

Structures, Reactivities, and Antibiotic Activities of the Marinopyrroles A-F

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General. Diazomethane was prepared according to the method of Arndt (Arndt, F. *Org. Synth.* **1943**, Coll. Vol. 2, 165). Ethyl 2,5-dioxopentanoate (**28**) was synthesized via ozonolysis of ethyl 2-oxohex-5-enoate (Macritchie, J. A.; Silcock, A.; Willis, C. L. *Tetrahedron Asym.* **1997**, 8, 3895-3902). Ethyl 3-amino-1*H*-pyrrole-2-carboxylate (**29**) was prepared according to literature procedures (Furneaux, R. H.; Tyler, P. C. *J. Org. Chem.* **1999**, 64, 8411-8412). Pyridine was distilled from calcium hydride prior to use, and tetrahydrofuran (THF) was distilled from potassium benzophenone ketyl. All other reagents and solvents were purchased commercially and were used without further purification. Column chromatography was performed using 60 Å 230-400 mesh silica gel. RP-HPLC purification was performed on a C8(2) Phenomenex® Luna column (5 µm, 250 x 10 mm) at 3 mL min⁻¹ with UV detection (254 nm). Unless otherwise stated, ¹H NMR spectra were recorded at 500 MHz in CDCl₃ (residual CHCl₃ referenced to 7.26 ppm), DMSO-*d*₆ (residual DMSO referenced to 2.50 ppm), or CD₃CN (residual CH₃CN referenced to 1.94 ppm) on a Varian Inova spectrometer. ¹³C NMR spectra were recorded at 75 MHz in CDCl₃ (referenced to 77.0 ppm) or DMSO-*d*₆ (referenced to 39.4 ppm) on a Varian Inova NMR spectrometer.

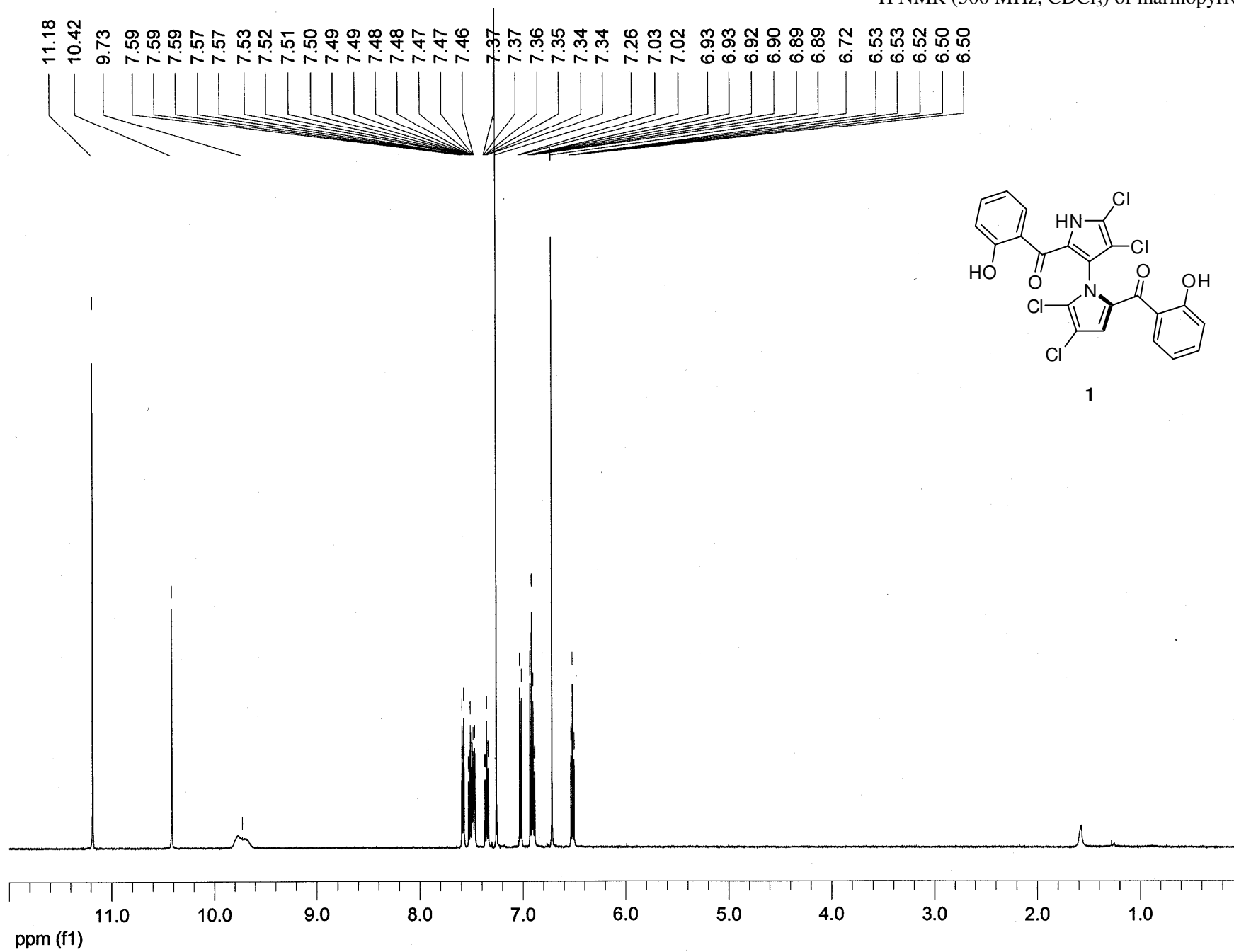
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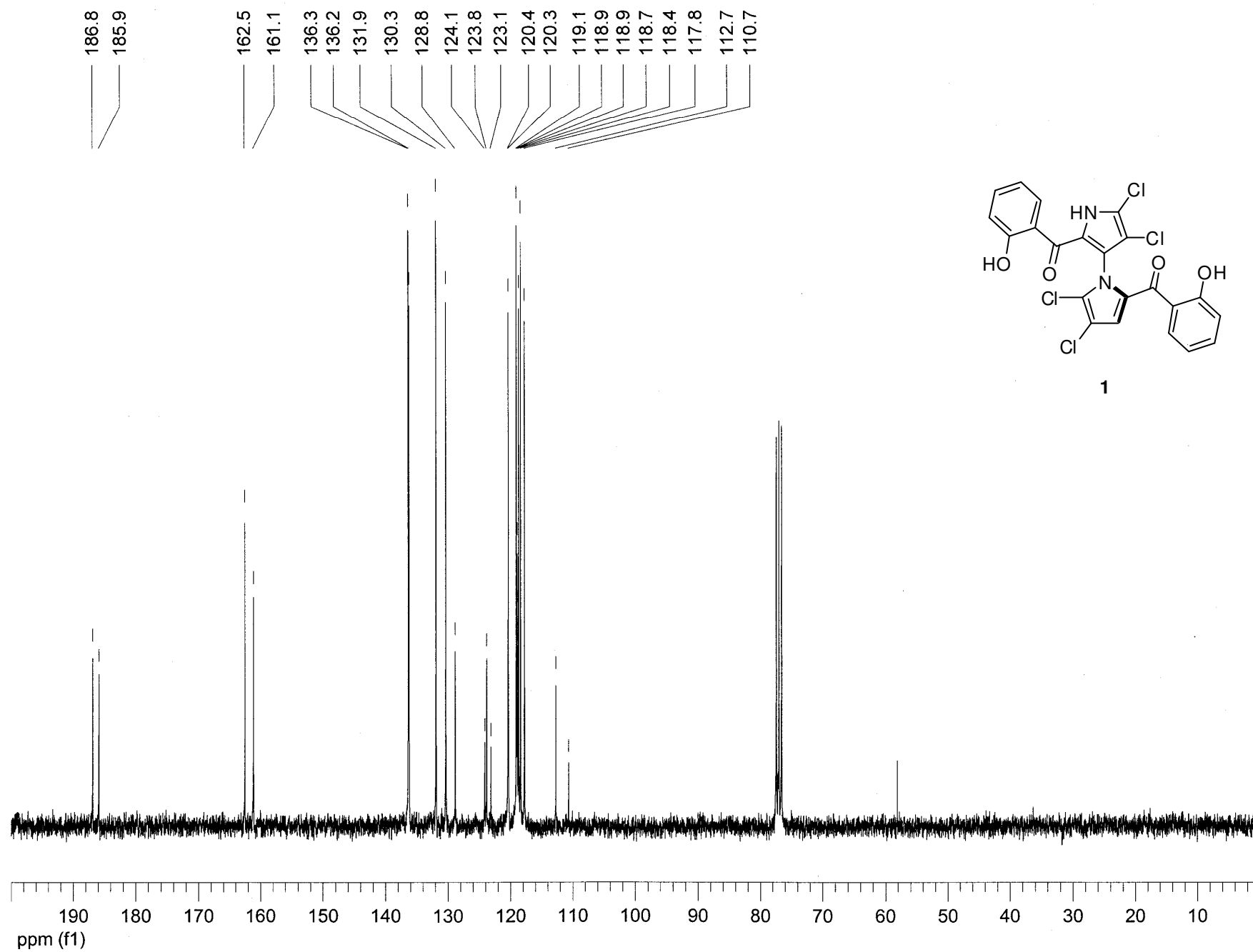
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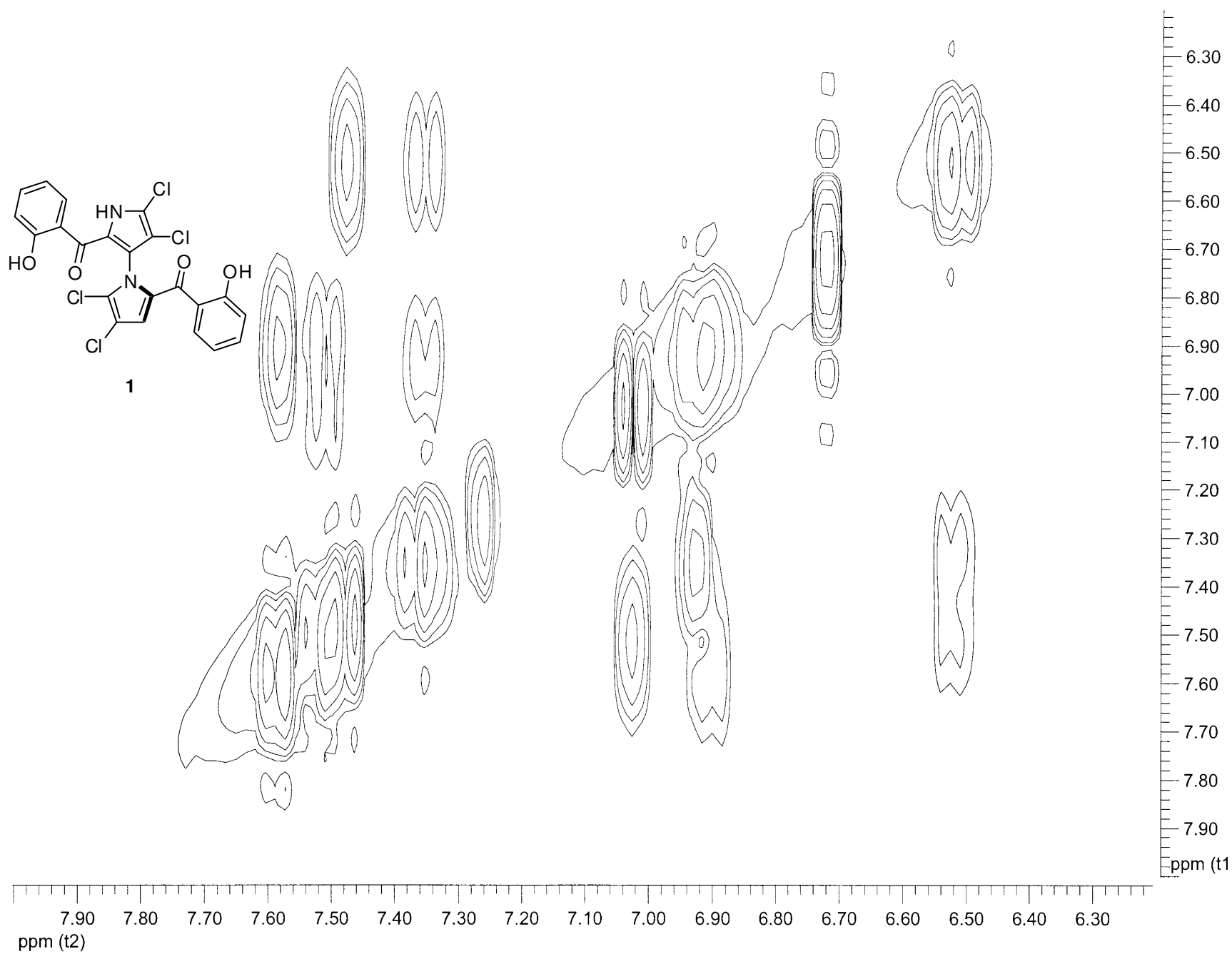
¹H NMR (500 MHz, CDCl₃) of marinopyrrole A (**1**)



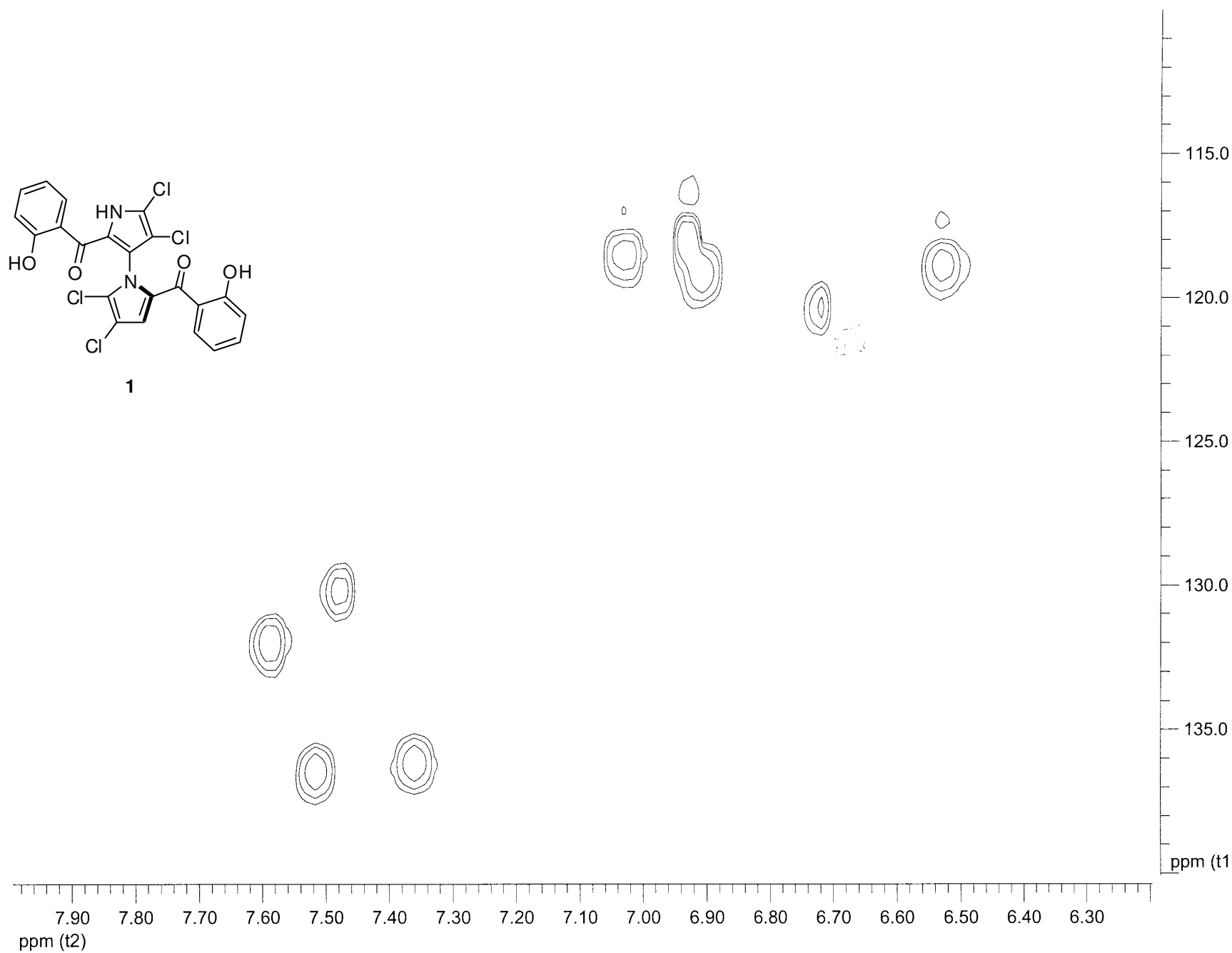
^{13}C NMR (75 MHz, CDCl_3) of marinopyrrole A (**1**)



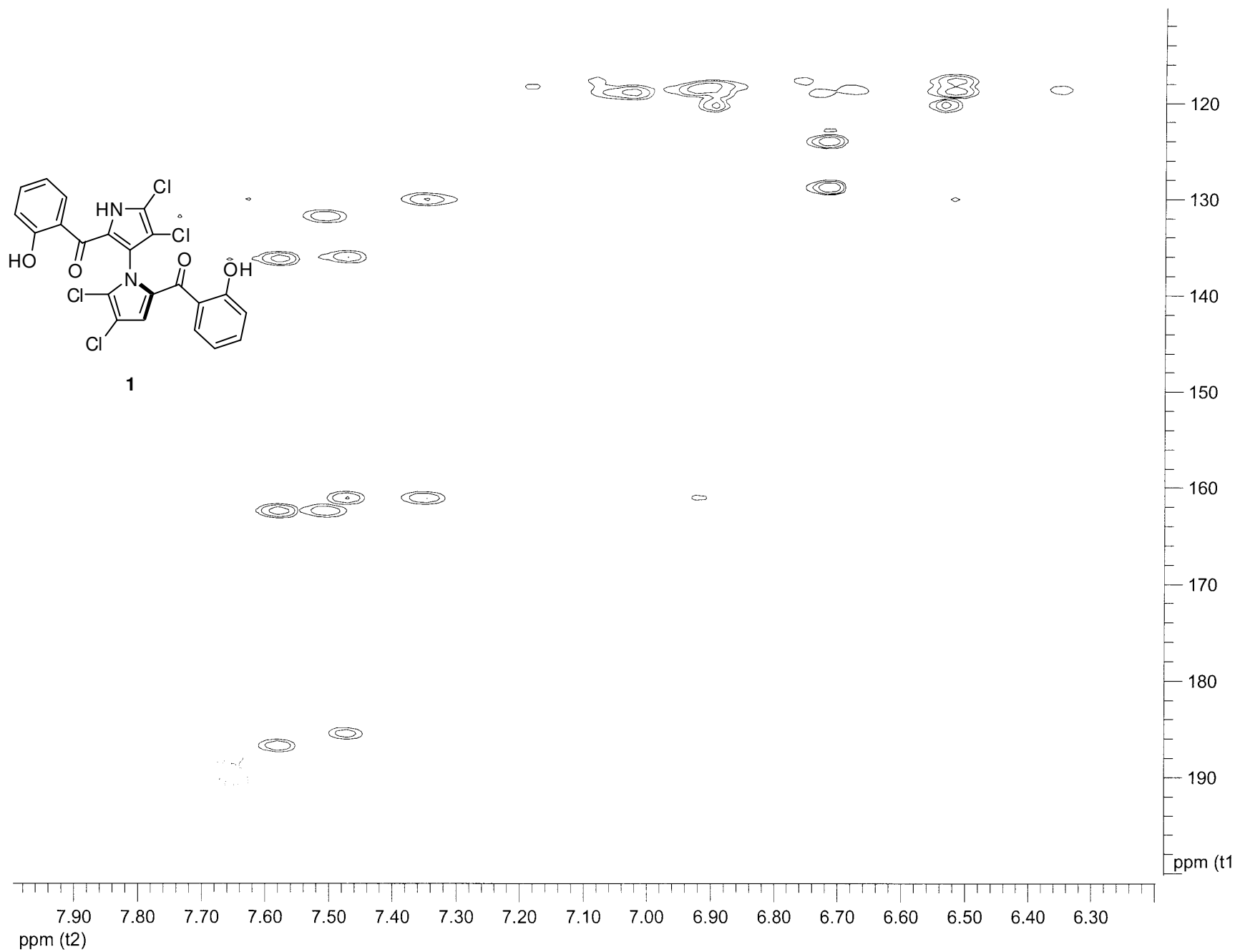
COSY (500 MHz, CDCl₃) of marinopyrrole A (**1**)



HSQC (500 MHz, CDCl₃) of marinopyrrole A (**1**)



HMBC (500 MHz, CDCl₃) of marinopyrrole A (**1**)

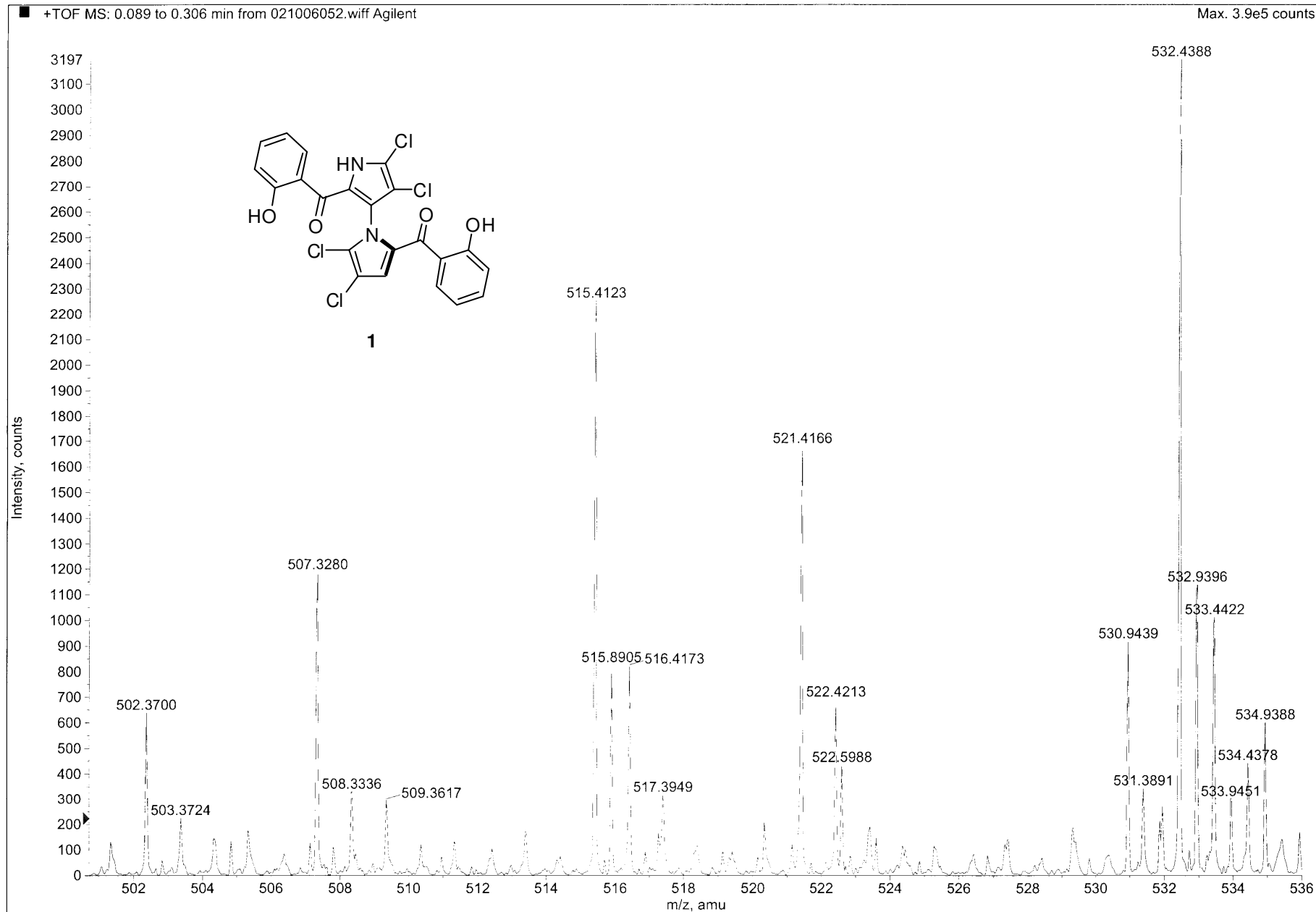


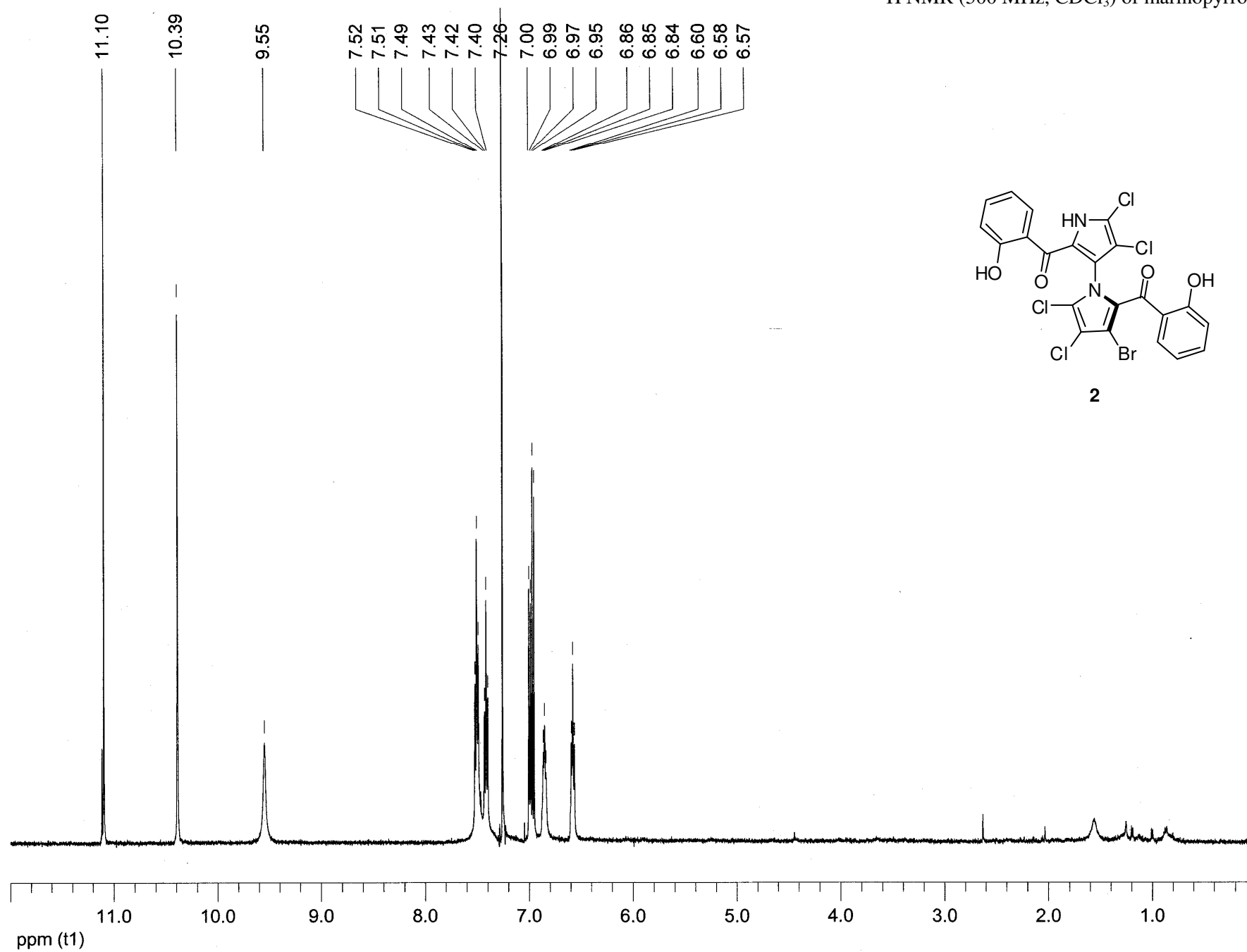
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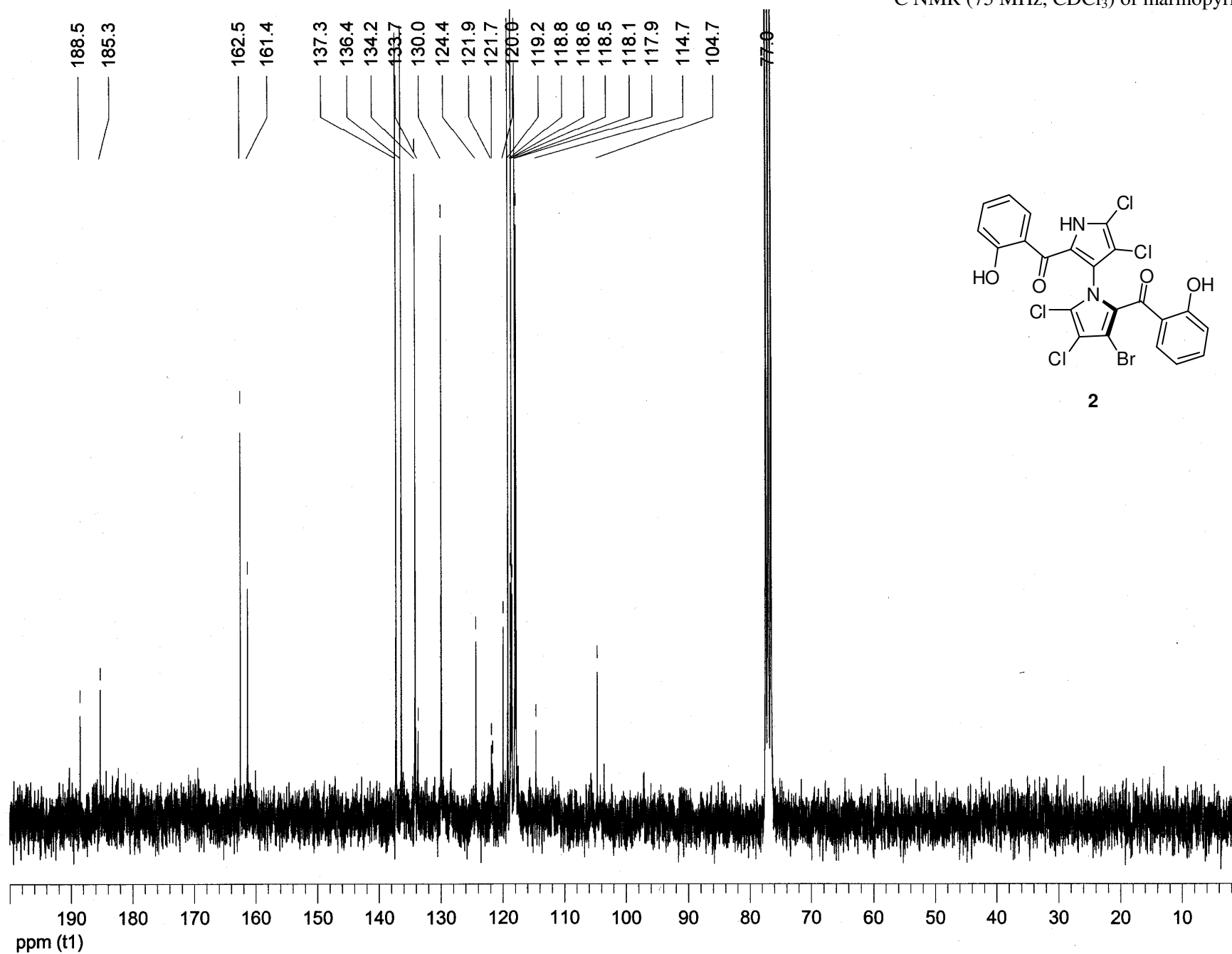
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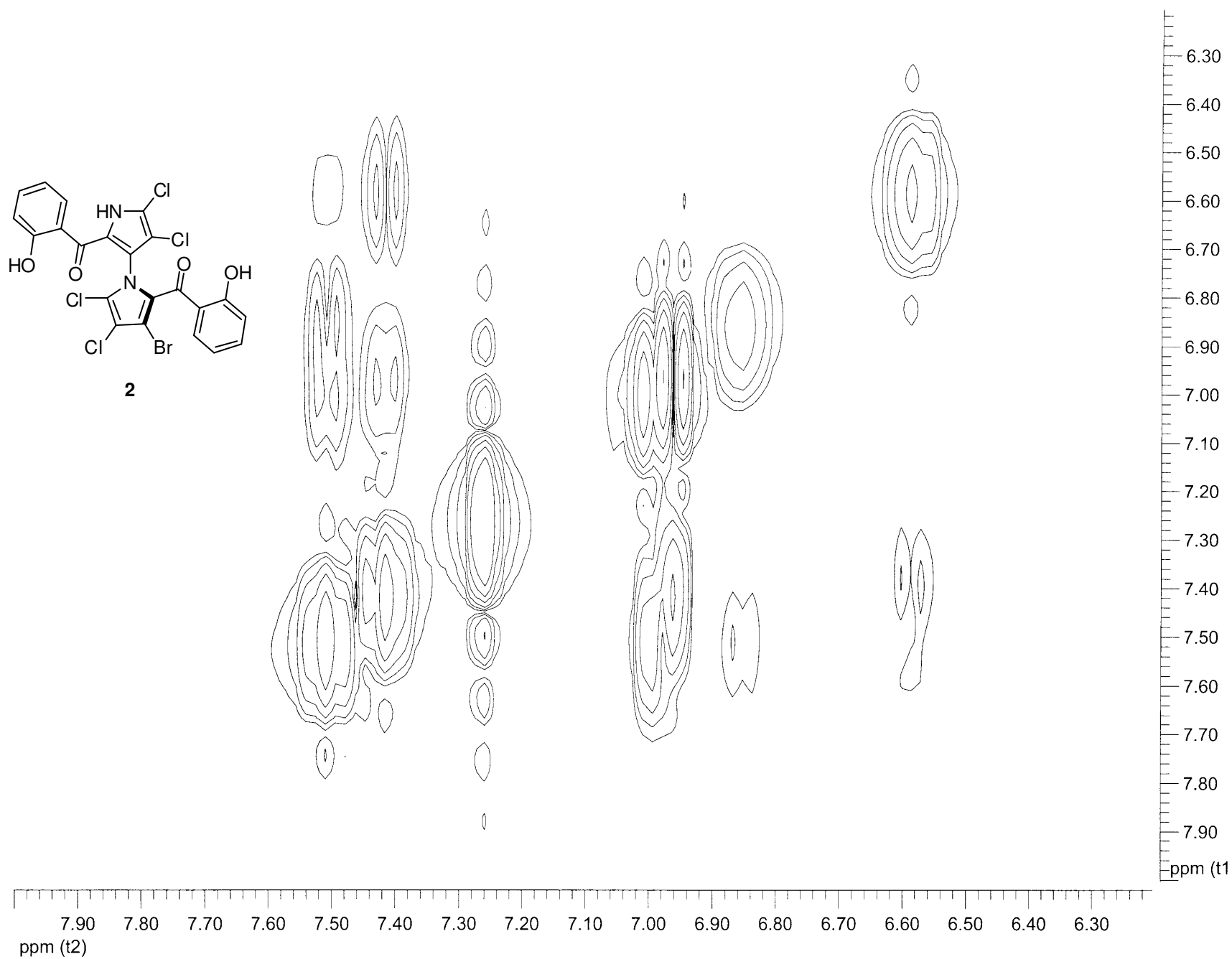


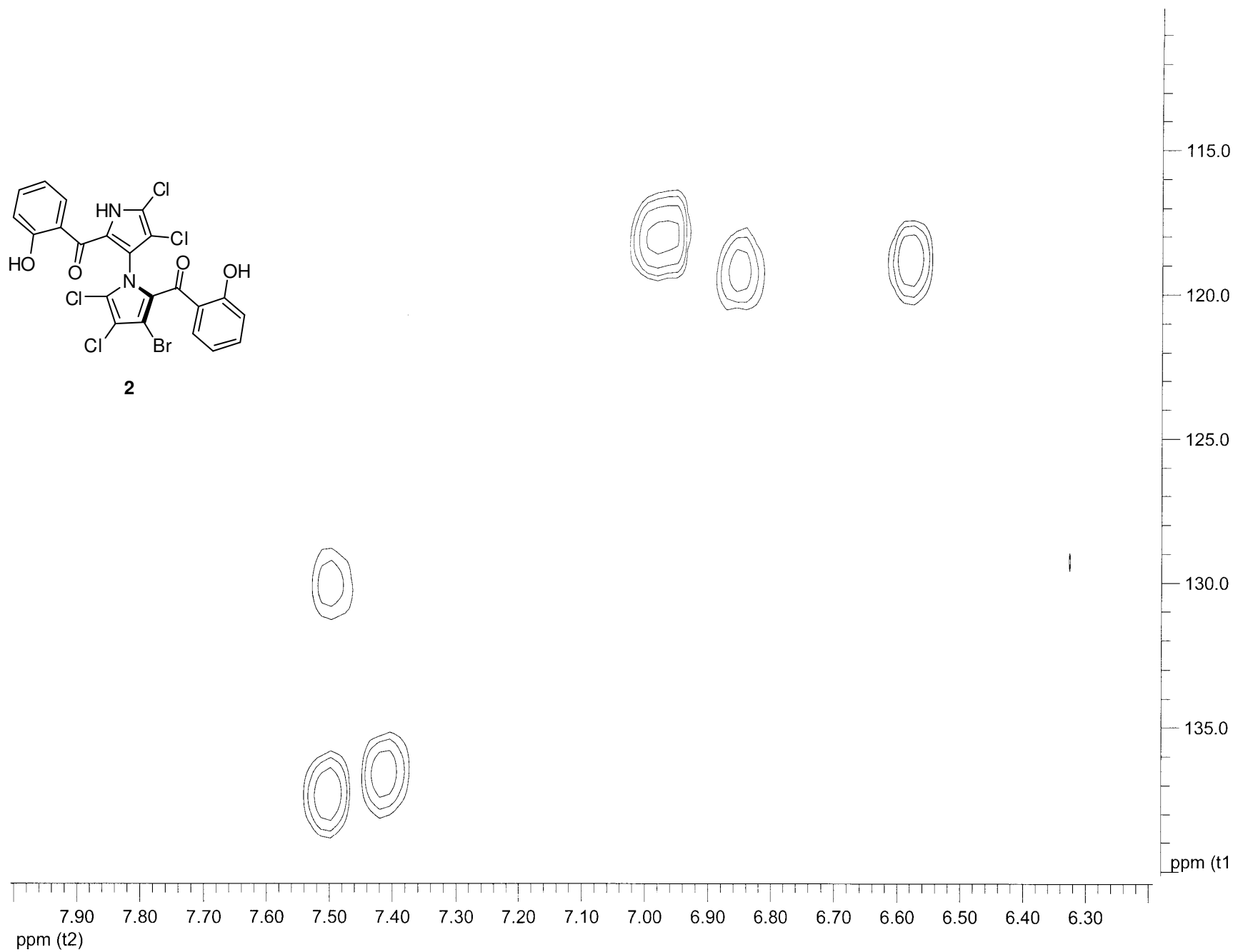


¹³C NMR (75 MHz, CDCl₃) of marinopyrrole B (2)

