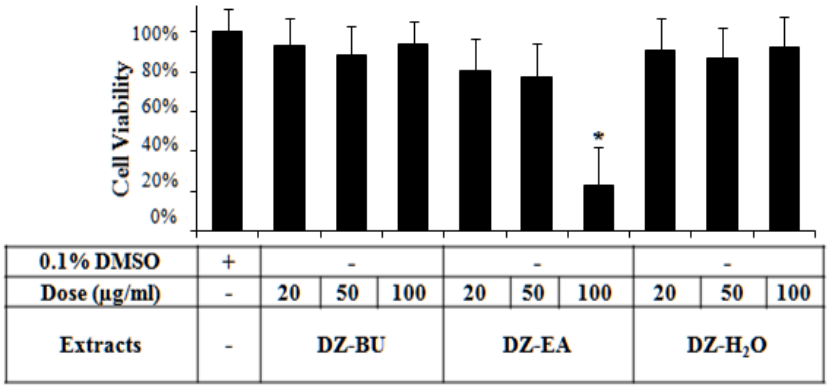
(a)



(b)

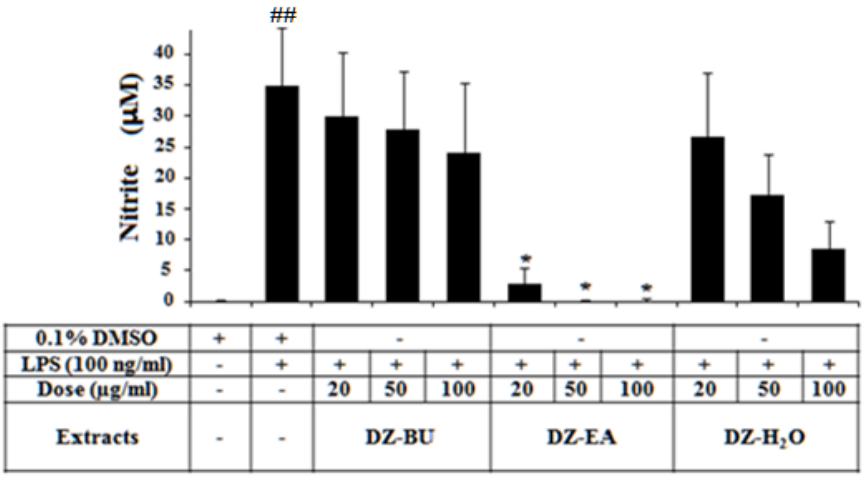
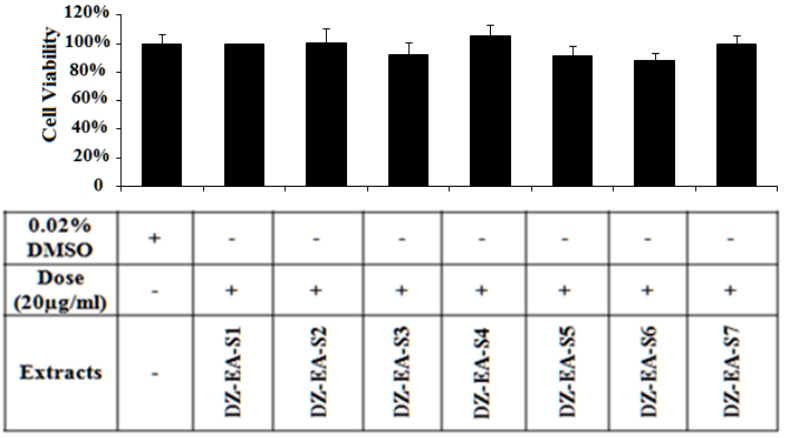


Figure S1 (a) Effect of DZ-BU, DZ-EA and DZ-H_2O on BV-2 cell viability. BV-2 cells were pretreated with 0.1 % DMSO or extracts (20, 50 and 100 μg/ml) for 48 hr. Cell viability of BV-2 cells was determined by MTT assays. All values are presented as mean ± S.D. of three independent experiments. *, $p < 0.05$; compared with DMSO group. **(b) Effect of DZ-BU, DZ-EA and DZ-H_2O on NO production in LPS-stimulated BV-2 cells.** BV-2 cells were pretreated with 0.1% DMSO or extracts (20, 50 and 100 μg/ml) for 2 hr, followed by the stimulation of LPS (100 ng/ml) for another 24 hr. The supernatants were collected for the nitrite measurement using Griess reagent. All data are presented as mean ± S.D. of three independent experiments. $^{\#\#}$, $p < 0.01$; compared with DMSO group; *, $p < 0.05$; compared with DMSO + LPS group.

