

***Combining Ring-Opening Multibranching and RAFT
Polymerization: Multifunctional Linear-
Hyperbranched Block Copolymers via
Hyperbranched Macro Chain Transfer Agents***

Supporting Information

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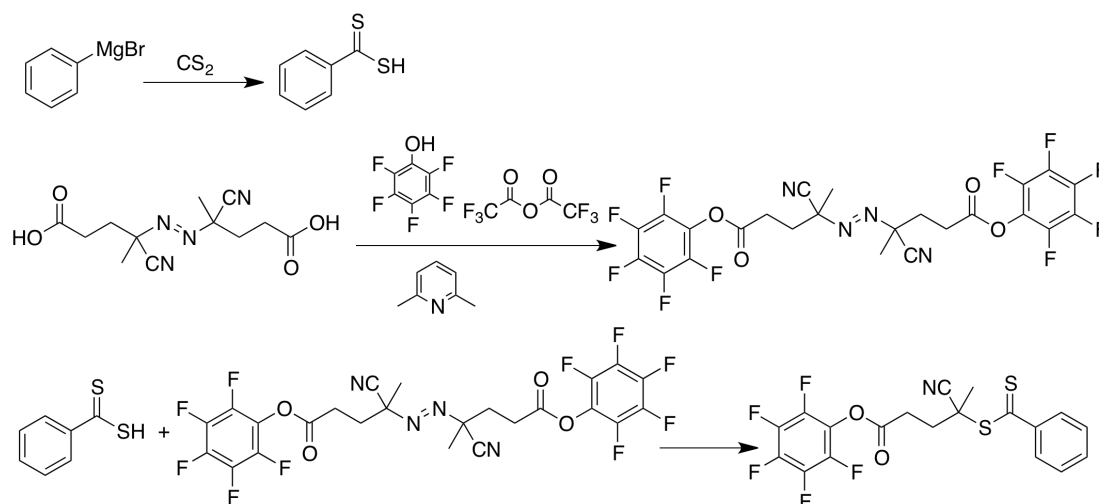
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Syntheses

1. Synthesis of Pentafluorophenyl 4-cyano-4-((phenylcarbonothioyl)thio)pentanoate PFP-CTA

The following procedure was slightly modified from a literature procedure¹:



Scheme S1: Synthesis of Pentafluorophenyl 4-cyano-4-((phenylcarbonothioyl)thio)pentanoate

Dithiobenzoic Acid

In a two-necked round bottom flask equipped with a reflux condenser, a dropping funnel and a magnetic stir bar 1M phenylmagnesium bromide (15 mL; 0.015 mol) was dissolved in anhydrous THF (75 mL) under nitrogen atmosphere. The dropping funnel was filled with carbon disulfide (1.64 mL; 0.027 mol) under nitrogen atmosphere and this component was then dropped slowly to the reaction mixture while stirring vigorously. The reaction mixture turned red brown and was stirred for another two hours at 45°C after complete addition of carbon disulfide, before it was pored on ice (110 g) with conc. HCl (50mL). The suspension changed its color to purple and was extracted with diethyl ether (100 mL) three times. All organic phases were combined, dried over MgSO₄ and concentrated by rotary evaporation affording dithiobenzoic acid (2.36 g; 0.015 mol; quantitative) as a violet liquid of characteristic odor.

¹H-NMR (dioxane-d⁸, 300 MHz): δ [ppm] = 8.07 (d, *J* = 6 Hz, 2H, o-ArH); 7.61 (t, *J* = 6 Hz, 1H, p-ArH); 7.41 (t, *J* = 6 Hz, 2H, m-ArH).

Bis(pentafluorophenyl) 4,4'-(diazene-1,2-diyl)bis(4-cyanopentanoate)

In a two-necked round bottom flask equipped with septums and a magnetic stir bar pentafluorophenol (17.2 g; 0.093 mol), azobis(4-cyanovaleric acid) (10.9 g; 0.039 mol) and 2,6-lutidine (34.4 mL; 0.295 mol) were dissolved in anhydrous dichloromethane (225 mL) under nitrogen atmosphere. While cooling with an ice bath and stirring vigorously trifluoroacetic anhydride (16.2 mL; 0.116 mol) was added slowly via syringe through the septum. After

complete addition the mixture was allowed to warm to room temperature within 12 h while stirring. It was subsequently extracted three times with water (100 mL) and the organic phase dried over MgSO_4 and concentrated by rotary evaporation at 25°C . The raw product was dissolved in dichloromethane and precipitated in cold n-hexane to remove remaining lutidine. This process had to be repeated six times affording bis(pentafluorophenyl) 4,4'-(diazene-1,2-diyl)bis(4-cyanopentanoate) (7.85 g; 0.013 mol; 33%) as a colorless powder.

$^1\text{H-NMR}$ (CDCl_3 , 300 MHz): δ [ppm] = 2.96-2.54 (m, 8H, CH_2); 1.80 (s, 6H, CH_3).

$^{19}\text{F-NMR}$ (CDCl_3 , 376 MHz): δ [ppm] = -152.85 (d, J = 15 Hz, 2F, o- ArF); -157.54 (t, J = 22 Hz, 1F, p- ArF); -162.19 (t, J = 19 Hz, 2F, m- ArF).

Pentafluorophenyl 4-cyano-4-((phenylcarbonothioyl)thio)pentanoate PFP-CTA

In a two-necked round bottom flask equipped with a reflux condenser, a septum and a magnetic stir bar dithiobenzoic acid (2.20 g; 0.014 mol) and bis(pentafluorophenyl) 4,4'-(diazene-1,2-diyl)bis(4-cyanopentanoate) (7.10 g; 0.011 mol) were dissolved in ethyl acetate (120 mL) under nitrogen atmosphere. The solution was bubbled with argon for one hour to deoxygenate the reaction, before it was refluxed for 12 h under nitrogen atmosphere. The reaction mixture was then concentrated by rotary evaporation and purified by column chromatography (elution chloroform: cyclohexane = 3:1) affording pentafluorophenyl 4-cyano-4-((phenylcarbonothioyl)thio)pentanoate **PFP-CTA** as a purple solid (1.69 g; 0.004 mol; 72%).

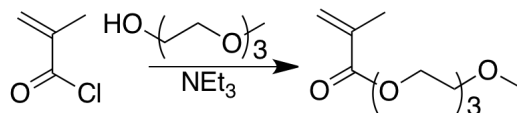
$^1\text{H-NMR}$ (CDCl_3 , 300 MHz): δ [ppm] = 7.93 (d, J = 6 Hz, 2H, o- ArH); 7.59 (t, J = 7.5 Hz, 1H, p- ArH); 7.42 (t, J = 7.5 Hz, 2H, m- ArH); 3.10-3.03 (m, 2H, $-\text{CH}_2-\text{CH}_2-\text{CO}-$); 2.82-3.72 (m, 1H, $-\text{CH}_2-\text{CH}_2-\text{CO}-$); 2.61-2.51 (m, 1H, $-\text{CH}_2-\text{CH}_2-\text{CO}-$); 1.99 (s, 3H, $-\text{CH}_3$).

$^{13}\text{C-NMR}$ (CDCl_3 , 75 MHz): δ [ppm] = 219.91 ($-\text{CS}_2-$); 167.92 ($-\text{COO}-$); 144.47 ($\text{H}_5\text{ArC-O}$); 142.96-142.81 (m- F_5ArC , multiplet); 141.70-140.34 ($\text{F}_5\text{ArC-O}$, multiplet); 139.74-139.47 (p- F_5ArC , multiplet); 136.38-136.22 and 134.16-133.37 (o- F_5ArC , multiplet); 133.37 (p- H_5ArC); 128.80 (m- H_5ArC); 126.86 (o- H_5ArC); 118.38 ($-\text{CN}$); 45.64 (C_{quat}); 33.15 ($-\text{CH}_2-\text{CH}_2-\text{CO}-$); 29.24 ($-\text{CH}_2-\text{CH}_2-\text{CO}-$); 24.44 ($-\text{CH}_3$).

