

Supporting Information for

A hot oxidant, 3-NO₂Y₁₂₂ radical, unmasking conformational gating in
ribonucleotide reductase

Kenichi Yokoyama[†], Ulla Uhlin[§], and JoAnne Stubbe^{†,‡,}*

Department of Chemistry and Biology, Massachusetts Institute of Technology, 77 Massachusetts Avenue,

Cambridge, MA 02139-4307 and Department of Molecular Biology, Swedish University of Agricultural

Science, Uppsala Biomedical Center, Box 590, SE-75124 Uppsala, Sweden

Table of Contents

- Figure S1. Microwave power saturation behavior of $\text{NO}_2\text{Y}\cdot$ and the new $\text{Y}\cdot$ EPR signal(s).
- Figure S2. dCDP formation by $\text{Y}_{122}\text{NO}_2\text{Y}-\beta 2$ under single turnover and steady state conditions.
- Figure S3. EPR spectra of the products generated when $\text{Y}_{122}\text{NO}_2\text{Y}-\beta 2$ or $\text{Y}_{122}[\beta\text{-}^2\text{H}_2]\text{NO}_2\text{Y}-\beta 2$ is incubated with wt- $\alpha 2$, ATP, and CDP.
- Figure S4. EPR spectra of the new radical species acquired from 9 to 55 ms.
- Figure S5. The stability of the new radical species.
- Figure S6. EPR spectra of the new radical species in mutants with a block in the PCET pathway (Figure 1).
- Figure S7. Tri-exponential fits for dCDP formation monitored by RCQ and new radical formation by RFQ-EPR using the rate constants for $\text{NO}_2\text{Y}_{122}^-$ formation.

Reference

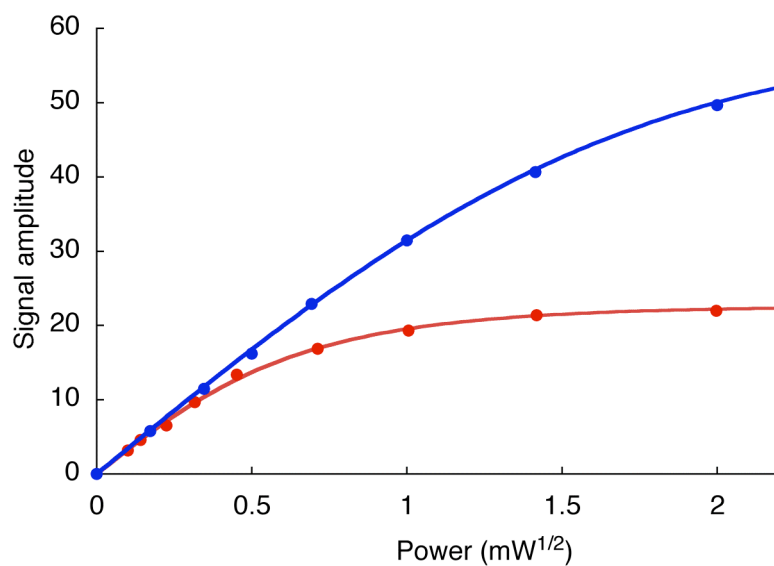


Figure S1. Microwave power saturation behavior of NO₂Y• (blue) and the new radical (red) signals. The lines are fits to Eq. 1 and yielded $P_{1/2}$ of 11.4 ± 0.5 mW ($b = 1.2 \pm 0.3$) and 0.54 ± 0.08 mW ($b = 1.1 \pm 0.2$) for NO₂Y₁₂₂• (blue) and the new Y• (red), respectively.