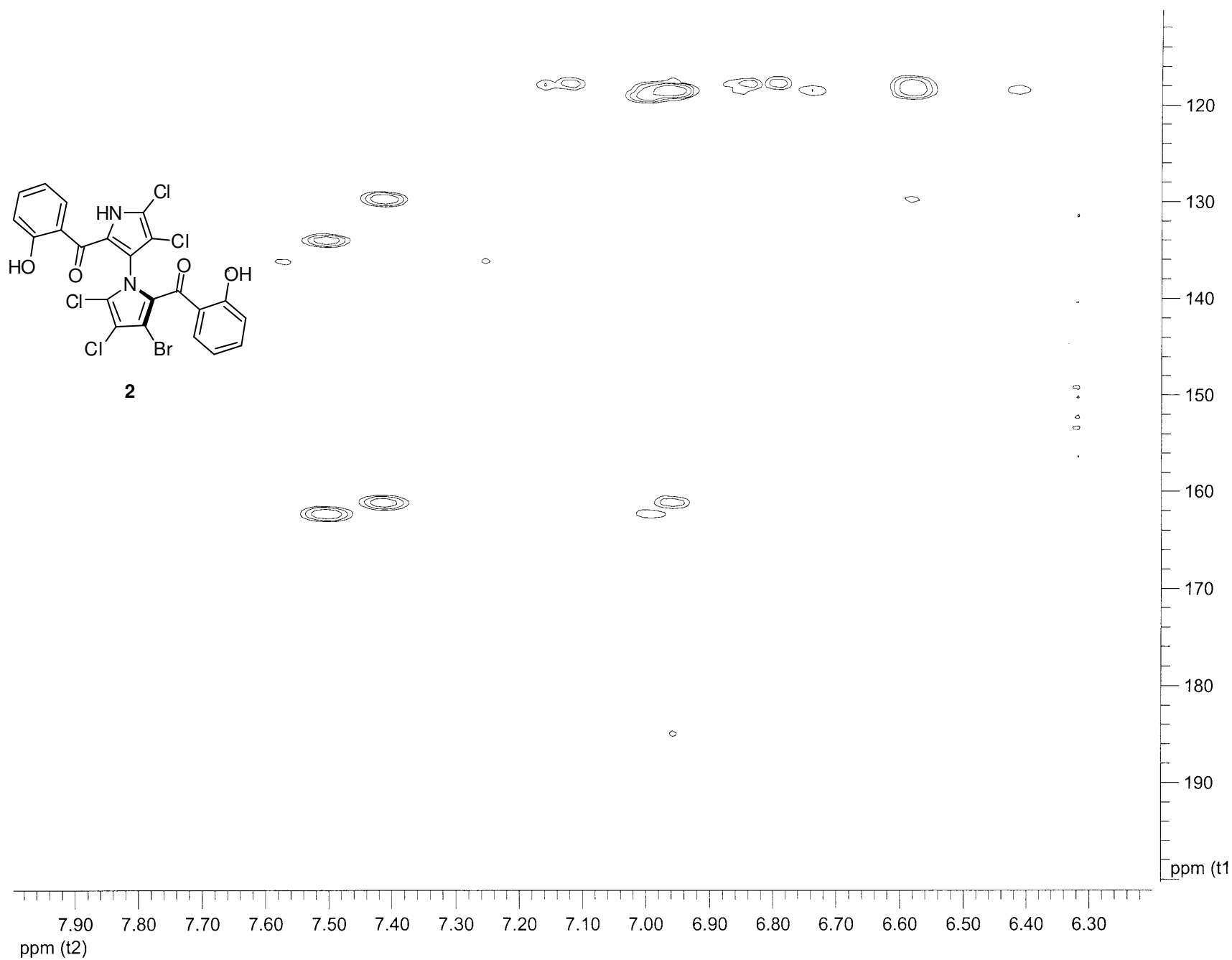


COSY (500 MHz, CDCl₃) of marinopyrrole B (**2**)





HMBC (500 MHz, CDCl₃) of marinopyrrole B (**2**)



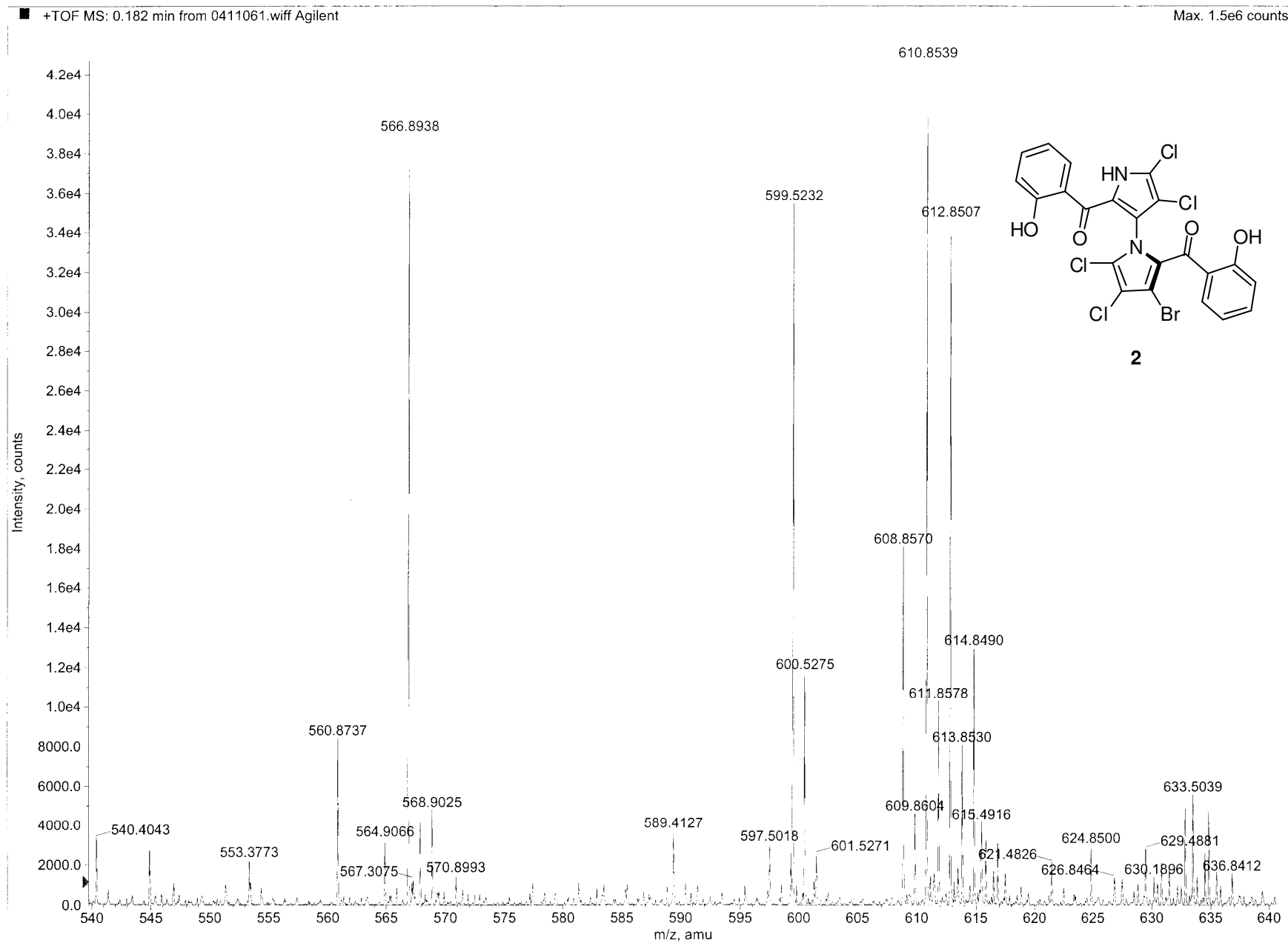
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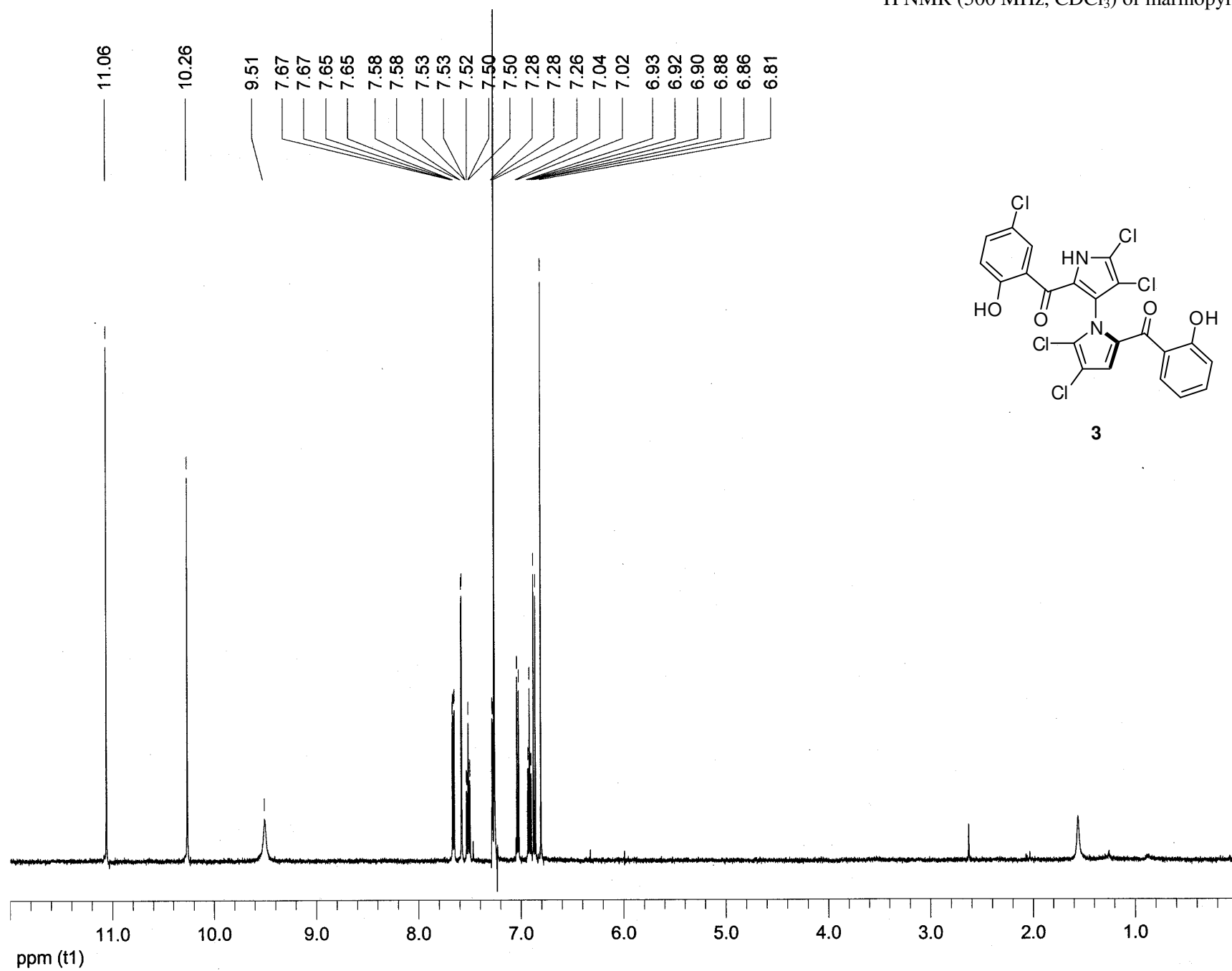
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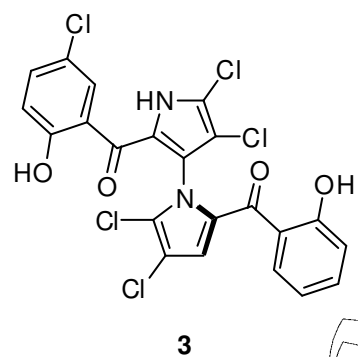
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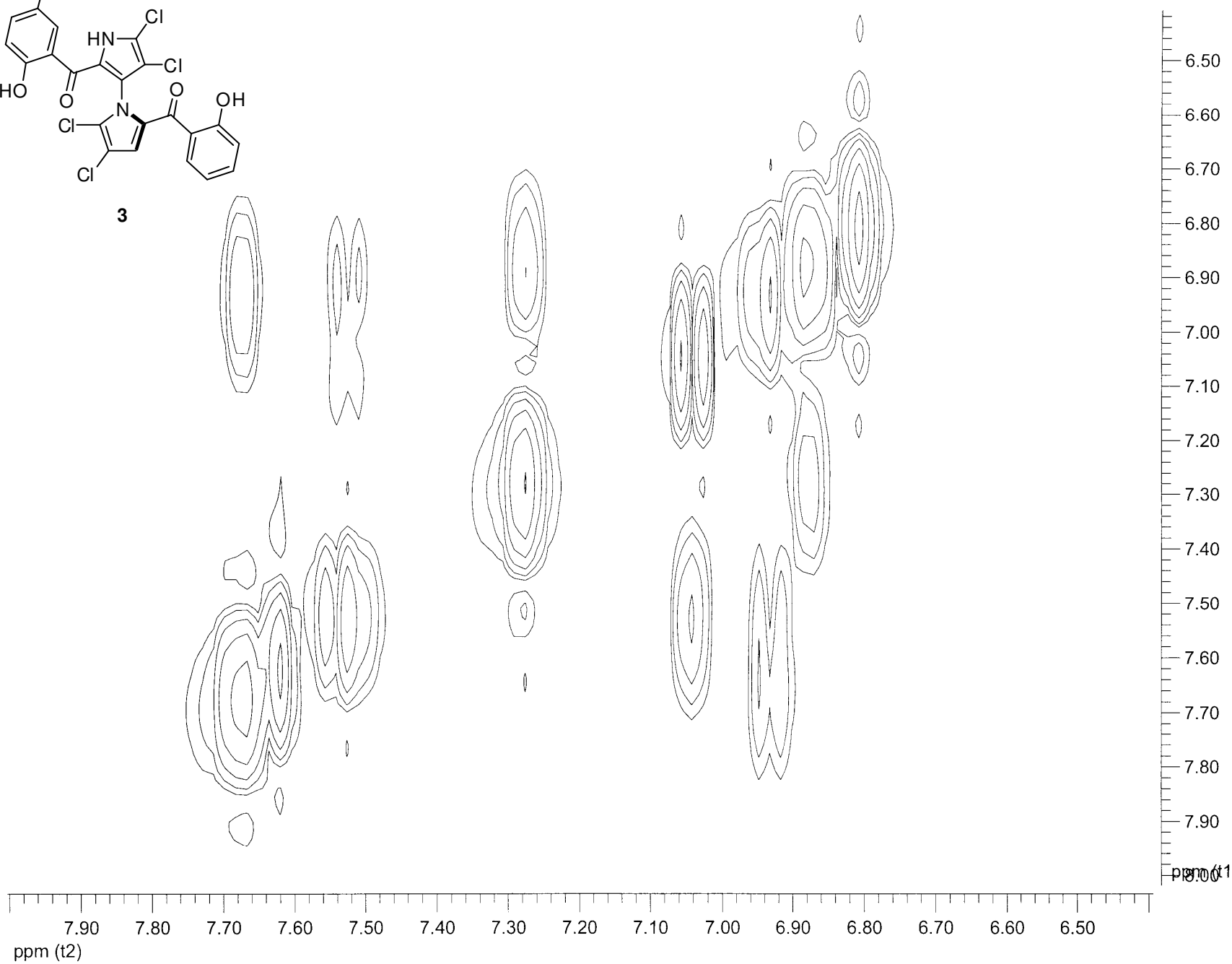
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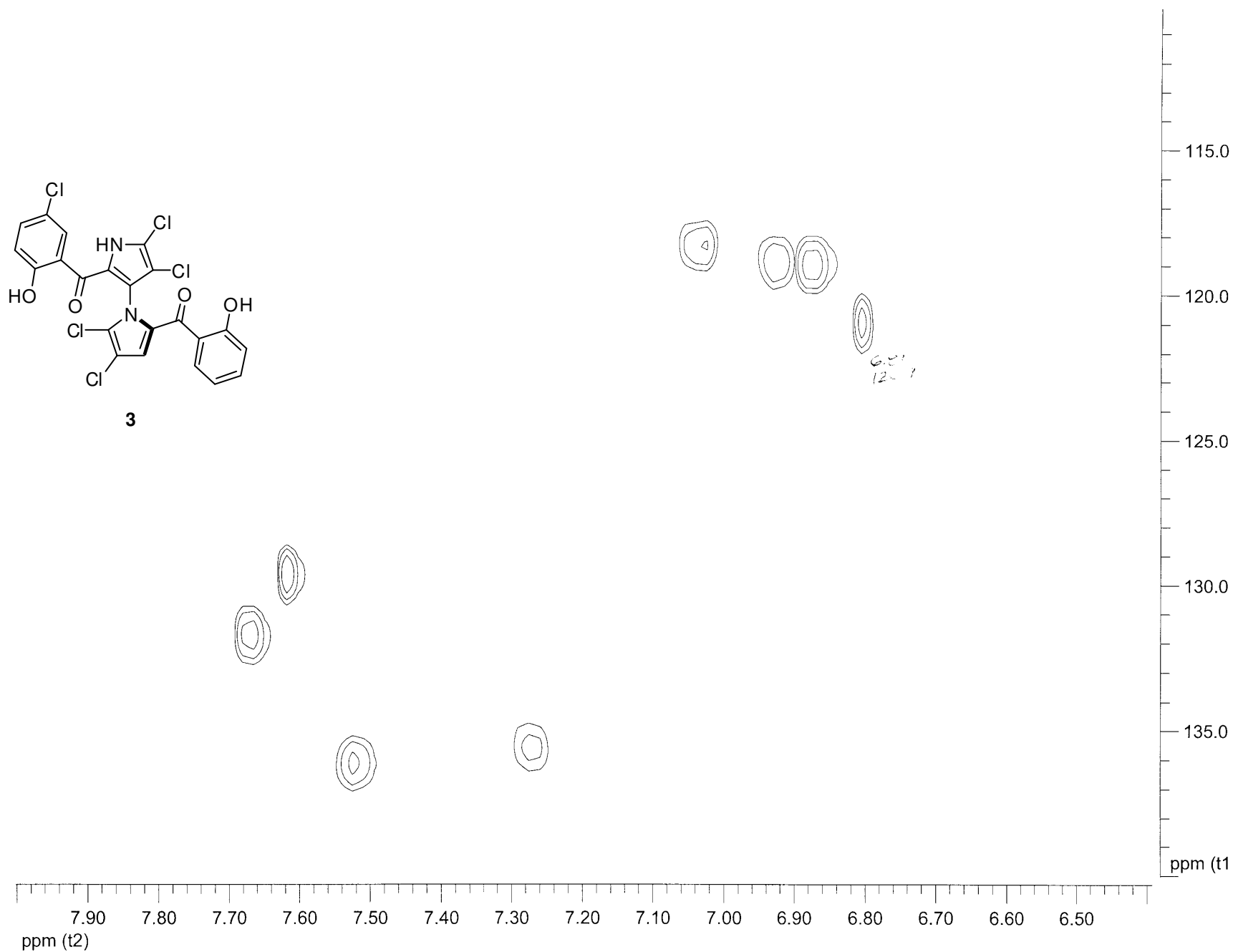




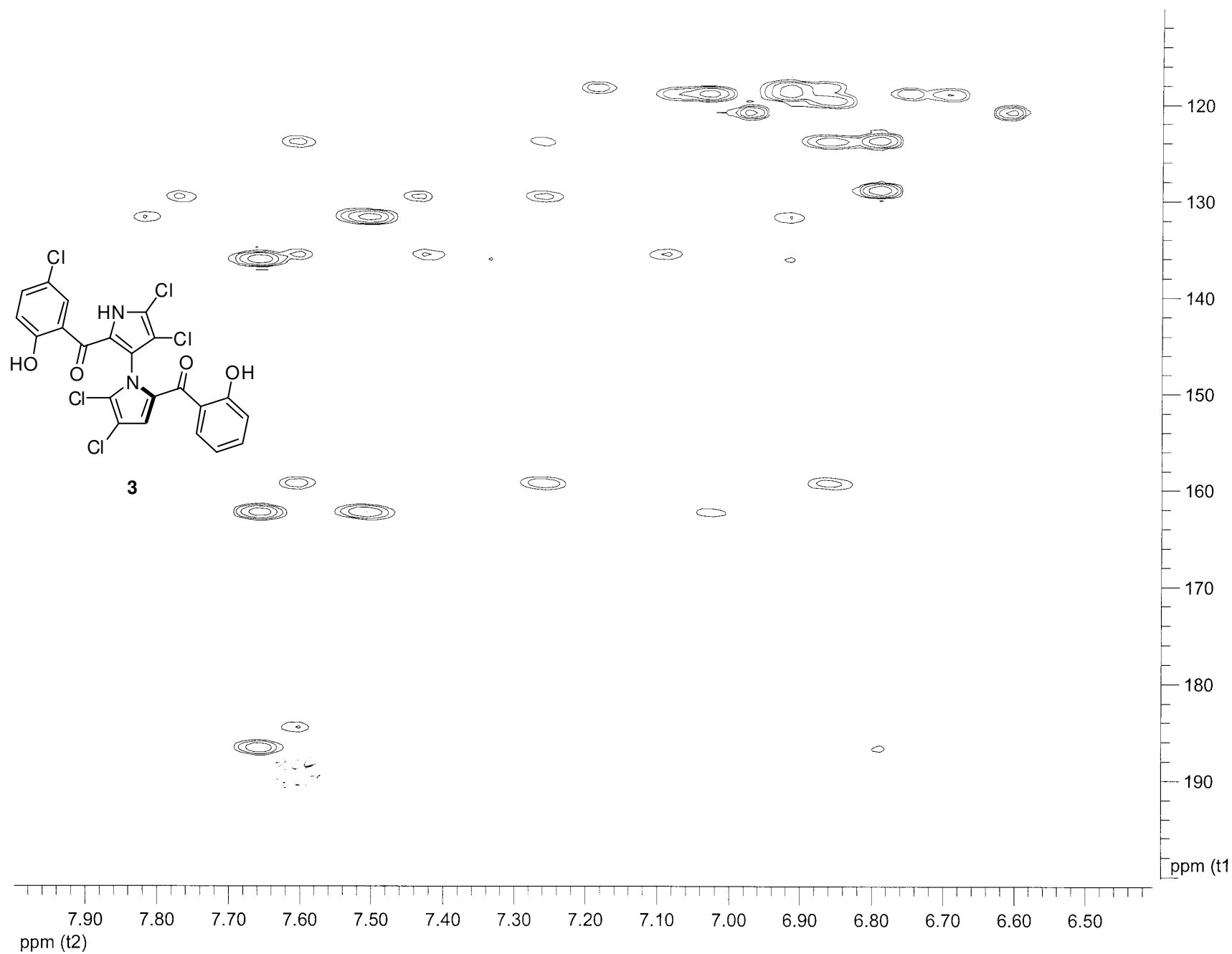
COSY (500 MHz, CDCl₃) of marinopyrrole C (**3**)



HSQC (500 MHz, CDCl₃) of marinopyrrole C (**3**)



HMBC (500 MHz, CDCl₃) of marinopyrrole C (**3**)

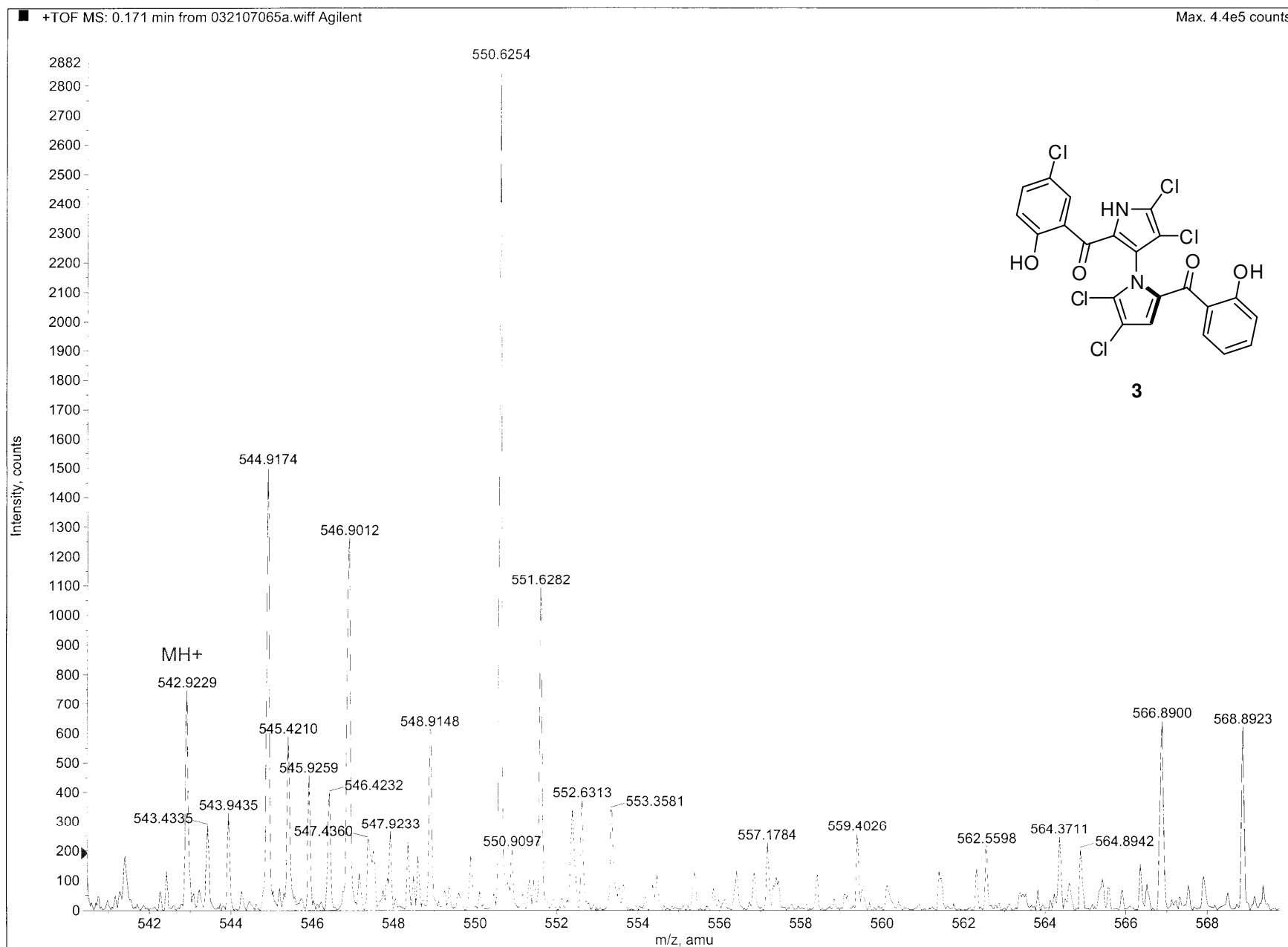


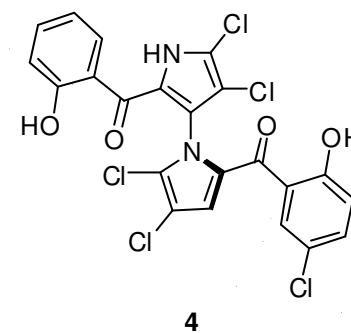
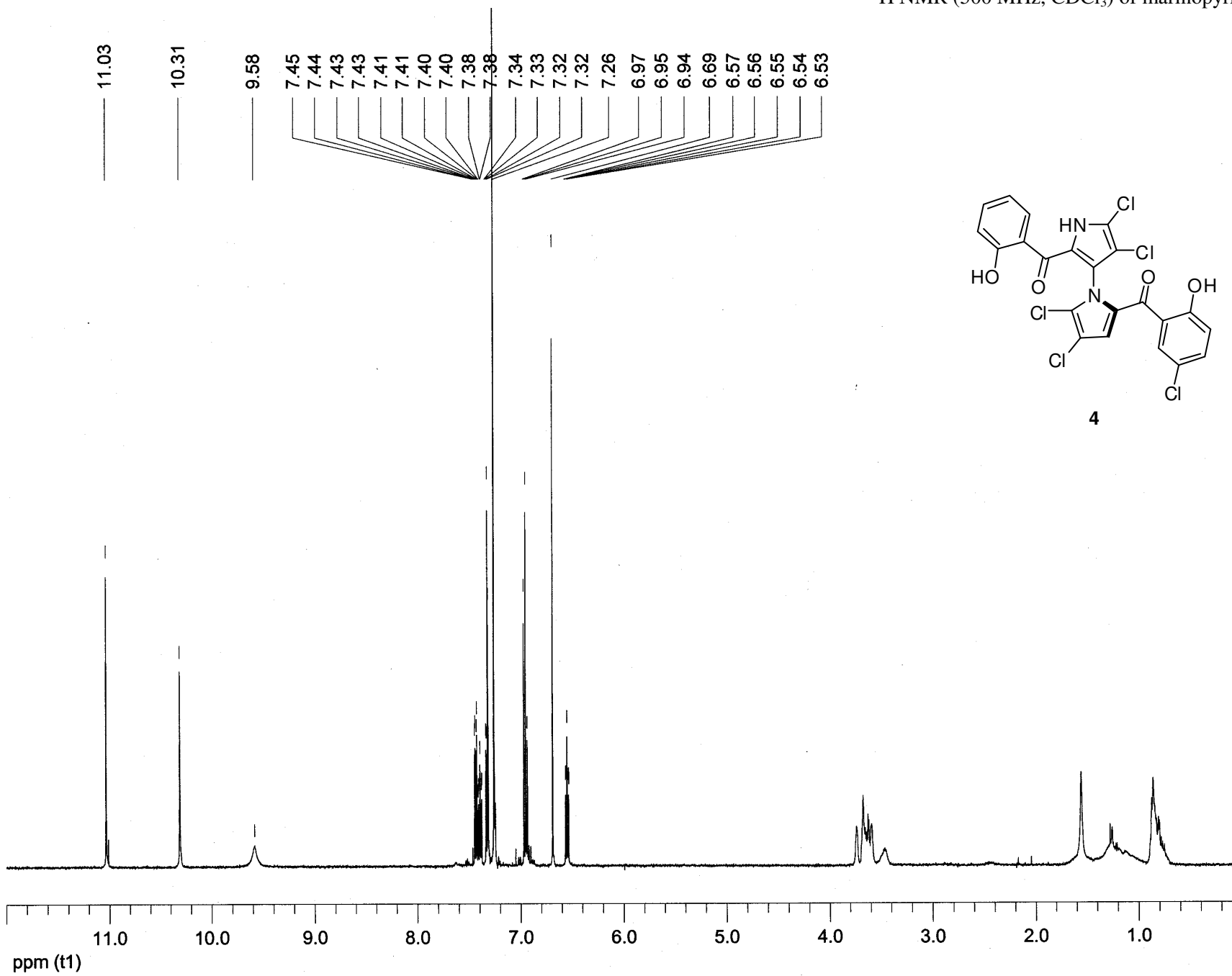
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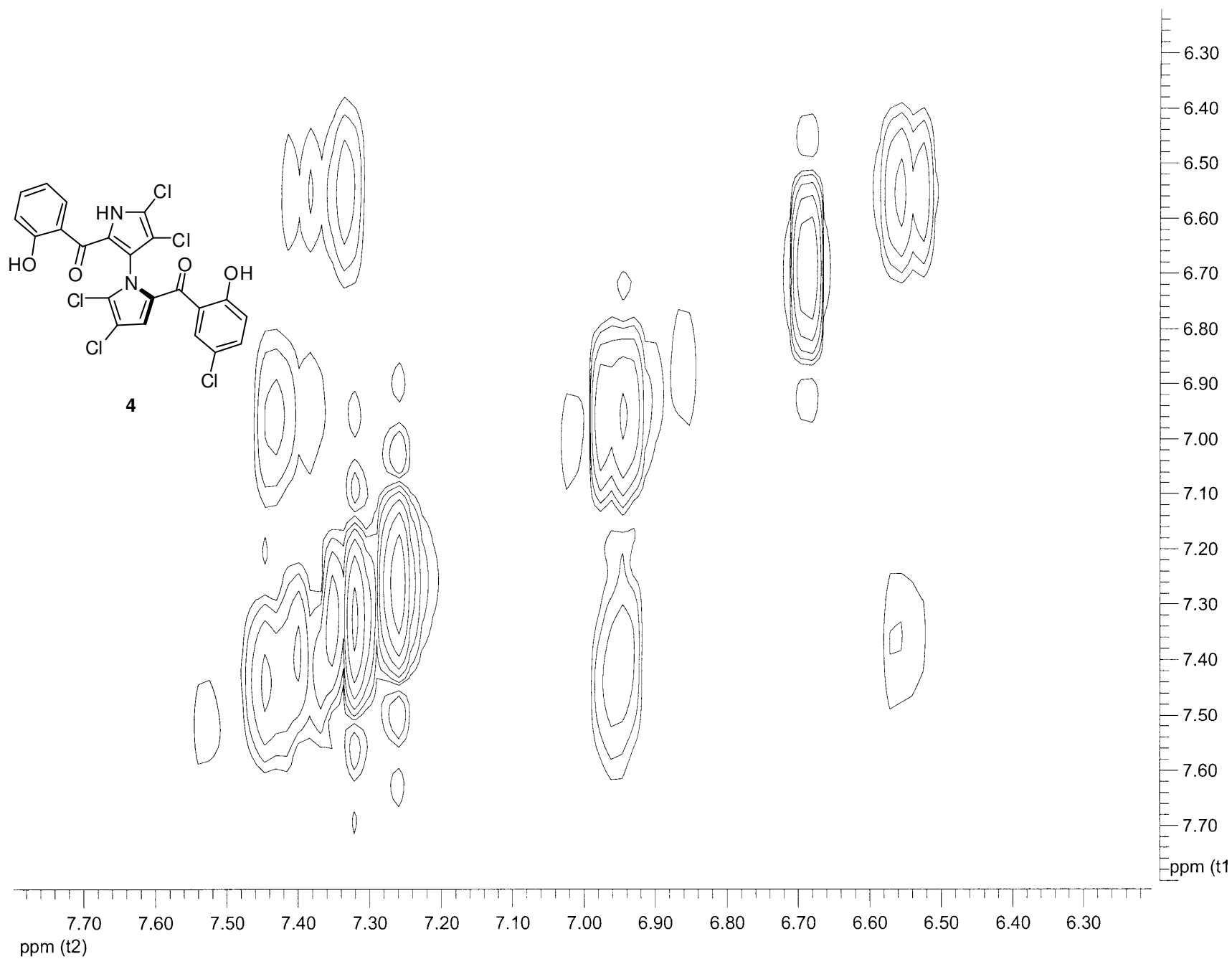
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Acq. Date: Wednesday, March 21, 2007

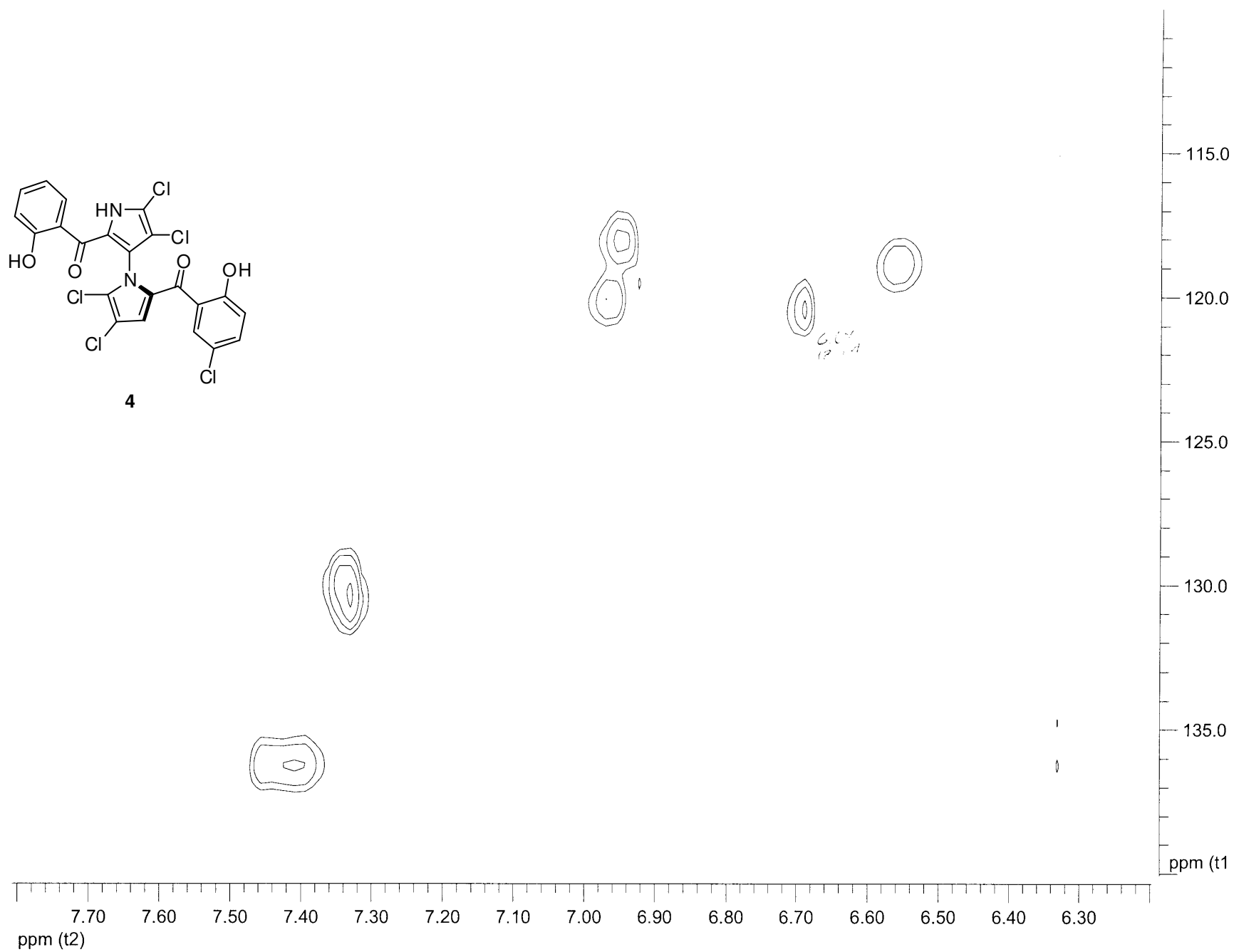
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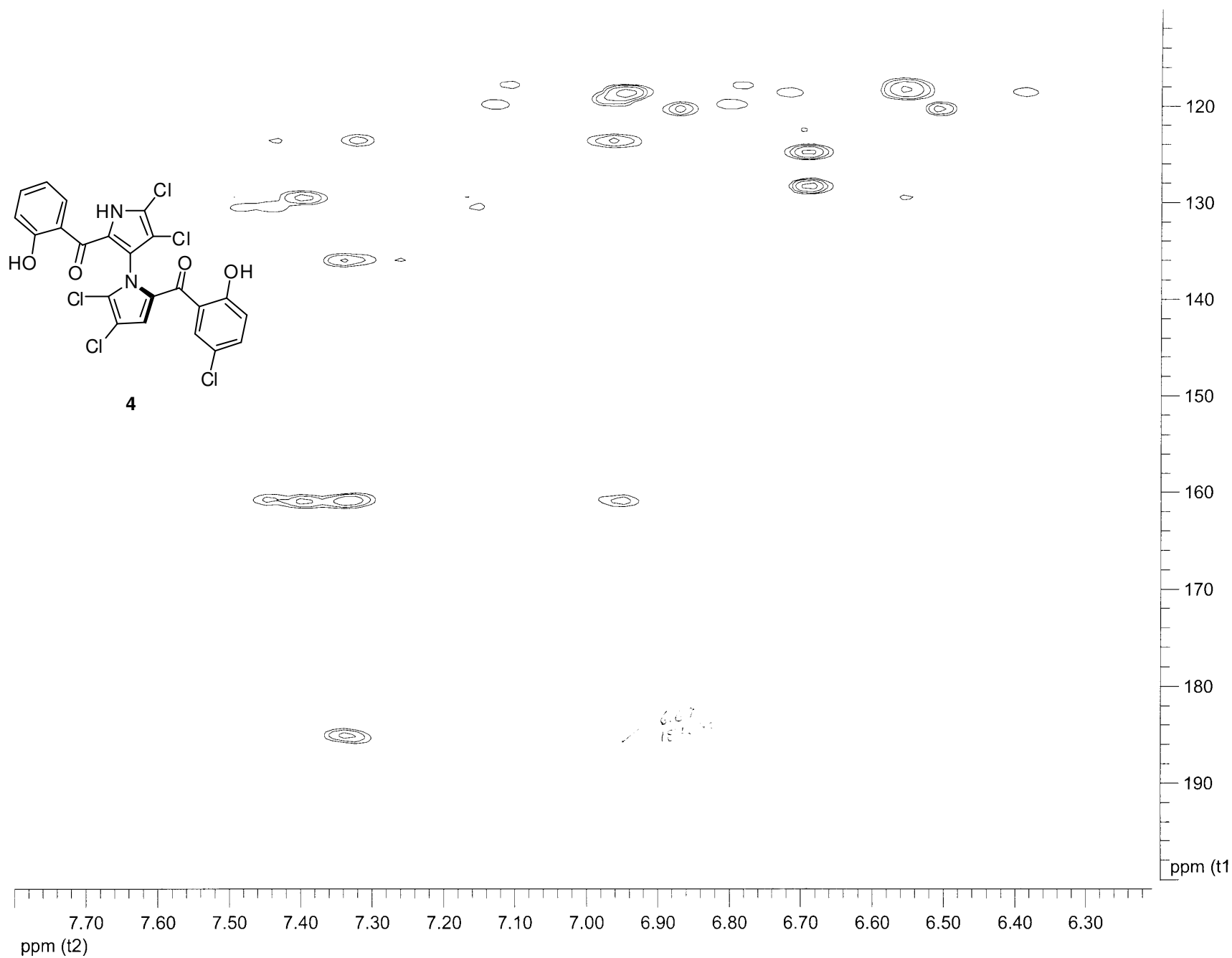
¹H NMR (500 MHz, CDCl₃) of marinopyrrole D (**4**)



HSQC (500 MHz, CDCl₃) of marinopyrrole D (**4**)



HMBC (500 MHz, CDCl₃) of marinopyrrole D (**4**)