$^{19}\text{F-NMR}$ (CDCl_3 , 376 MHz): δ [ppm] = -152.85 (d, J = 19 Hz, 2F, o- ArF); -157.59 (t, J = 21 Hz, 1F, p- ArF); -162.20 (t, J = 19 Hz, 2F, m- ArF).

2. Synthesis of Tri(ethylene glycol) methyl ether methacrylate MEO₃MA

The following procedure was slightly modified from a literature procedure²:



Scheme S2: Synthesis of Tri(ethylene glycol) methyl ether methacrylate

Tri(ethylene glycol) methyl ether methacrylate MEO₃MA

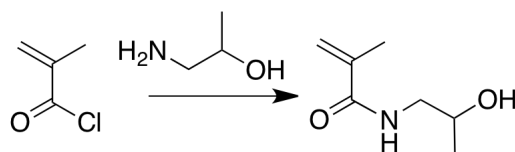
In a round bottom flask equipped with a dropping funnel and a magnetic stir bar tri(ethylene glycol) monomethyl ether (29.4 mL; 0.188 mol) was dissolved in anhydrous dichloromethane (90 mL) under nitrogen atmosphere. Triethylamine (50.5 mL; 0.363 mol) was added via syringe through a septum. The dropping funnel was filled with methacryloyl chloride (10.6 mL; 0.125 mol) under nitrogen atmosphere and this component was then dropped slowly to the reaction mixture while stirring vigorously and cooling with an ice bath. After complete addition the reaction mixture was cooled for further 15 min and then slowly warmed to room temperature for about 12 h. It was filtered through filter paper to remove precipitated salts and then concentrated by rotary evaporation. The crude mixture was purified by column chromatography (gradient elution cyclohexane : ethyl acetate from 6:1 to 5:1) affording tri(ethylene glycol) methyl ether methacrylate **MEO₃MA** as a colorless liquid (11.0 g; 0.047 mol; 38%). Due to educt contamination di(ethylene glycol) methyl ether methacrylate could be isolated as a byproduct (2.0 g; 0.011; 9%).

¹H-NMR (CDCl₃, 300 MHz): δ [ppm] = 6.08 (m, 1H, C=CH₂-*cis*); 5.52 (m, 1H, C=CH₂ *trans*); 4.25 (t, *J* = 5 Hz, 2H, COO-CH₂-CH₂-); 3.70 (t, *J* = 5 Hz, 2H, COO-CH₂-CH₂-); 3.63 -3.58 (m, 6H, O-CH₂-CH₂-O-CH₂-CH₂-O-CH₃); 3.49 (t, *J* = 6 Hz, 2H, CH₂-CH₂-O-CH₃); 3.32 (s, 3H, O-CH₃); 1.89 (s, 3H, =C-CH₃).

¹³C-NMR (CDCl₃, 75 MHz): δ [ppm] = 167.32 (-CO₂-); 136.17 (C=CH₂); 125.70 (C=CH₂); 71.93 (CH₂-CH₂-O-CH₃); 70.65 (O-CH₂-CH₂-O-CH₂-CH₂-O-CH₃); 70.63 (O-CH₂-CH₂-O-CH₂-CH₂-O-CH₃); 70.59 (O-CH₂-CH₂-O-CH₂-CH₂-O-CH₃); 69.14 (COO-CH₂-CH₂-); 63.88 (COO-CH₂-CH₂-); 59.02 (O-CH₃); 18.30 (=C-CH₃).

3. Synthesis of 2-Hydroxypropyl methacrylamide HPMA

The following procedure was slightly modified from a literature procedure³:



Scheme S3: Synthesis of 2-Hydroxypropyl methacrylamide

2-Hydroxypropyl methacrylamide HPMA

In a round bottom flask equipped with a dropping funnel and a magnetic stir bar 1-aminopropan-2-ol (10.0 mL; 0.129 mol) was dissolved in anhydrous dichloromethane (100 mL) under nitrogen atmosphere. The dropping funnel was filled with methacryloyl chloride (5.8 mL; 0.059 mol) under nitrogen atmosphere and this component was then dropped slowly to the reaction mixture while stirring vigorously and cooling with an ice bath. After complete addition the reaction mixture was slowly warmed to room temperature for about 12 h. It was then filtered through filter paper to remove precipitated salts and then concentrated by rotary evaporation. The crude mixture was purified by column chromatography (elution chloroform : methanol = 10:1) affording 2-hydroxypropyl methacrylamide **HPMA** (7.0 g; 0.049 mol; 83%) as colorless powder.

¹H-NMR (CDCl₃, 300 MHz): δ [ppm] = 6.43 (br, 1H, -CO-NH-); 5.72 (m, 1H, C=CH₂ *trans*); 5.34 (m, 1H, C=CH₂ *trans*); 3.94 (dq, J = 7.6, 6.3, 3.0 Hz, 1H, -CH(OH)-); 3.49 (ddd, J = 13.9, 6.5, 3.0 Hz, 1H, -CO-NH-CH₂-); 3.16 (ddd, J = 13.9, 7.6, 5.1 Hz, 1H, -CO-NH-CH₂-); 3.04 (br, 1H, -OH); 1.96 (s, 3H, =C-CH₃); 1.19 (d, J = 6.3 Hz, 1H, CH(OH)-CH₃).

¹³C-NMR (CDCl₃, 75 MHz): δ [ppm] = 169.60 (-CO₂-); 139.72 (C=CH₂); 120.27 (C=CH₂); 67.56 (CH(OH)); 47.30 (-CO-NH-CH₂-); 21.12 (CH(OH)-CH₃); 18.78 (=C-CH₃).

Characterization Data

1. Spectroscopic Data of Pentafluorophenyl 4-cyano-4-((phenylcarbonothioyl)thio)pentanoate PFP-CTA

¹H-NMR PFP-CTA

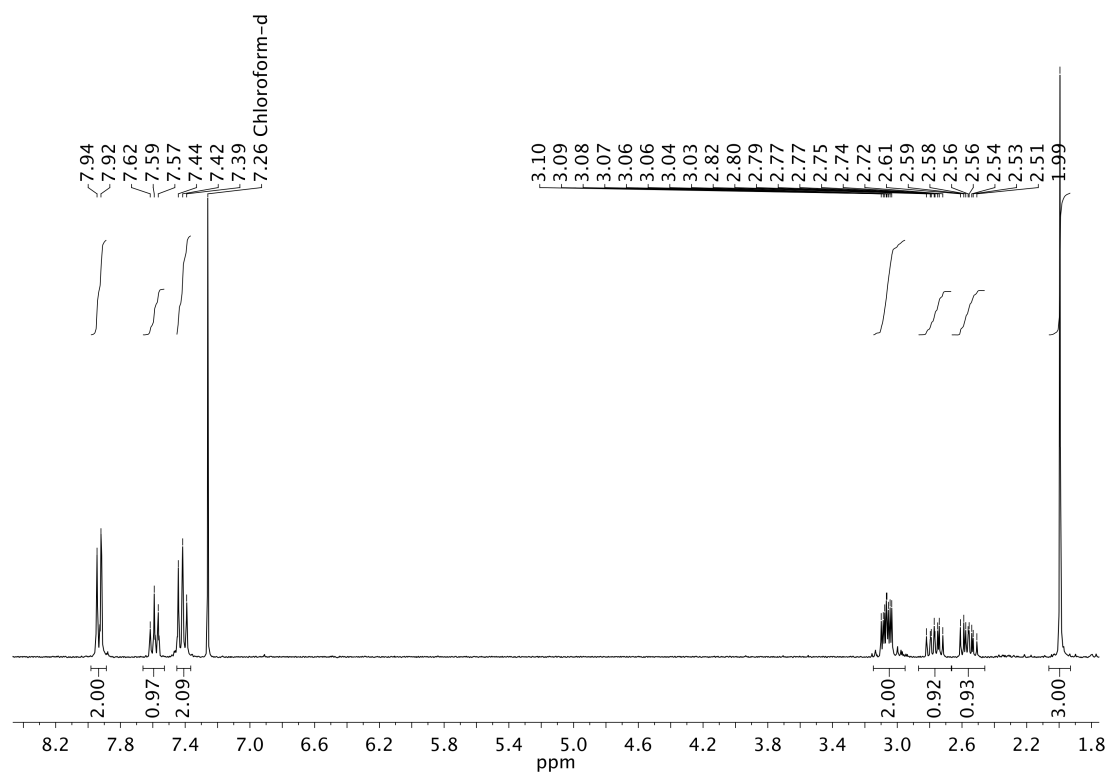


Figure S1: ¹H-NMR (CDCl₃, 300 MHz) Pentafluorophenyl 4-cyano-4-((phenylcarbonothioyl)thio)pentanoate