(a)



(b)

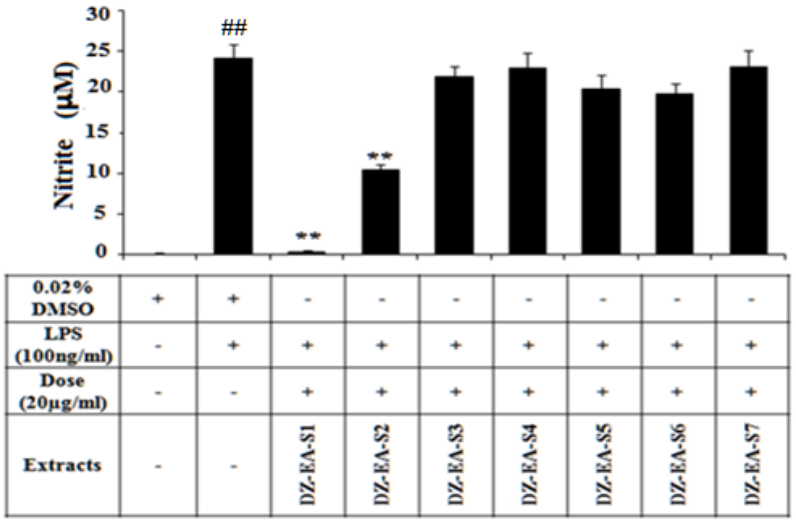
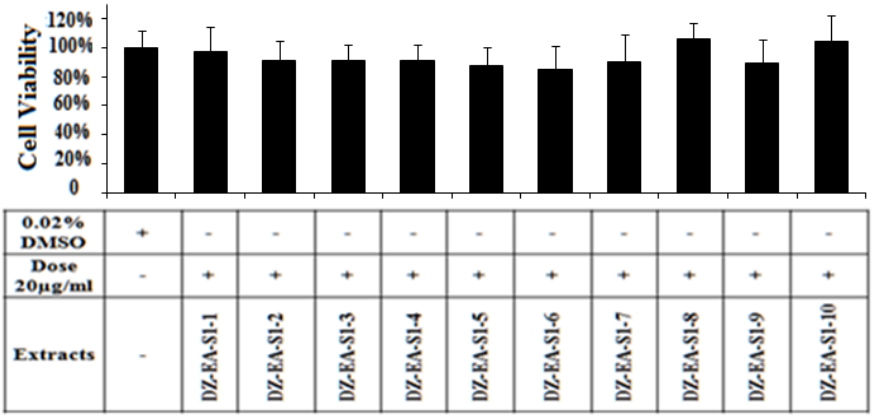


Figure S2 (a) Effect of subfractions of DZ-EA on BV-2 cell viability. BV-2 cells were pretreated with 0.02 % DMSO or subfractions of DZ-EA (20 μg/ml) for 48 hr. Cell viability of BV-2 cells was determined by MTT assays. All values are presented as mean ± S.D. of three independent experiments. **(b) Effect of subfractions of DZ-EA on NO production in LPS-stimulated BV-2 cells.** BV-2 cells were pretreated with 0.02% DMSO or subfractions of DZ-EA (20 μg/ml) for 2 hr, followed by the stimulation of LPS (100 ng/ml) for another 24 hr. The supernatants were collected for the nitrite measurement using Griess reagent. All data are presented as mean ± S.D. of three independent experiments. $^{\#\#}$, $p < 0.01$; compared with DMSO group. **, $p < 0.01$; compared with DMSO + LPS group.