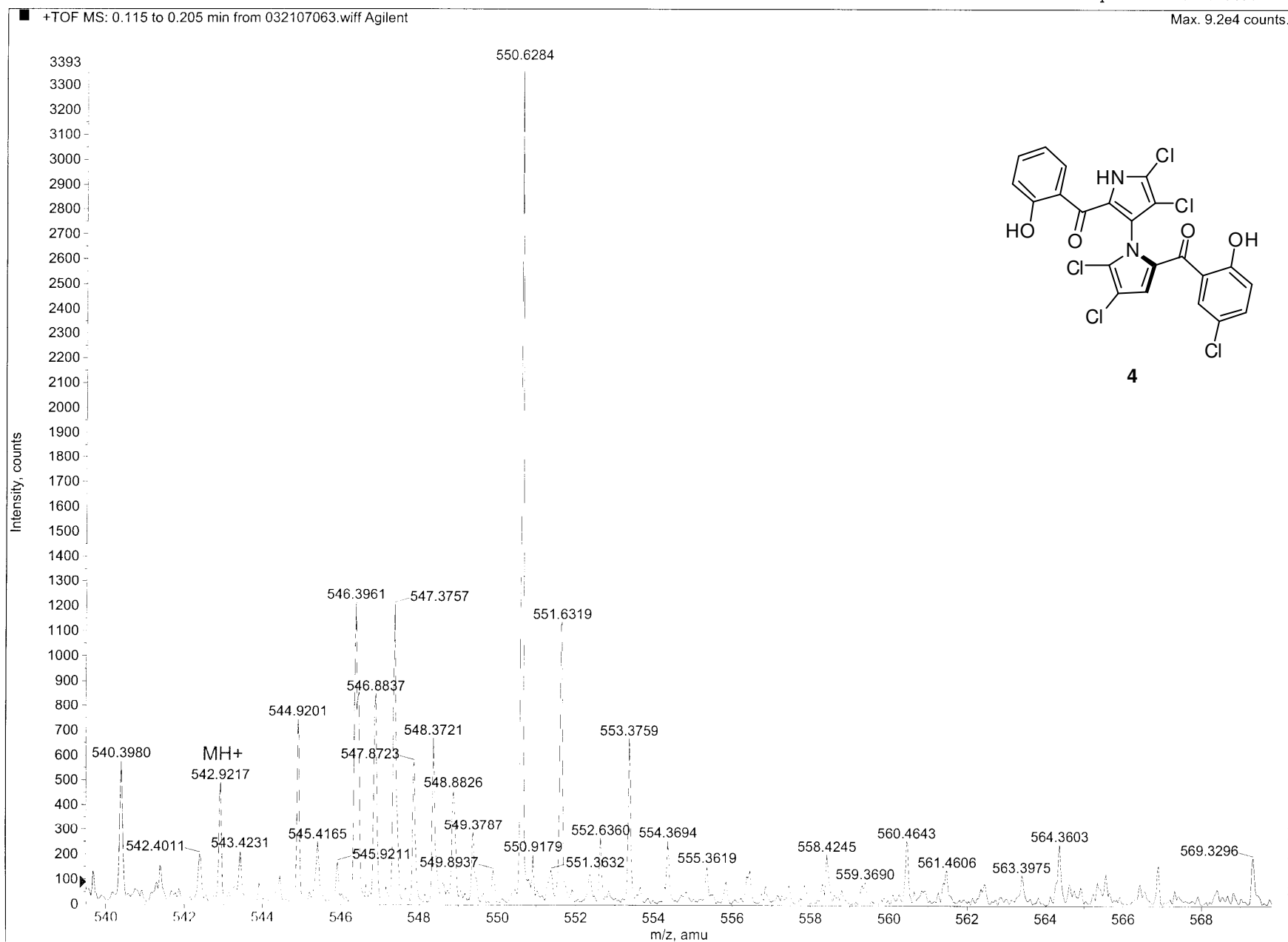


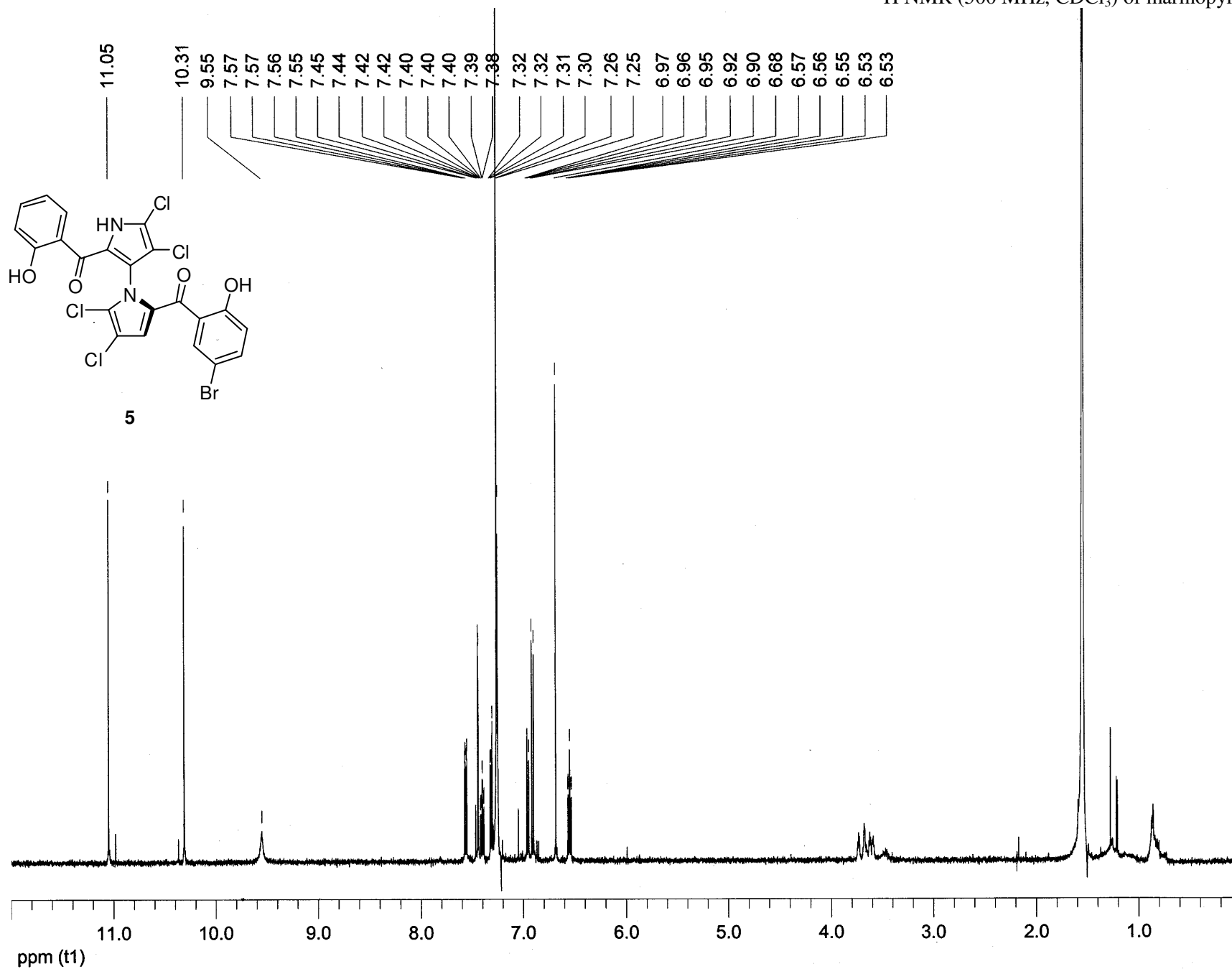
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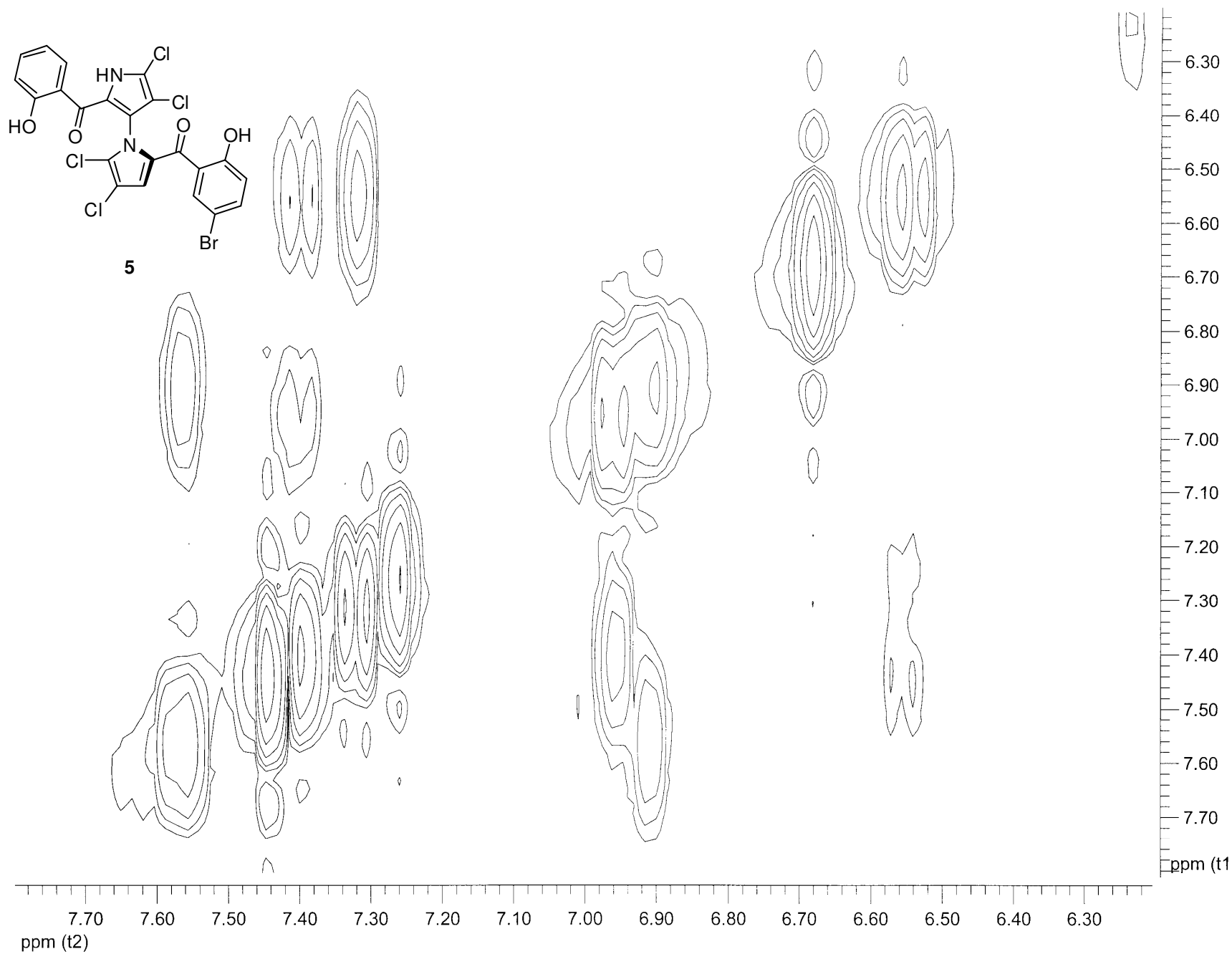
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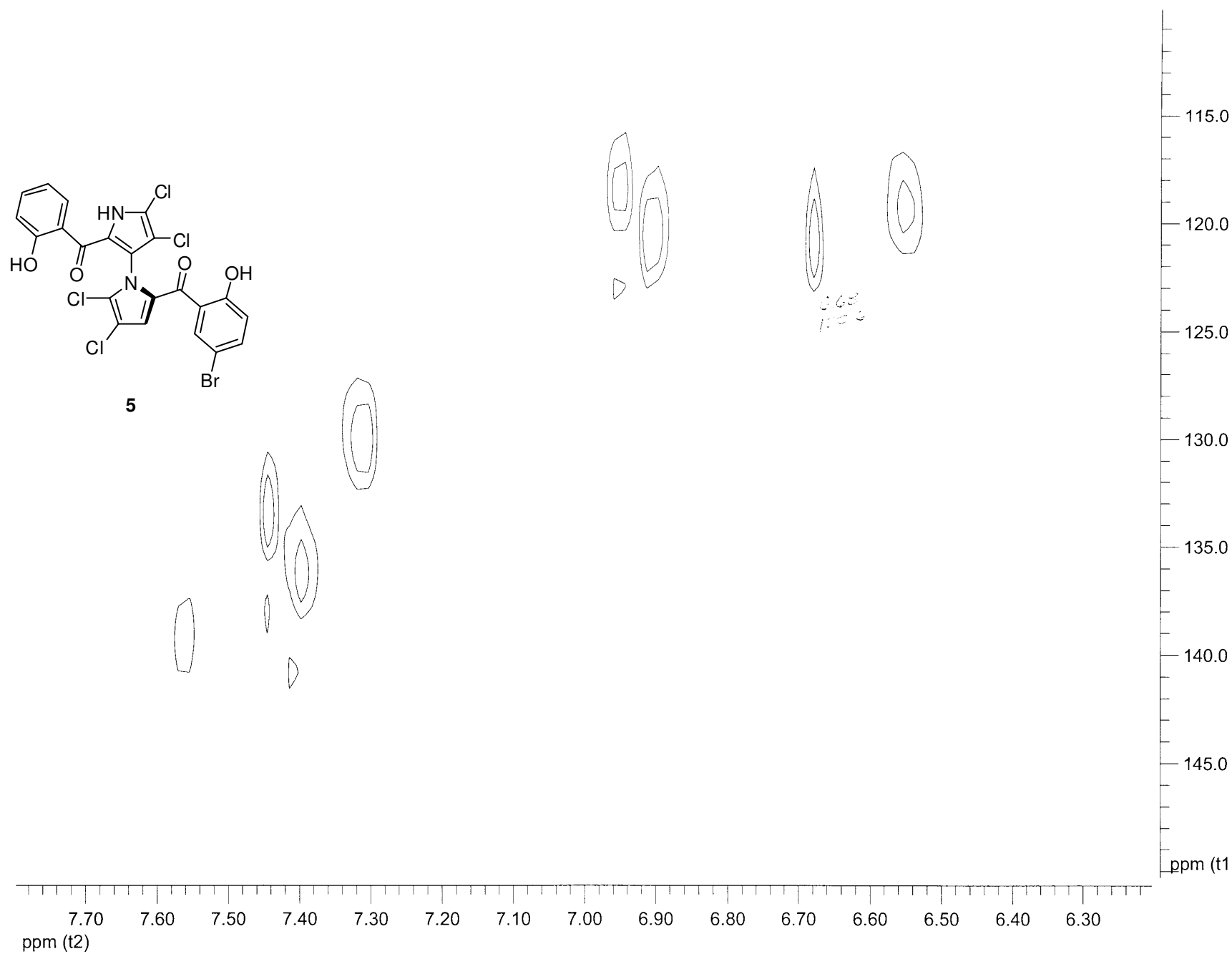




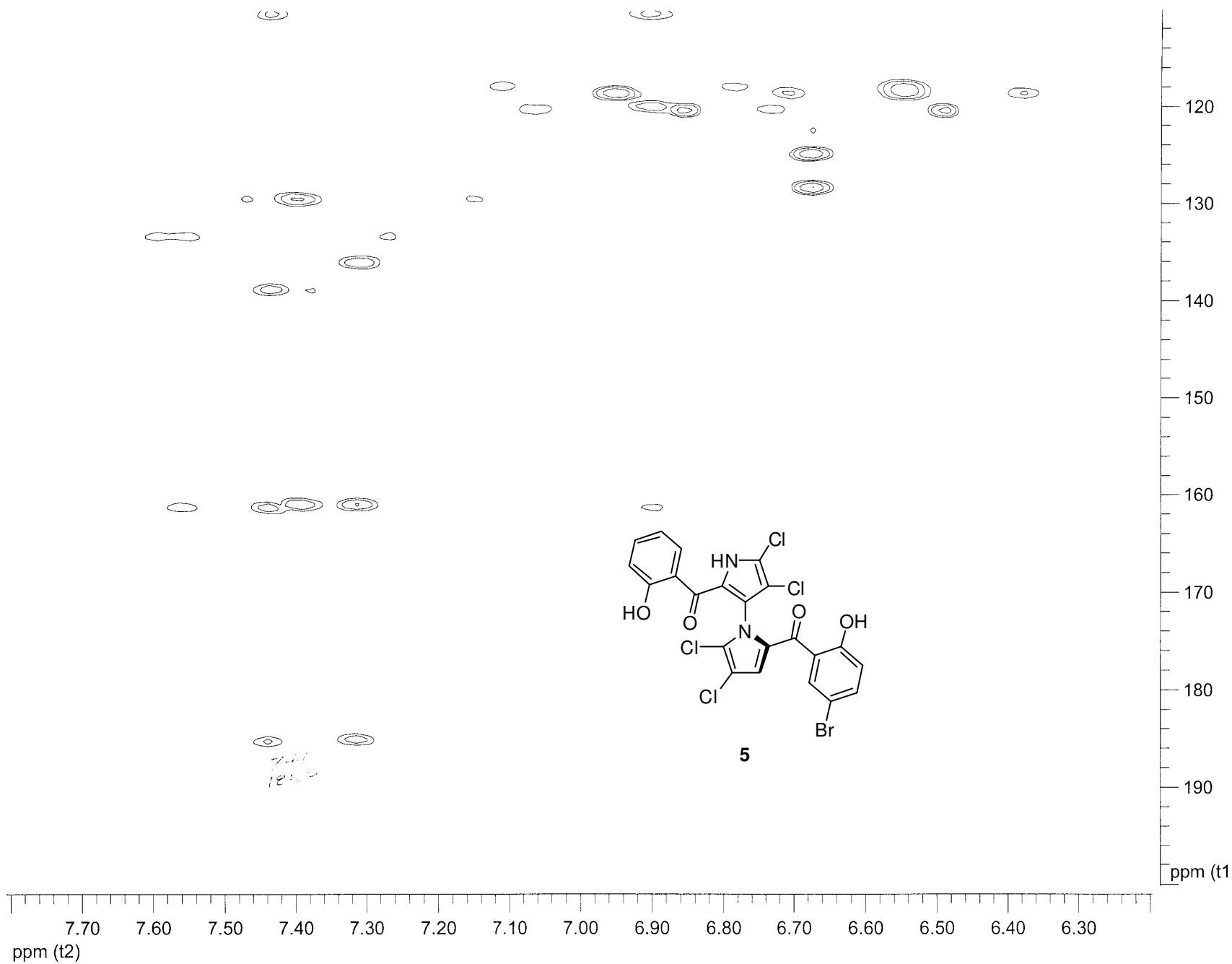
COSY (500 MHz, CDCl₃) of marinopyrrole E (**5**)



HSQC (500 MHz, CDCl₃) of marinopyrrole E (**5**)



HMBC (500 MHz, CDCl₃) of marinopyrrole E (**5**)



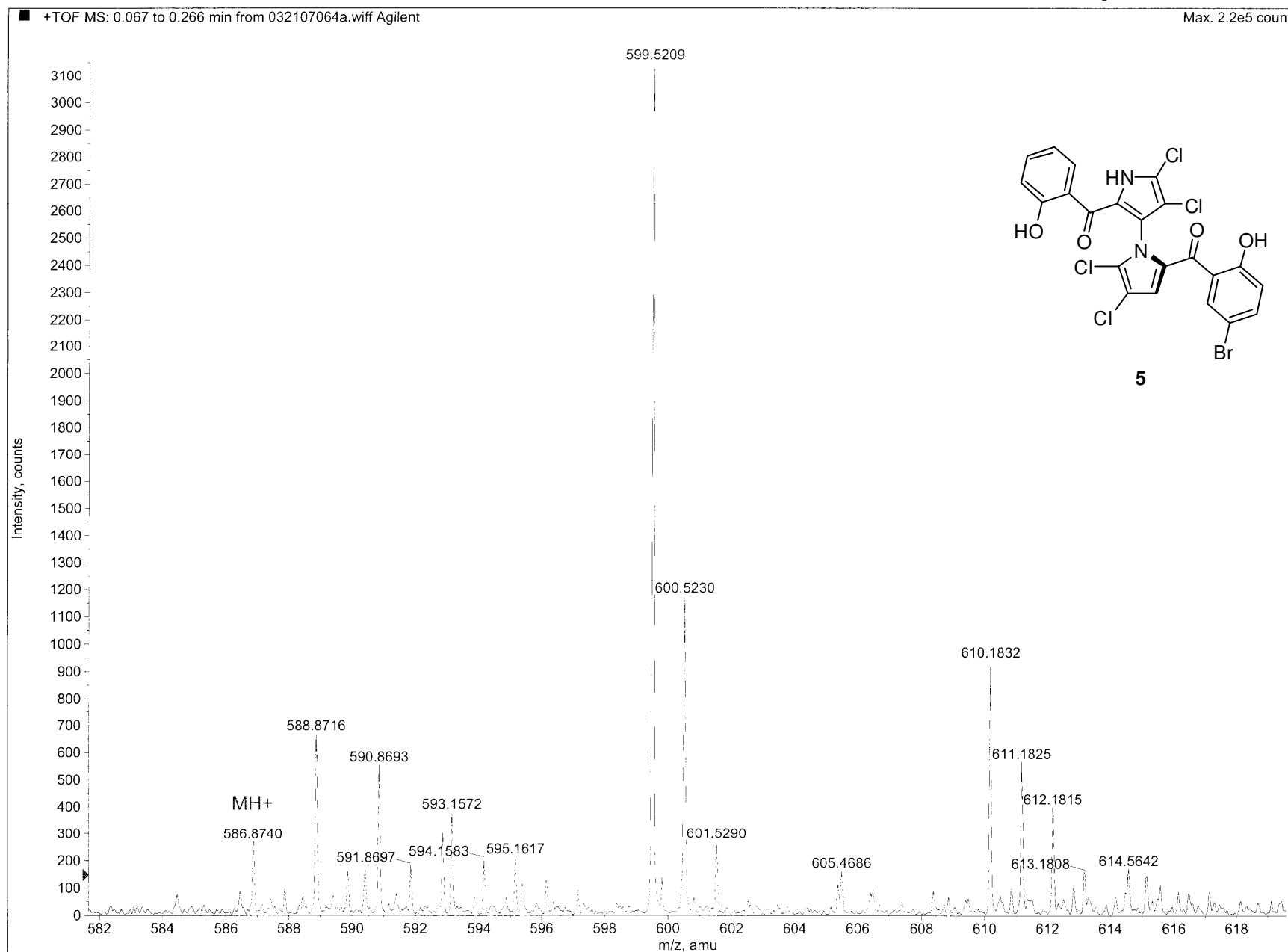
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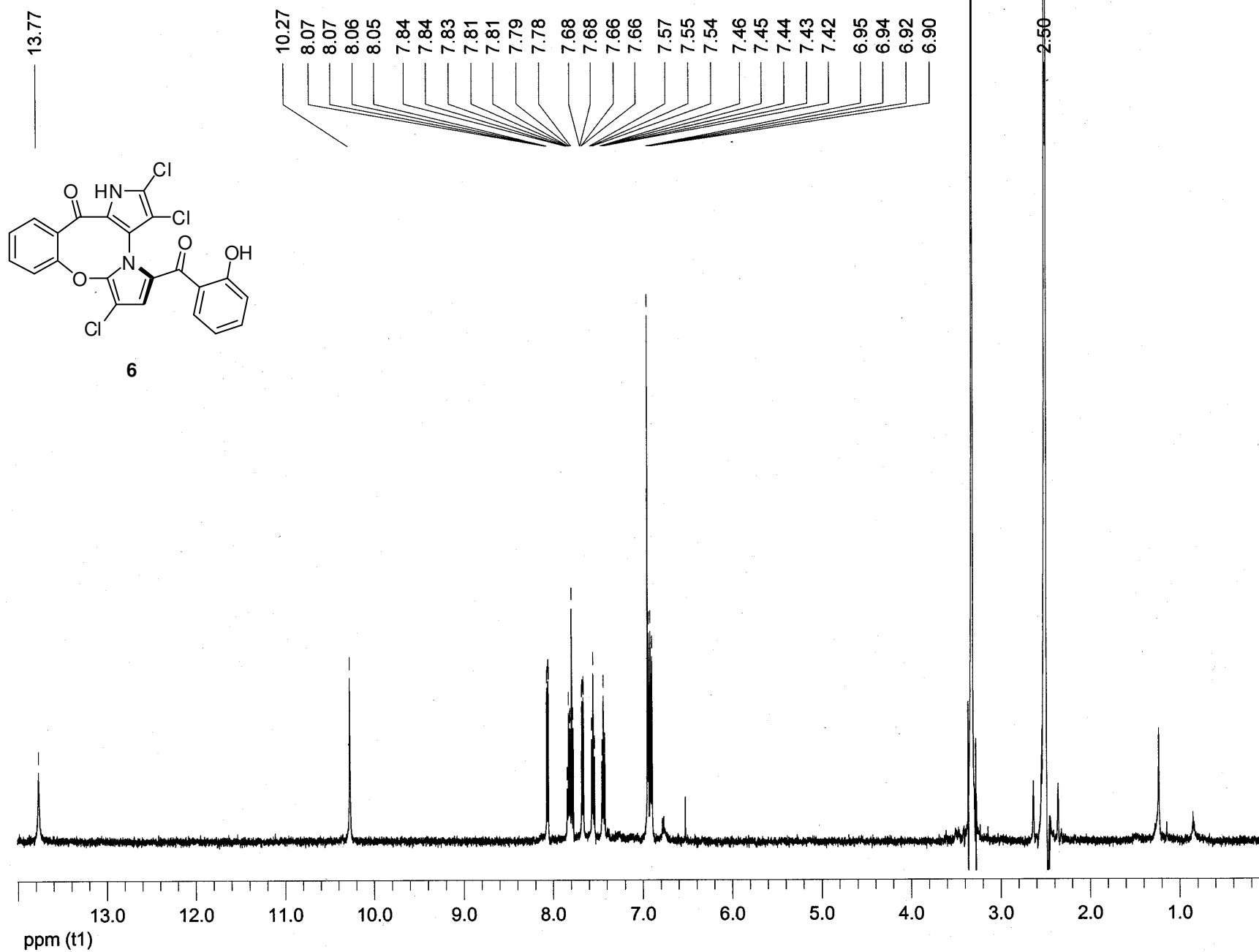
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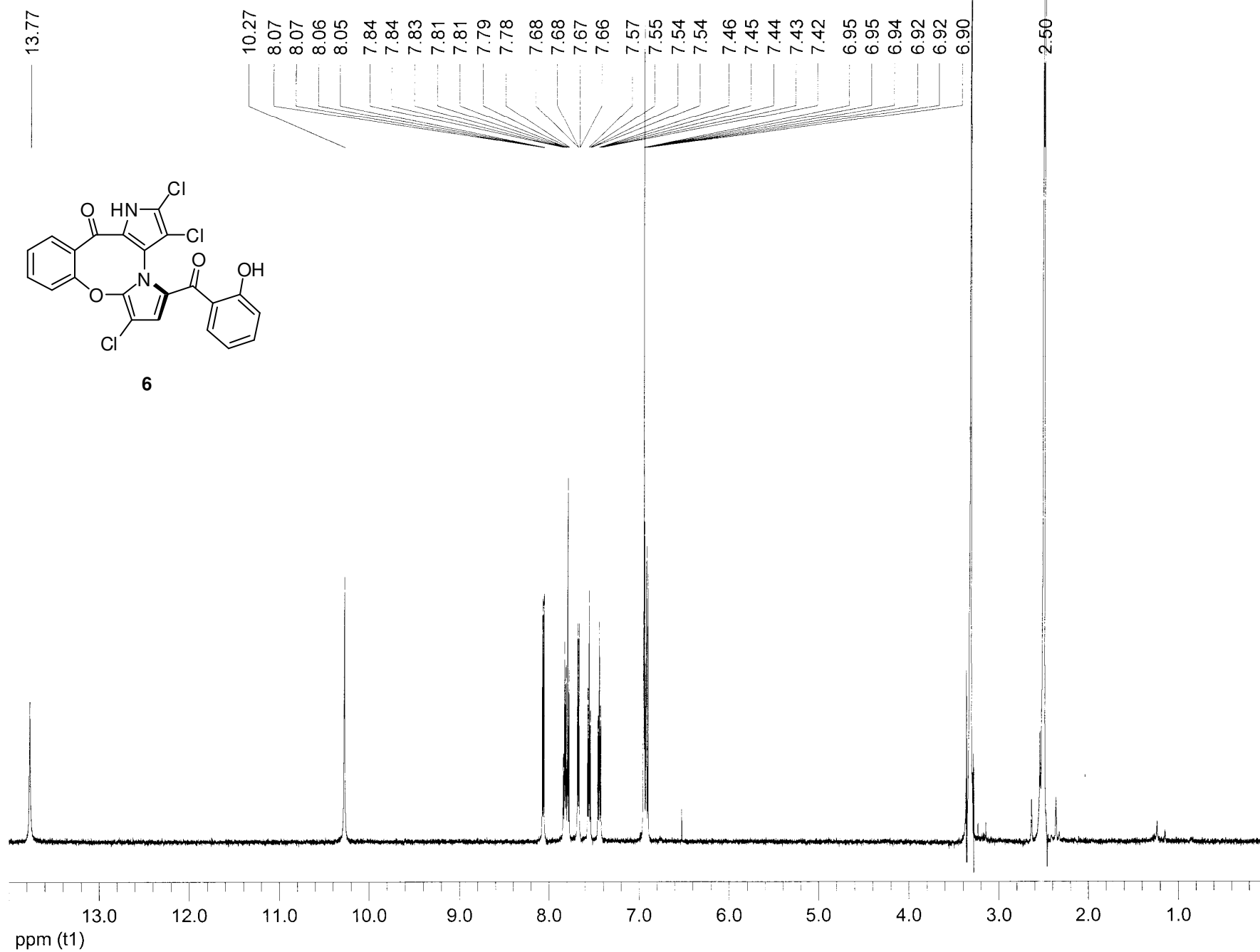
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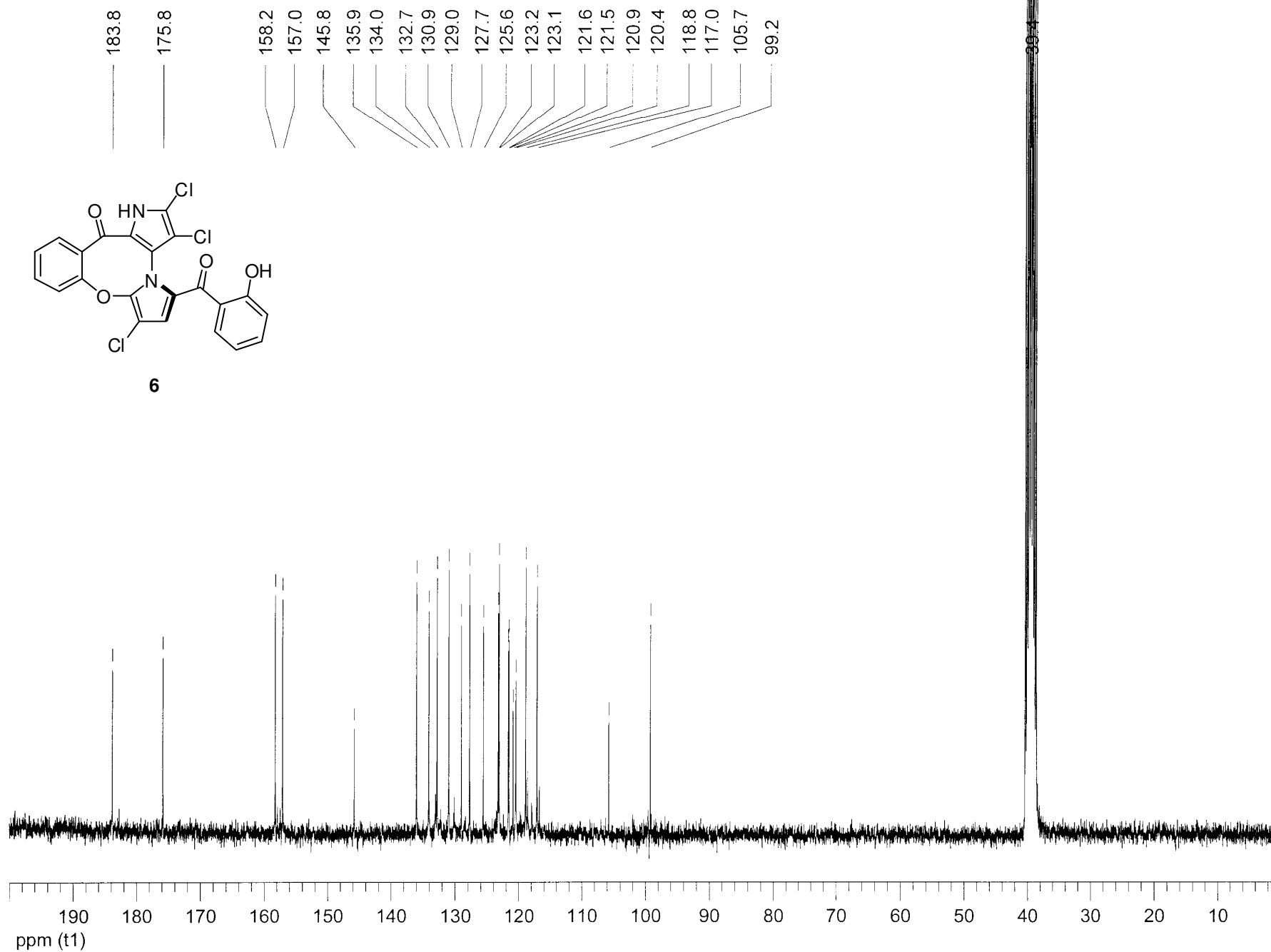
¹H NMR (500 MHz, DMSO-*d*₆) of marinopyrrole F (**6**)



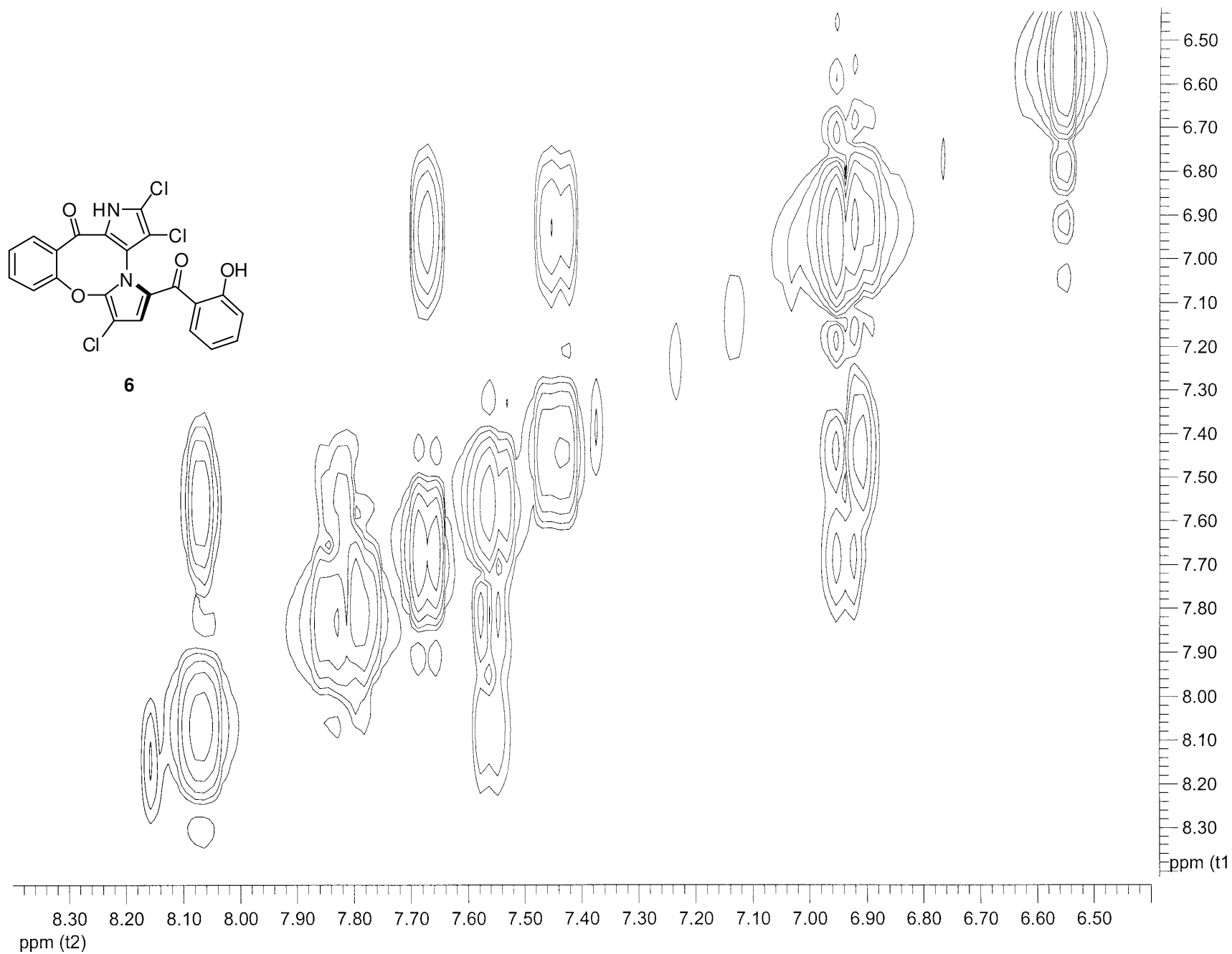
¹H NMR (500 MHz, DMSO-*d*₆) of synthetic marinopyrrole F (**6**)



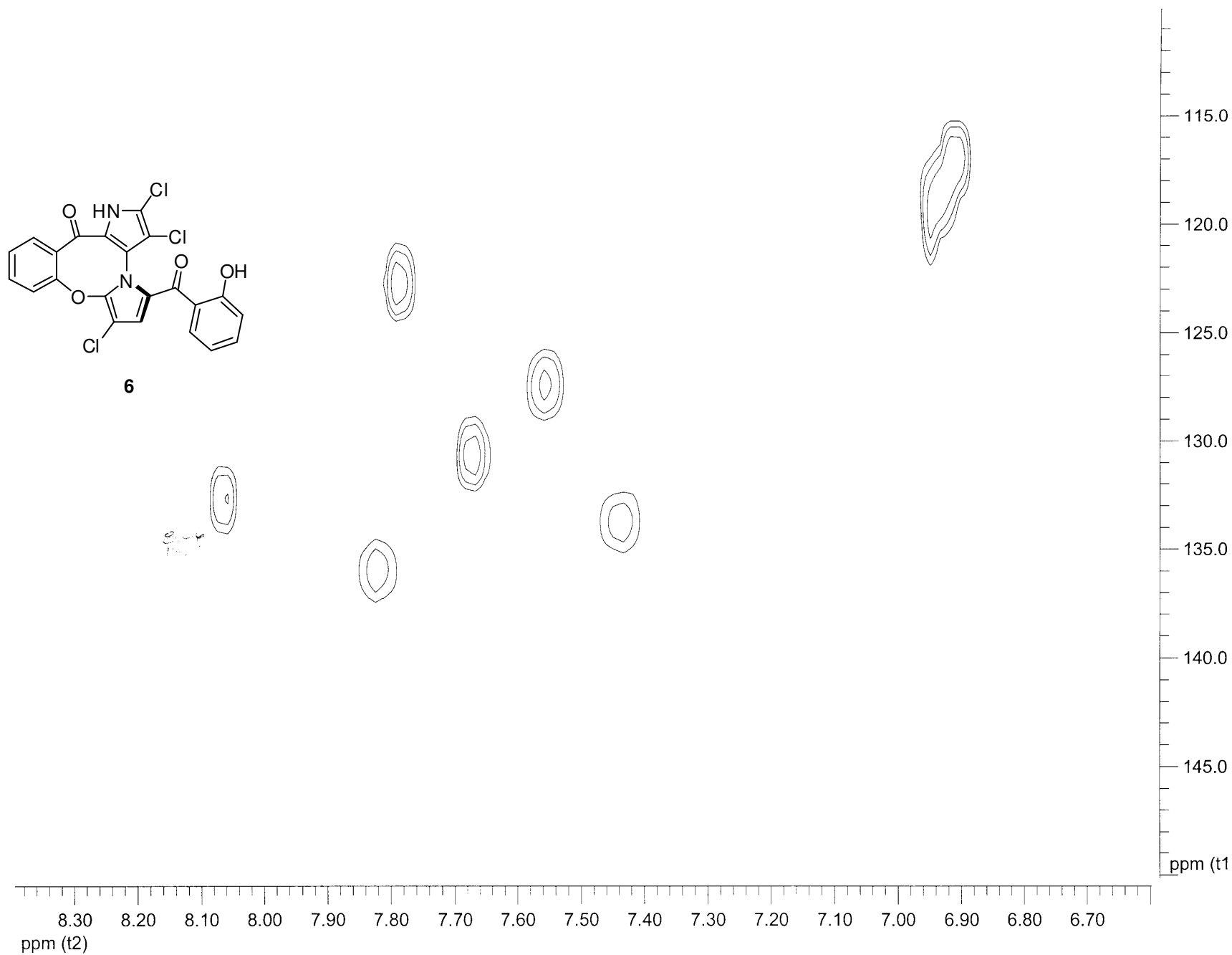
¹³C NMR (75 MHz, DMSO-*d*₆) of synthetic marinopyrrole F (**6**)