^{13}C -NMR PFP-CTA

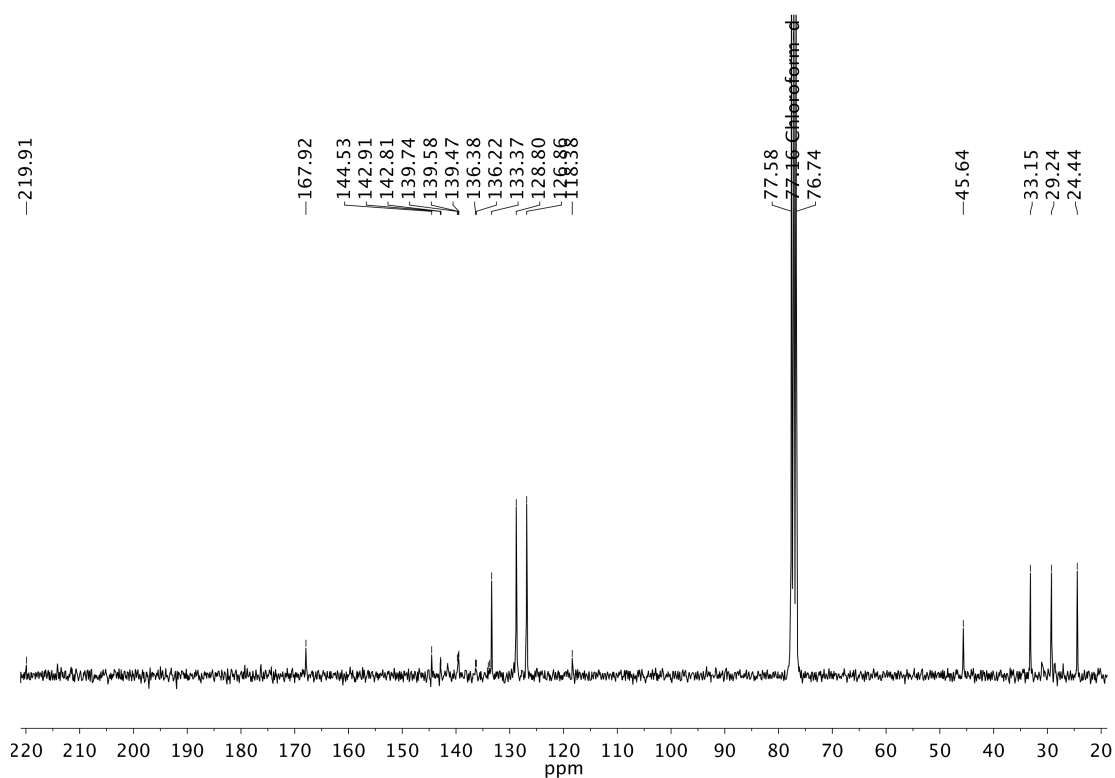


Figure S2: ^{13}C -NMR (CDCl_3 , 75 MHz) Pentafluorophenyl 4-cyano-4-((phenylcarbonothioyl)thio)pentanoate

^{19}F -NMR PFP-CTA

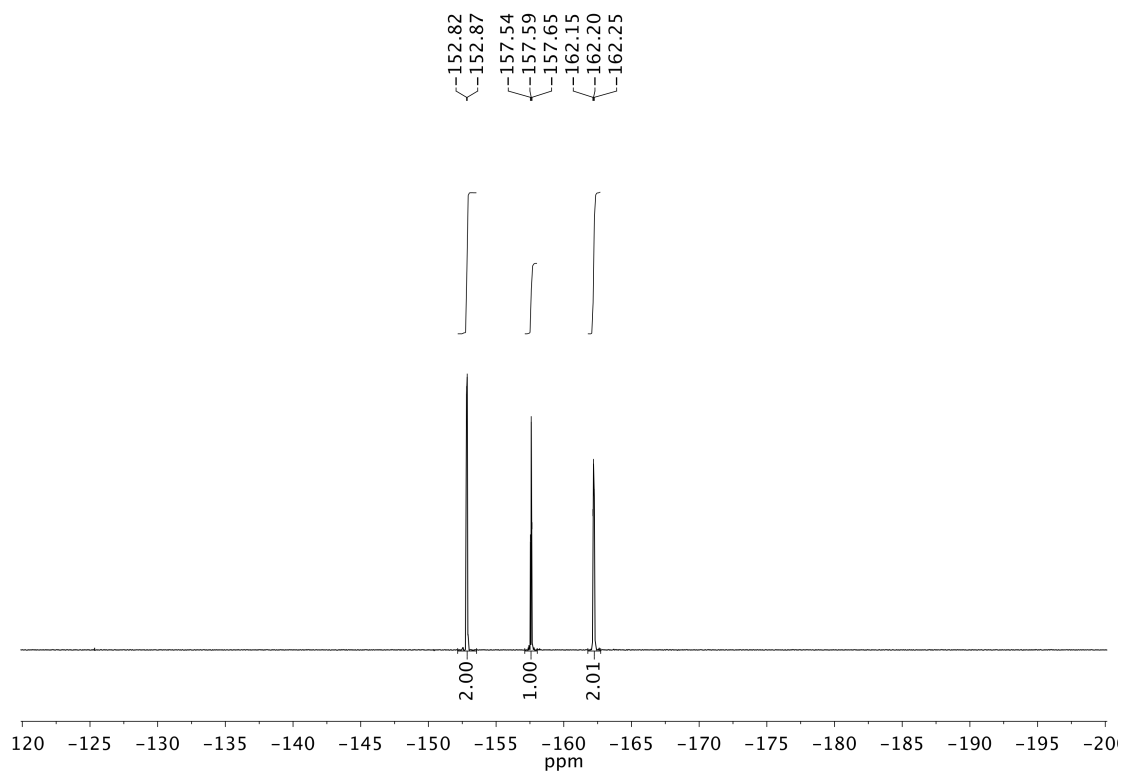


Figure S3: ^{19}F -NMR (CDCl_3 , 376 MHz) Pentafluorophenyl 4-cyano-4-((phenylcarbonothioyl)thio)pentanoate