(a)



(b)

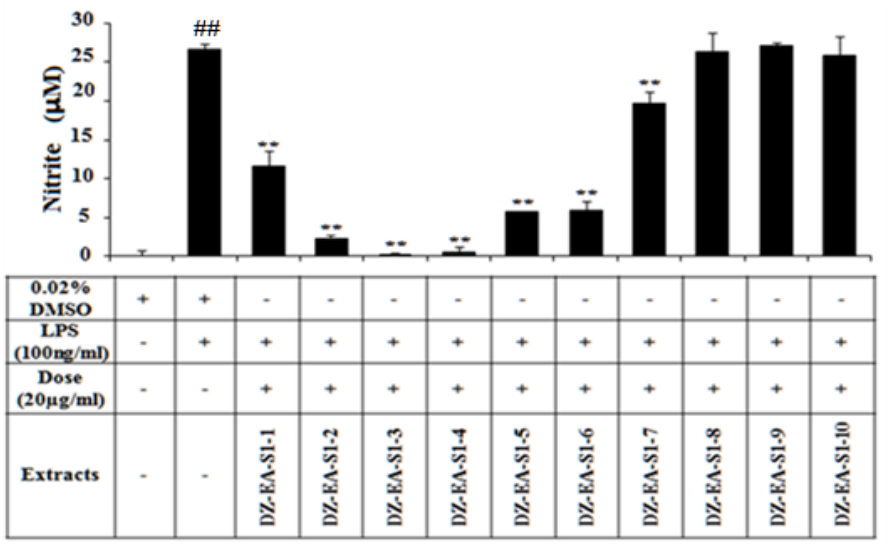


Figure S3 (a) Effect of subfractions of DZ-EA-S1 on BV-2 cell viability. BV-2 cells were pretreated with 0.02 % DMSO or subfractions of DZ-EA-S1 (20 μg/ml) for 48 hr. Cell viability of BV-2 cells was determined by MTT assays. All values are presented as mean ± S.D. of three independent experiments. **(b) Effect of subfractions of DZ-EA-S1 on NO production in LPS-stimulated BV-2 cells.** BV-2 cells were pretreated with 0.02% DMSO or subfractions of DZ-EA-S1 (20 μg/ml) for 2 hr, followed by the stimulation of LPS (100 ng/ml) for another 24 hr. The supernatants were collected for the nitrite measurement using Griess reagent. All data are presented as mean ± S.D. of three independent experiments. $^{\#\#}$, $p < 0.01$; compared with DMSO group. **, $p < 0.01$; compared with DMSO + LPS group.