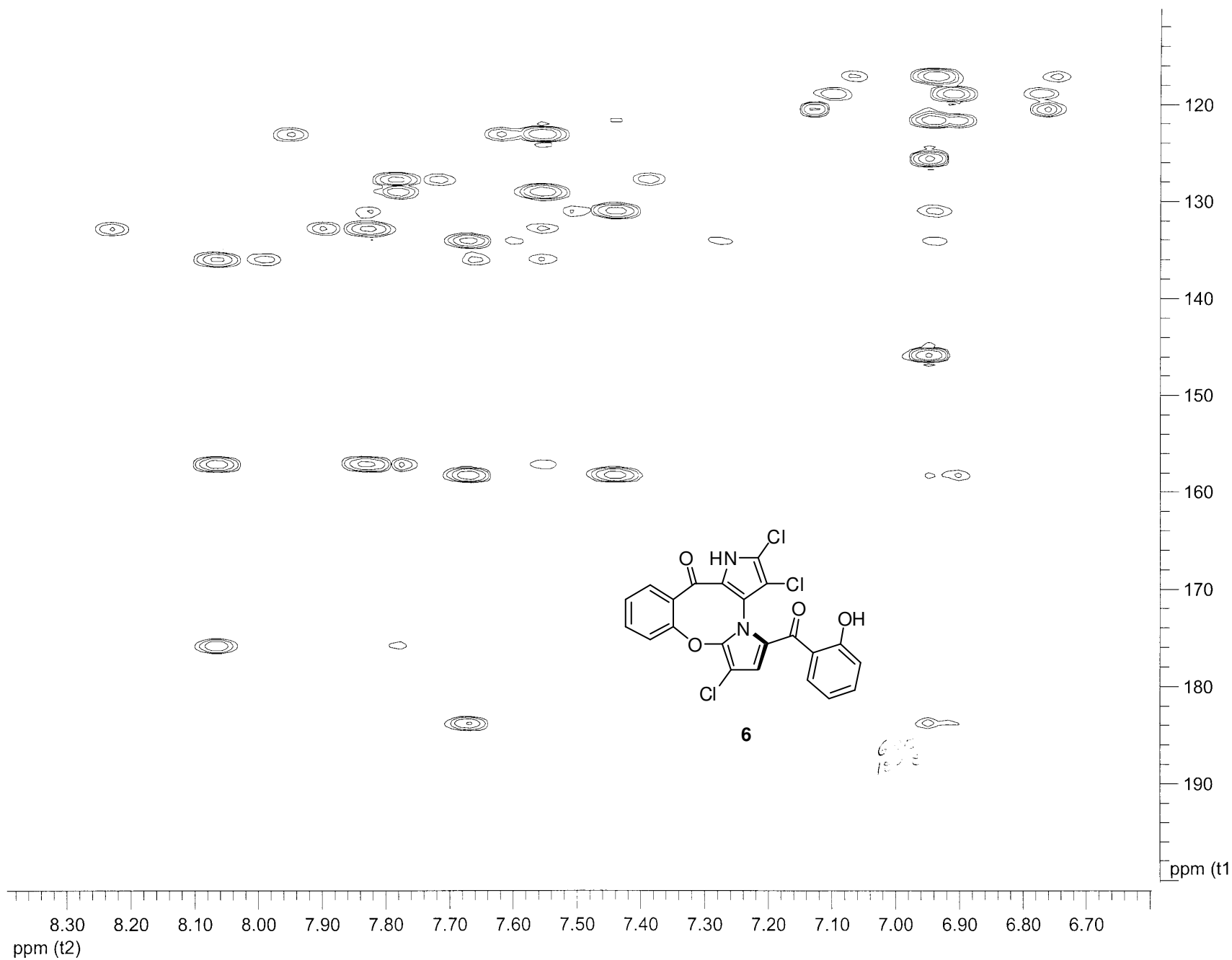
COSY NMR (500 MHz, DMSO-*d*₆) of synthetic marinopyrrole F (**6**)



HSQC NMR (500 MHz, DMSO-*d*₆) of synthetic marinopyrrole F (**6**)



HMBC NMR (500 MHz, DMSO-*d*₆) of synthetic marinopyrrole F (**6**)



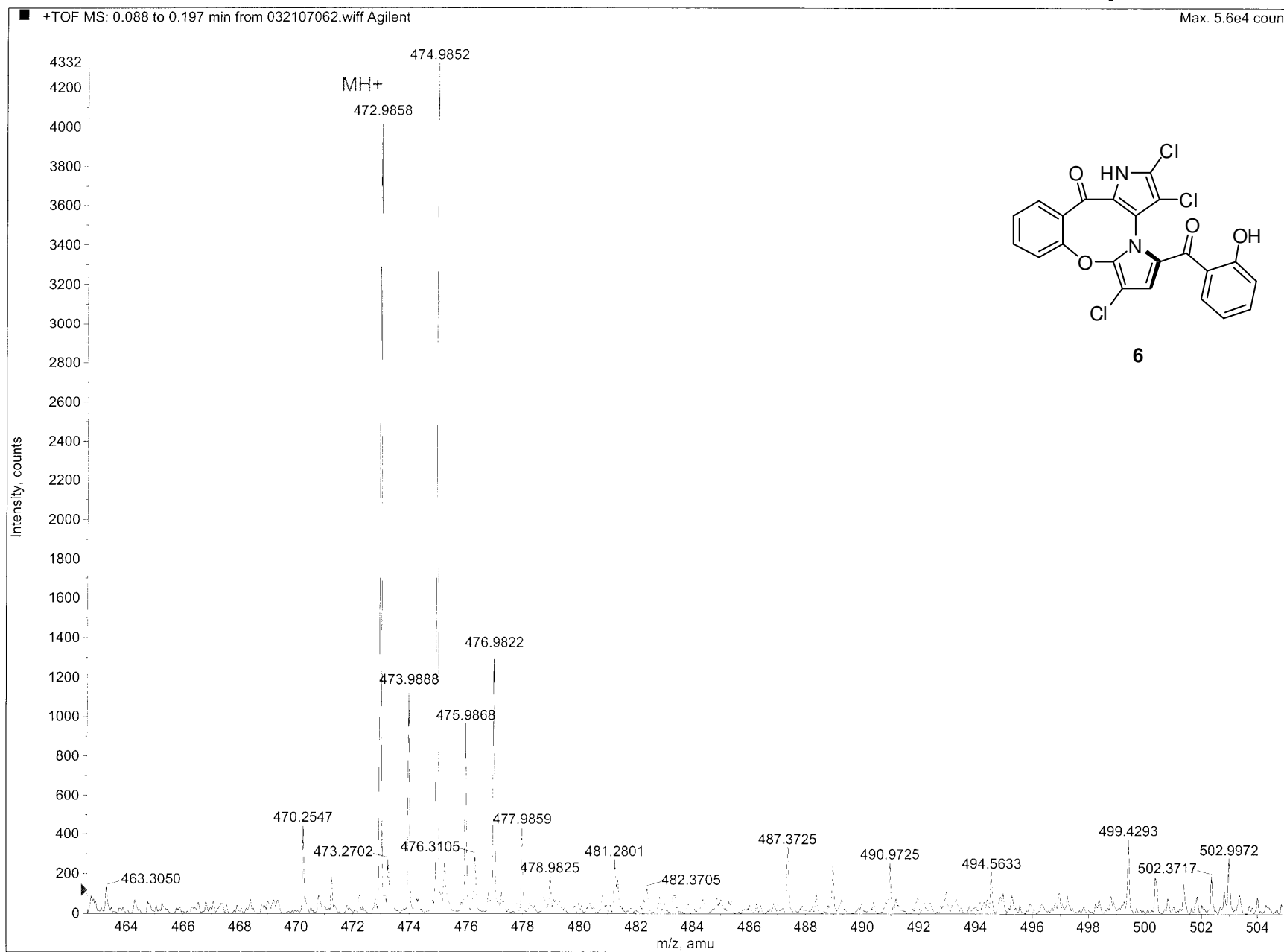
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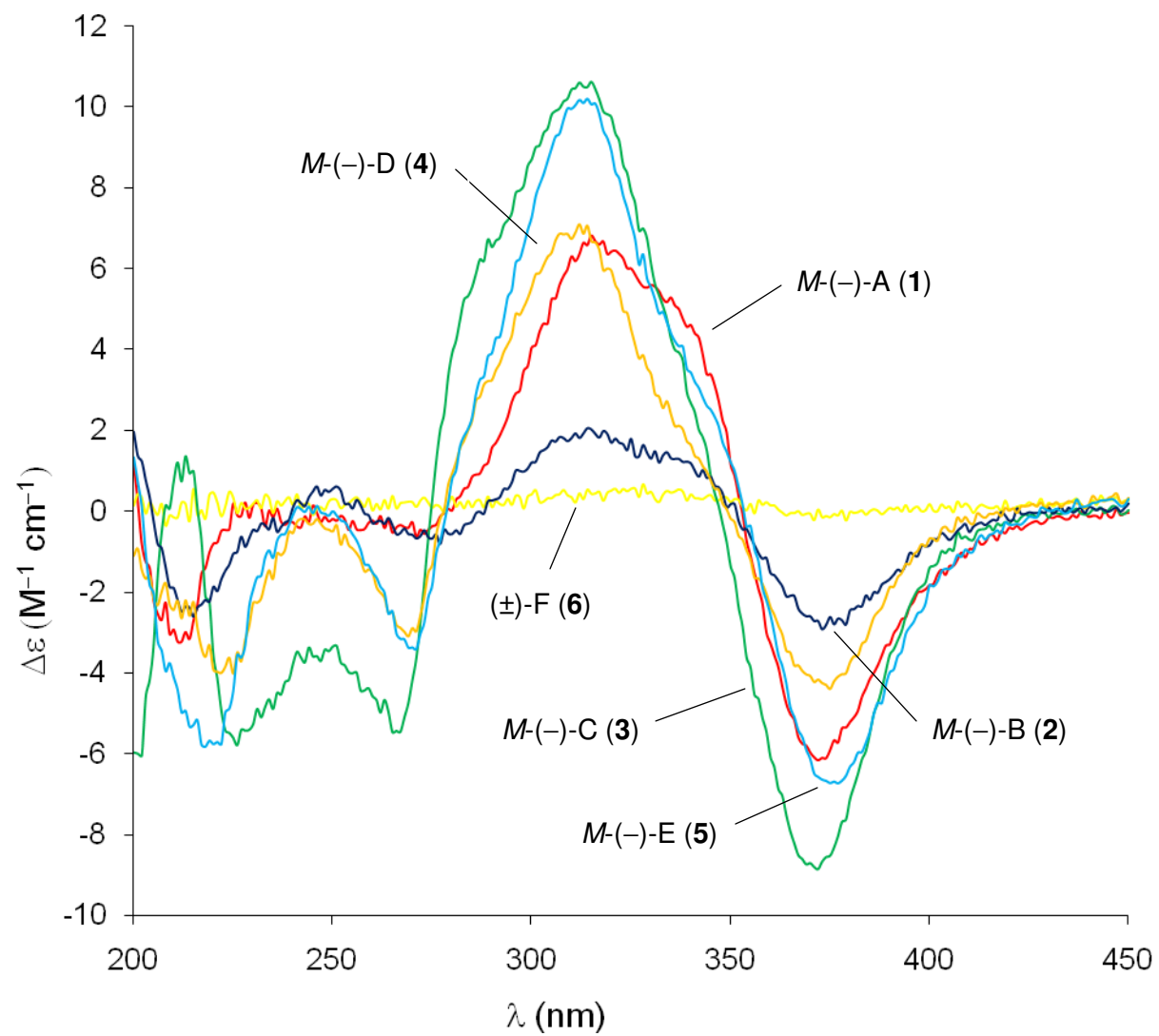
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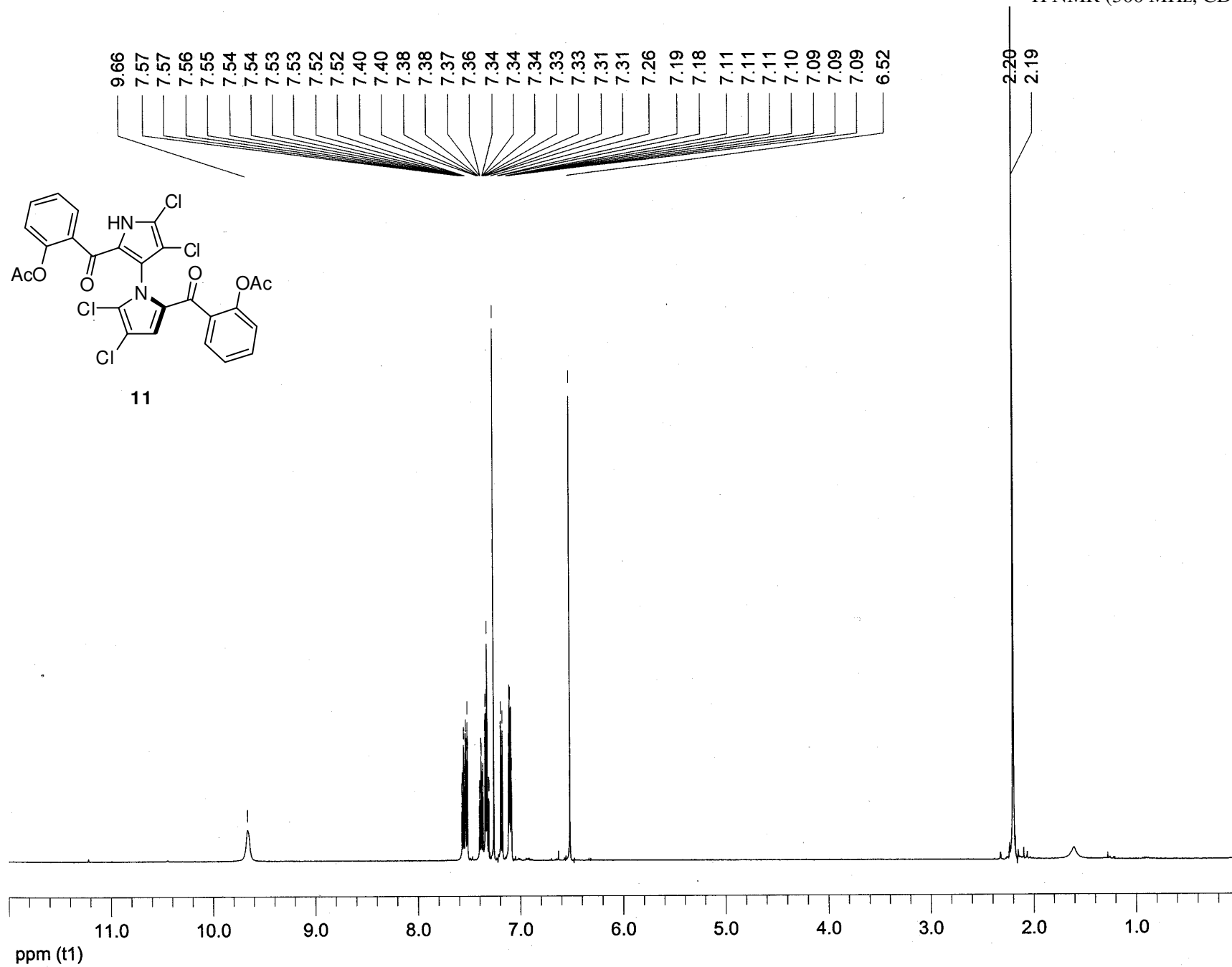
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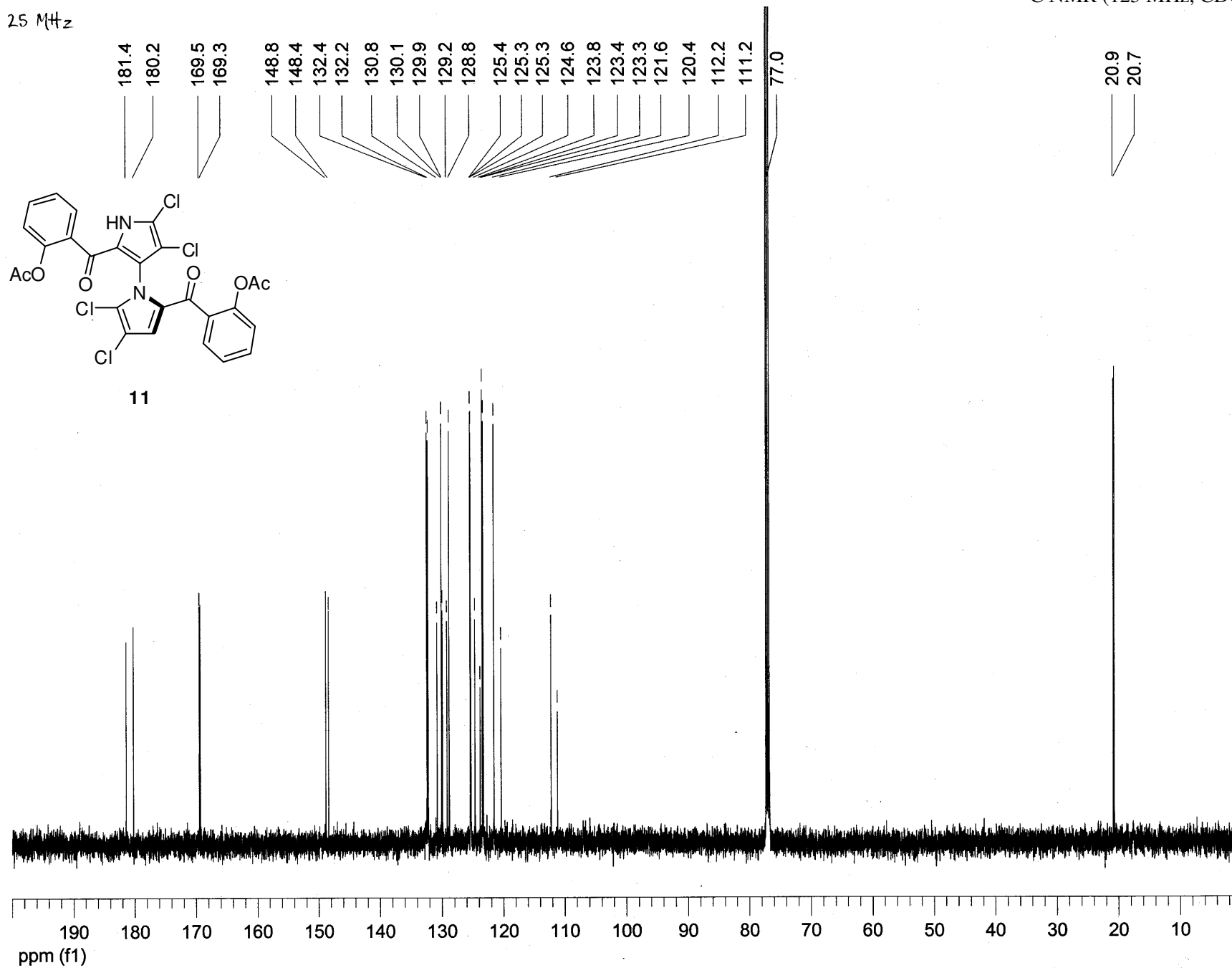
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25 MHz

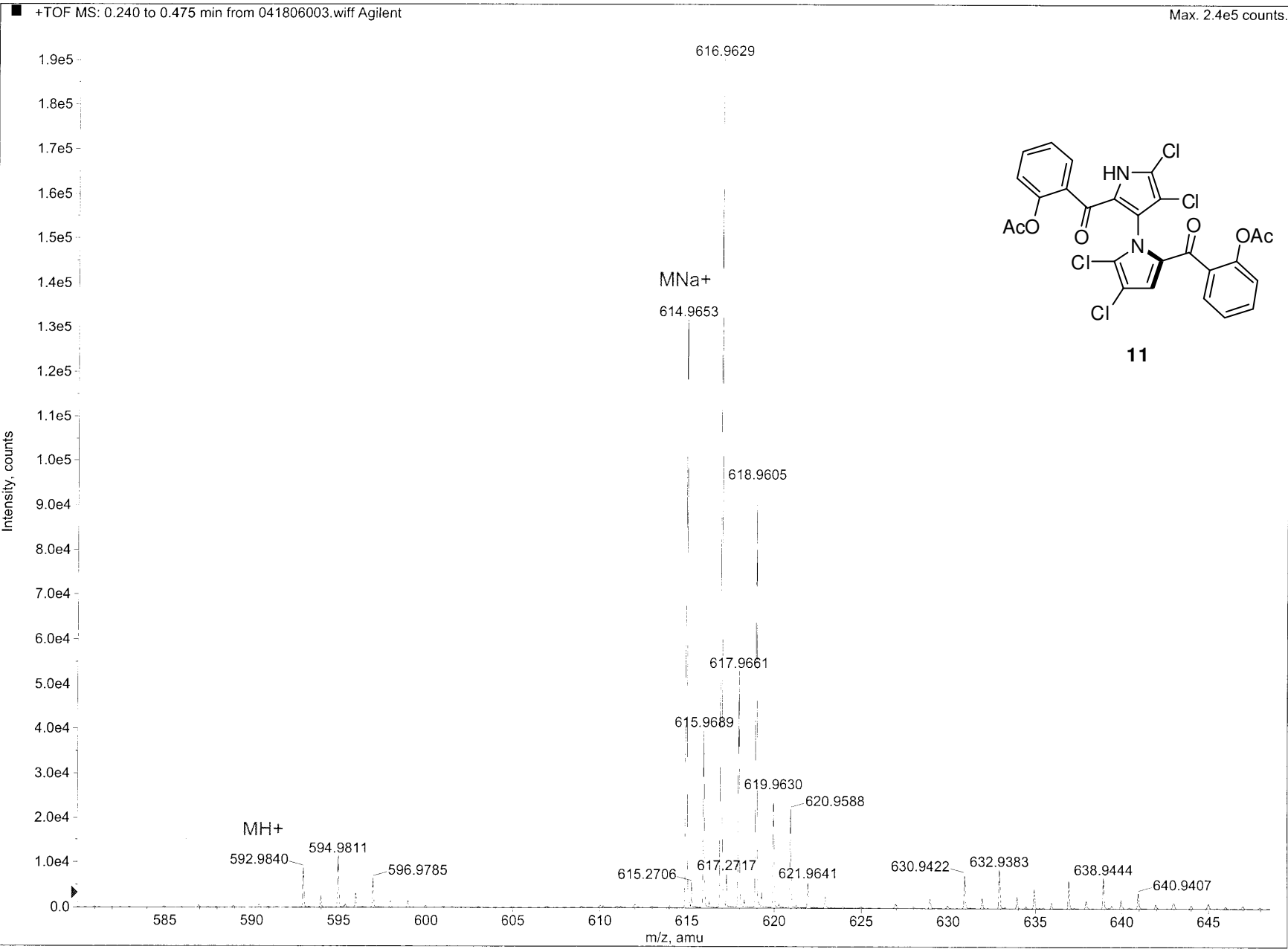


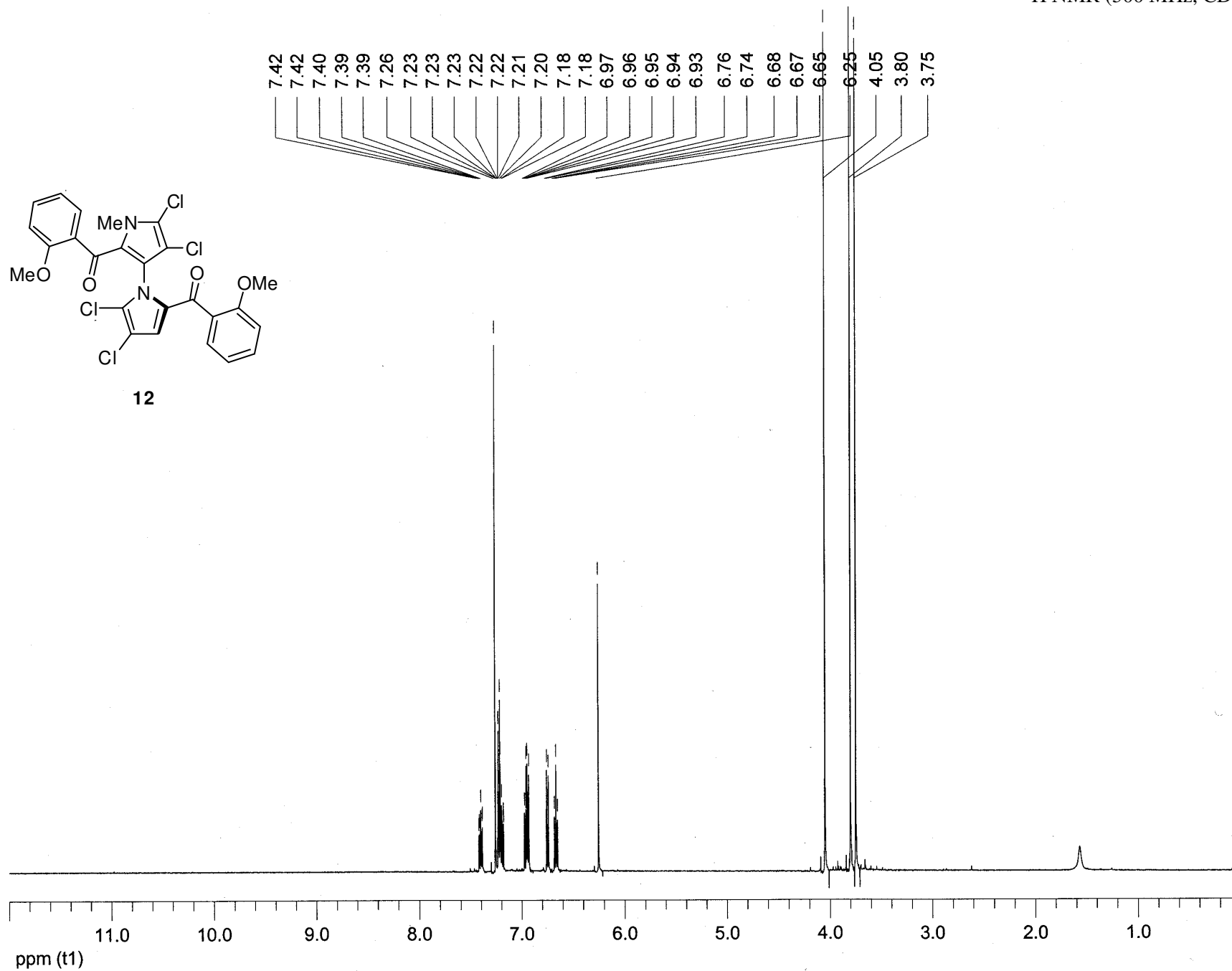
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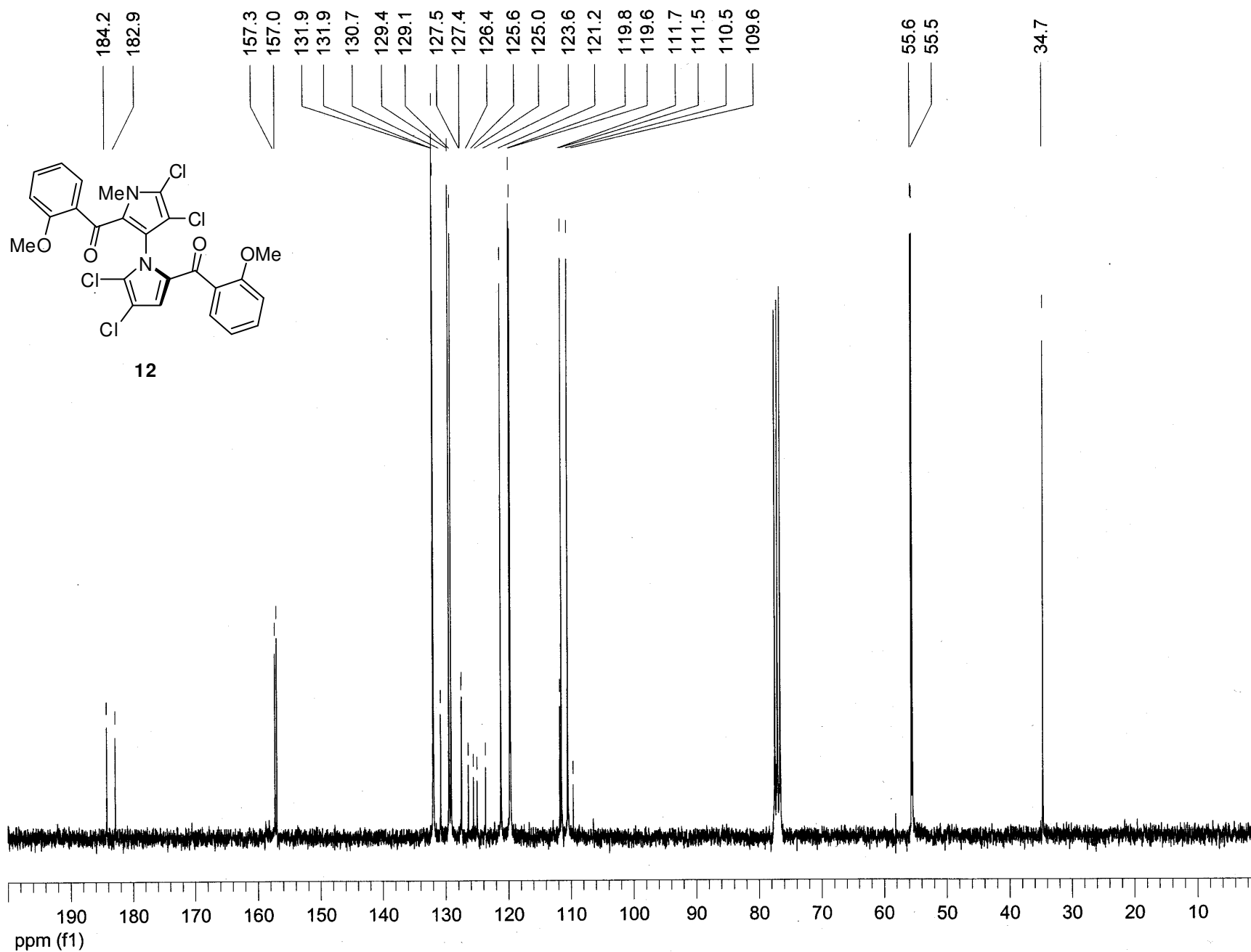
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Acq. File: 041806003.wiff





5 MHz

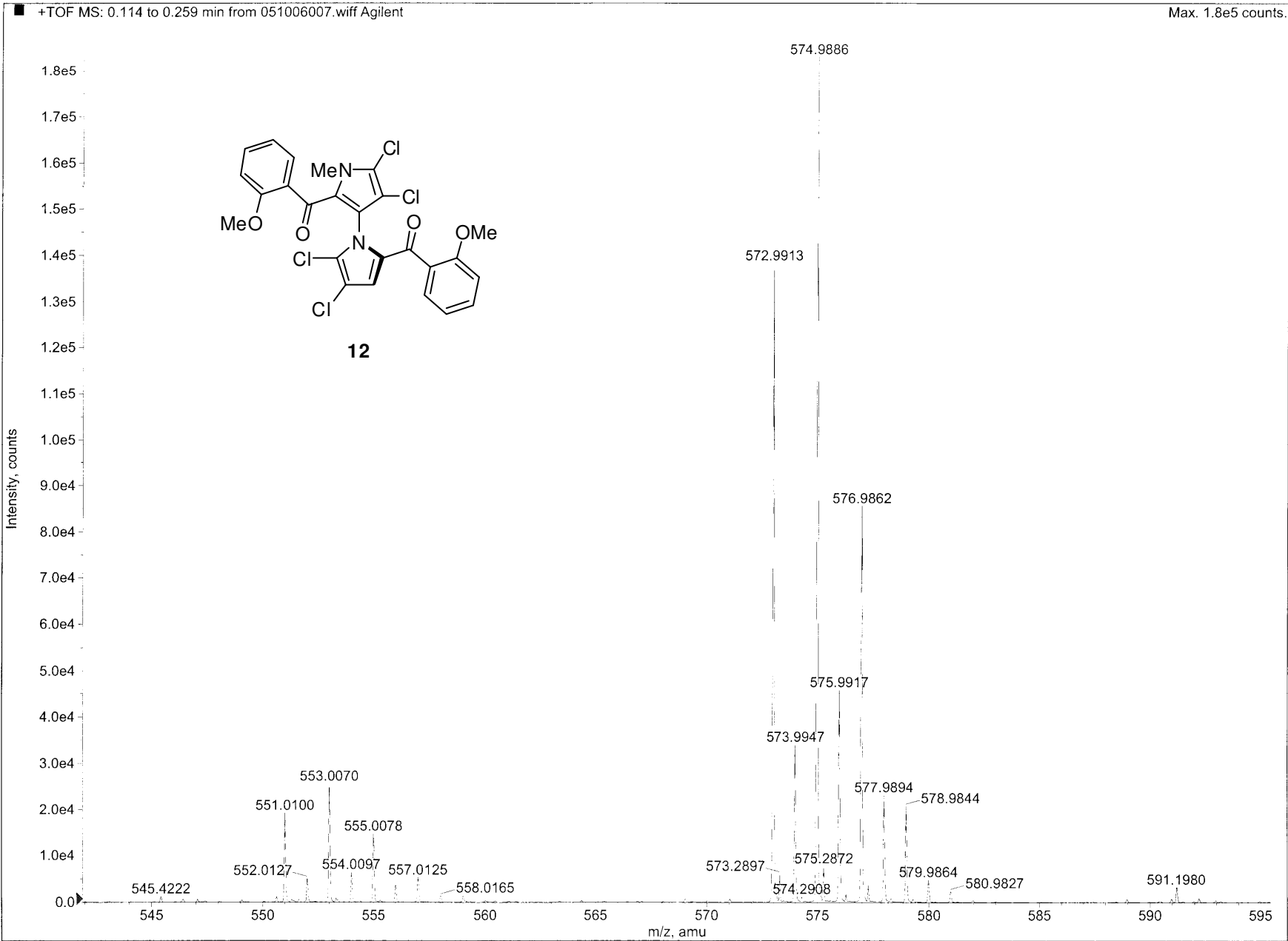


Polarity/Scan Type: Positive

Sample Name: flch3956

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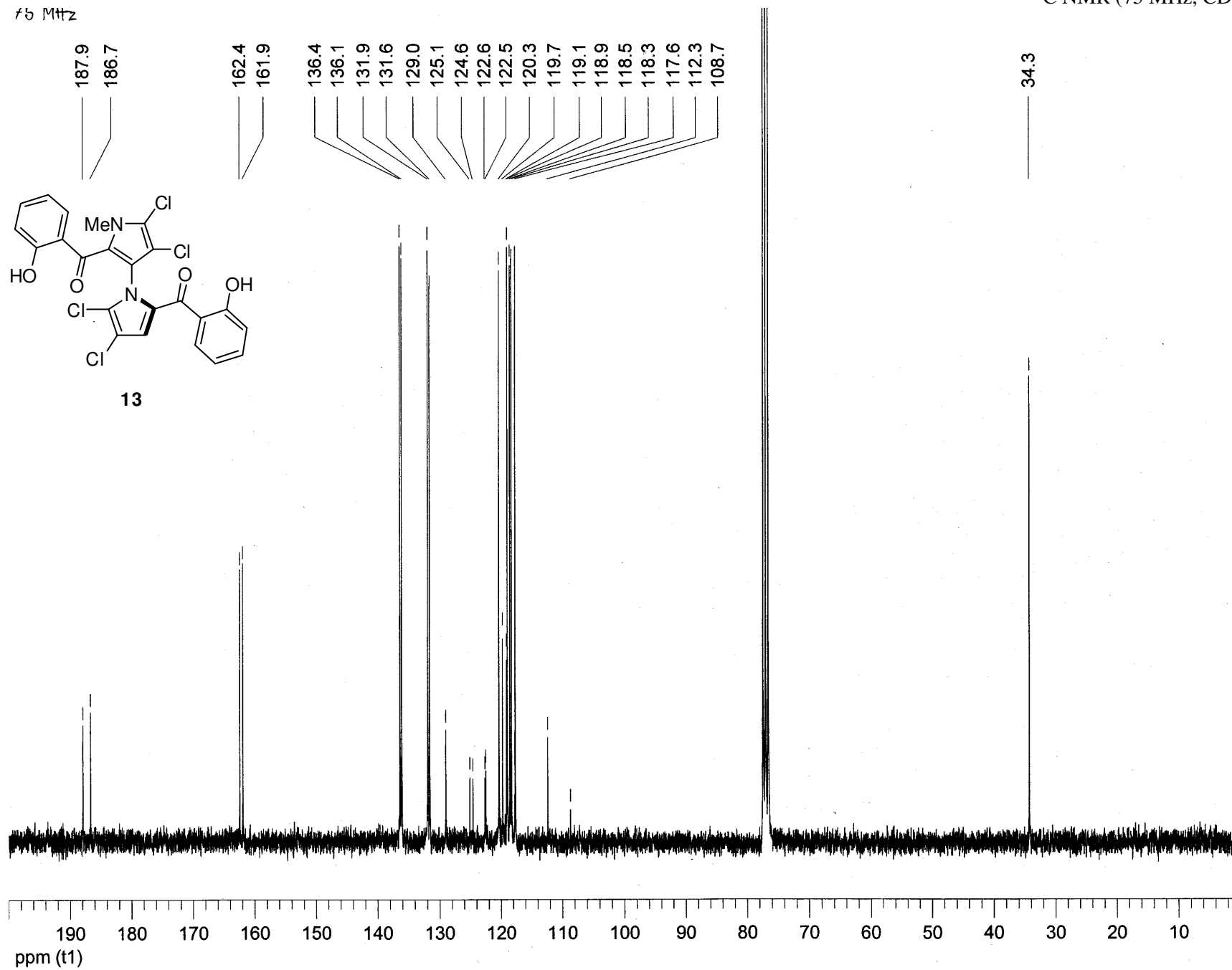
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34.3