2. Spectroscopic Data of Tri(ethylene glycol) methyl ether methacrylate MEO₃MA

¹H-NMR MEO₃MA

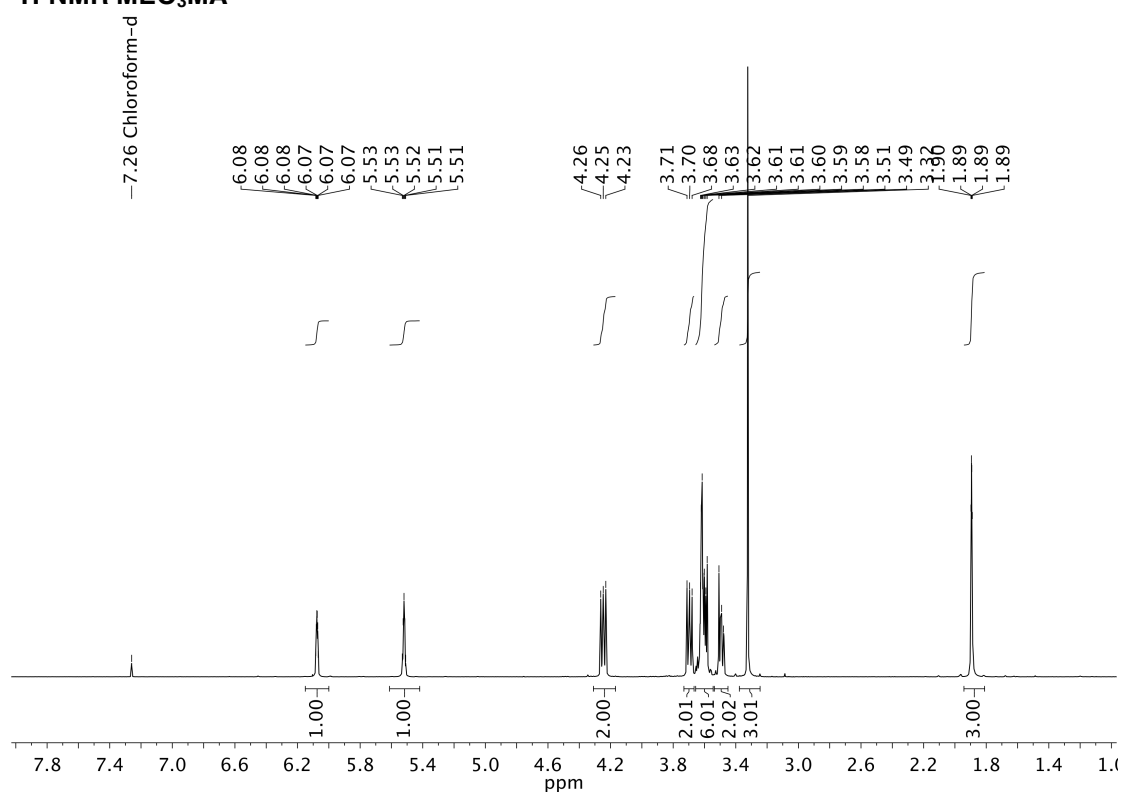


Figure S4: ¹H-NMR (CDCl₃, 300 MHz) Tri(ethylene glycol) methyl ether methacrylate

¹³C-NMR MEO₃MA

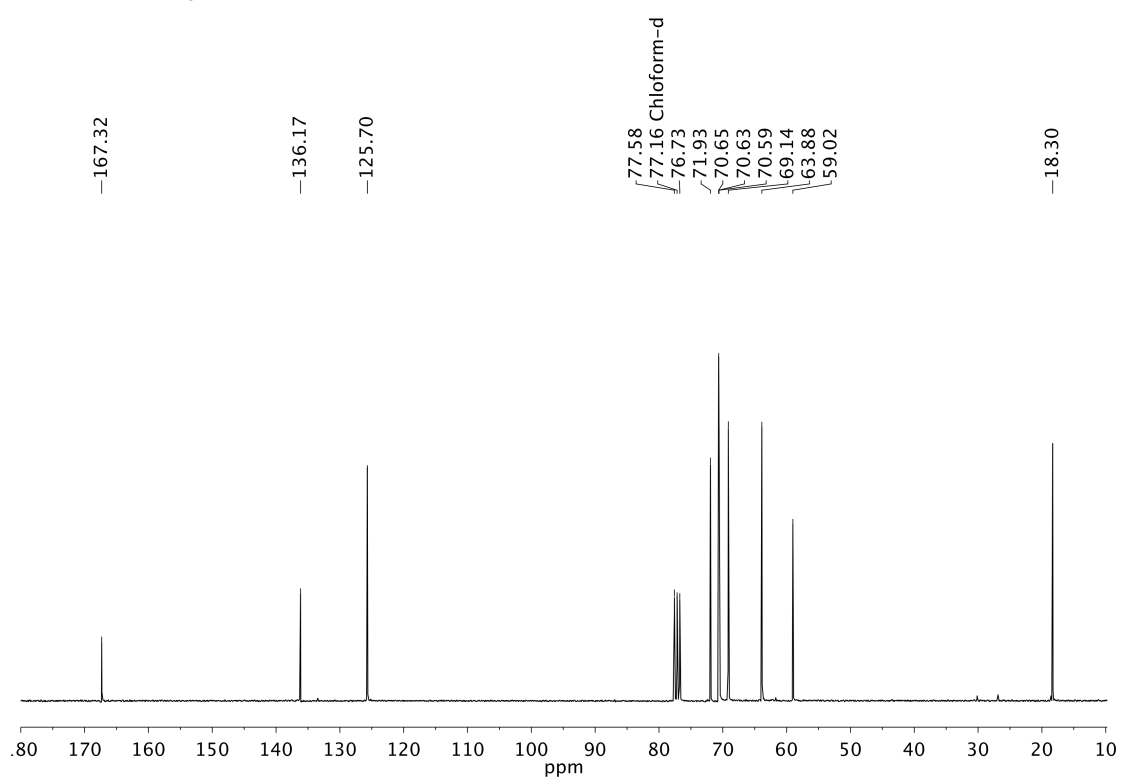


Figure S5: ¹³C-NMR (CDCl₃, 75 MHz) Tri(ethylene glycol) methyl ether methacrylate