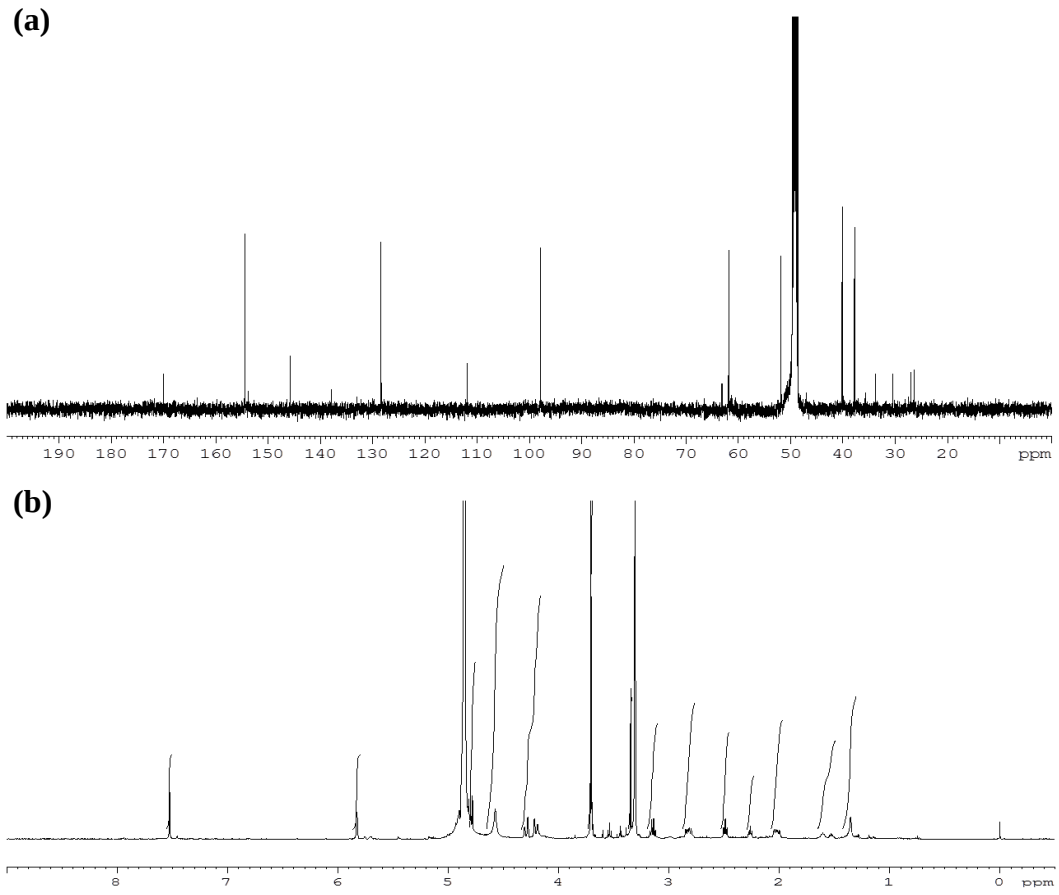


Figure S4 (a) ^{13}C NMR and (b) ^{1}H NMR spectra of compound C. The structure of compound C was elucidated using a Bruker 500-MH_Z PRX NMR spectrometer. Methanol-*d* was used as the solvent. The ^{13}C NMR (125 MHz, CD_3OD) spectrum showed signals at δ170.0 (-CO_2-), 97.8 (C-1), 154.4 (C-3), 111.8 (C-4), 51.8 (C-5), 40.1 (C-6), 128.3 (C-7), 145.7 (C-8), 48.5 (C-9), 61.8 (C-10), 37.7 (C-11). The ^{1}H NMR (500 MHz, CD_3OD) spectrum showed signals at δ4.65 (d, H-1), 7.52 (s, H-3), 3.13 (ddd, H-5), 2.1 (ddt, H-6α), 2.86 (ddt, H-6β), 5.83 (s, H-7), 2.5 (ddd, H-9), 4.25 (d, H-10) and 3.74 (s, H-11).

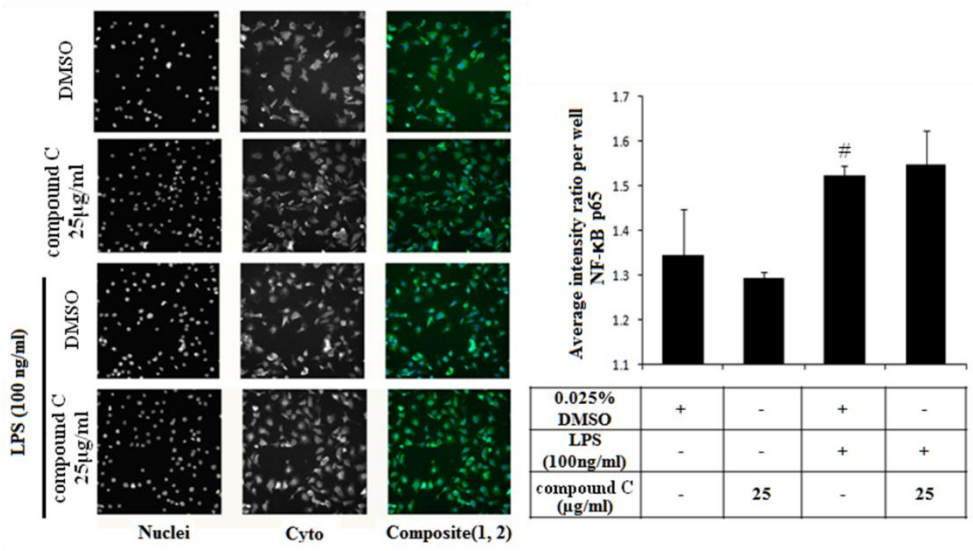


Figure S5 Effect of compound C on NF-κB p65 nuclear translocation in LPS-stimulated BV-2 cells. BV-2 cells were pretreated with compound C (25 μg/ml) for 2 hr, followed by the stimulation of LPS (100 ng/ml) for another 30 min. ICC staining was then performed. Subcellular location of NF-κB p65 subunits were visualized using Cellomics ArrayScan VTI (Thermo Scientific, USA). DAPI was used for nuclei staining. All data are presented as mean ± S.D. of three independent experiments. $^{\#}$, $p < 0.05$; compared with DMSO group.