13

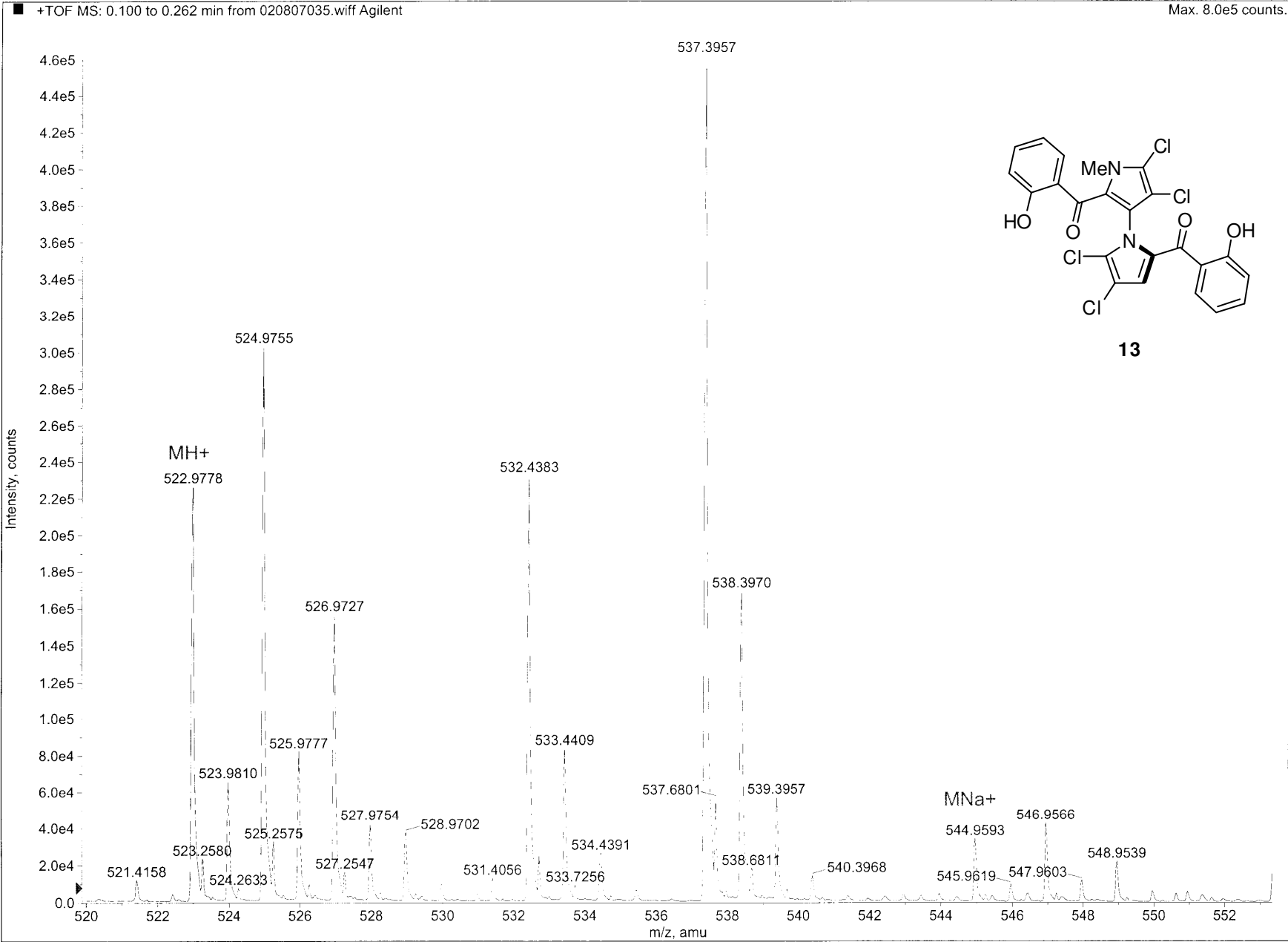


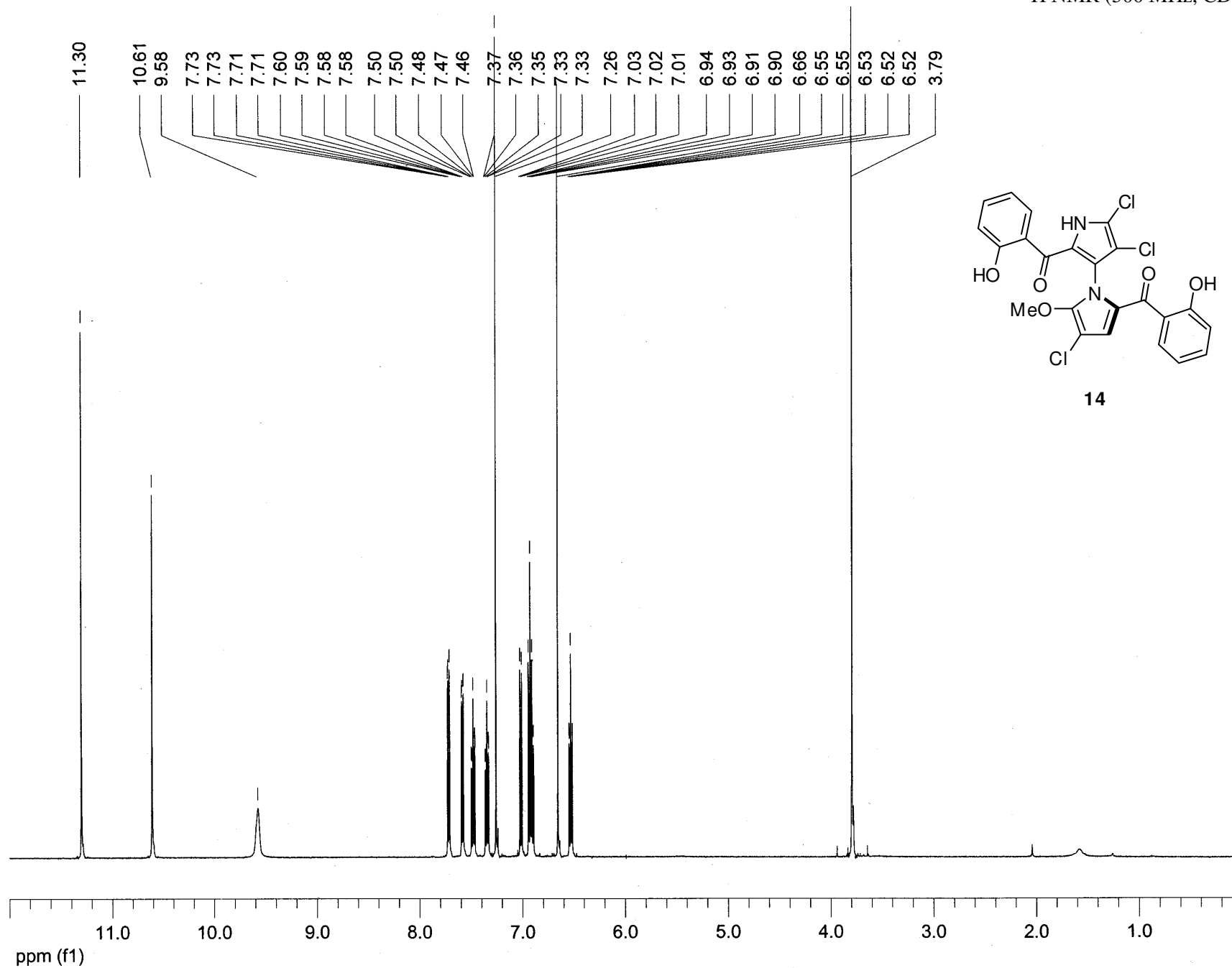
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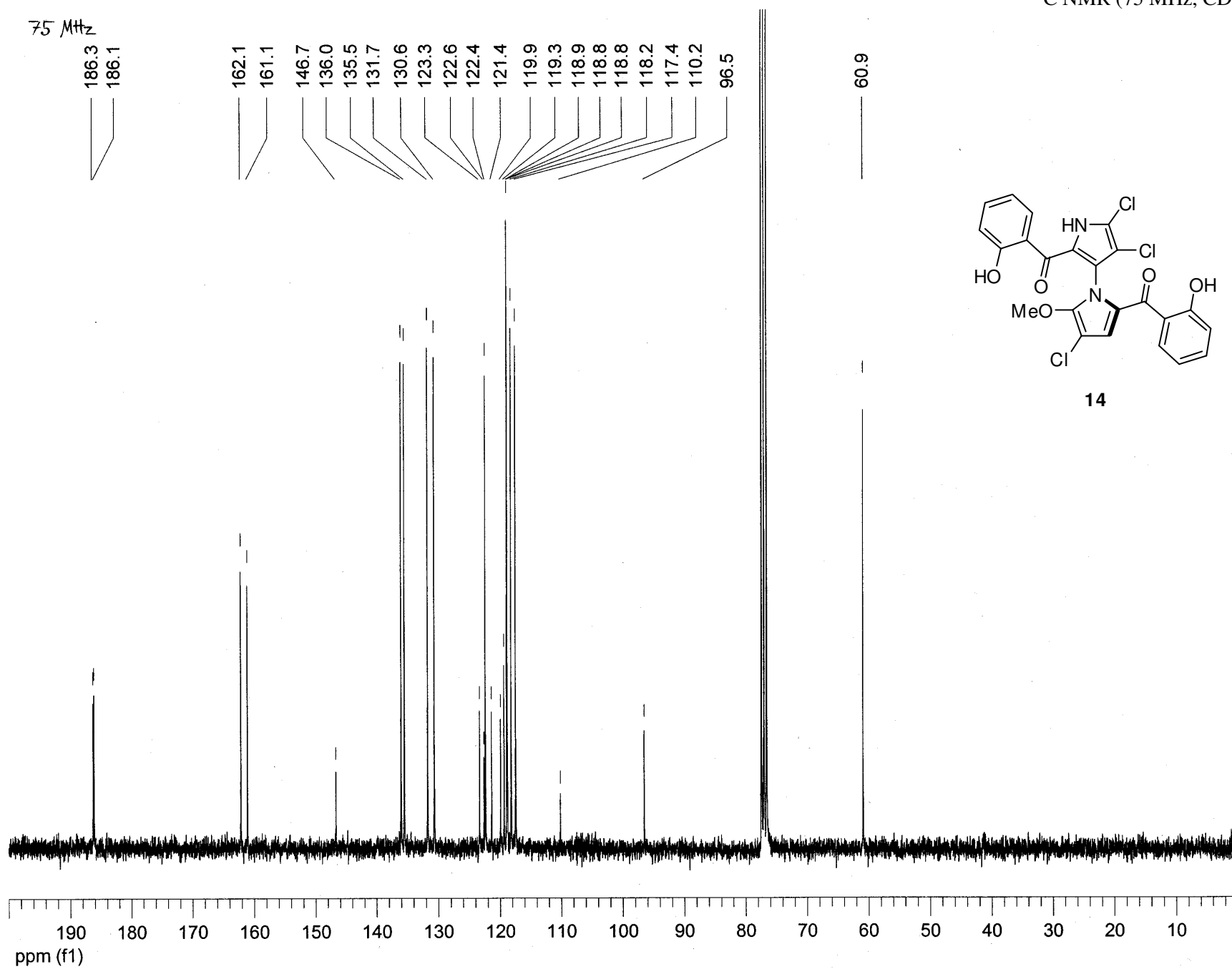
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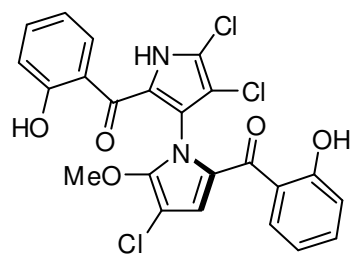
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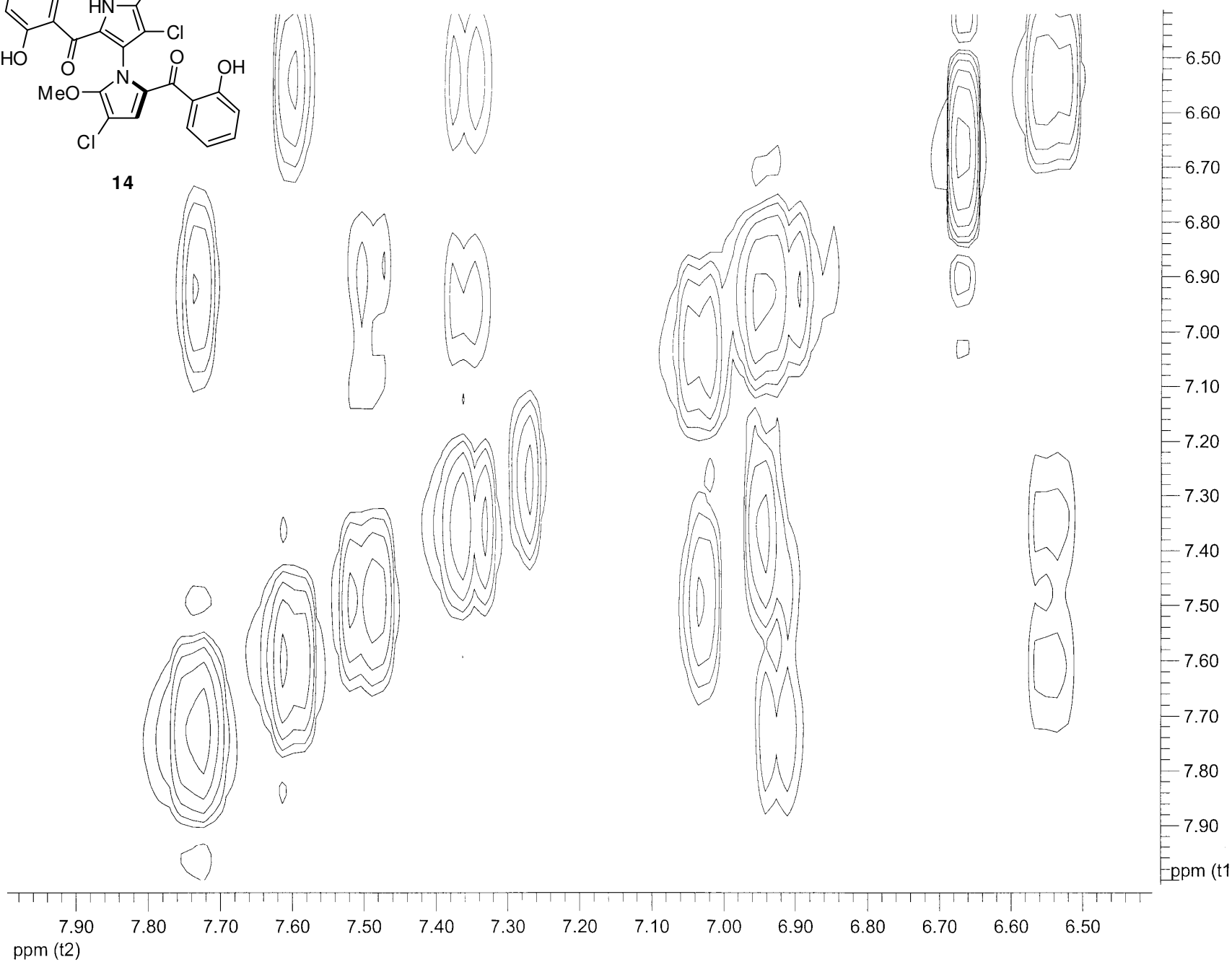


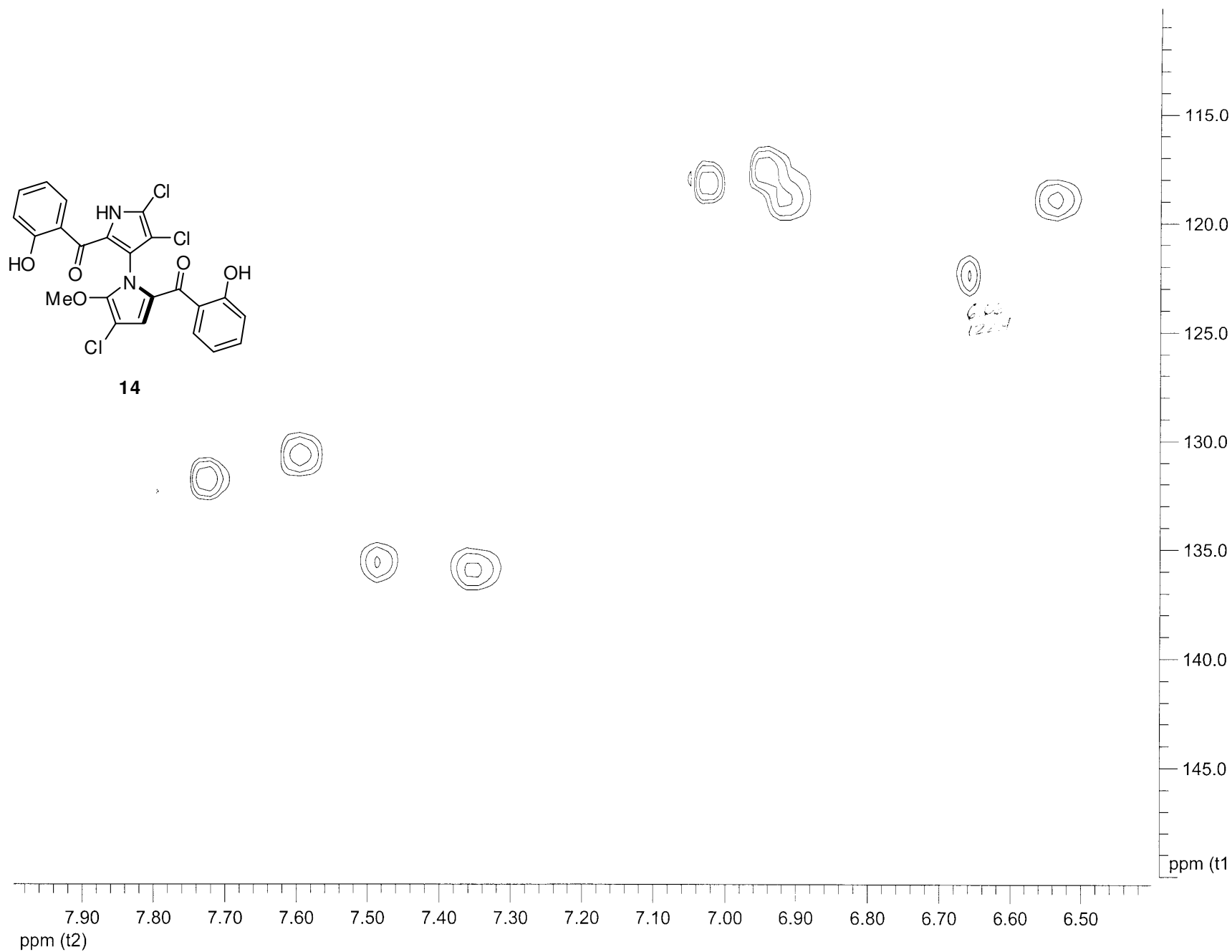


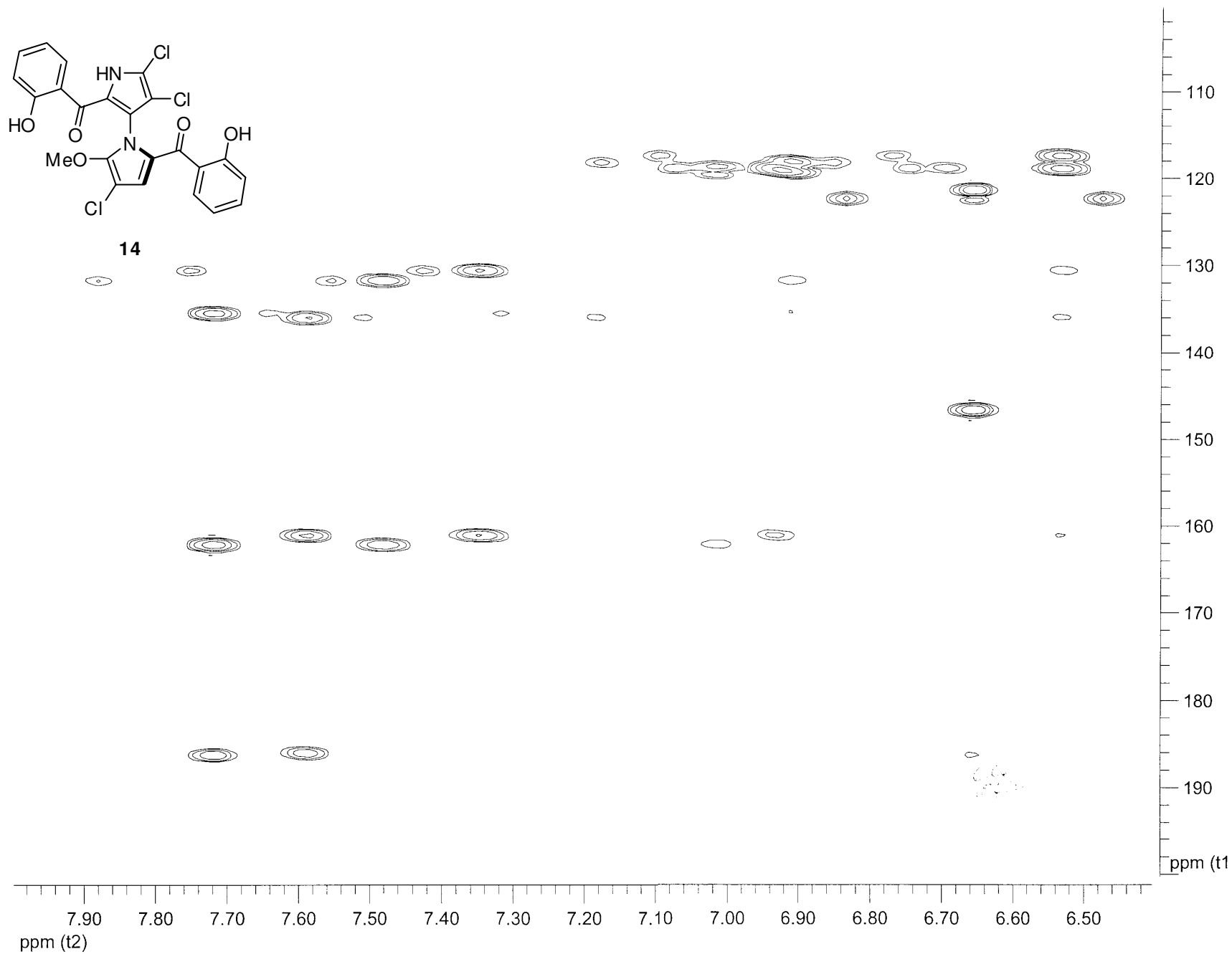


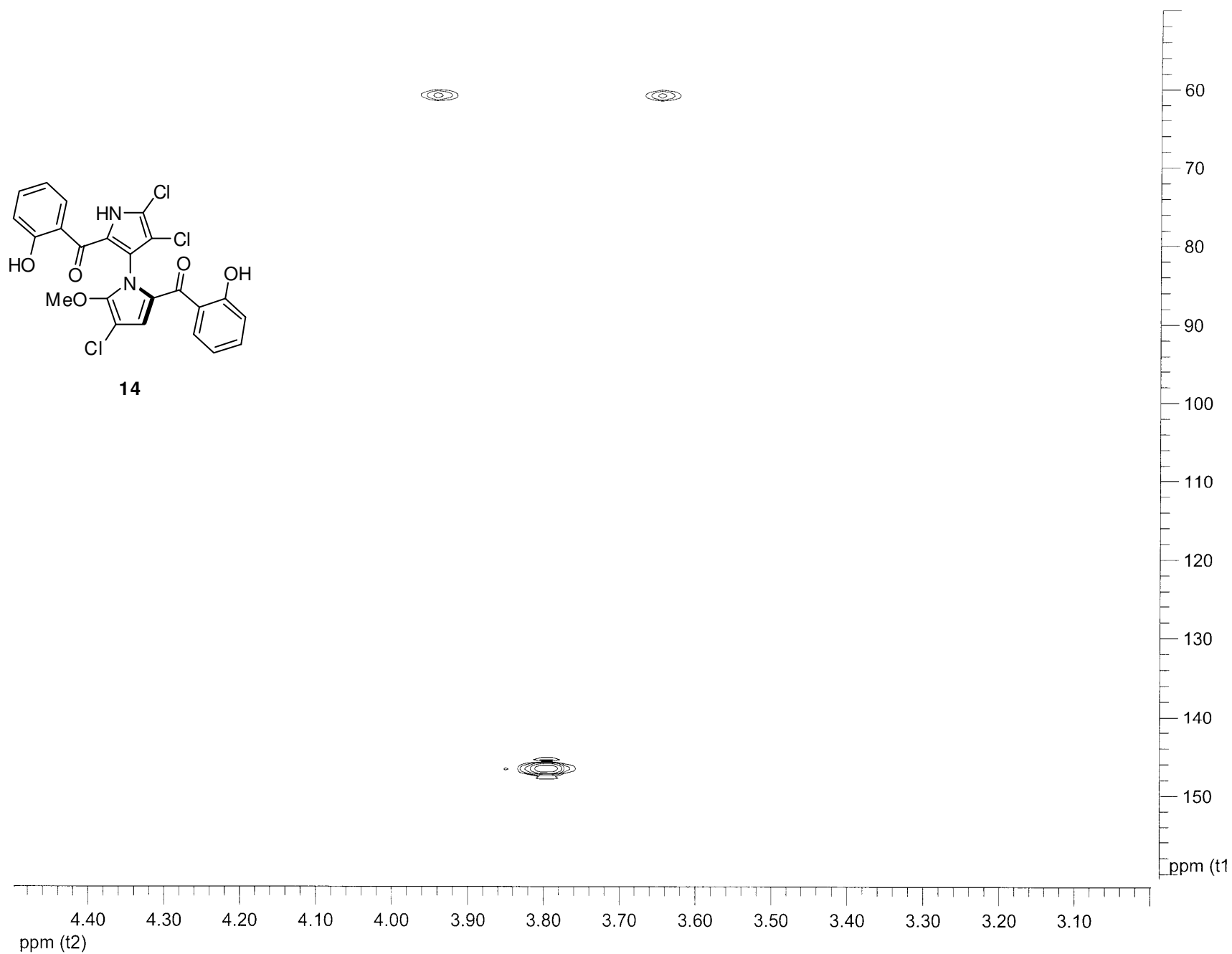
14

COSY (500 MHz, CDCl₃) of **14**







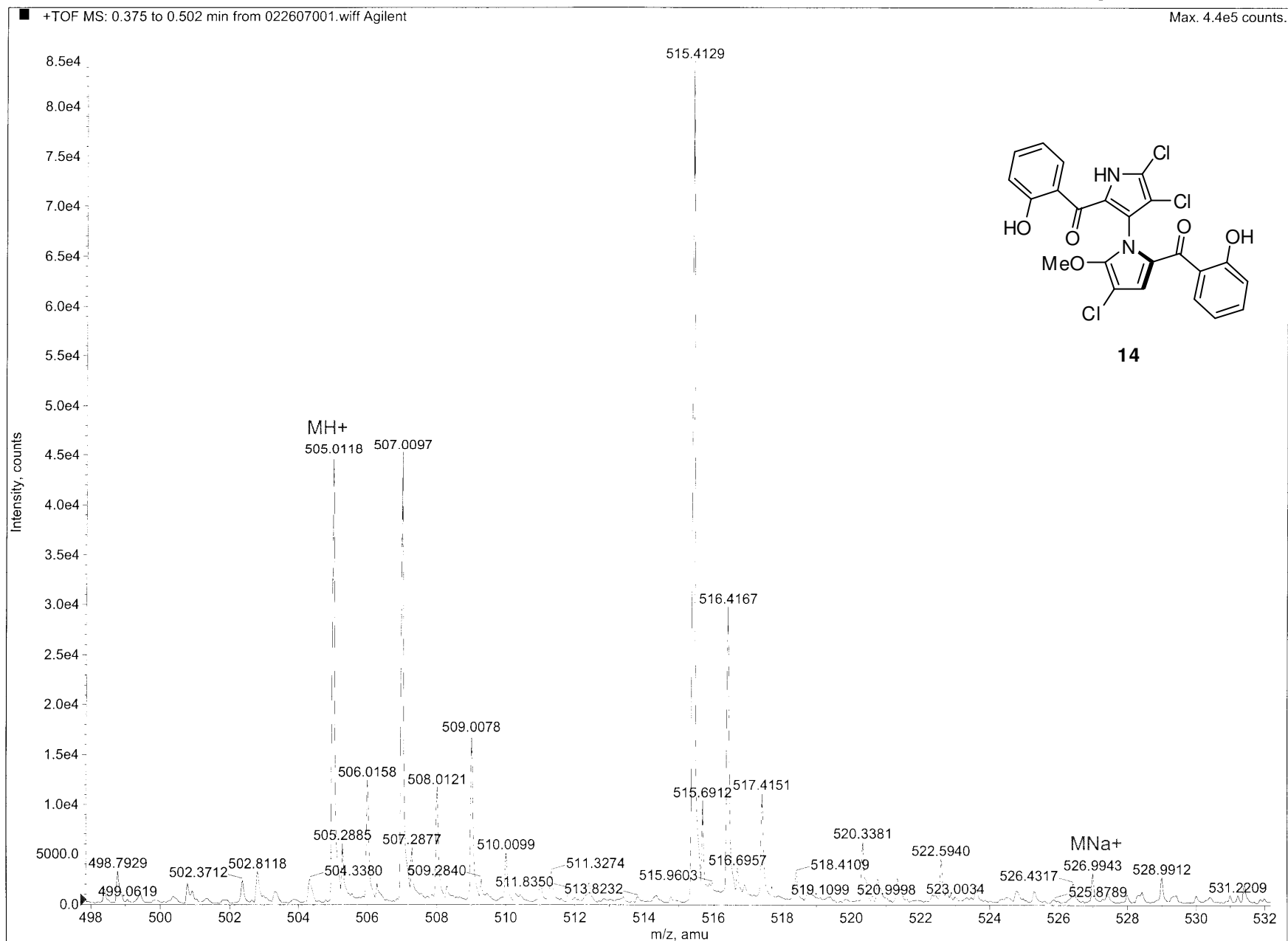


Polarity/Scan Type: Positive

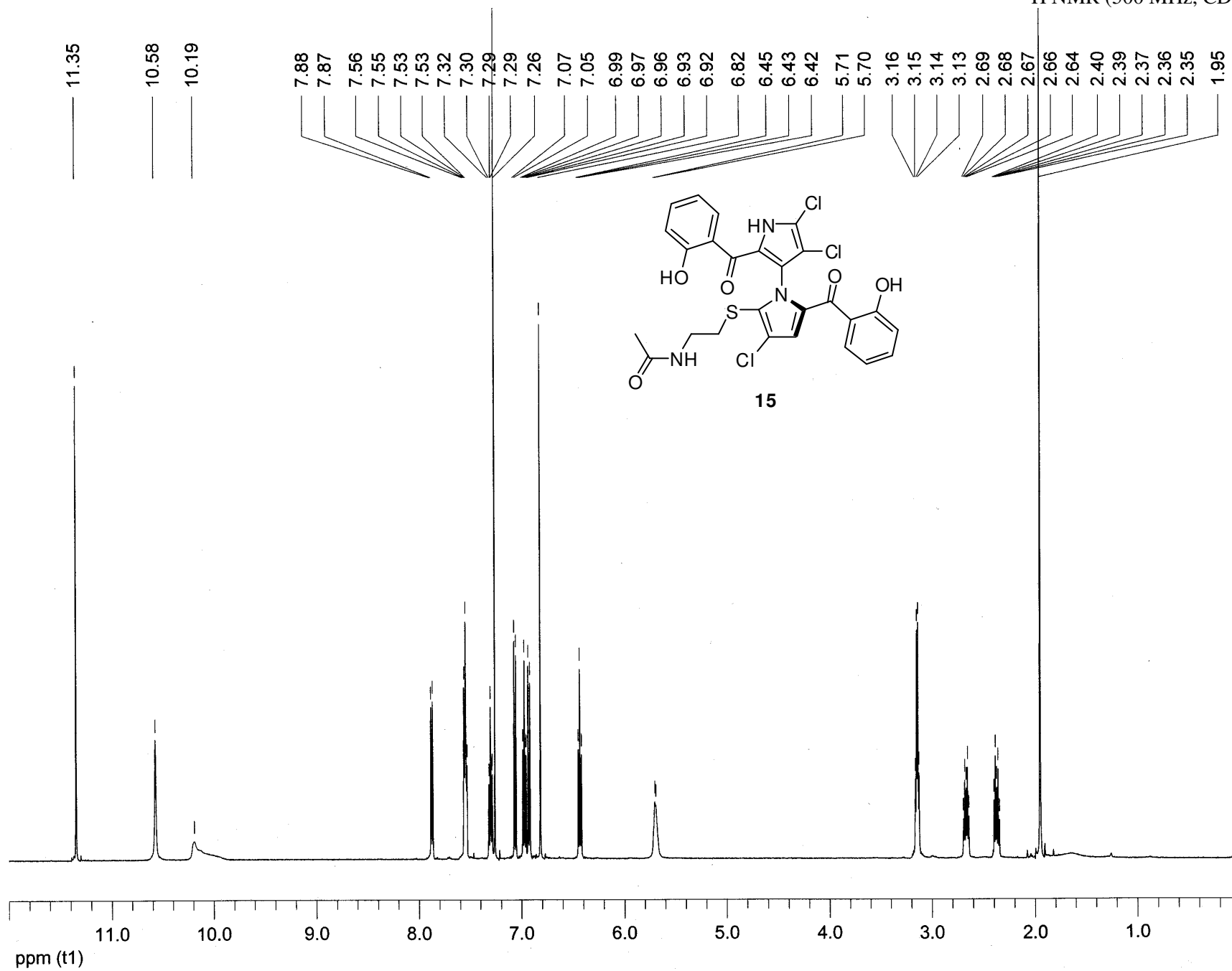
Sample Name: flch2488

Acq. Date: Monday, February 26, 2007

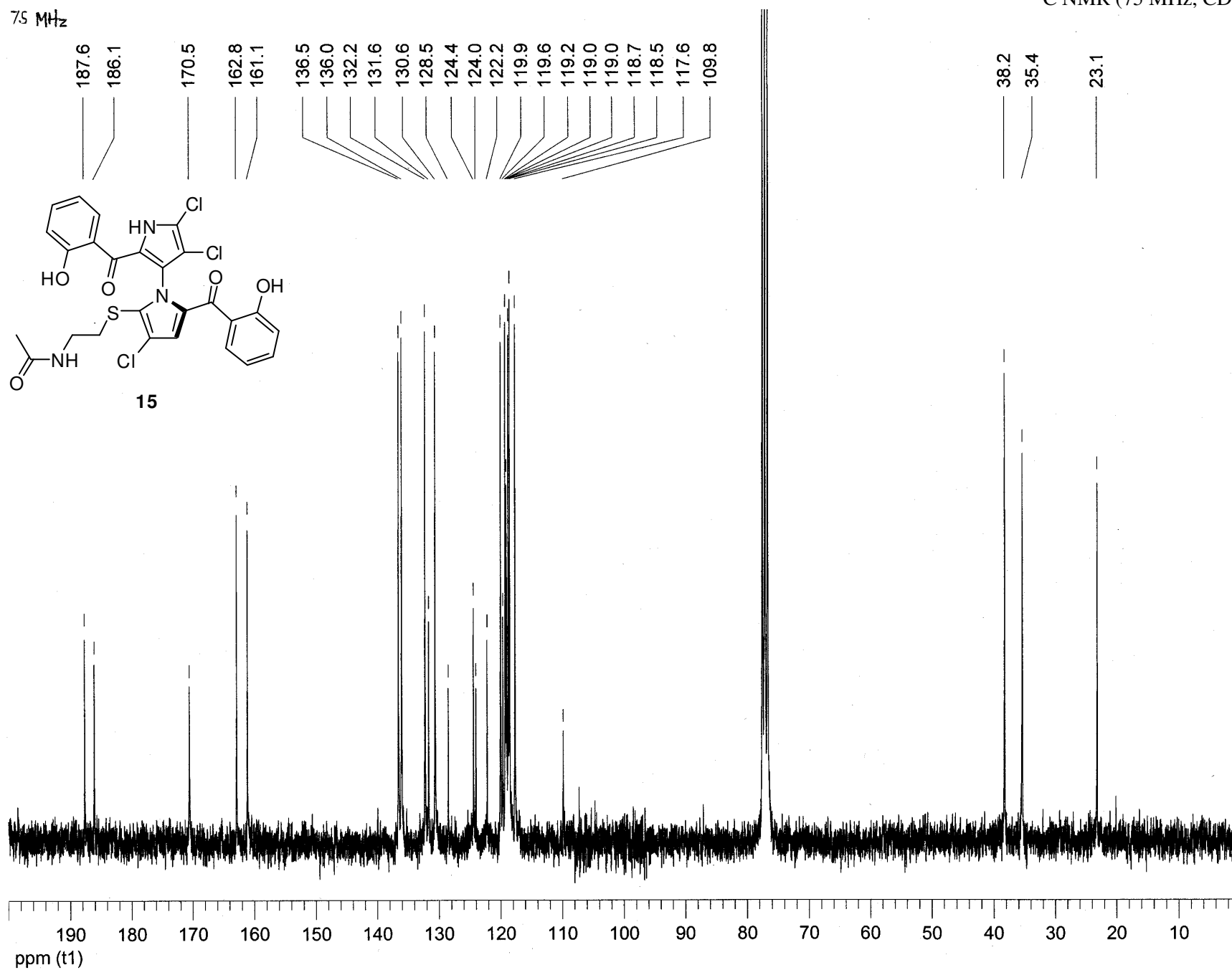
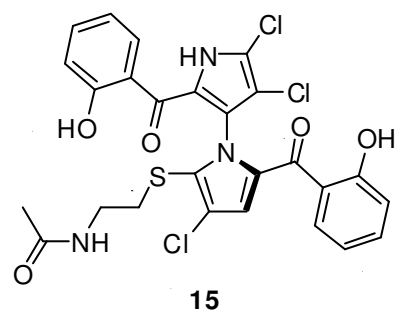
Acq. File: 022607001.wiff