3. Spectroscopic Data of 2-Hydroxypropyl methacrylamide HPMA

¹H-NMR HPMA

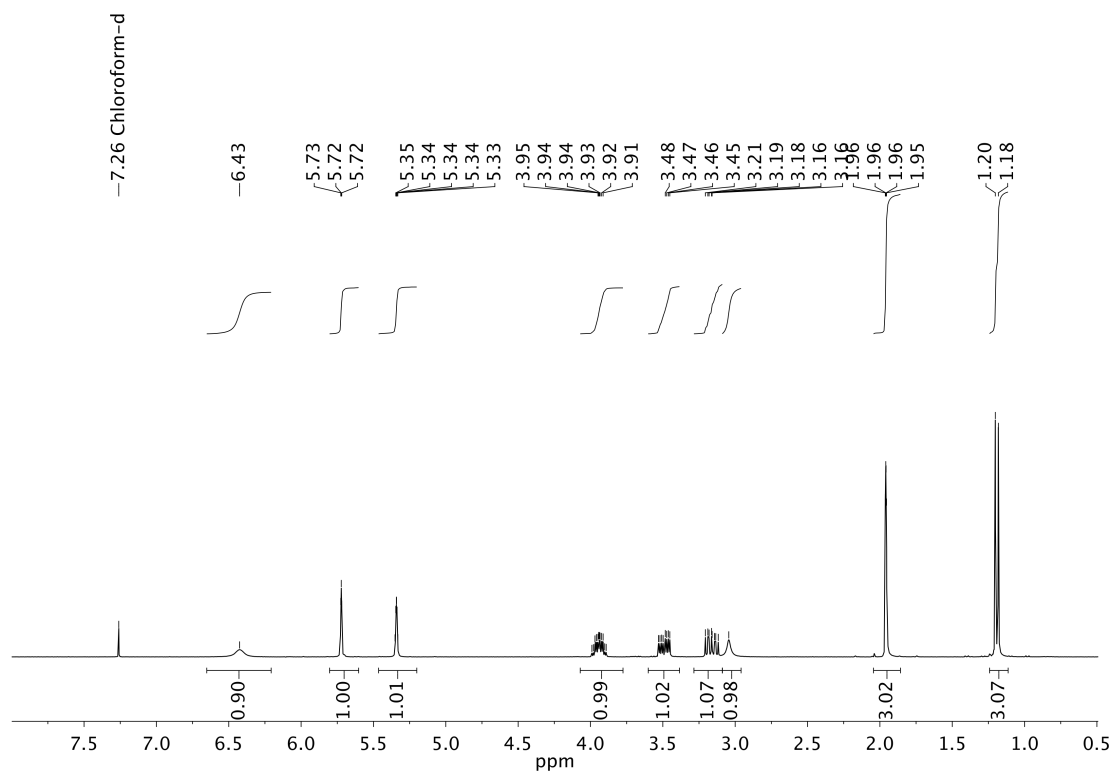


Figure S6: ¹H-NMR (CDCl₃, 300 MHz) 2-Hydroxypropyl methacrylamide

¹³C-NMR HPMA

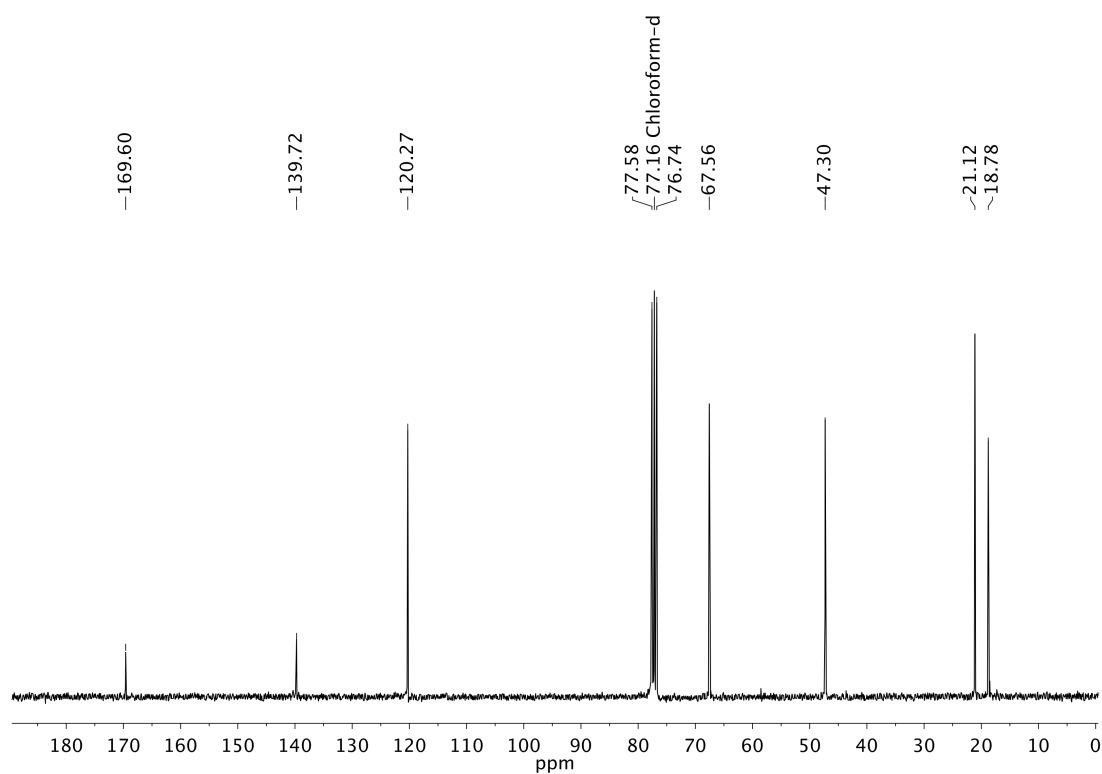


Figure S7: ¹³C-NMR (CDCl₃, 75 MHz) 2-Hydroxypropyl methacrylamide

4. Spectroscopic Data of Hyperbranched Polyglycerol Dendrons

$^1\text{H-NMR}$ hbPG-NBn₂

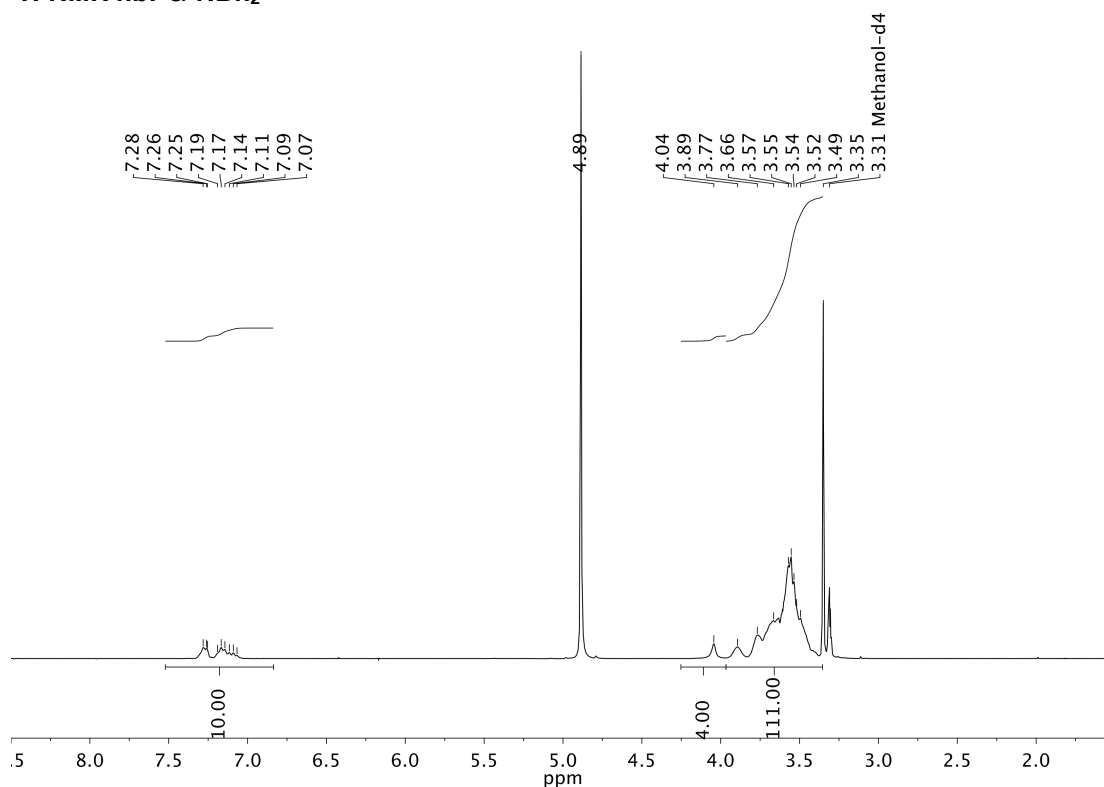


Figure S8: $^1\text{H-NMR}$ (CD₃OD, 400 MHz) hbPG₂₁-NBn₂

Inverse Gated $^{13}\text{C-NMR}$ hbPG-NBn₂

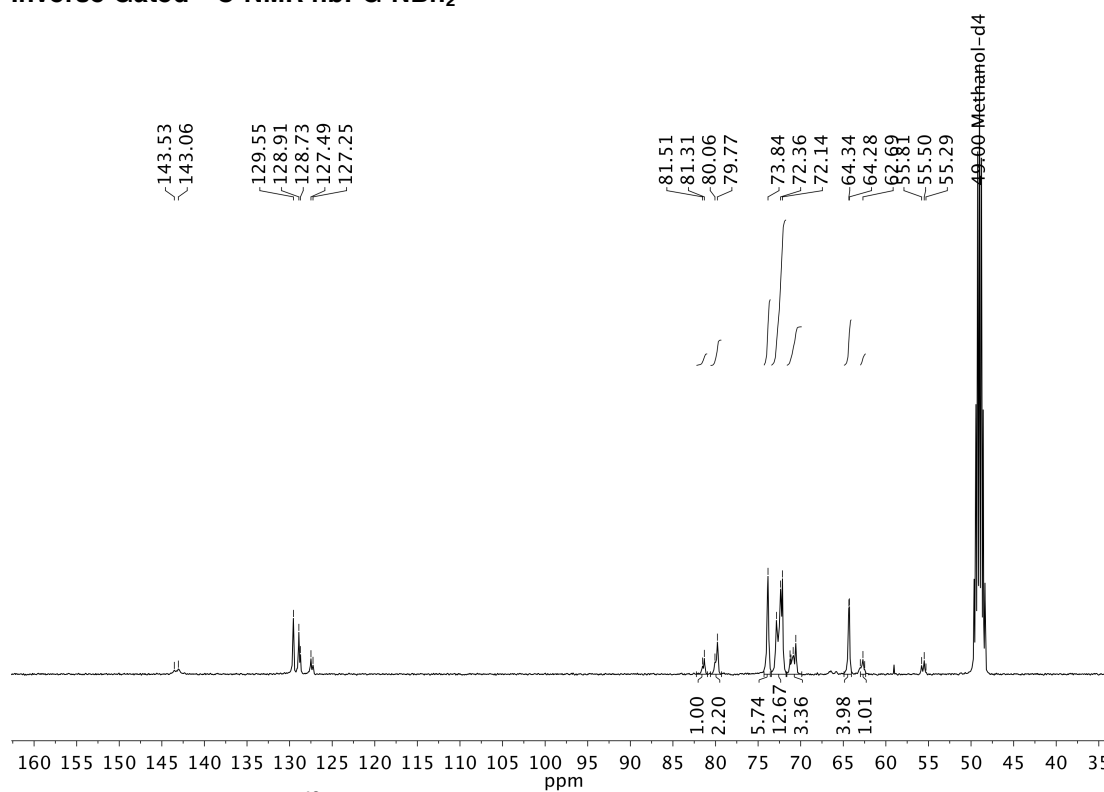


Figure S9: Inverse Gated $^{13}\text{C-NMR}$ (CD₃OD, 100 MHz) hbPG₂₁-NBn₂

Degree of Branching by Inverse Gated ^{13}C -NMR of hbPG-NBn₂

Polymer	L₁₃	L₁₄	D	T	DB*
Bn ₂ TRIS-hbPG ₂₁	0.10	0.29	0.22	0.39	0.53