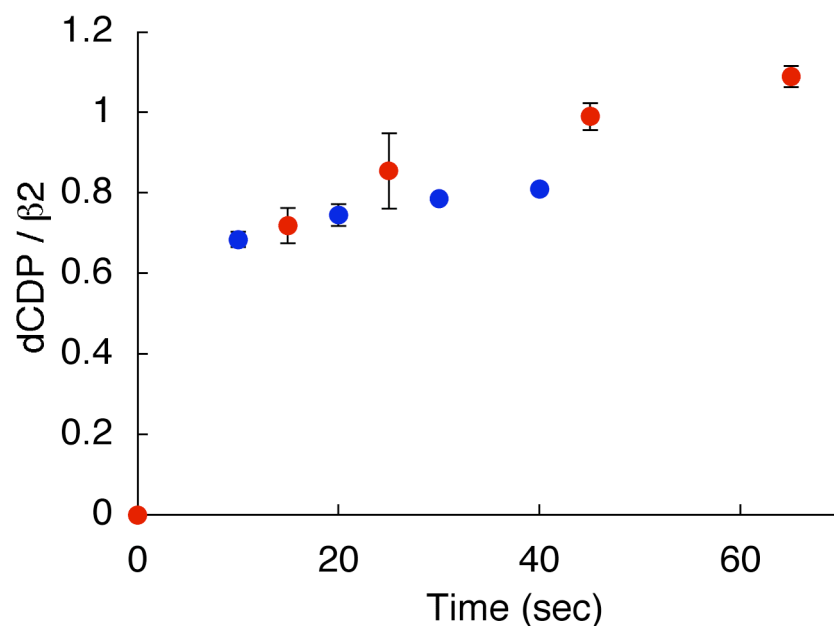
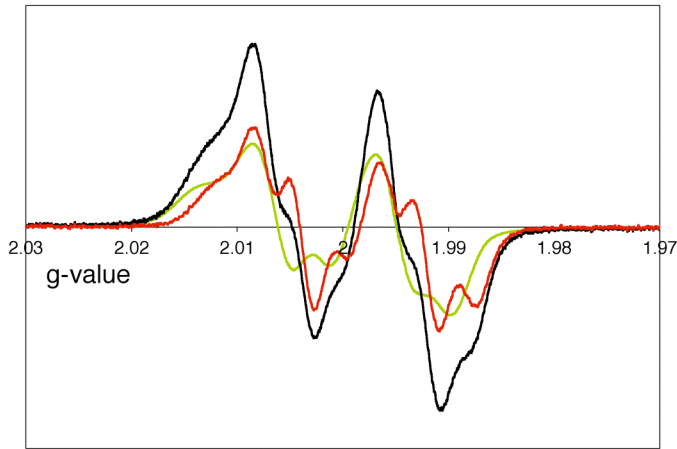
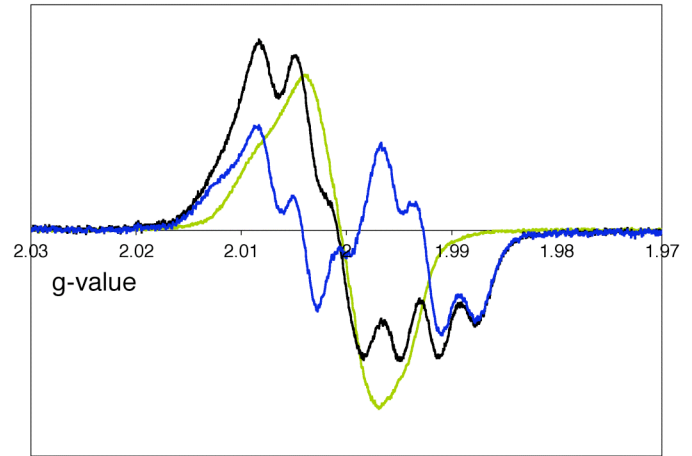


Figure S2. dCDP formation by $[\text{NO}_2\text{Y}\cdot]\text{-}\beta 2$ ($0.95 \pm 0.05 \text{ NO}_2\text{Y}\cdot/\beta 2$) incubated with $\alpha 2$, ATP and CDP under steady state conditions using TR/TRR/NADPH as reductant (red circles) or under single turnover conditions with no reductant (blue circles). The slow phase after 10 s in the steady-state assay corresponds to 6.9 nmol/min/mg, 0.1% that of wt- $\beta 2$, consistent with our earlier characterization of dCDP formation by contaminating wt- $\beta 2$.¹