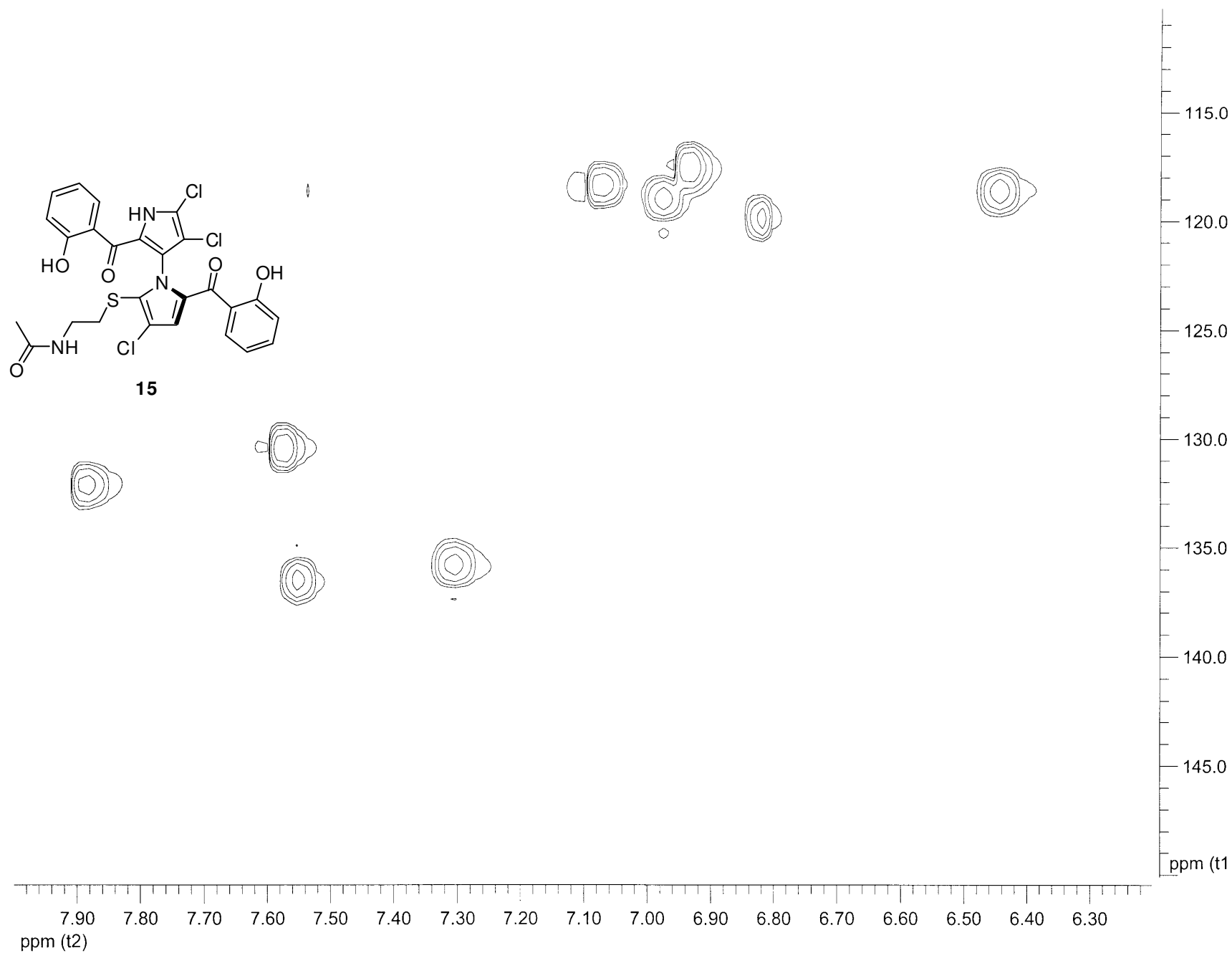
¹H NMR (500 MHz, CDCl₃) of **15**

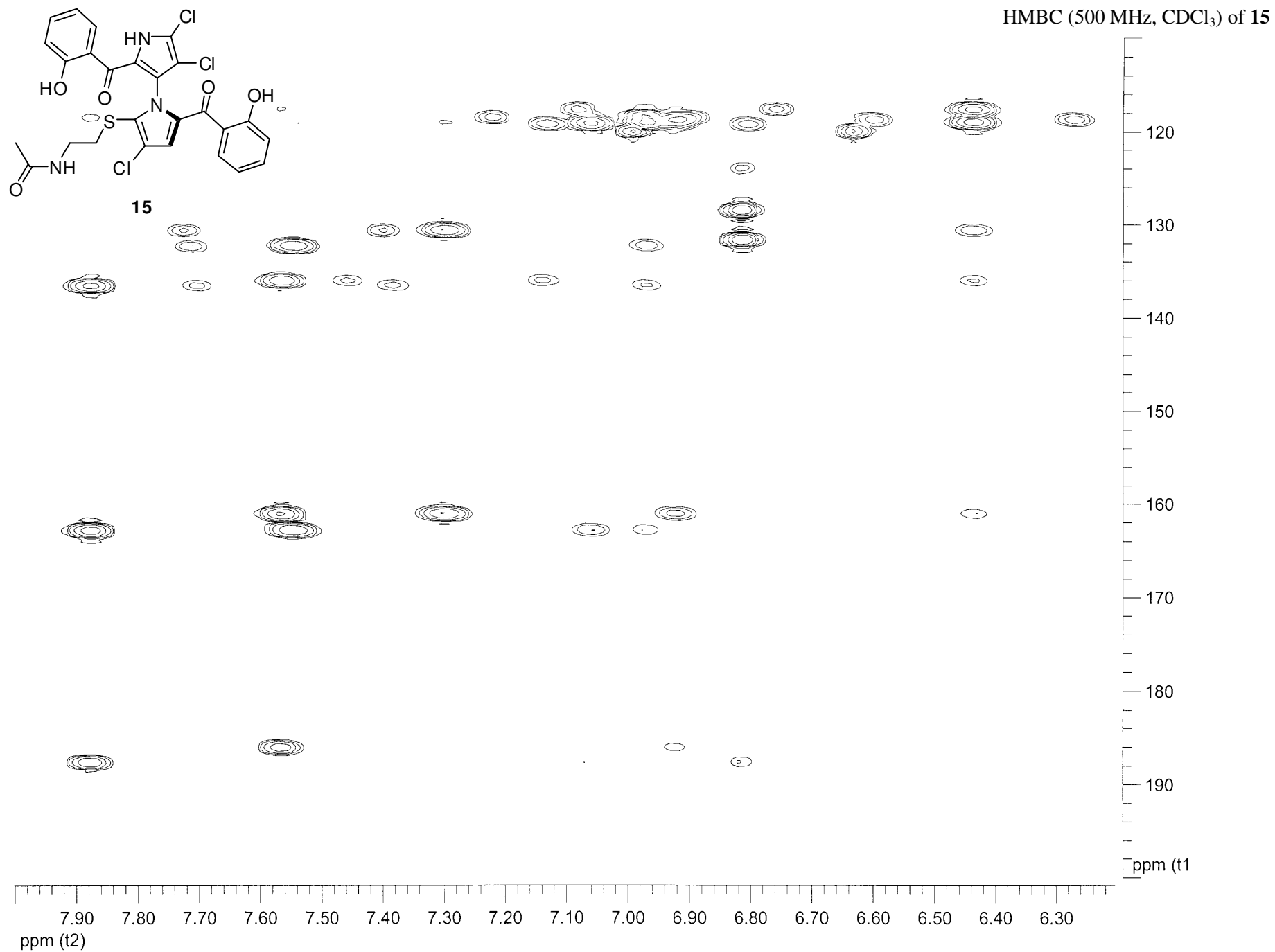


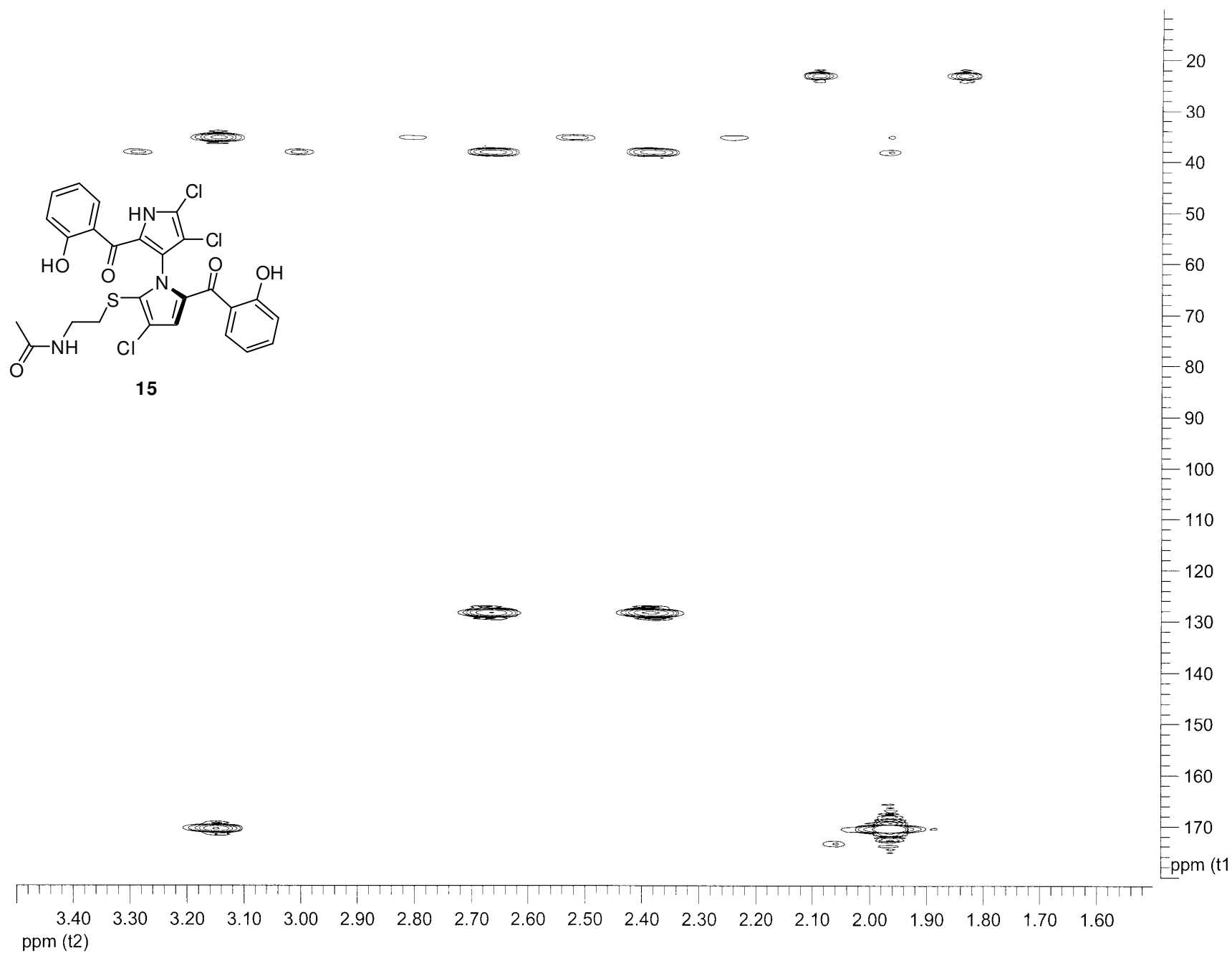
75 MHz









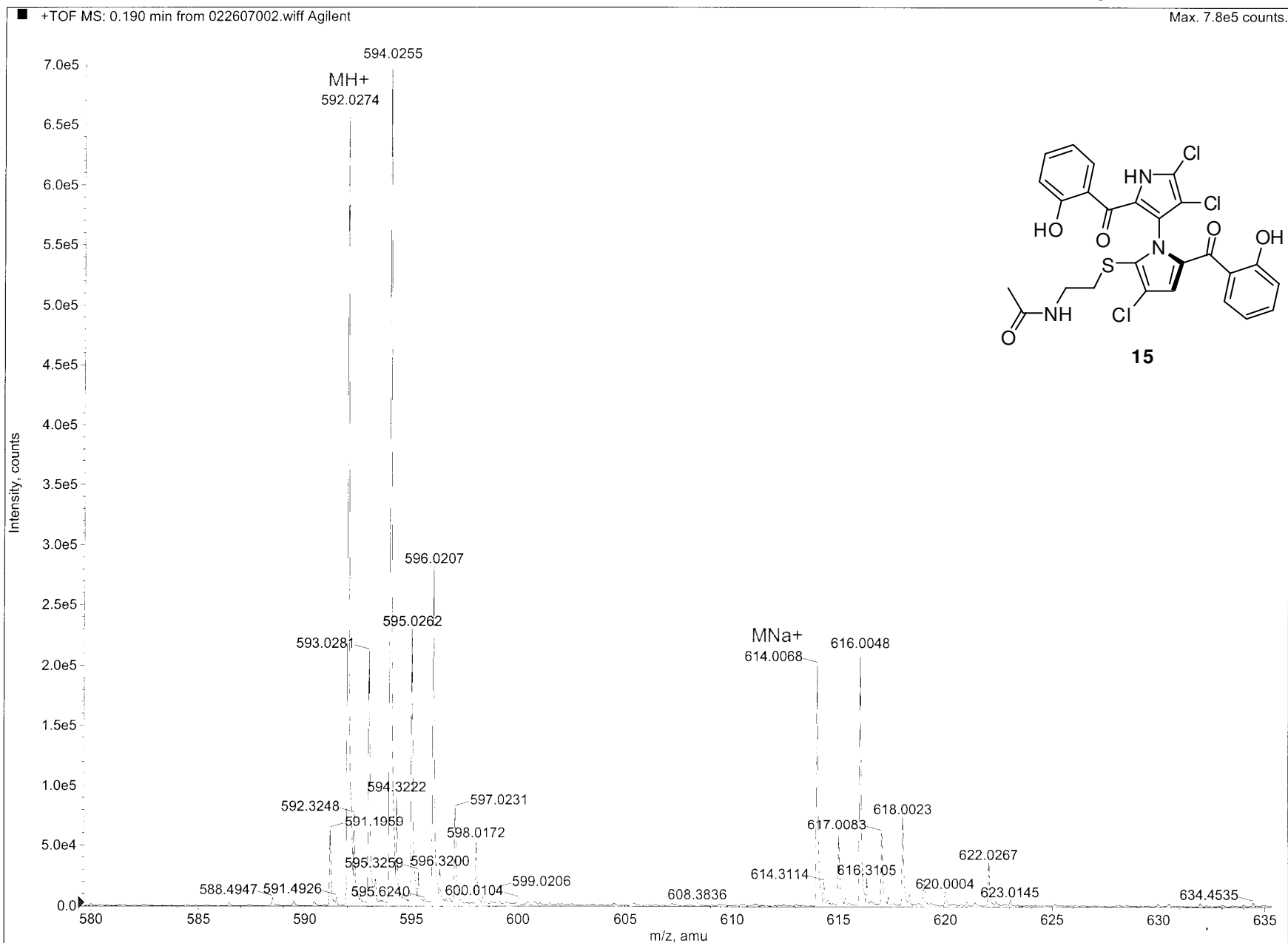


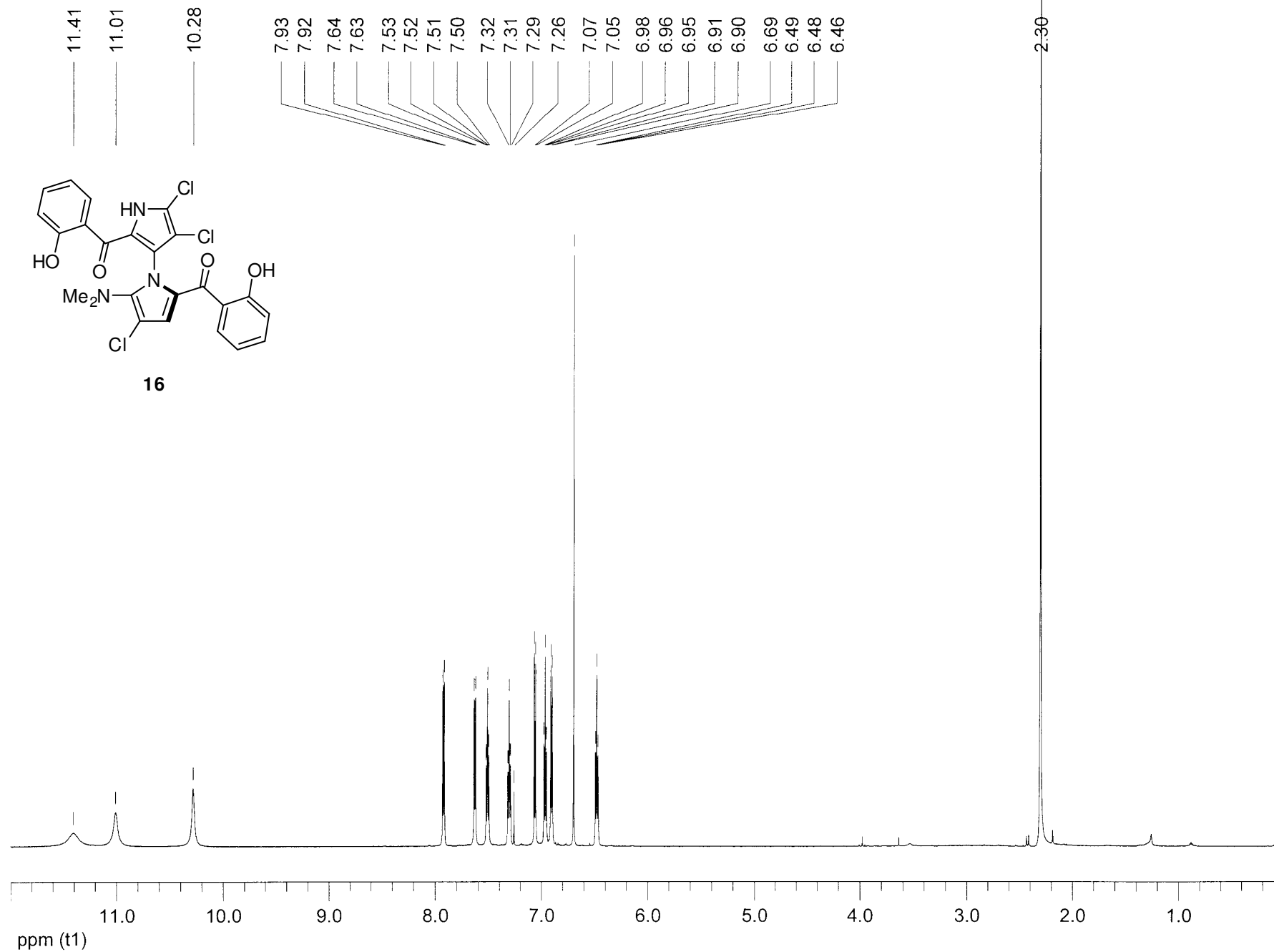
Polarity/Scan Type: Positive

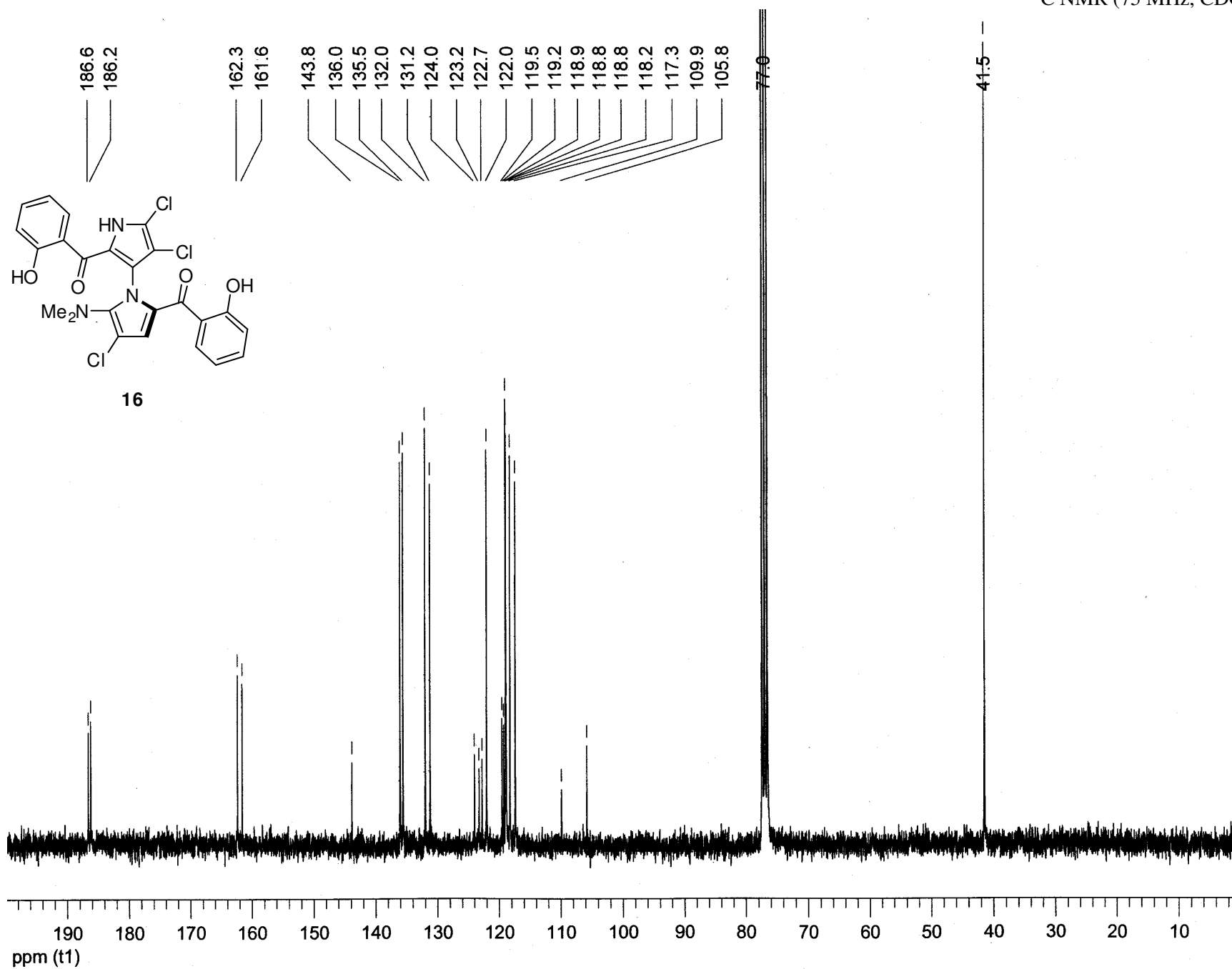
Sample Name: flch2489

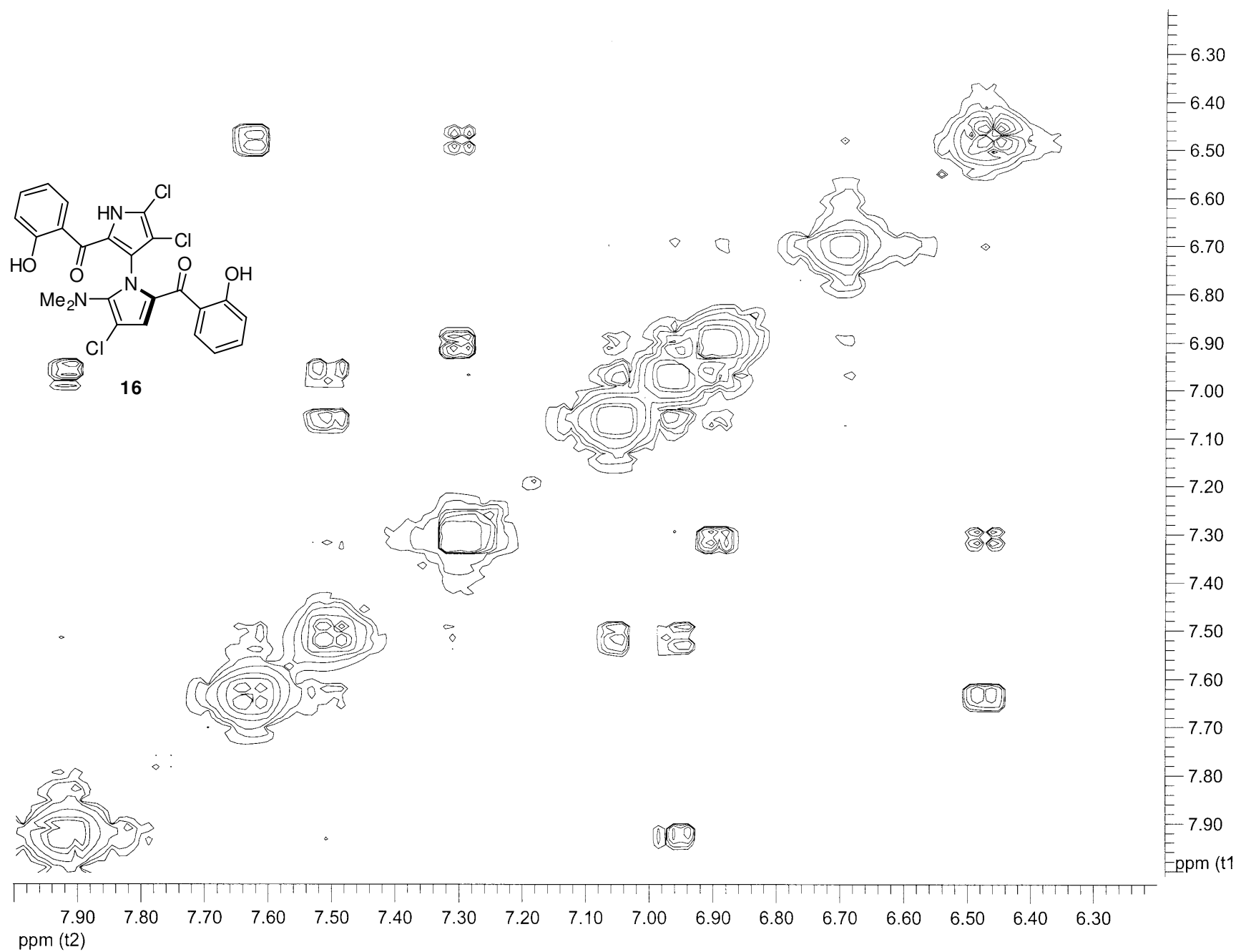
Acq. Date: Monday, February 26, 2007

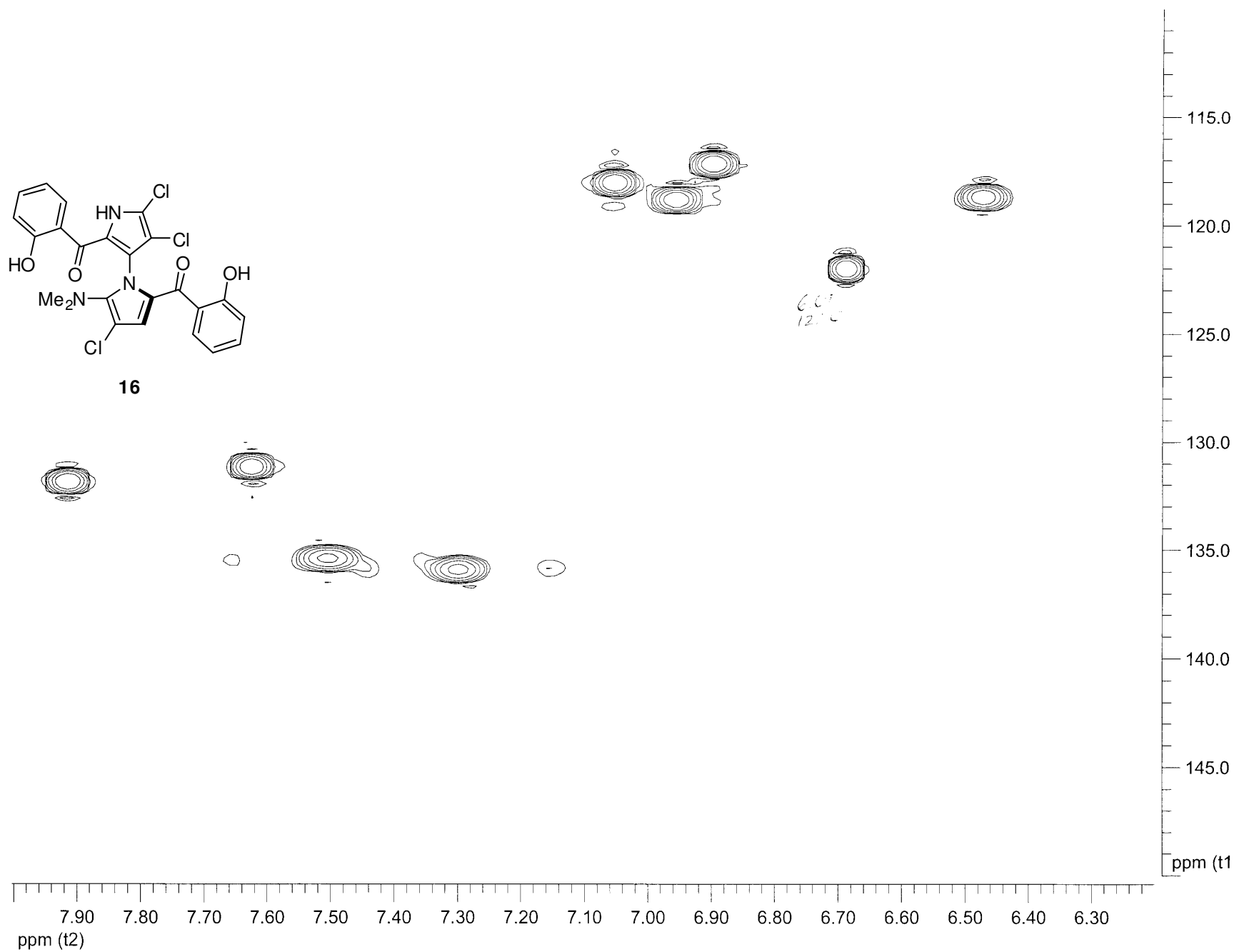
Acq. File: 022607002.wiff

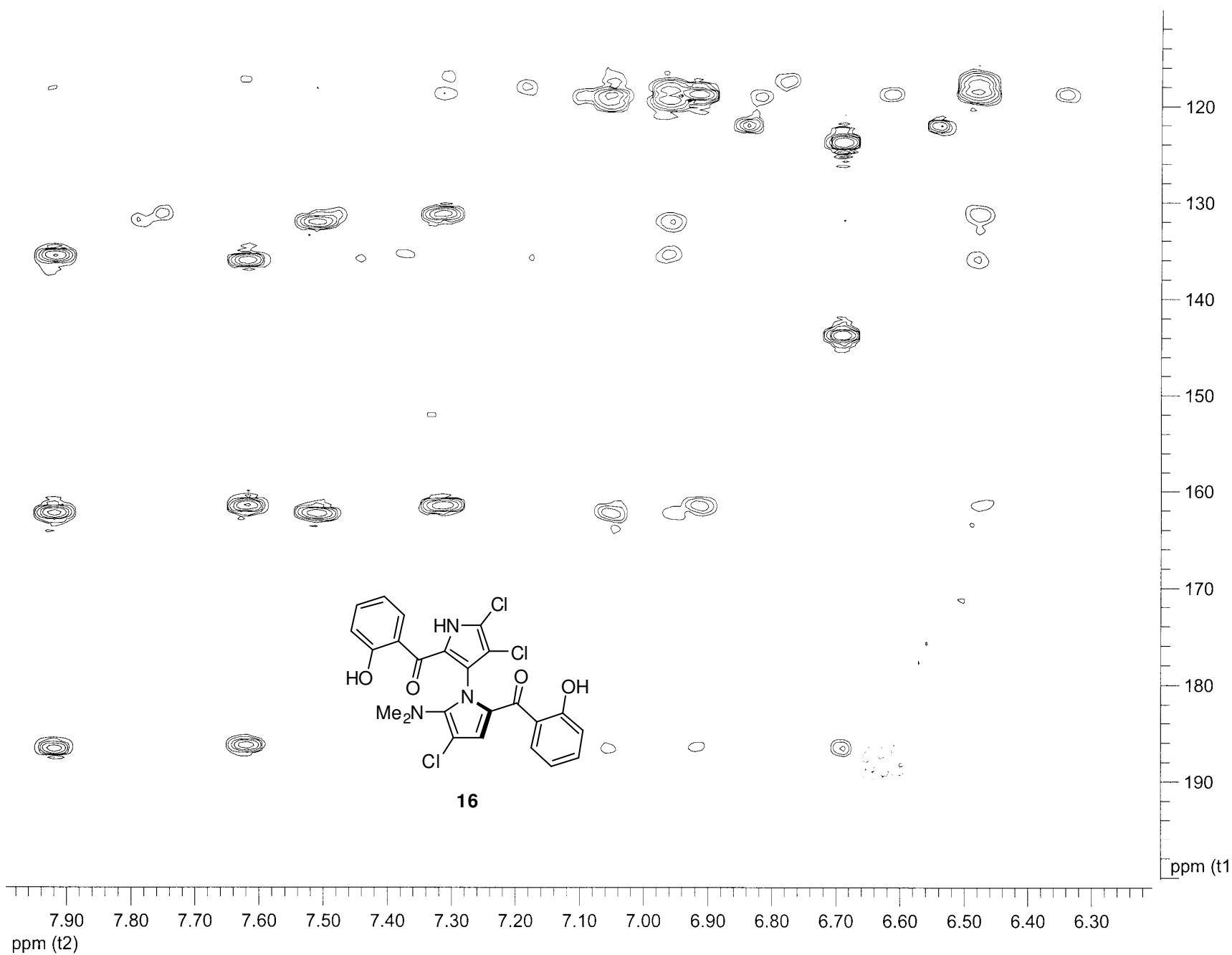


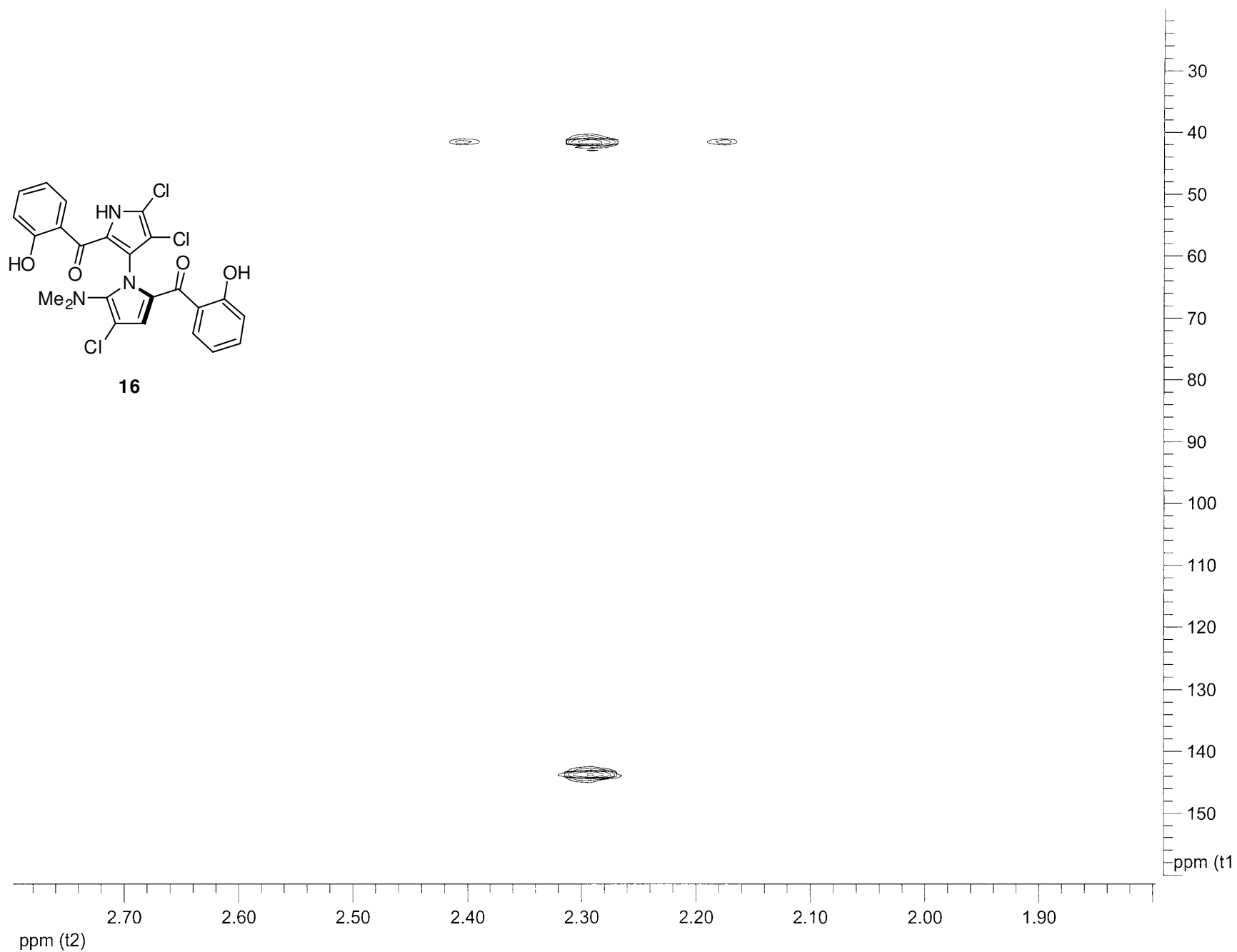












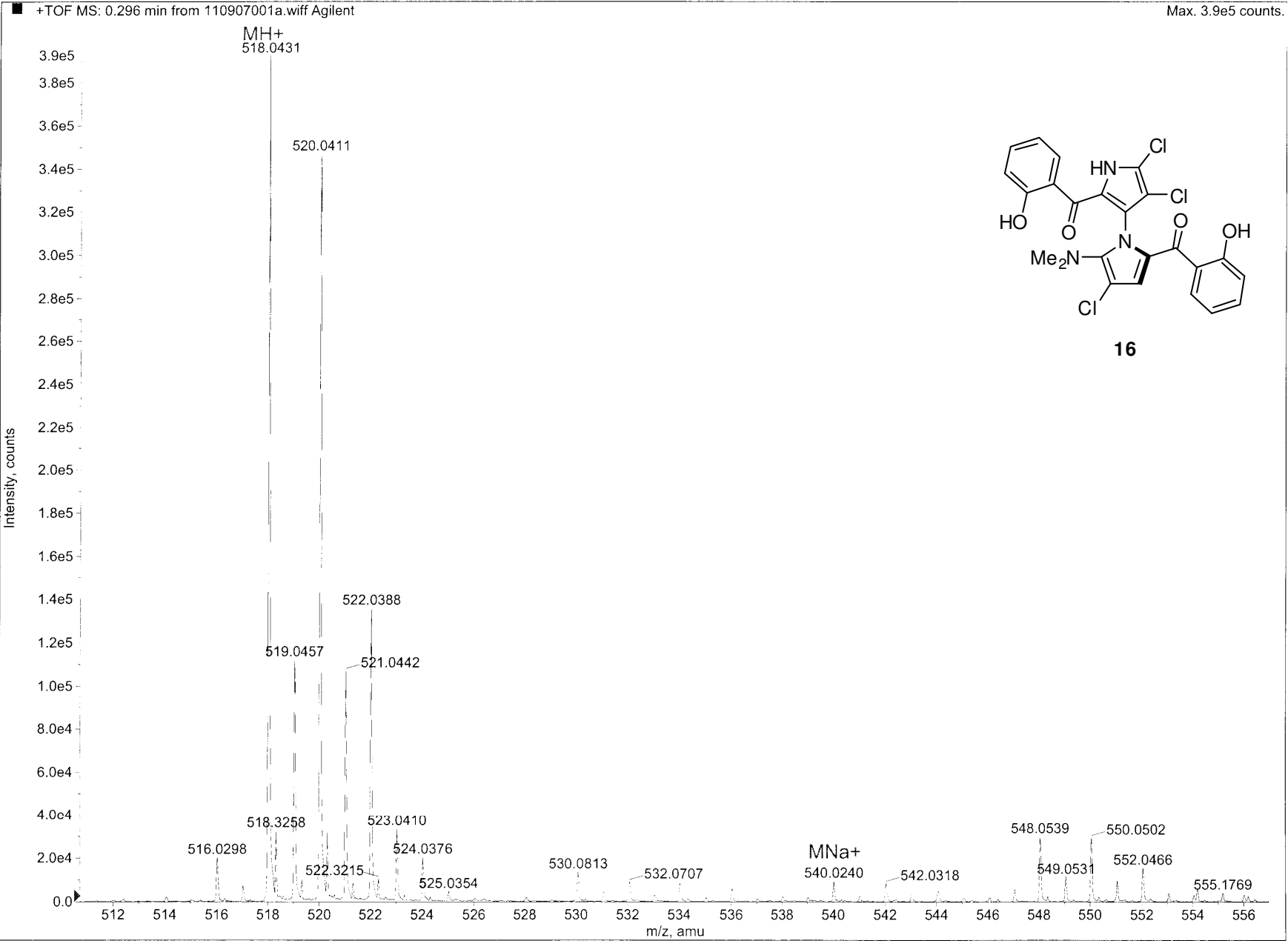
Polarity/Scan Type: Positive

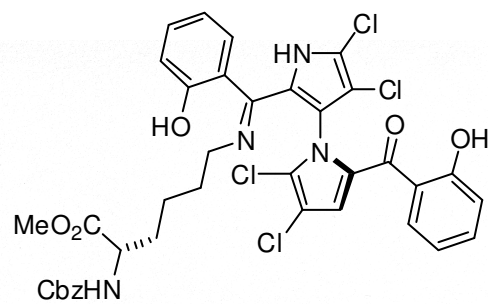
Sample Name: flch8669

Acq. Date: Friday, November 09, 2007

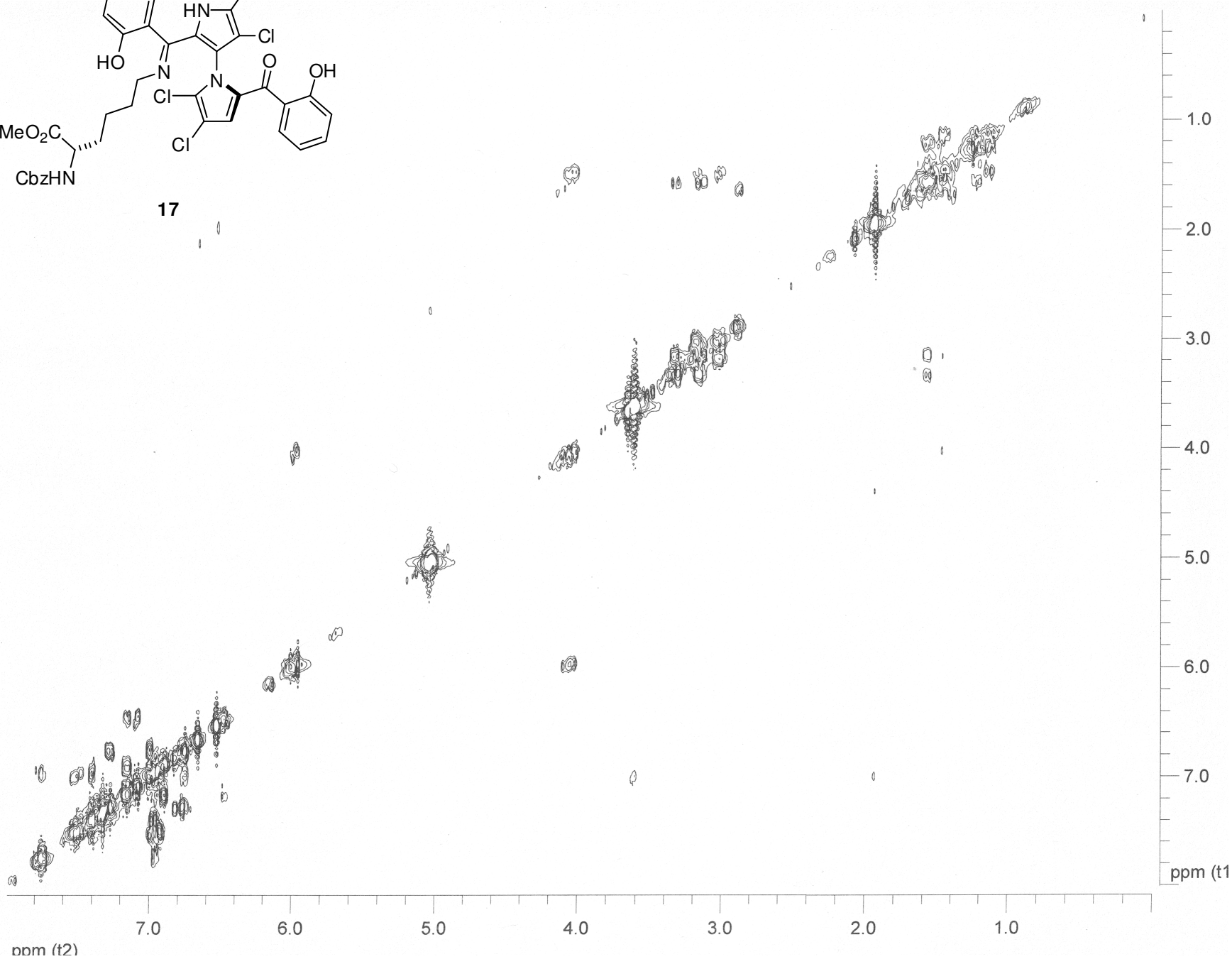
Acq. File: 110907001a.wiff

Max. 3.9e5 counts.