Bn ₂ TRIS-hbPG ₃₂	0.10	0.28	0.27	0.35	0.59
Bn ₂ TRIS-hbPG ₉₁	0.10	0.28	0.26	0.36	0.58

Table S1: Inverse Gated ^{13}C NMR (100MHz, CD₃OD) characterization data of benzyl protected hyperbranched polyglycerol dendron analogues – Repeat Units: L₁₃=linear-1,3, L₁₄: linear-1,4, D: dendritic, T: terminal.⁴ *: Degree of branching (DB) determined using the established equation $\text{DB} = 2\text{D}/(2\text{D} + \text{L}_{13} + \text{L}_{14})$.⁵

$^1\text{H-NMR}$ hbPG-NH₂

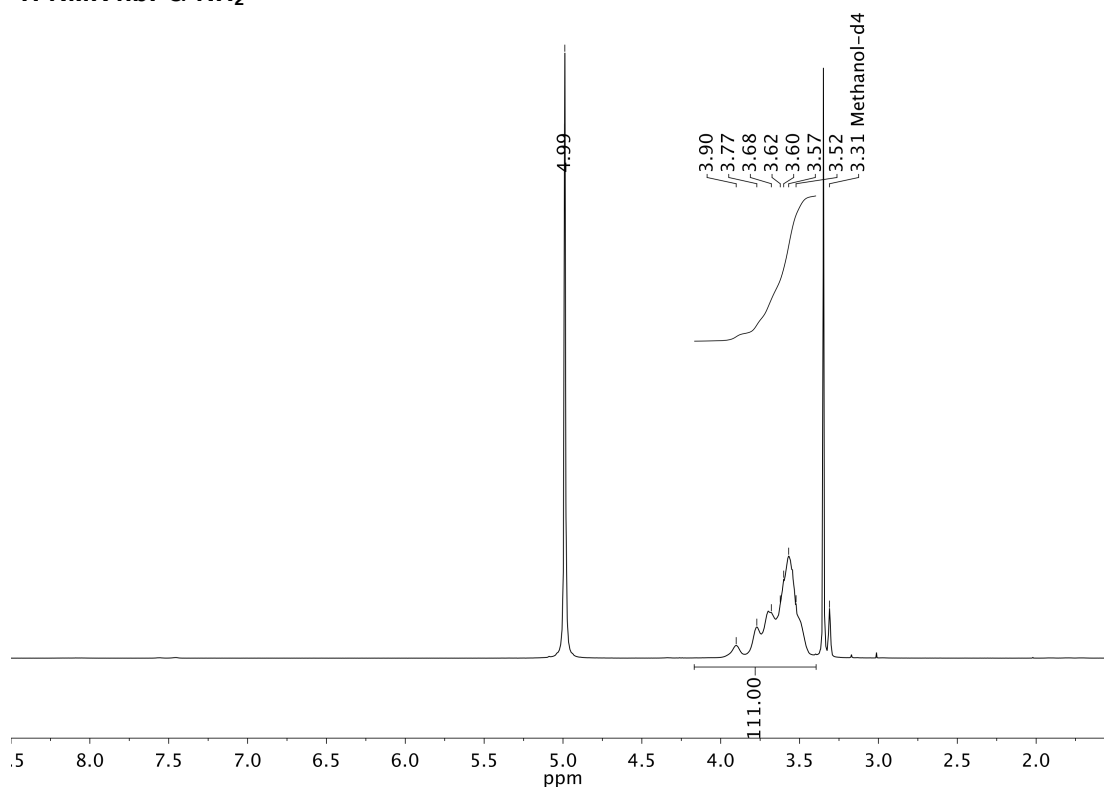


Figure S10: $^1\text{H-NMR}$ (CD₃OD, 400 MHz) hbPG₂₁-NH₂

$^1\text{H-NMR}$ hbPG-CTA

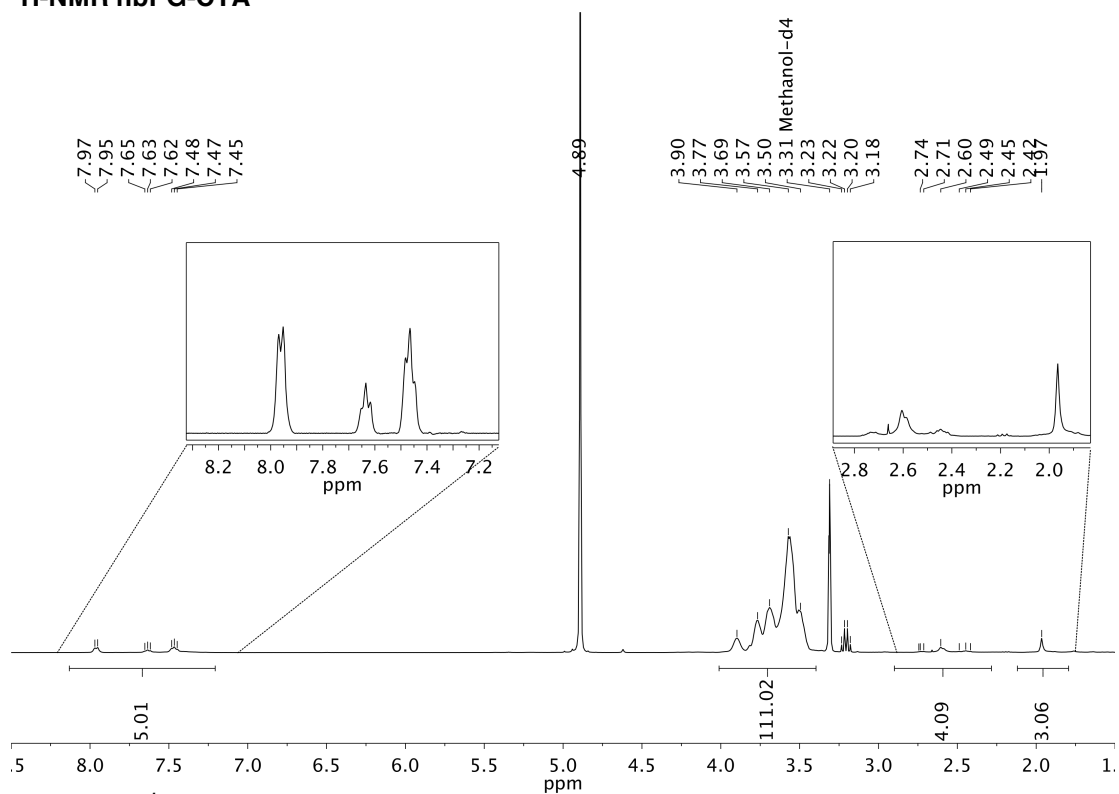


Figure S11: $^1\text{H-NMR}$ (CD₃OD, 400 MHz) hbPG₂₁-CTA

5. Chromatographic and Spectroscopic Data of Linear-Hyperbranched Polyglycerol Block Copolymers

¹H-NMR hbPG-*b*-P(MEO₃MA)