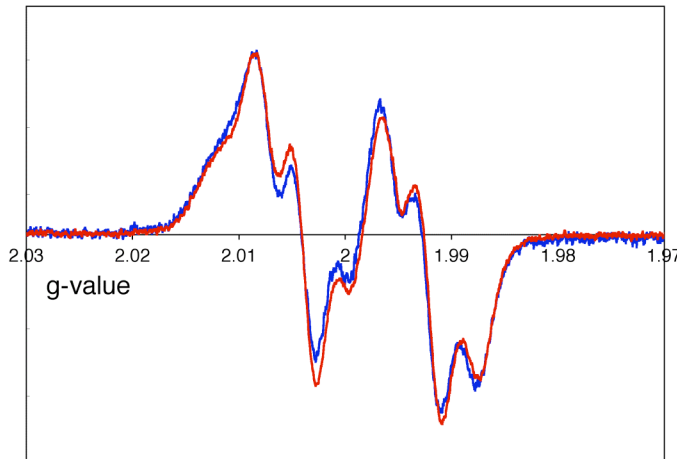
(A) $[\text{NO}_2\text{Y}\cdot]\text{-}\beta 2/\alpha 2/\text{ATP}/\text{CDP}$



(B) $[\beta\text{-}^2\text{H}_2]\text{NO}_2\text{Y}\cdot\text{-}\beta 2/\alpha 2/\text{ATP}/\text{CDP}$



(C) Overlay of new radicals



(D) Difference between trapped radical spectra

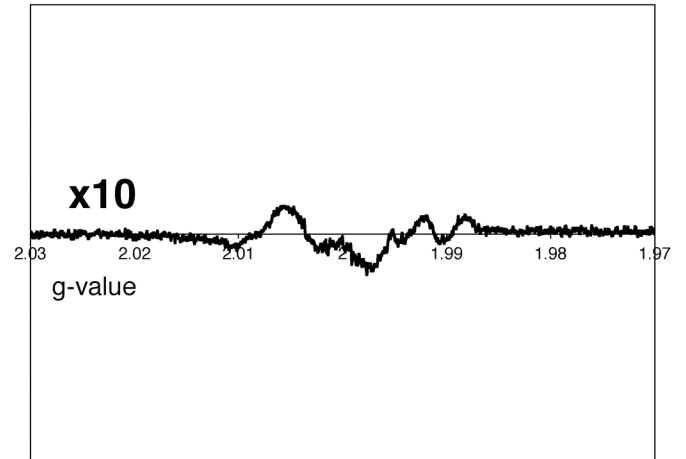


Figure S3. EPR spectra of (A) $\text{Y}_{122}\text{NO}_2\text{Y}\text{-}\beta 2$ or (B) $\text{Y}_{122}[\beta\text{-}^2\text{H}_2]\text{NO}_2\text{Y}\text{-}\beta 2$ incubated with wt- $\alpha 2$, ATP, and CDP freeze-quenched at 120 ms (black traces). Fifty percent of the spectrum of the $\text{NO}_2\text{Y}\cdot$ (A) or $[\beta\text{-}^2\text{H}_2]\text{NO}_2\text{Y}\cdot$ (B) (green) was subtracted to give the spectrum of new radical species (red and blue traces in (A) and (B), respectively). (C) Overlay of the red and blue spectra from (A) and (B). (D) Difference spectrum of the new radical species, red and blue traces in (C). The spectrum is magnified 10 folds.

