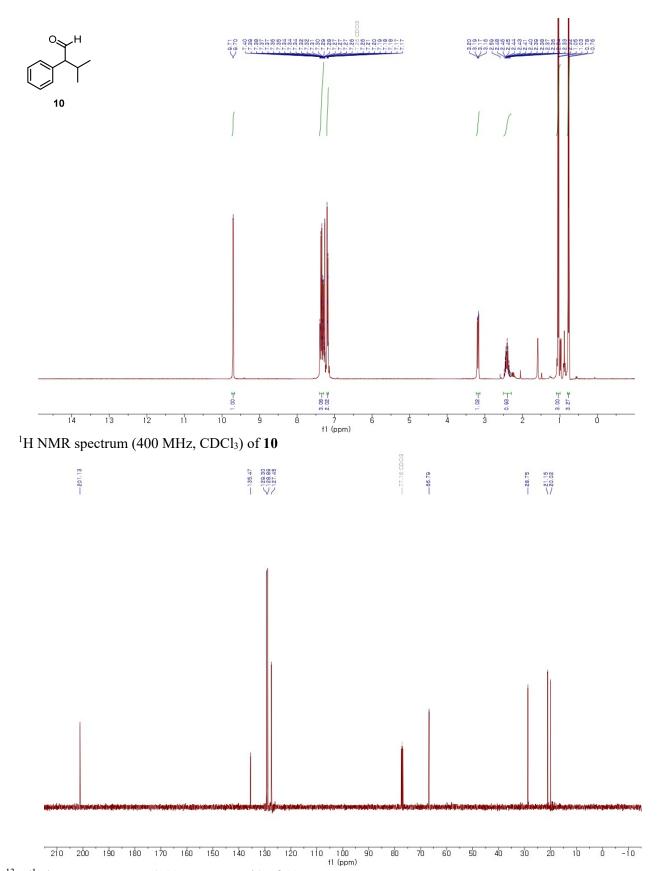




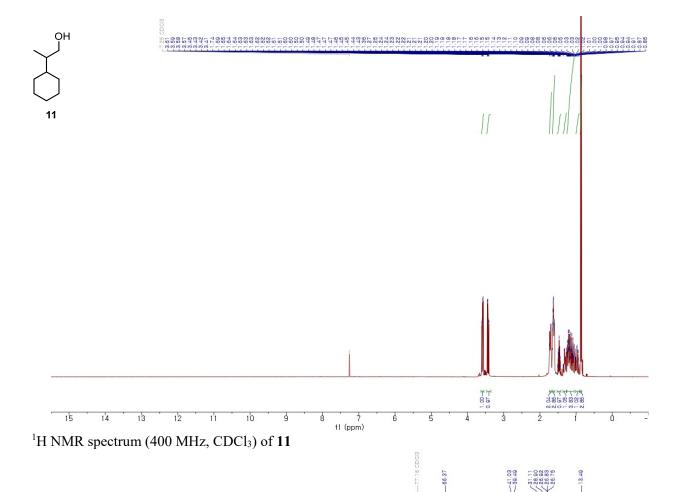
 $^{13}C\{^1H\}$ NMR spectrum (100 MHz, CDCl₃) of $\boldsymbol{8}$