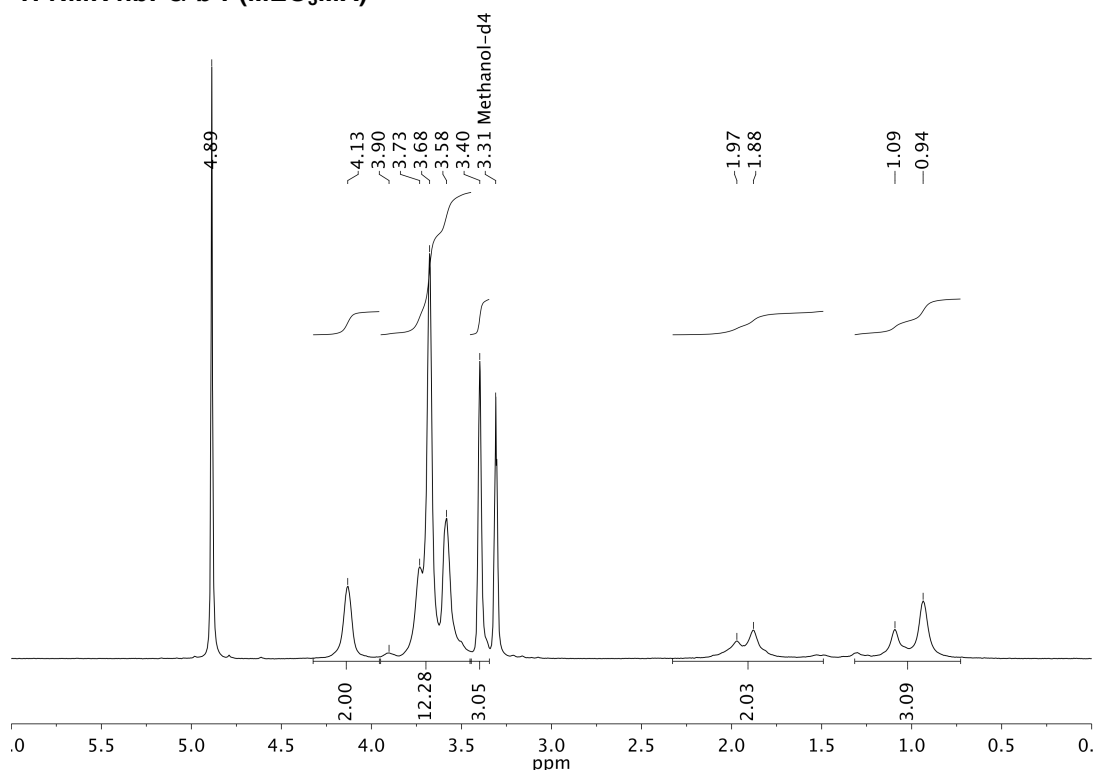


Figure S12: ¹H-NMR (CD₃OD, 400 MHz) hbPG₂₁-*b*-P(MEO₃MA)₄₆

¹H-NMR hbPG-*b*-P(HPMA)