17



[illegible]

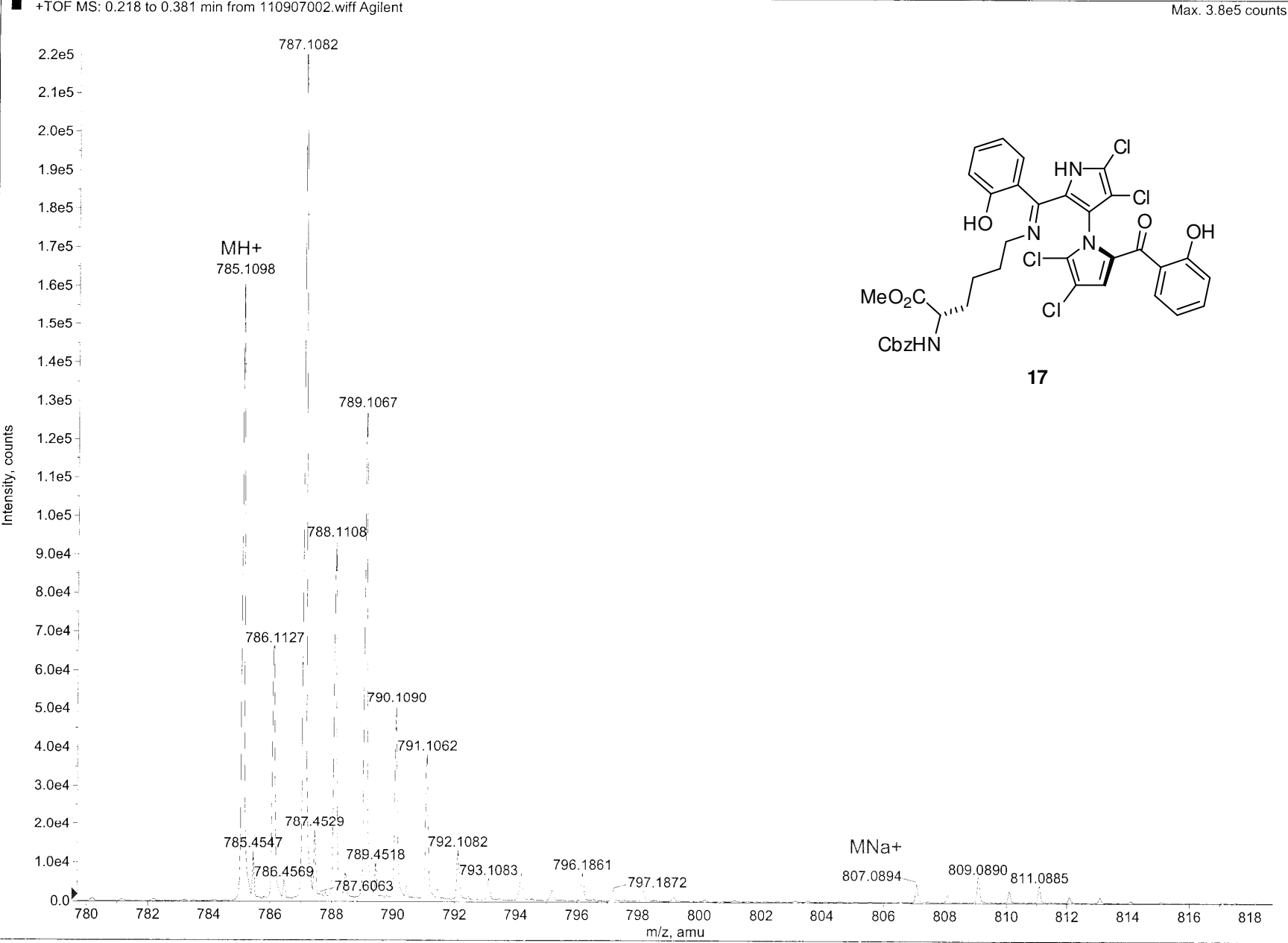
Polarity/Scan Type: Positive

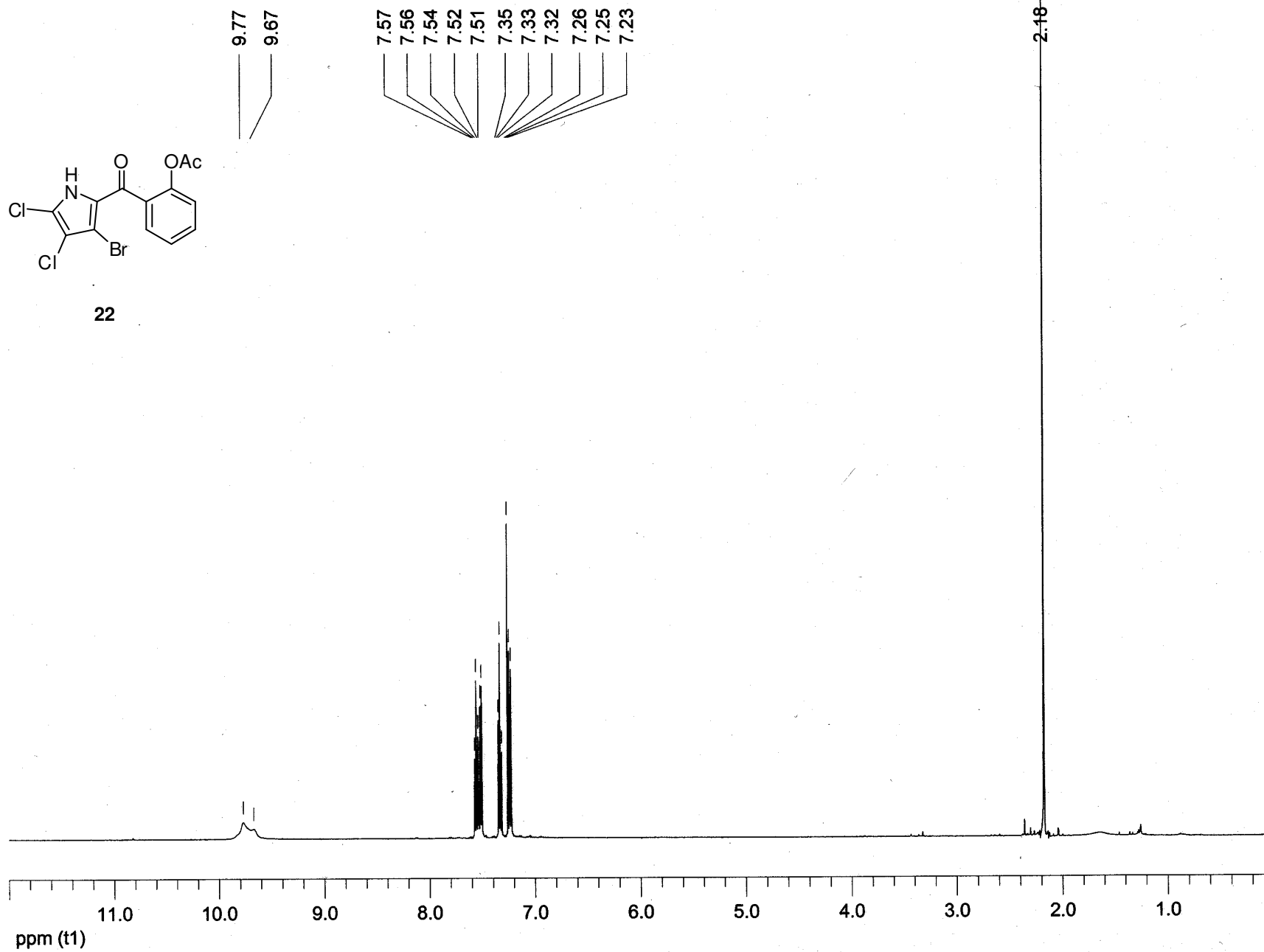
Sample Name: flch8670

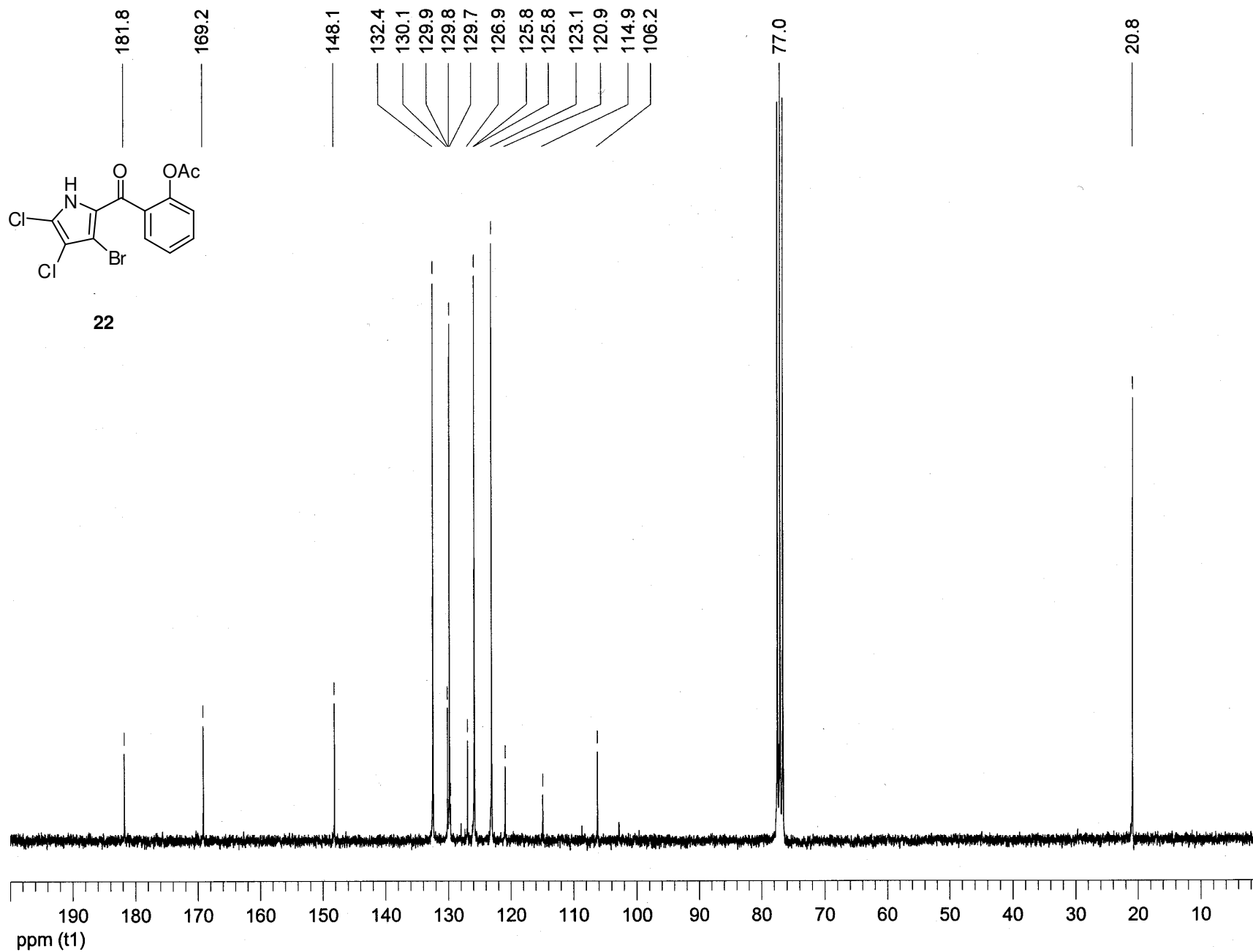
Acq. Date: Friday, November 09, 2007

Acq. File: 110907002.wiff

Max. 3.8e5 counts.





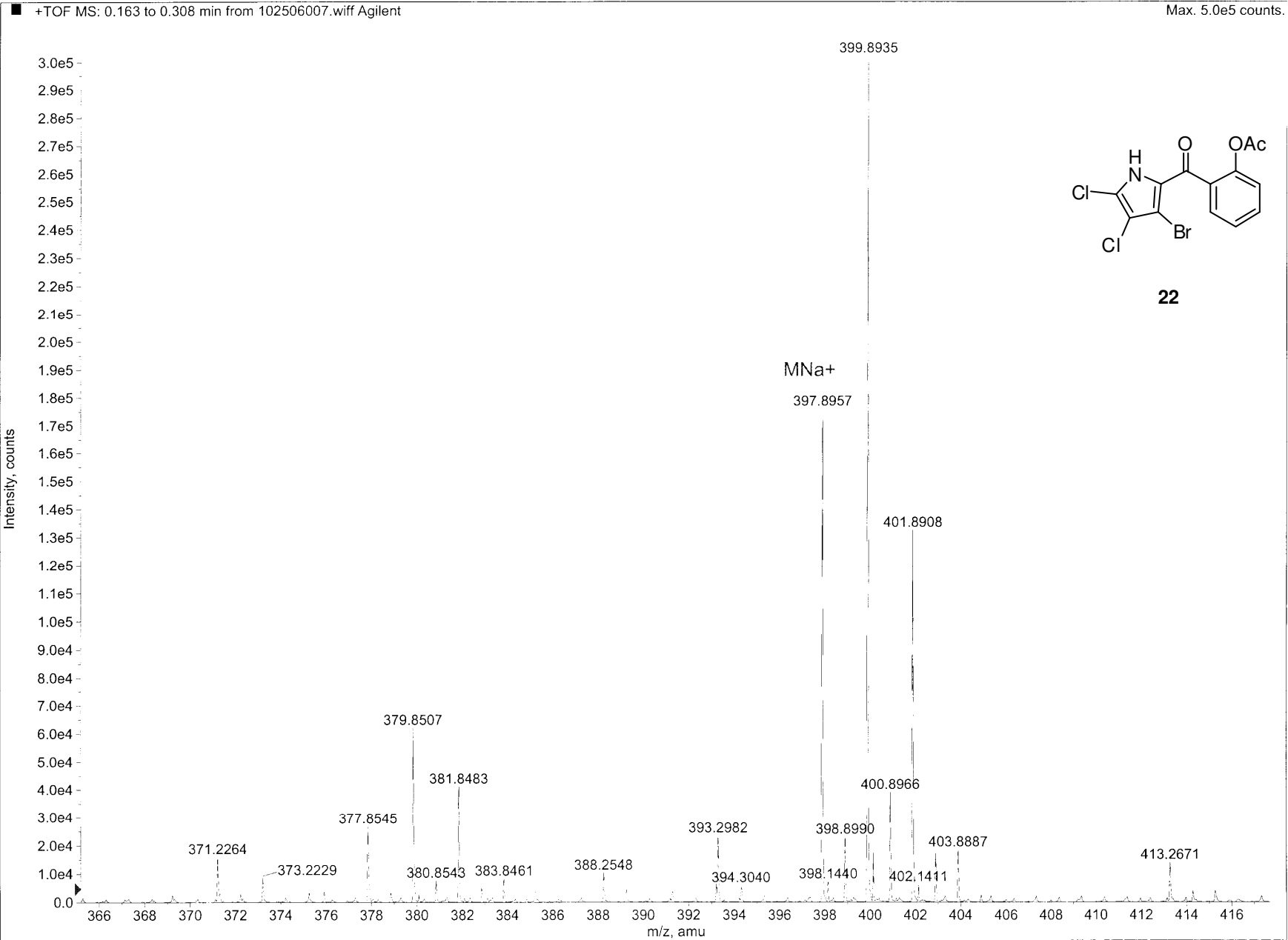


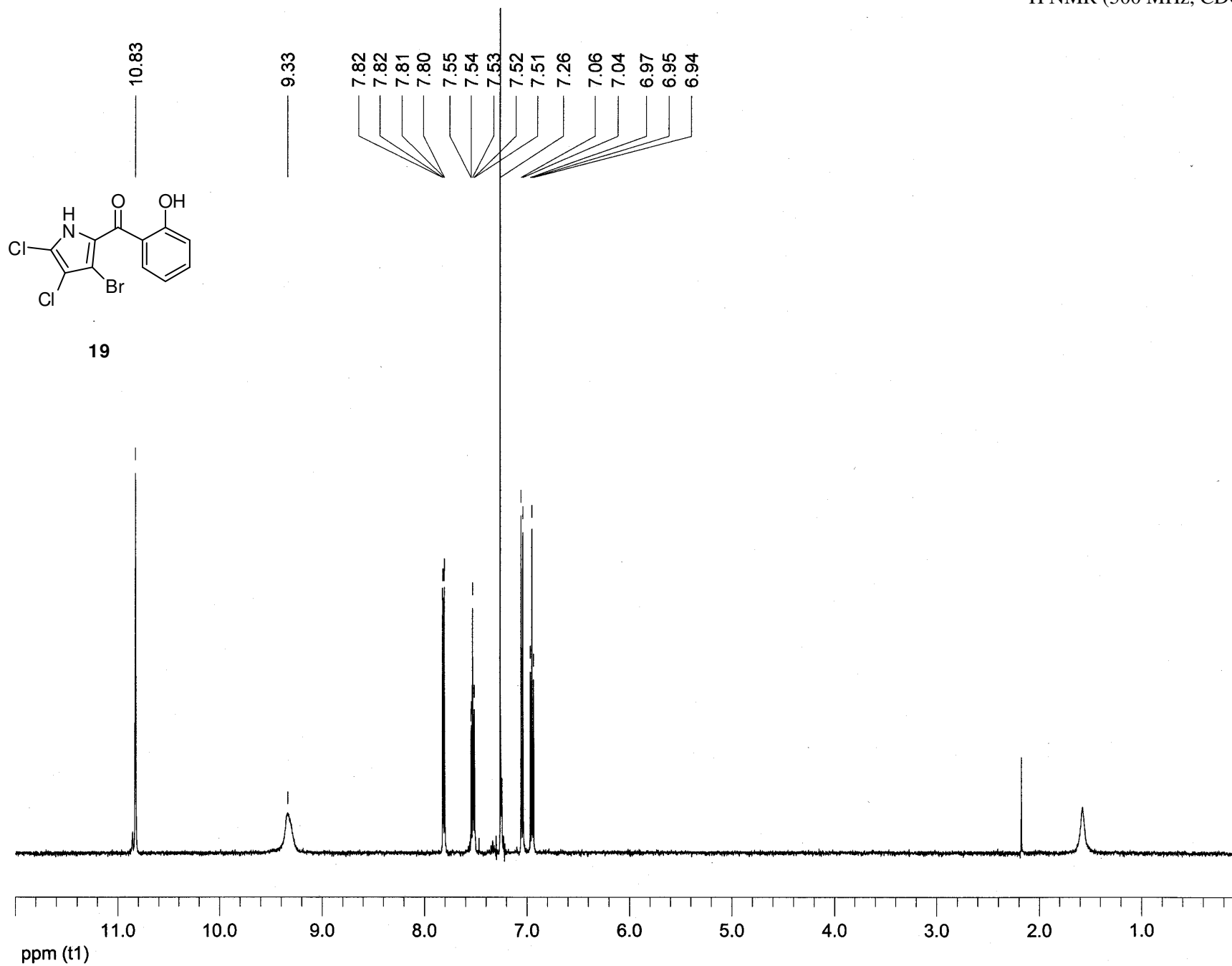
Polarity/Scan Type: Positive

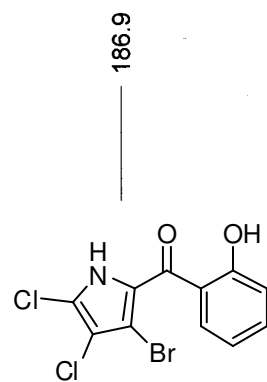
Sample Name: flch8550

Acq. Date: Wednesday, October 25, 2006

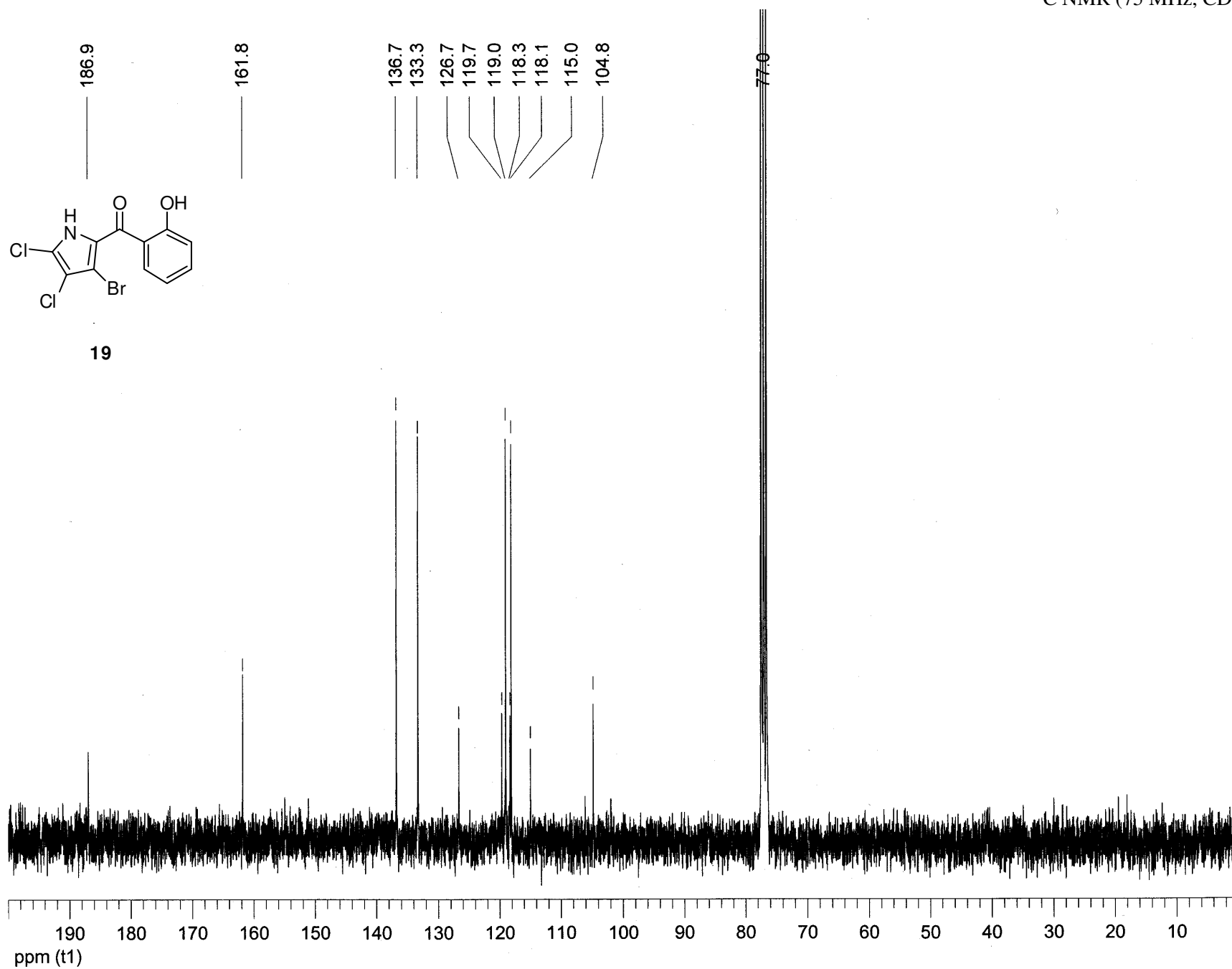
Acq. File: 102506007.wiff







19



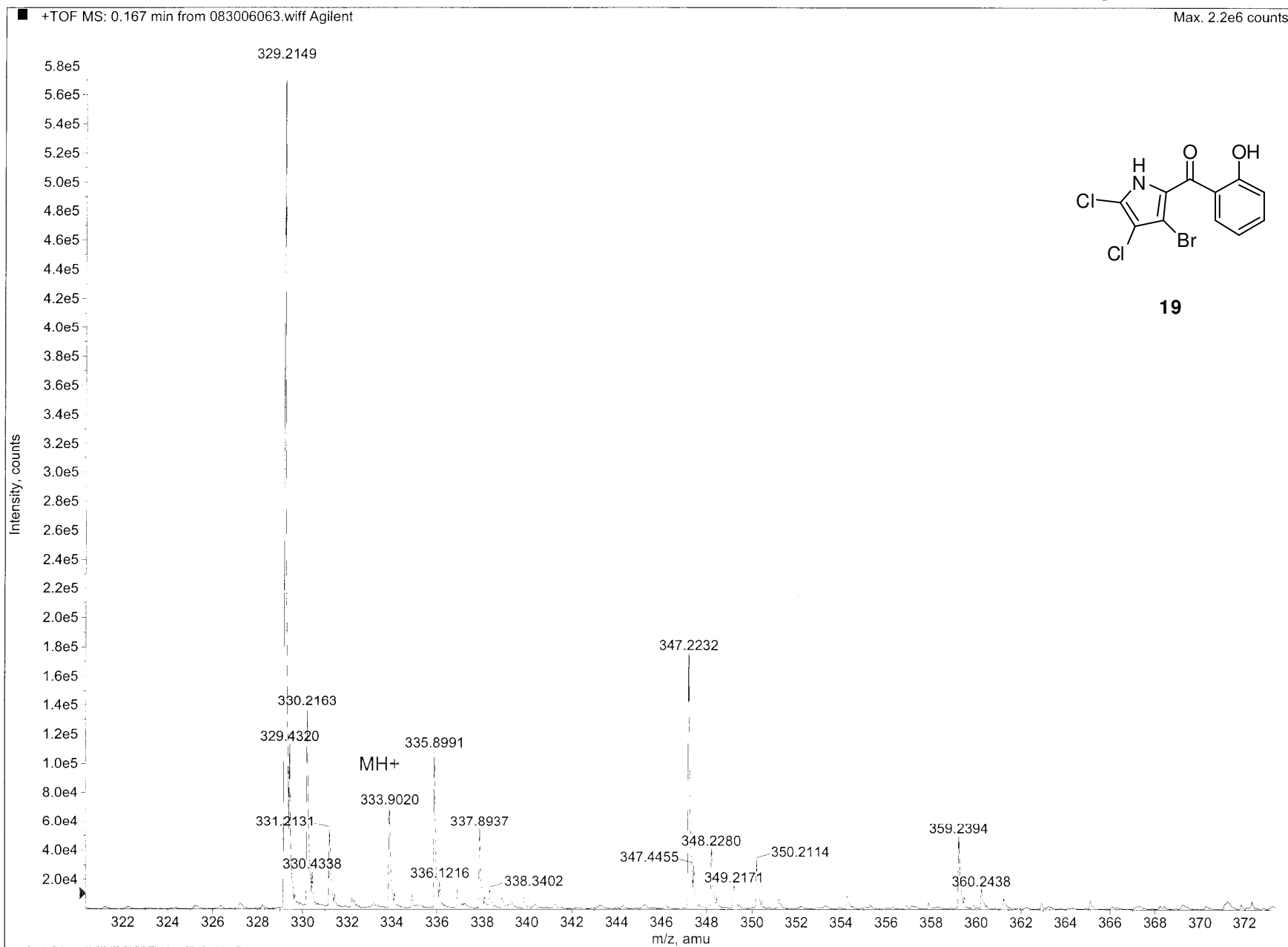
Polarity/Scan Type: Positive

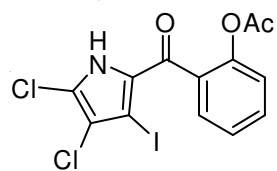
Sample Name: flch6844

Acq. Date: Wednesday, August 30, 2006

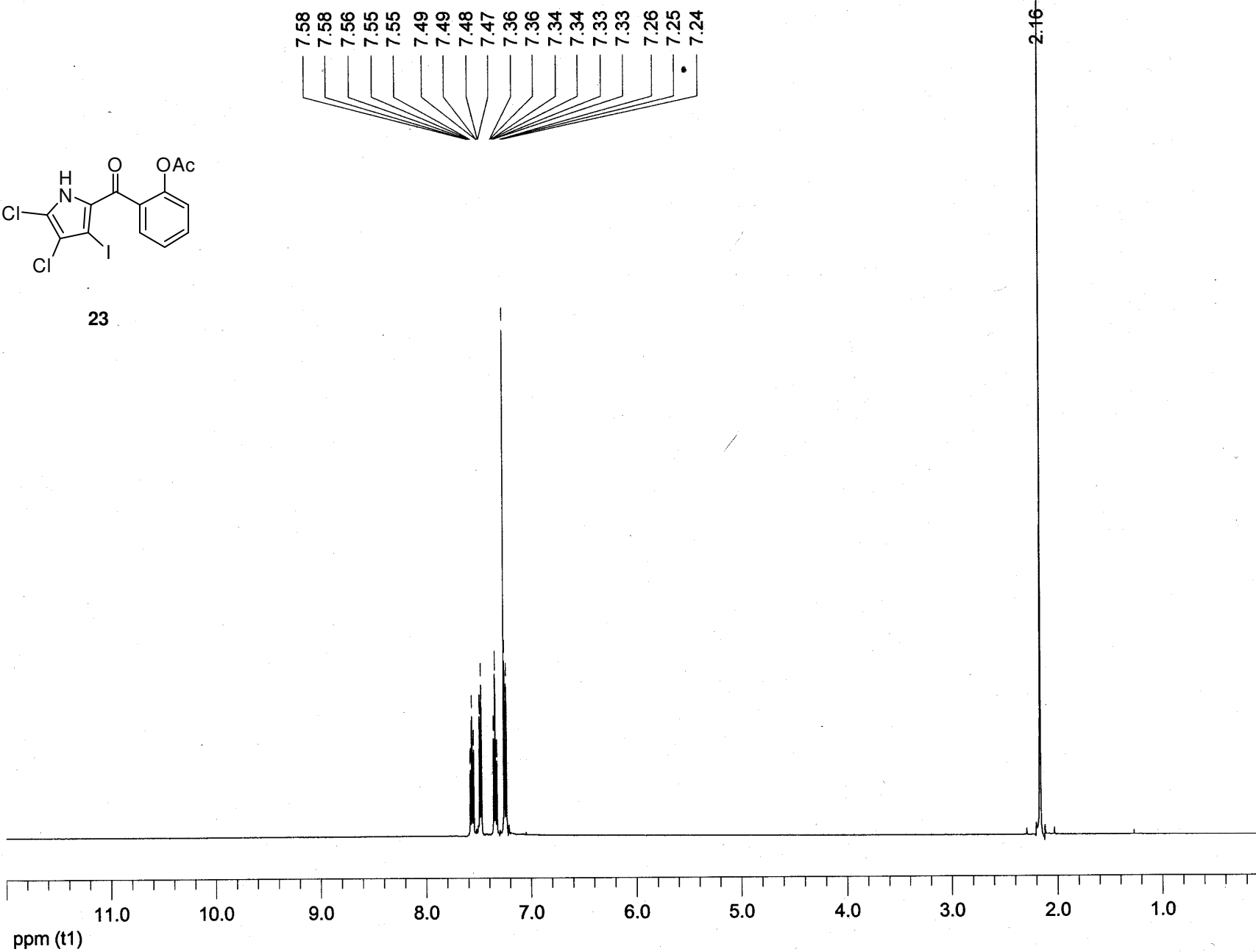
Acq. File: 083006063.wiff

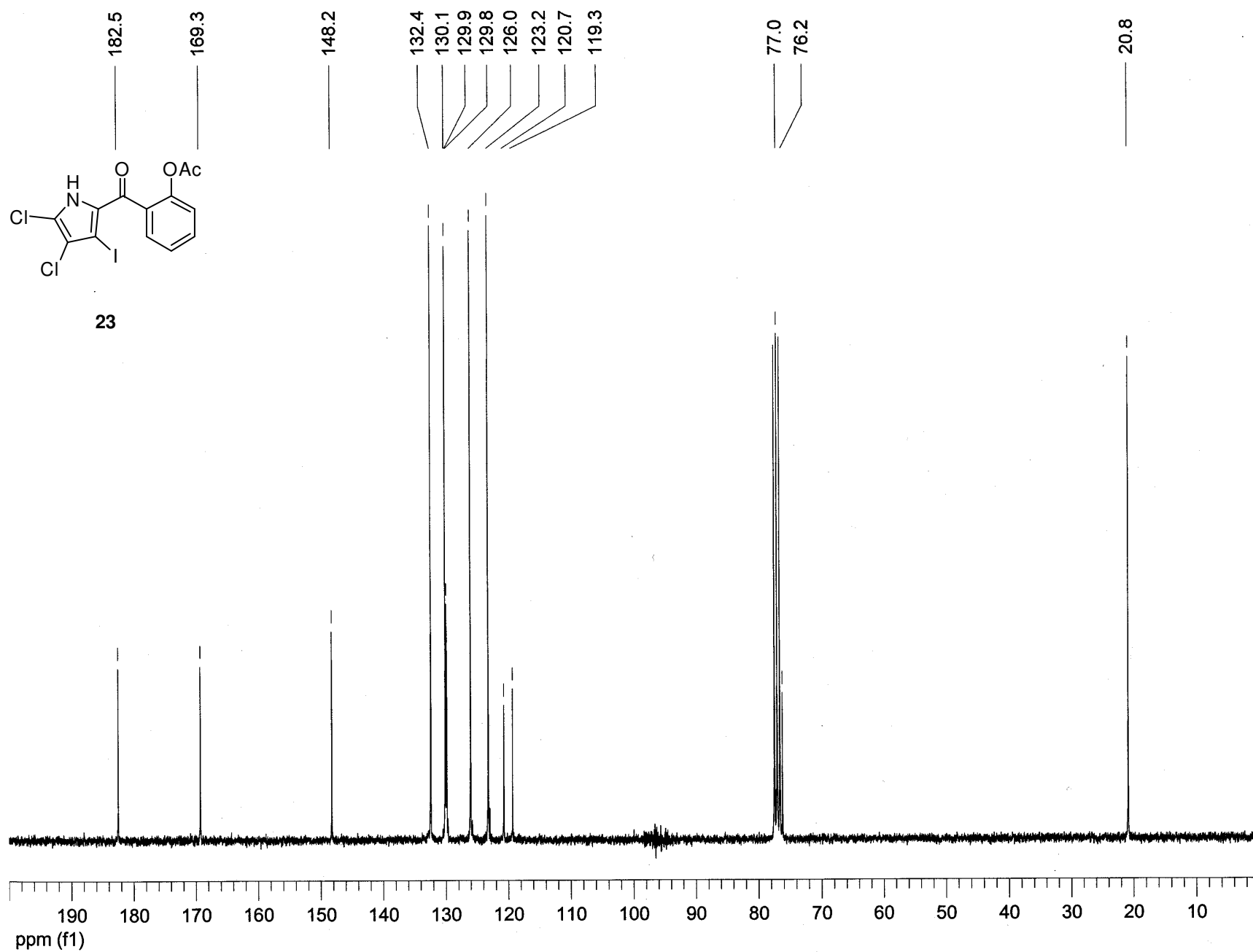
Max. 2.2e6 counts.