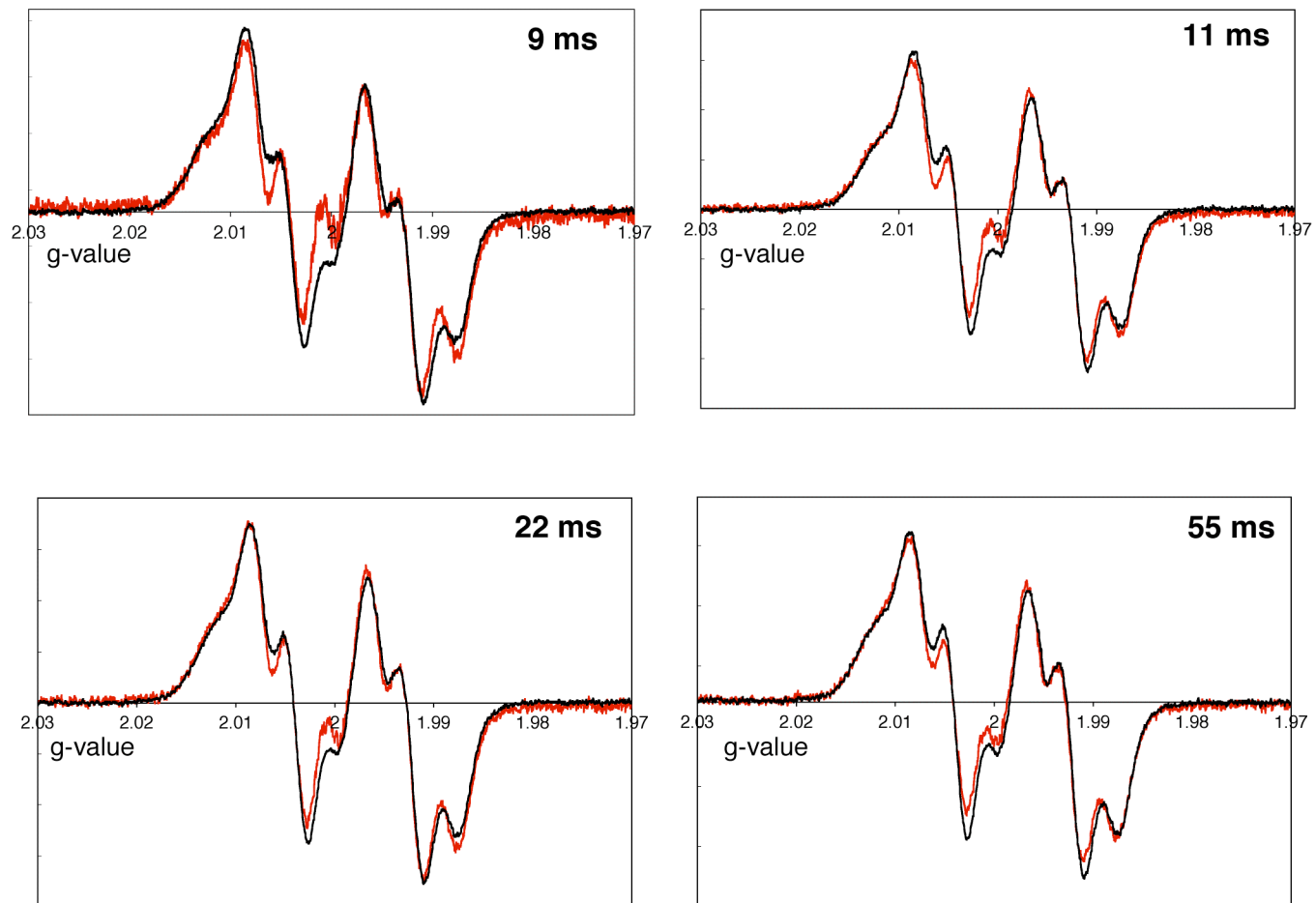


Figure S4. EPR spectra of the new radical acquired at the indicated time points. The spectrum of the new radical obtained as described in Figure S3 with $Y_{122}NO_2Y-\beta_2$ (black trace) is overlaid with that obtained from $Y_{122}[\beta\text{-}^2H_2]NO_2Y-\beta_2$ (red trace) at the same time point.