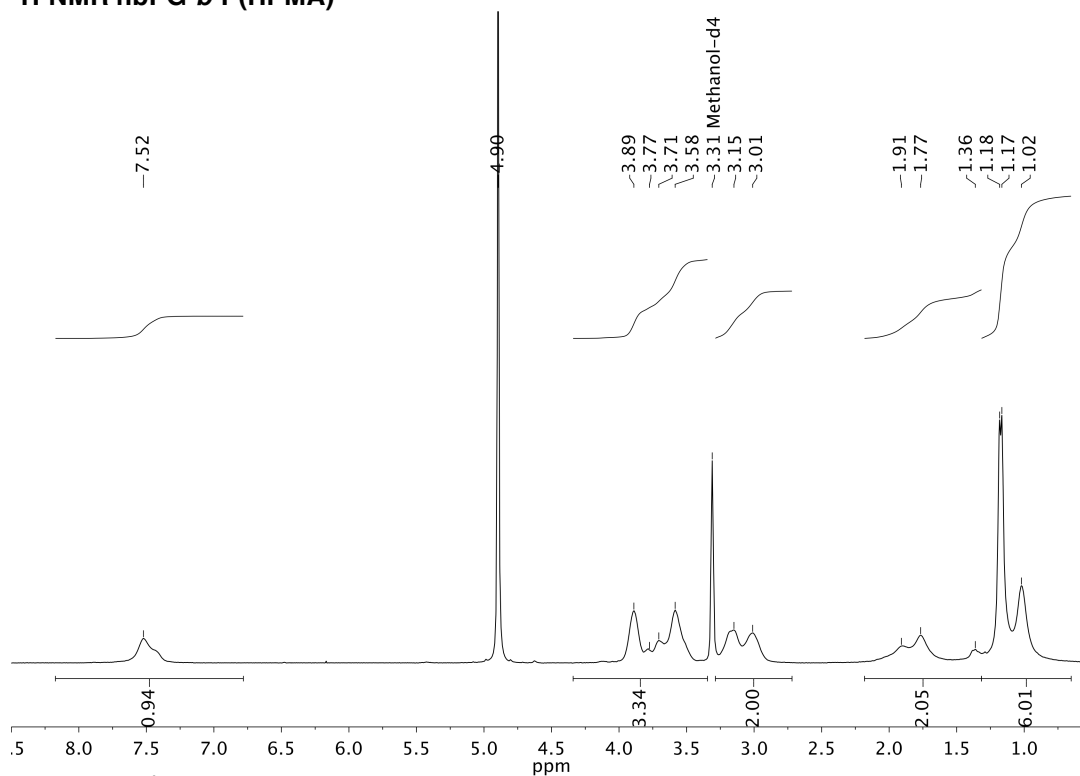


Figure S13: ¹H-NMR (CD₃OD, 400 MHz) hbPG₉₁-*b*-P(HPMA)₁₉₄

¹H-NMR hbPG-*b*-P(HPMA)-Biotin

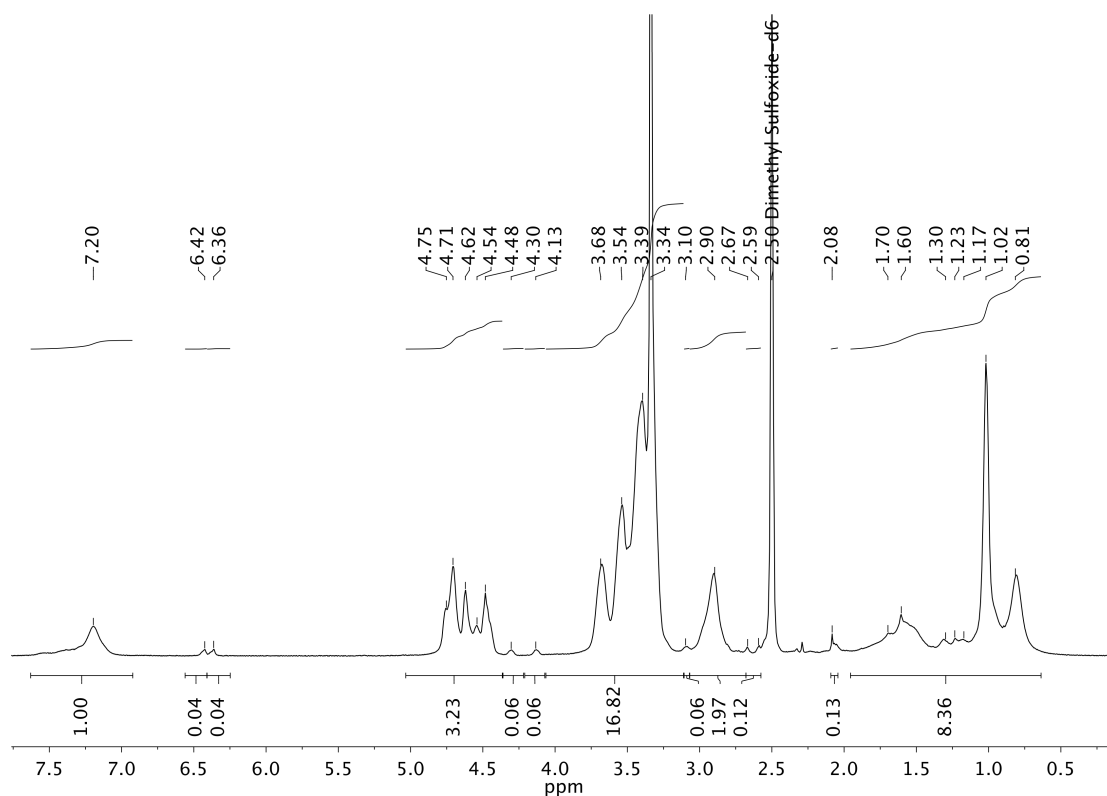


Figure S14: ¹H-NMR (DMSO-*d*₆, 400 MHz) hbPG₂₁-*b*-P(HPMA)₉-Biotin

Size Exclusion Chromatography of hbPG-*b*-P(HPMA)-Biotin

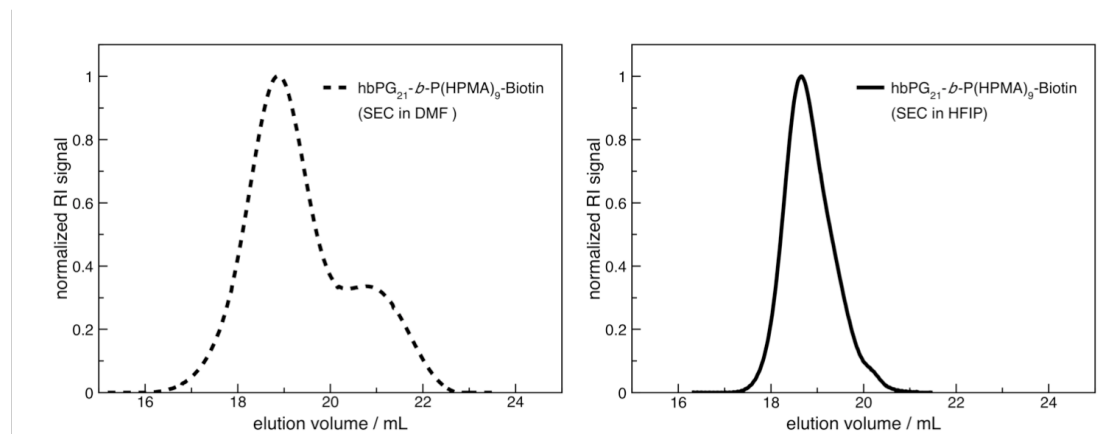


Figure S15: Elugram size exclusion chromatography of hbPG₂₁-*b*-P(HPMA)₉-Biotin in DMF (left) and HFIP (right).

¹H-NMR hbPG-*b*-P(HPMA)-TxRed

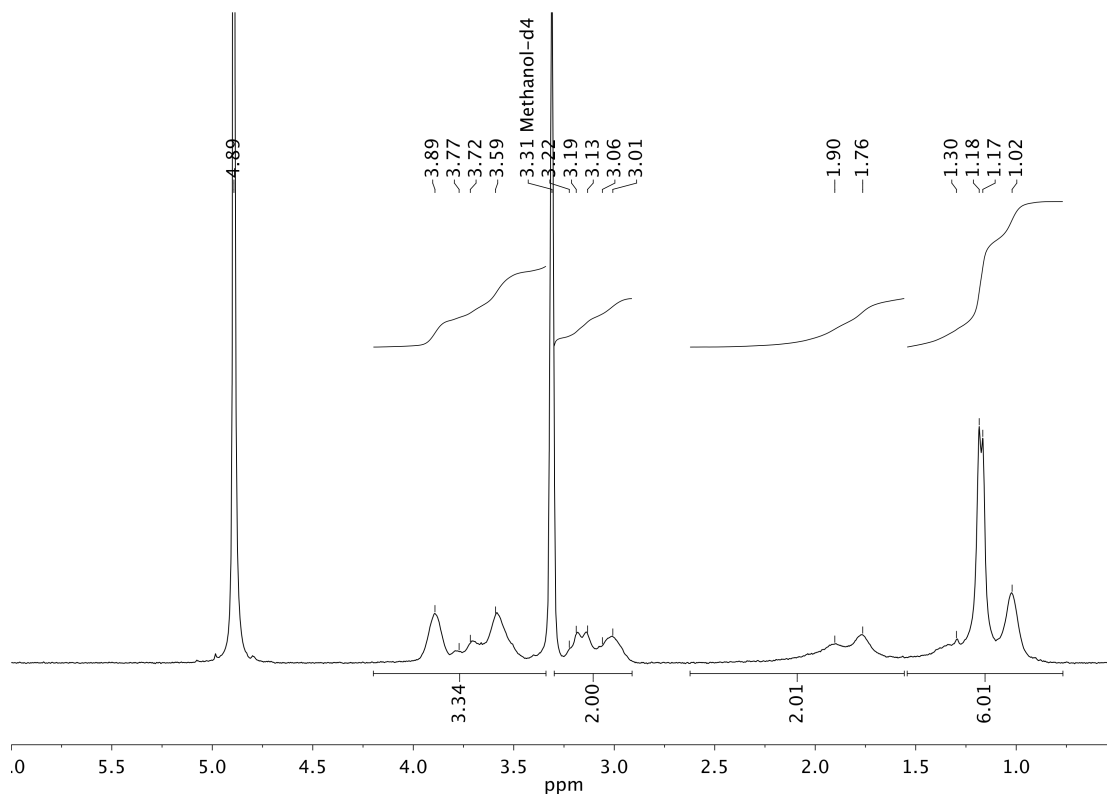


Figure S16: ¹H-NMR (CD₃OD, 400 MHz) hbPG₉₁-*b*-P(HPMA)₁₉₄-TxRed

Size Exclusion Chromatography of hbPG-*b*-P(HPMA)-TxRed

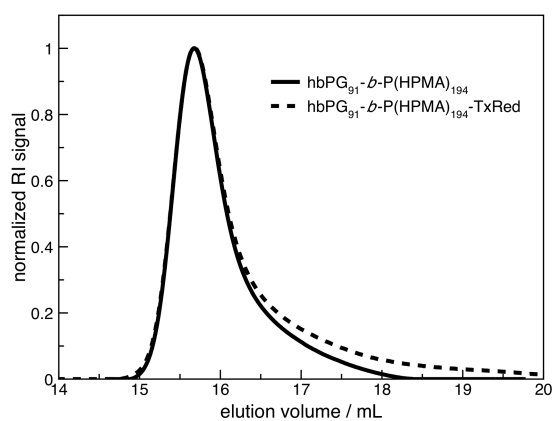


Figure S17: Elugram size exclusion chromatography of hbPG₉₁-*b*-P(HPMA)₁₉₄ and hbPG₉₁-*b*-P(HPMA)₁₉₄-TxRed in DMF.

References

- (1) Roth, P. J.; Wiss, K. T.; Zentel, R.; Theato, P. *Macromolecules* **2008**, *41*, 8513-8519.
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