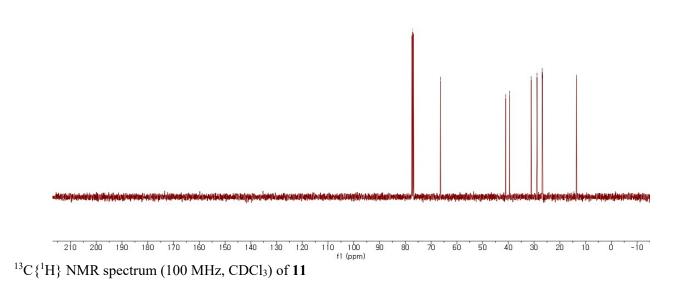


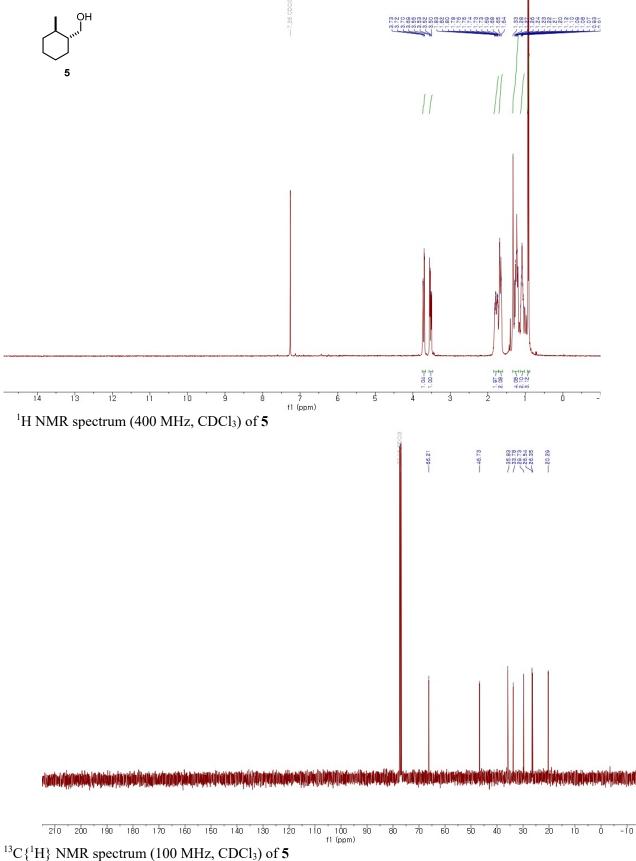


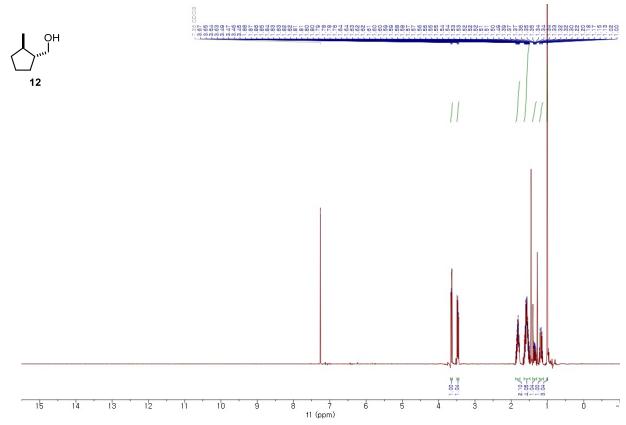


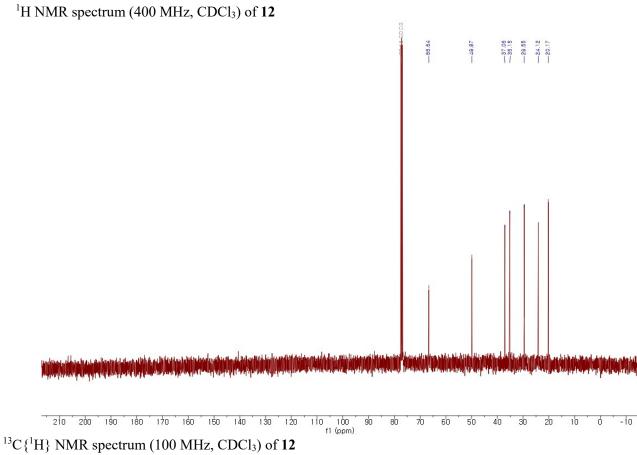
 $^{13}C\{^1H\}$ NMR spectrum (100 MHz, CDCl3) of $\boldsymbol{10}$

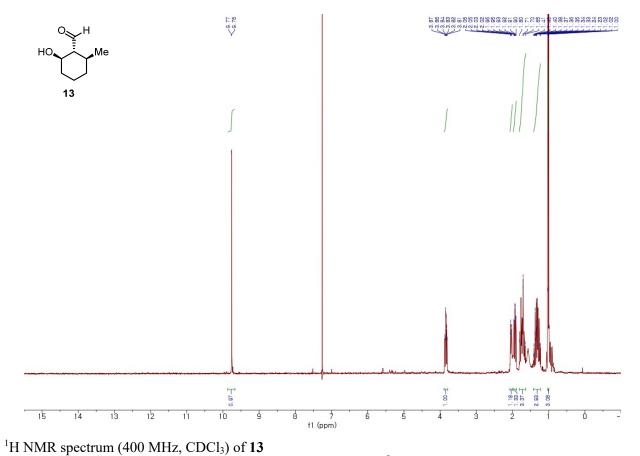


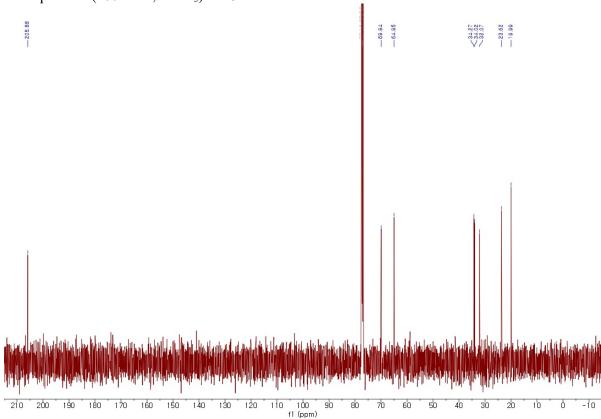


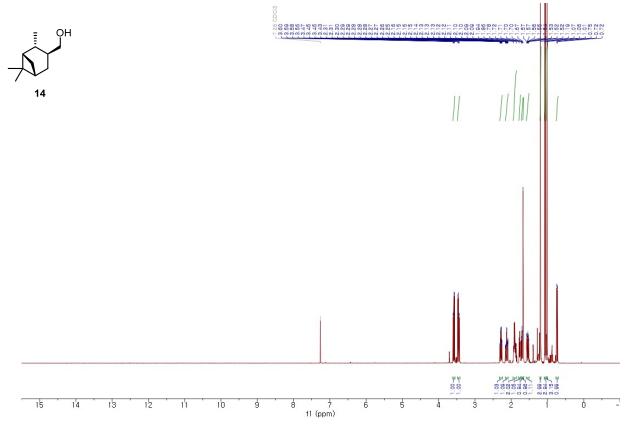






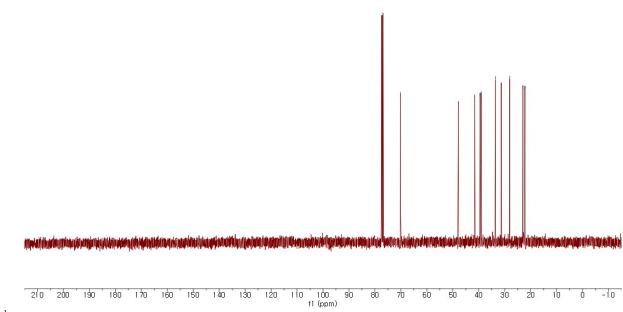






 ^{1}H NMR spectrum (400 MHz, CDCl₃) of **14**





 $^{13}C\{^1H\}$ NMR spectrum (100 MHz, CDCl3) of $\boldsymbol{14}$