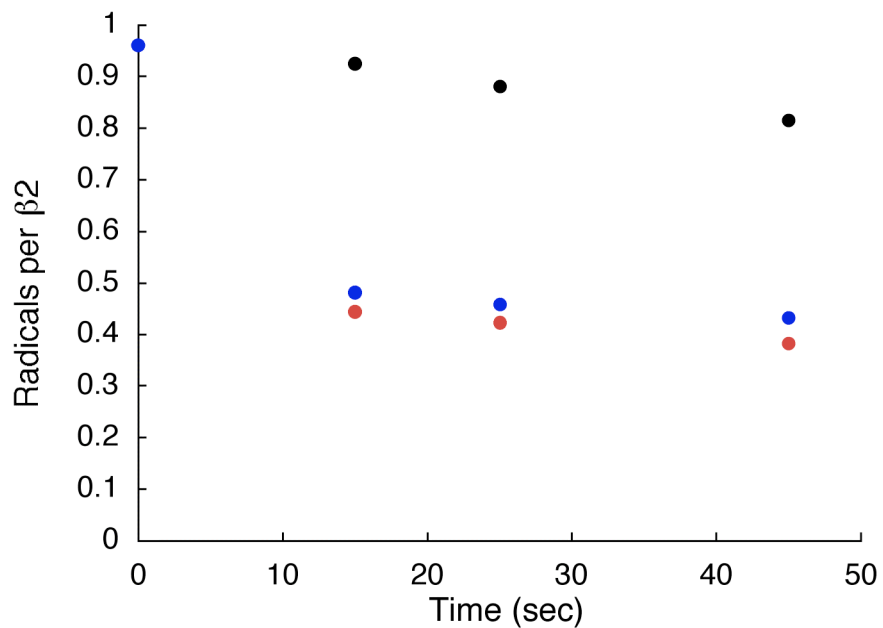


Figure S5. The stability of the new radical and NO₂Y•. Fe(II)₂-Y₁₂₂NO₂Y-β2 was manually mixed at 25 °C with O₂-saturated reconstitution buffer containing CDP and incubated for 10 s to generate 0.95 NO₂Y•/β2 (t = 0 time point). This mixture was then incubated with α2 and ATP. The reaction was quenched by hand in liquid N₂. Black circles (total radical at each time point); blue circles (NO₂Y•) and red circles (new radical).

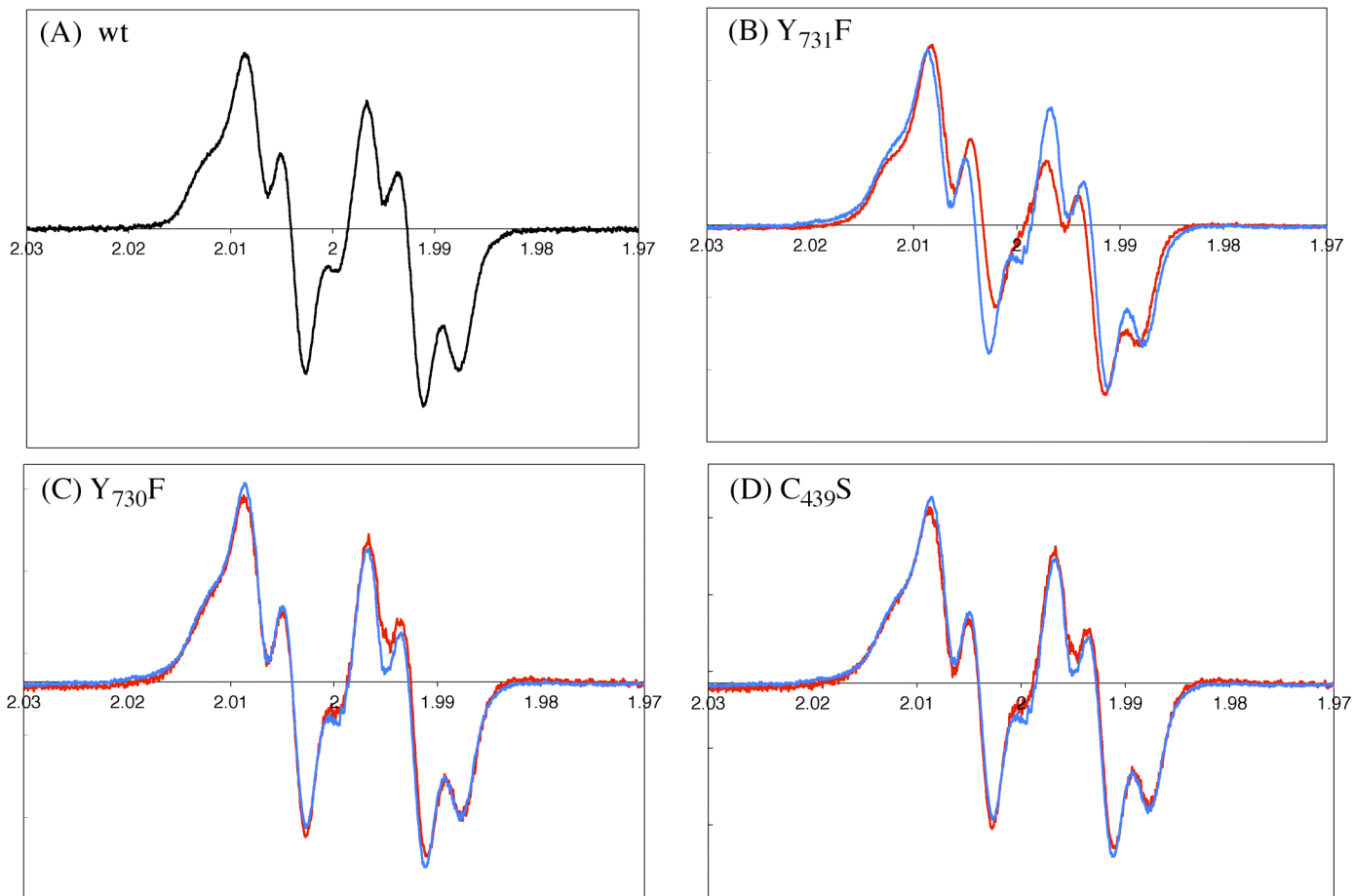


Figure S6. EPR spectra of the new radical observed in (A) wt- $\alpha 2$ (B) Y₇₃₁F- $\alpha 2$, (C) Y₇₃₀F- $\alpha 2$ and (D) C₄₃₉S- $\alpha 2$ after incubation with Y₁₂₂NO₂Y- $\beta 2$, ATP, and CDP. The EPR spectrum of NO₂Y• was subtracted from each spectrum as described in Methods. The new radical spectrum observed in wt- $\alpha 2$ (blue traces) was overlaid with the spectrum observed in each mutant (red traces).

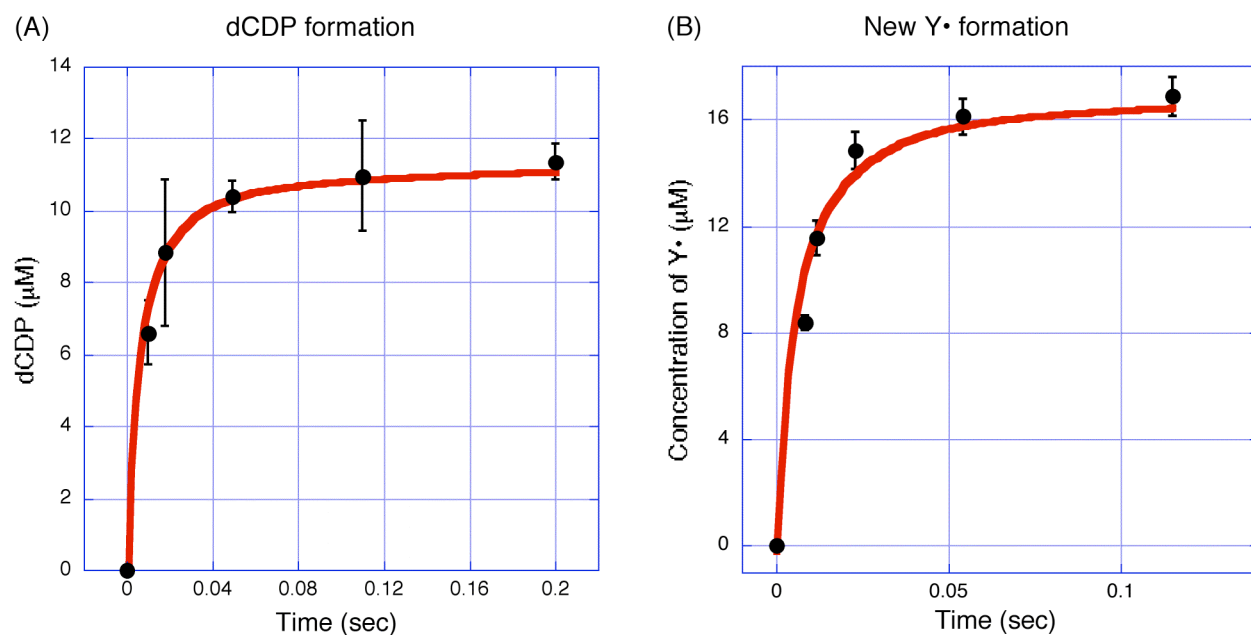


Figure S7. Refitting of experimental data for dCDP formation measured by RCQ (Figure 8) and for new radical formation by RFQ EPR (Figure 11) with tri-exponential fitting using the parameters obtained from the SF experiments measuring phenolate formation (Figure 9B, Table 2). The rate constants ($k_1 = 283$; $k_2 = 65$; and $k_3 = 10$) and amplitudes ($A_1, 48\%$; $A_2, 43\%$; $A_3, 9\%$) are fixed. The red line is the result of fitting with (A) 11.2 μM and (B) 16.9 μM for total dCDP and Y• formation, respectively.

Reference

- (1) Yokoyama, K.; Uhlin, U.; Stubbe, J. *J. Am. Chem. Soc.* **2010**, *132*, 8385-8397.