23



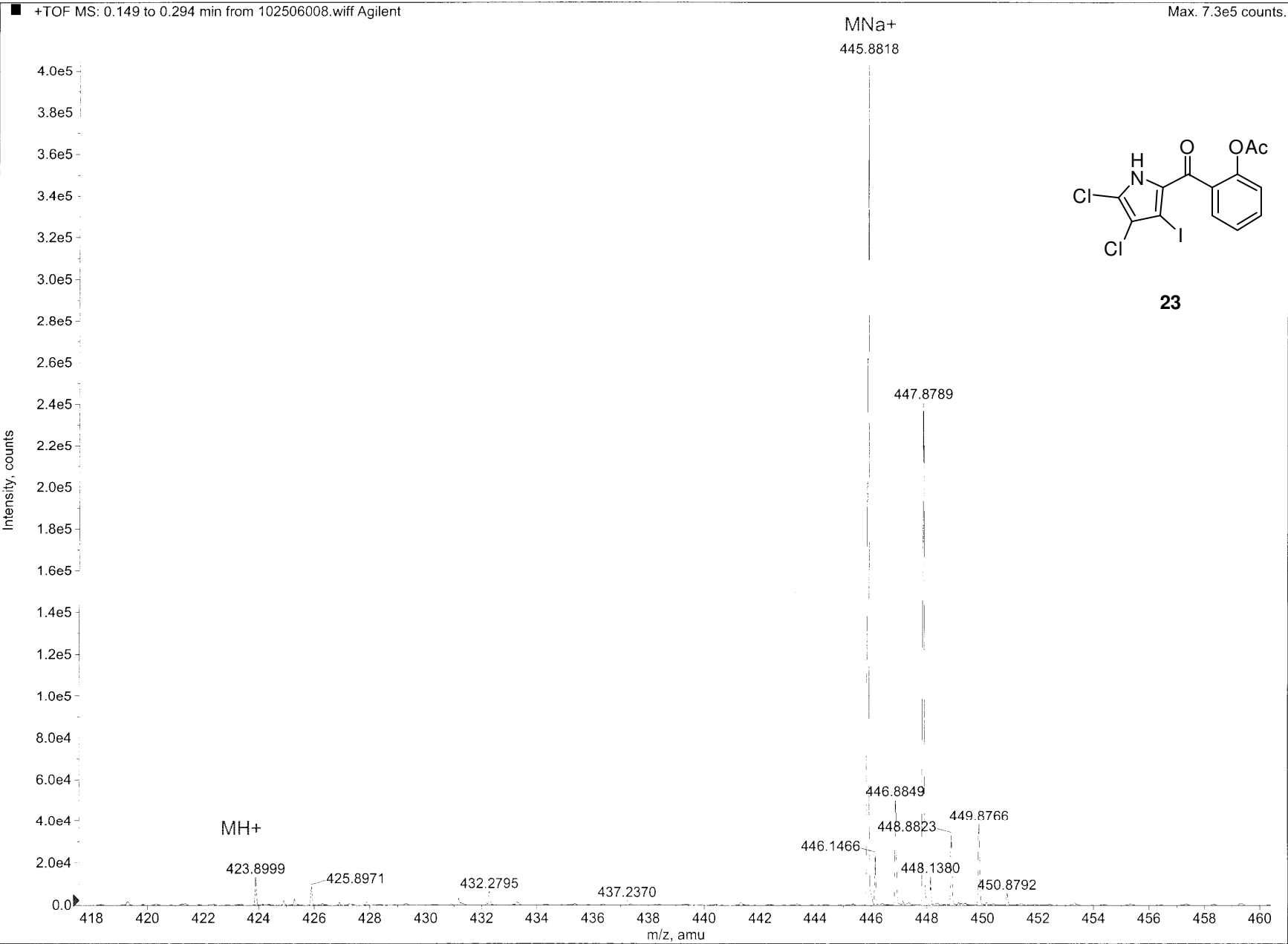


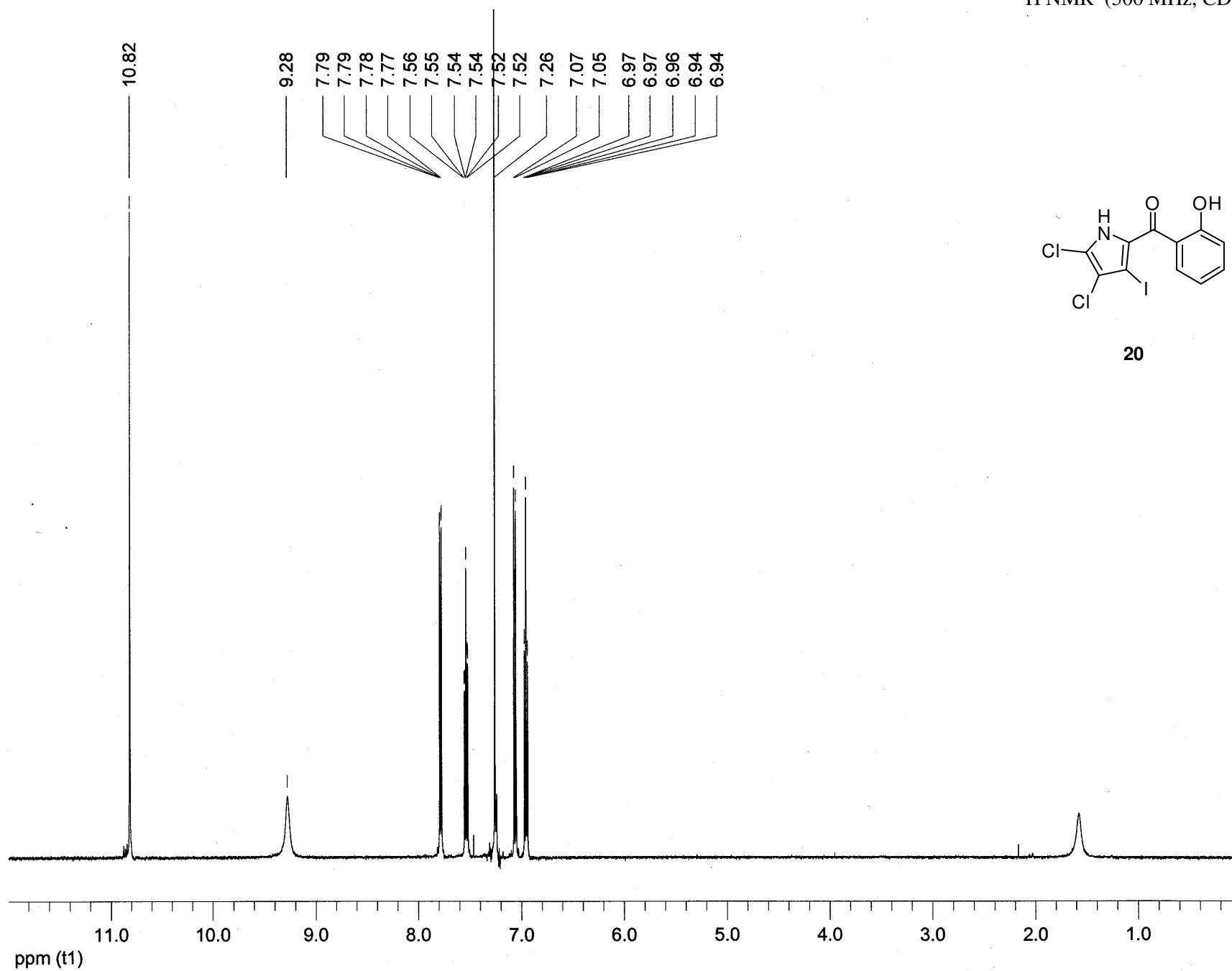
Polarity/Scan Type: Positive

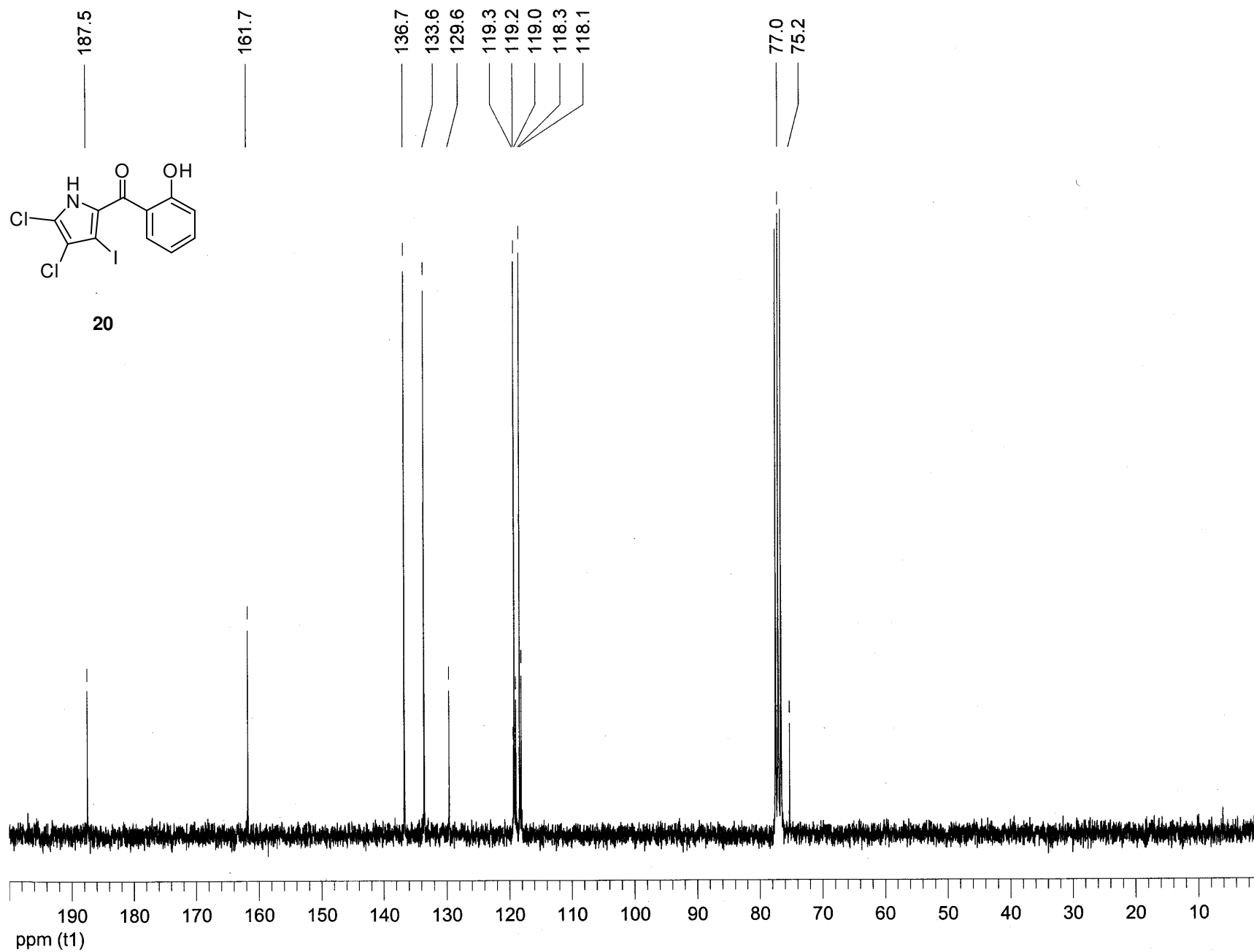
Sample Name: flch8551

Acq. Date: Wednesday, October 25, 2006

Acq. File: 102506008.wiff





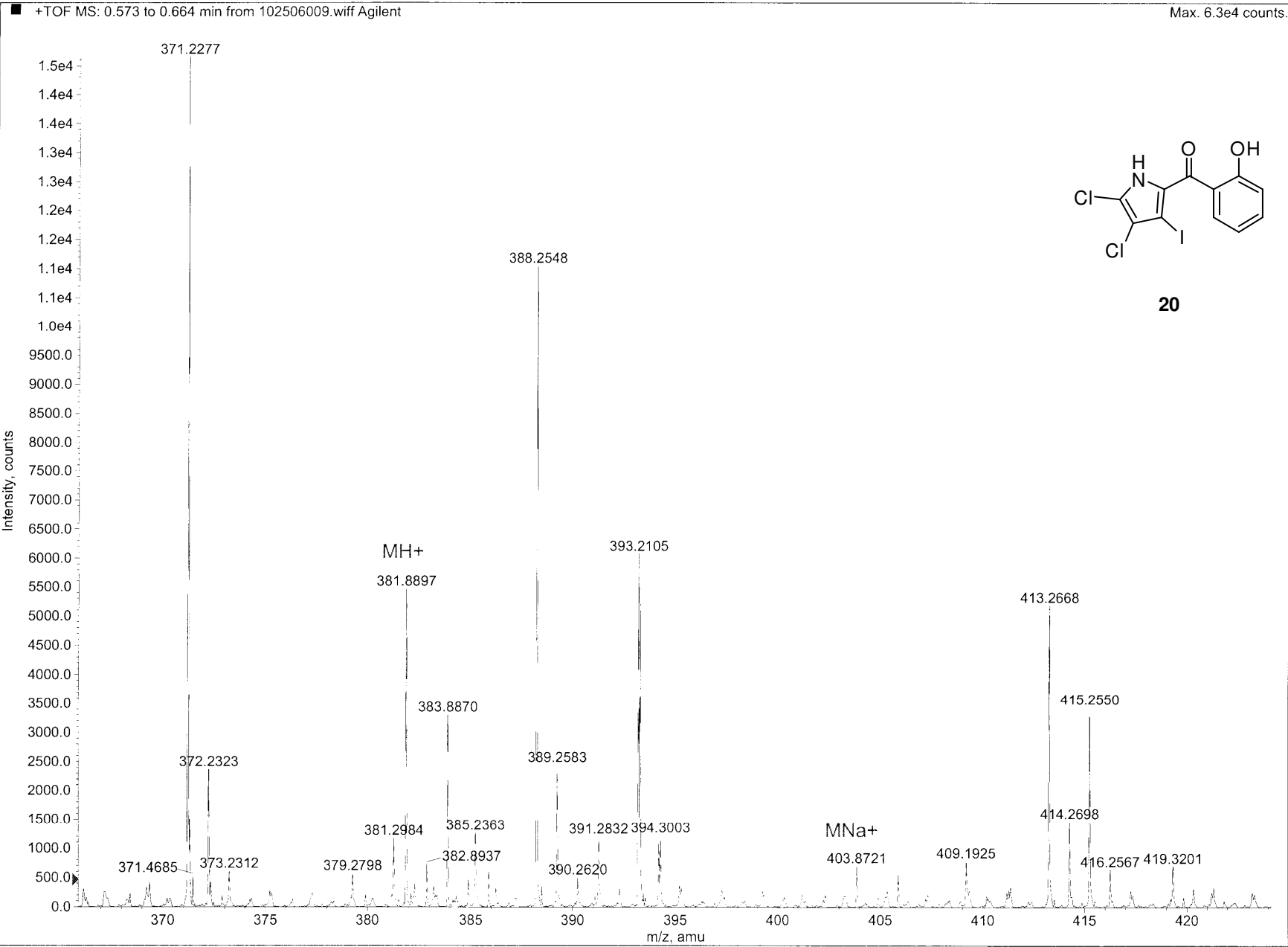


Polarity/Scan Type: Positive

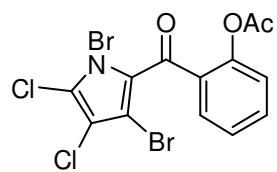
Sample Name: flch8552

Acq. Date: Wednesday, October 25, 2006

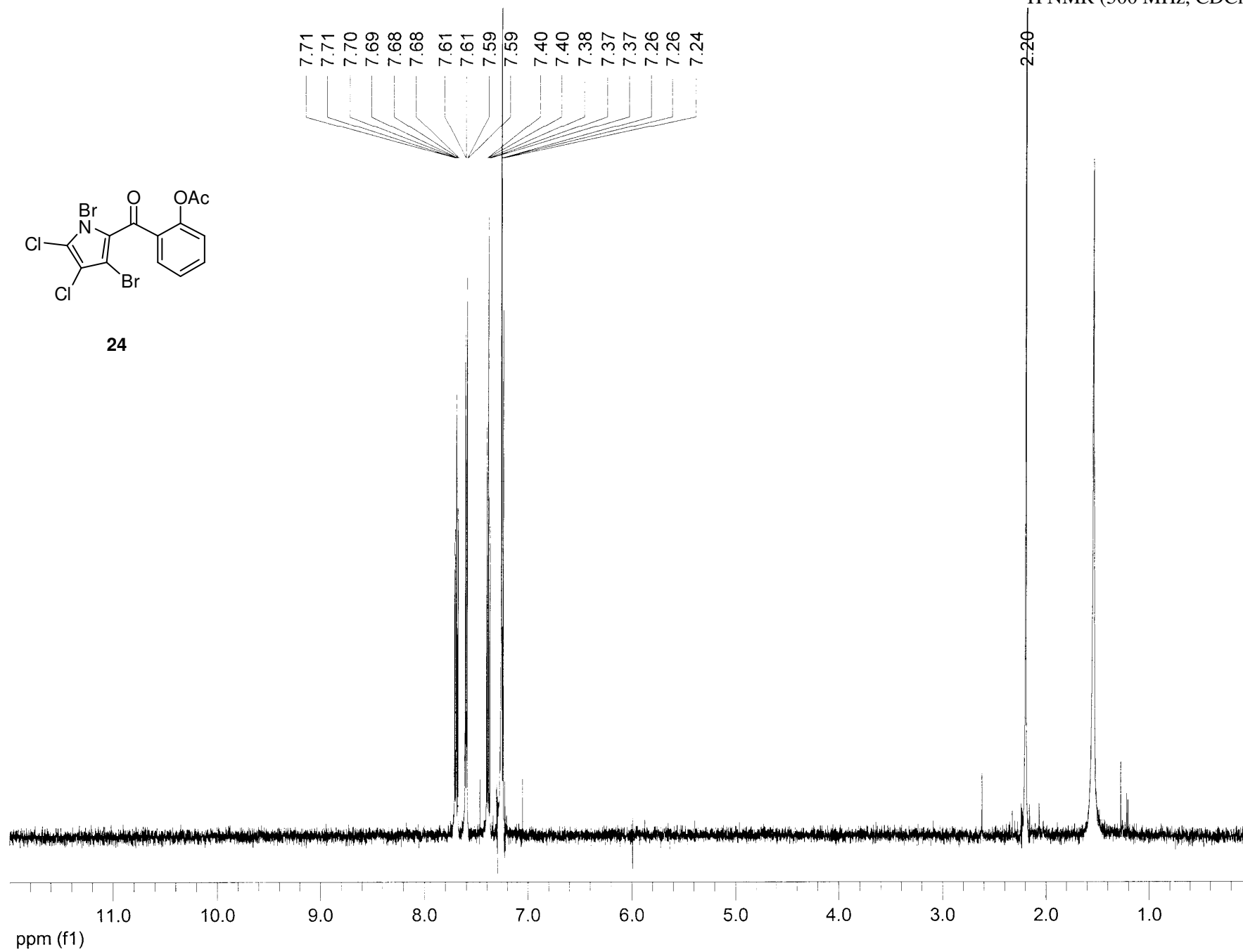
Acq. File: 102506009.wiff



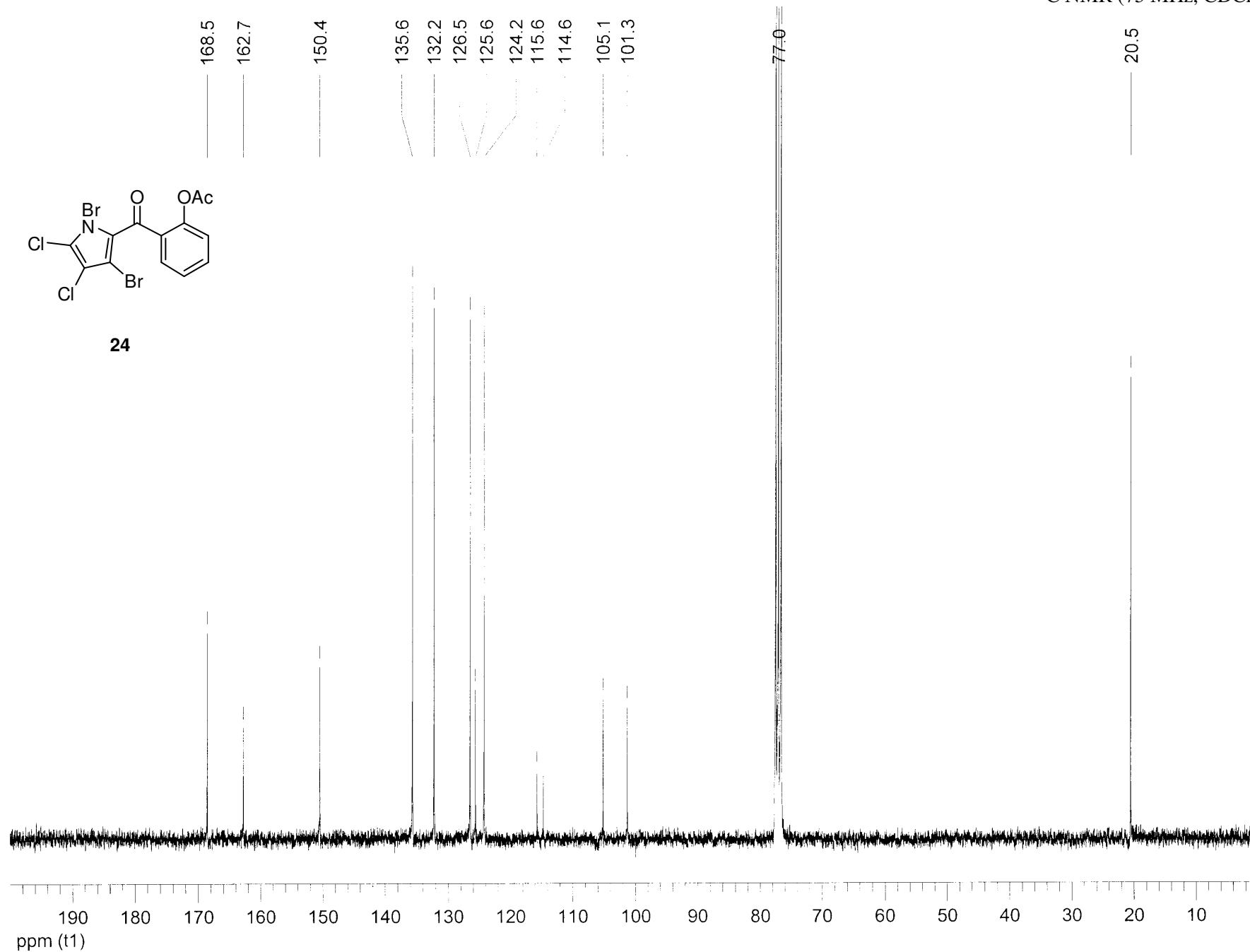
¹H NMR (500 MHz, CDCl₃) of **24**



24



¹³C NMR (75 MHz, CDCl₃) of **24**

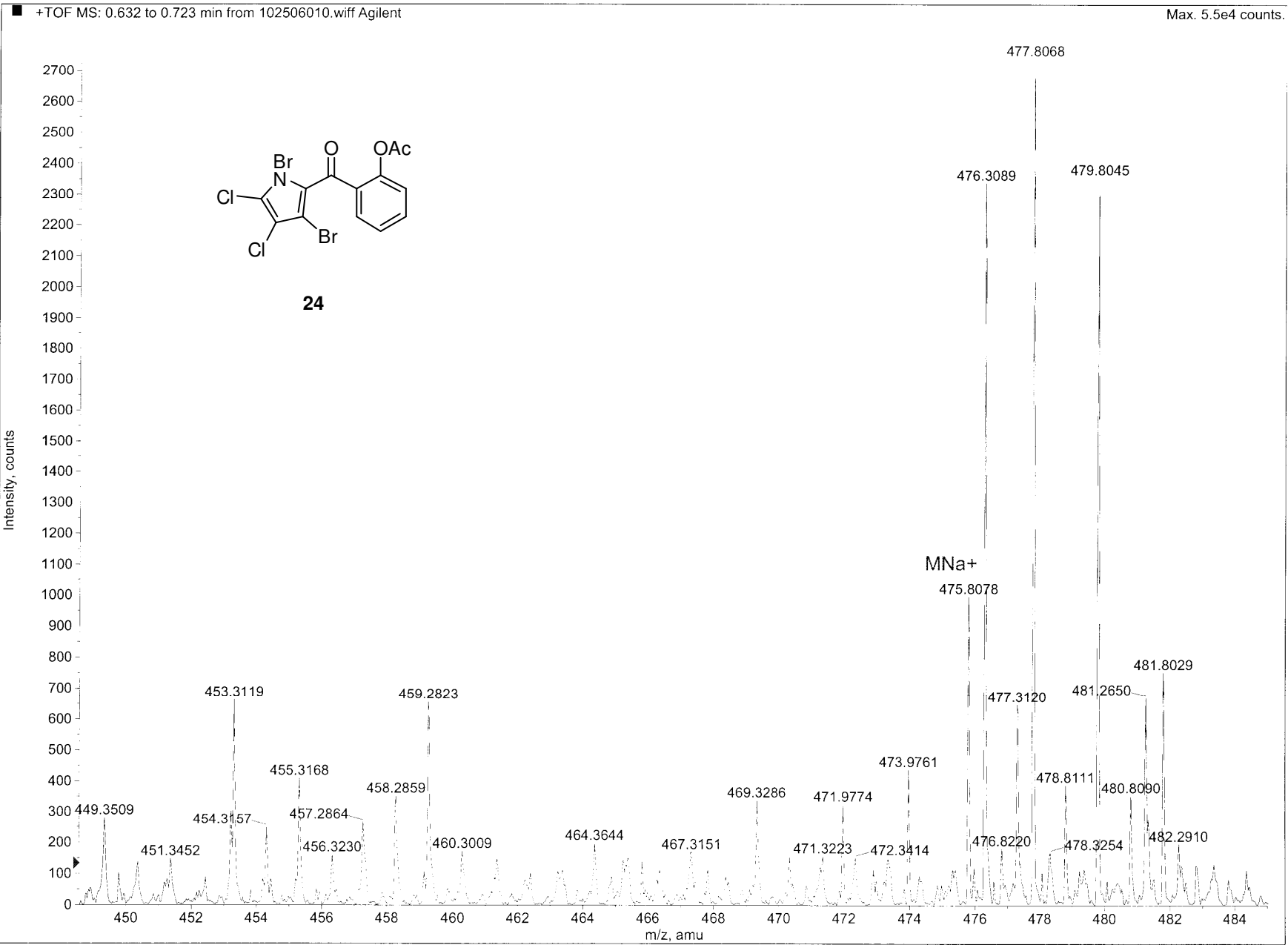


Polarity/Scan Type: Positive

Sample Name: flch8553

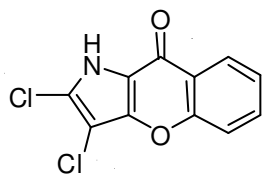
Acq. Date: Wednesday, October 25, 2006

Acq. File: 102506010.wiff



¹H NMR (500 MHz, DMSO-*d*₆) of **25**

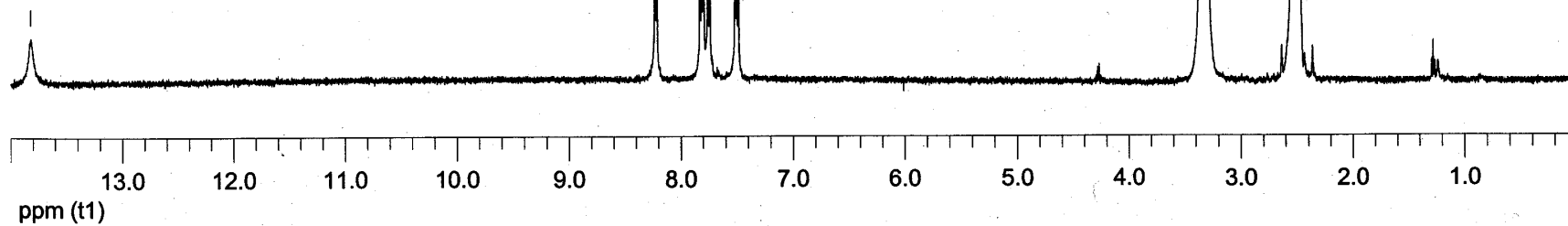
13.82

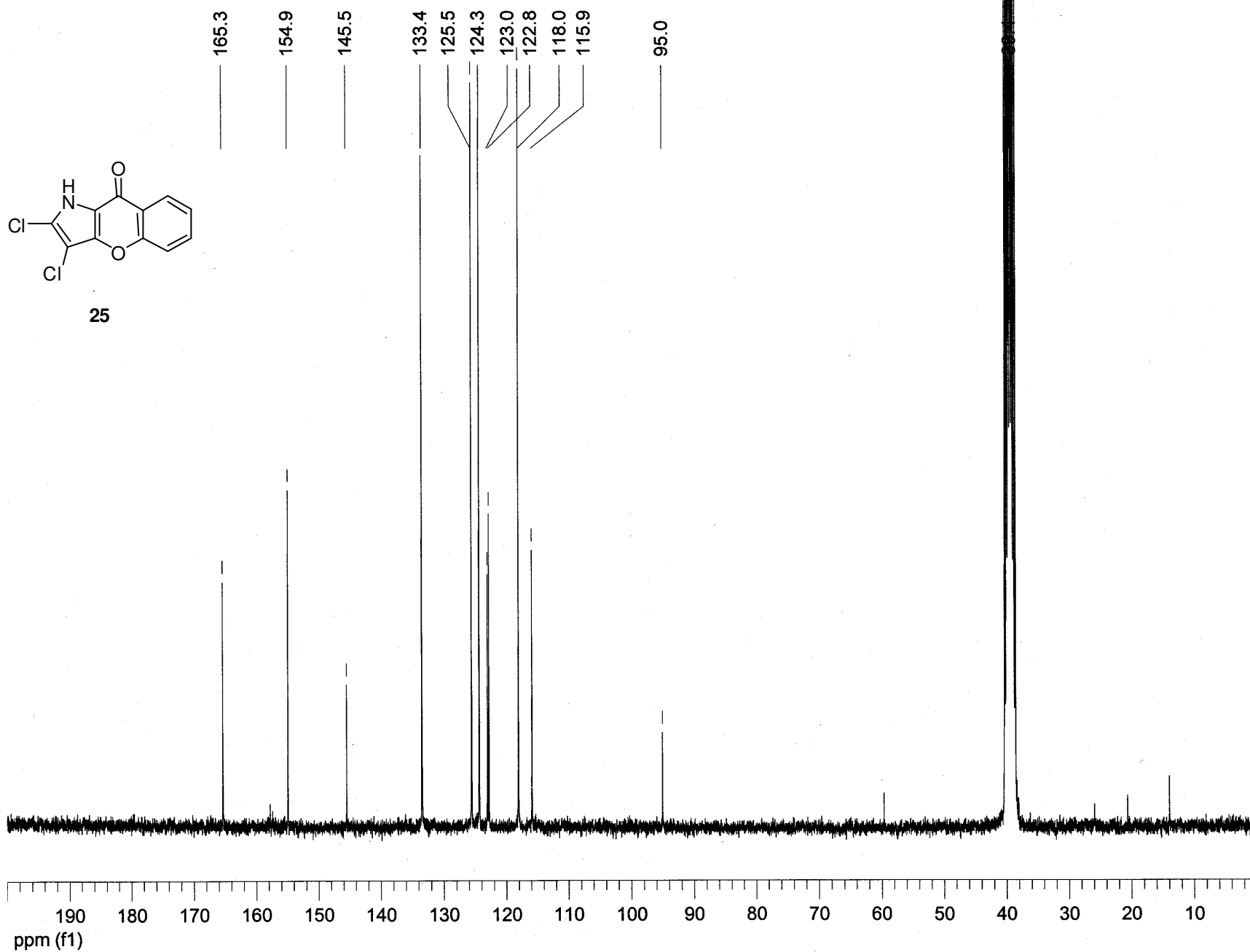


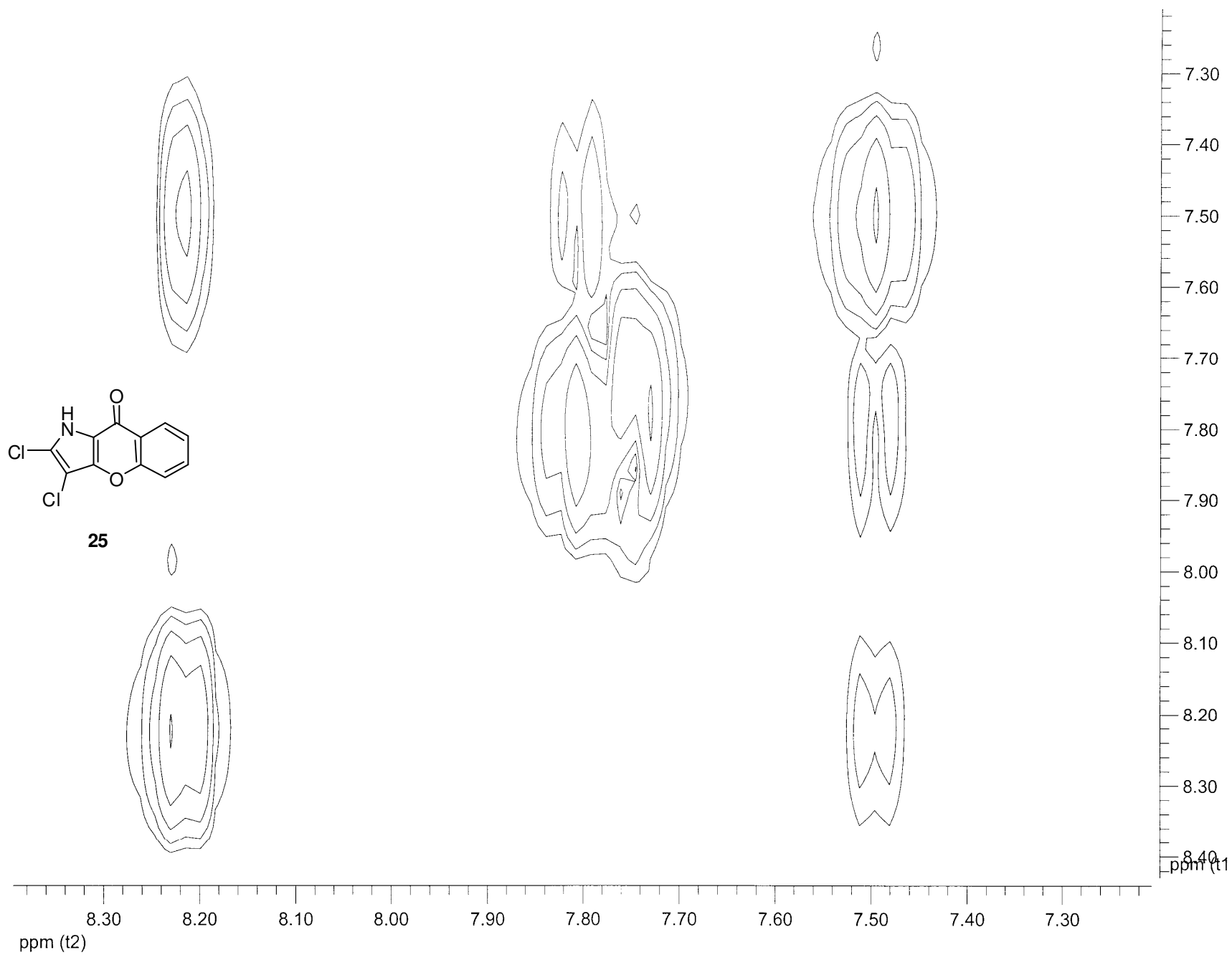
25

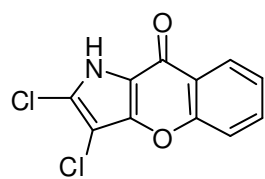
8.23
8.23
8.22
8.21
7.83
7.83
7.81
7.81
7.80
7.79
7.76
7.74
7.51
7.51
7.50
7.48
7.48

2.50

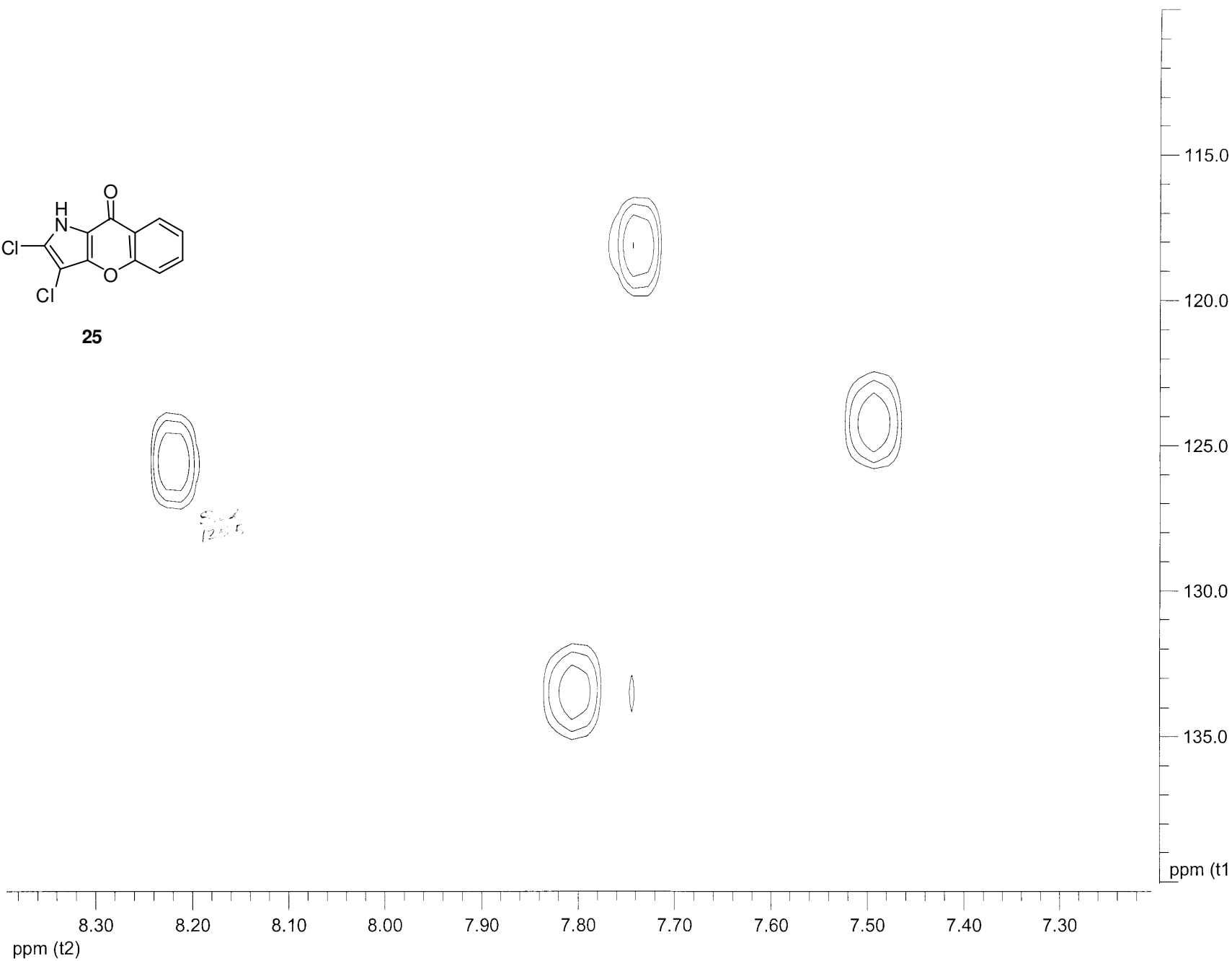


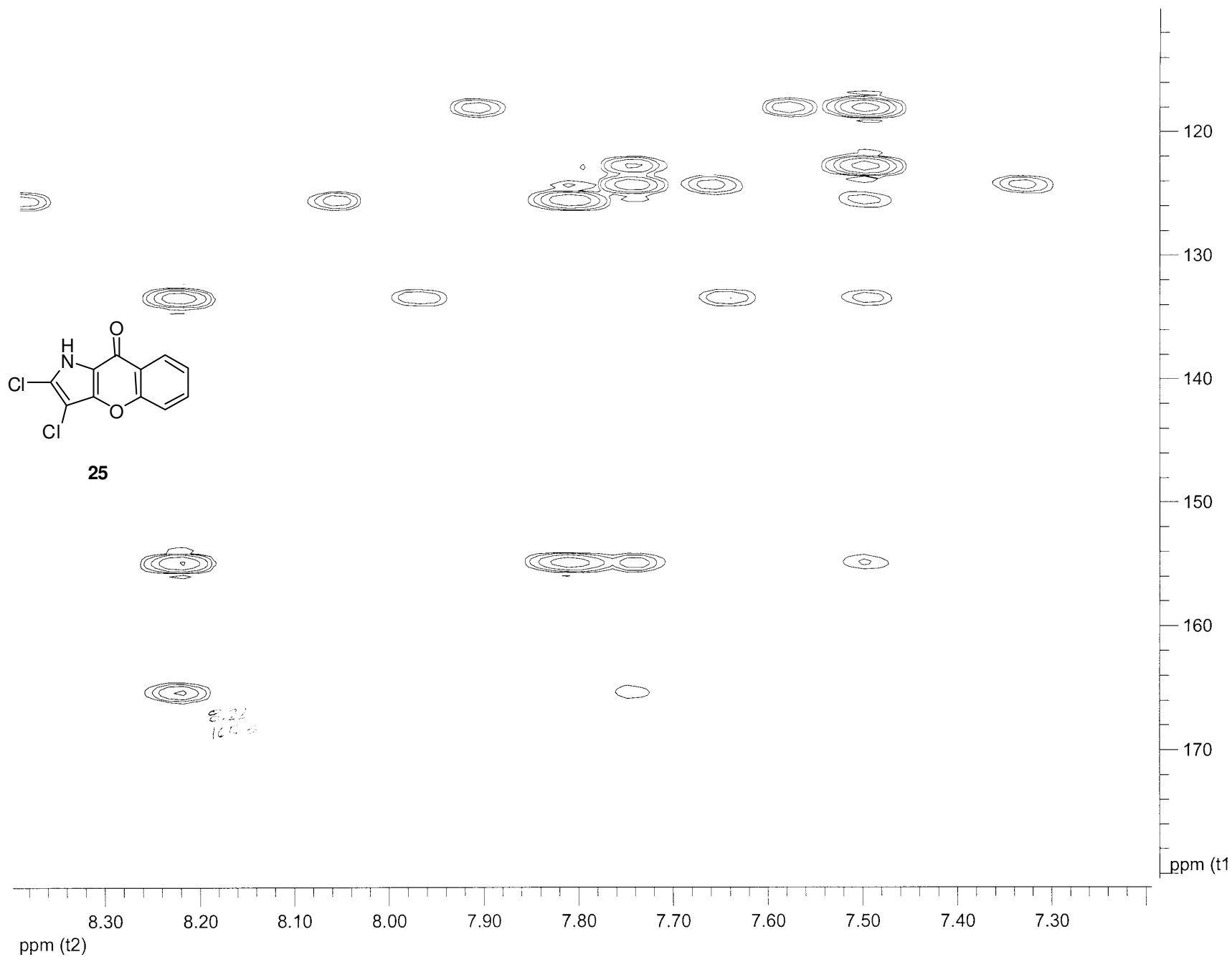






25





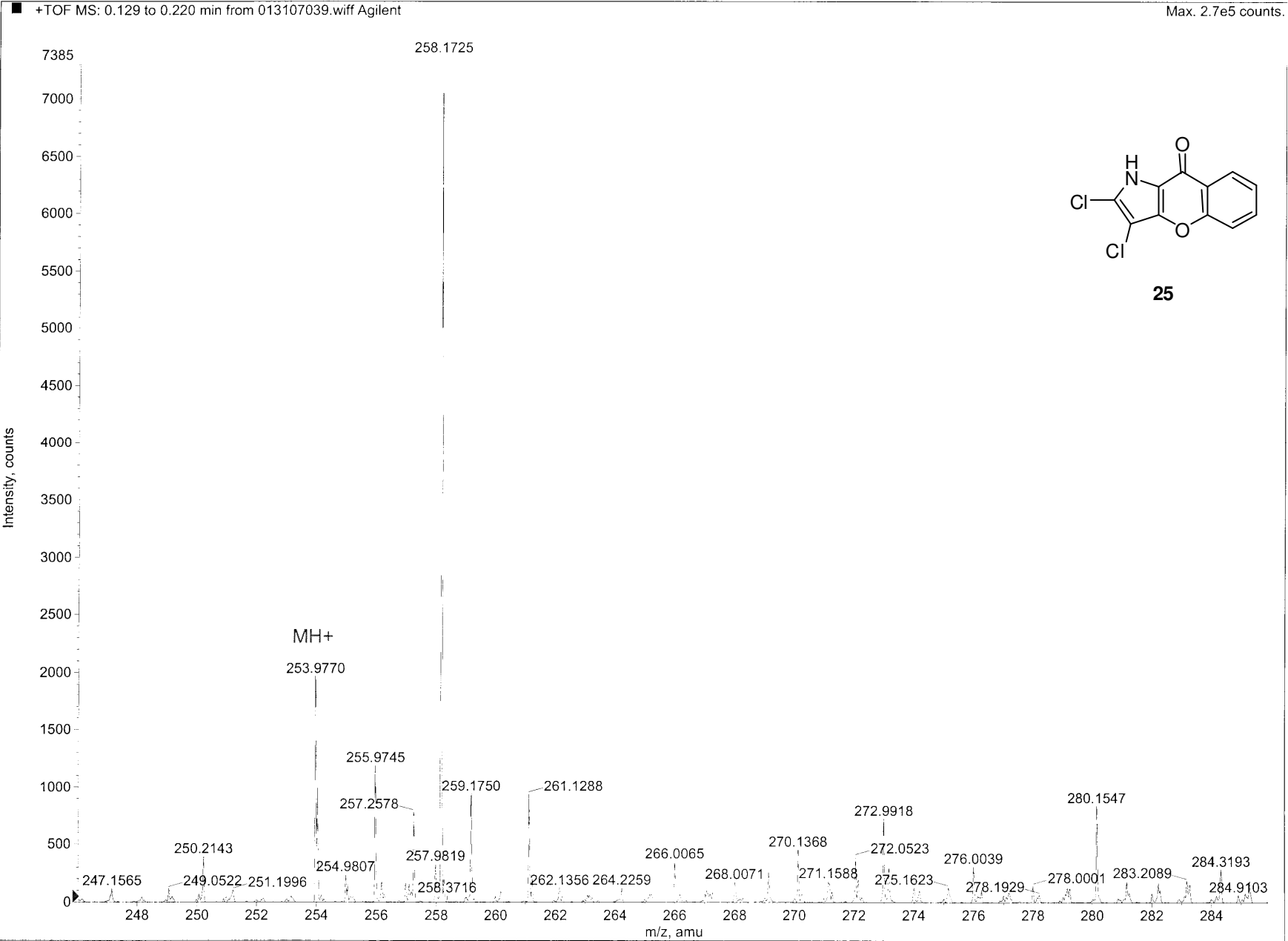
Polarity/Scan Type: Positive

Sample Name: flch2013

Acq. Date: Wednesday, January 31, 2007

Acq. File: 013107039.wiff

Max. 2.7e5 counts.



¹H, ¹³C, and HMBC NMR Spectral Data for **30** (CDCl₃)

position	δ _c ^a	δ _H , mult. (J, Hz) ^b	HMBC
1		9.42, br s	
2	129.7		
3	117.5		
4	109.9	6.29, m	3,5
5	120.4	6.89, m	2-4
6	160.0		
7	60.3	4.12, q (7.2)	6,8
8	13.9	1.10, t (7.2)	7
2'	124.6		
3'	117.8	7.06, m	2',4'-6'
4'	108.5	6.25, m	2',3',5'
5'	130.0	6.88, m	2'-4'
6'	160.6		
7'	59.6	4.16, q (7.2)	6',8'
8'	14.2	1.22, t (7.2)	7'

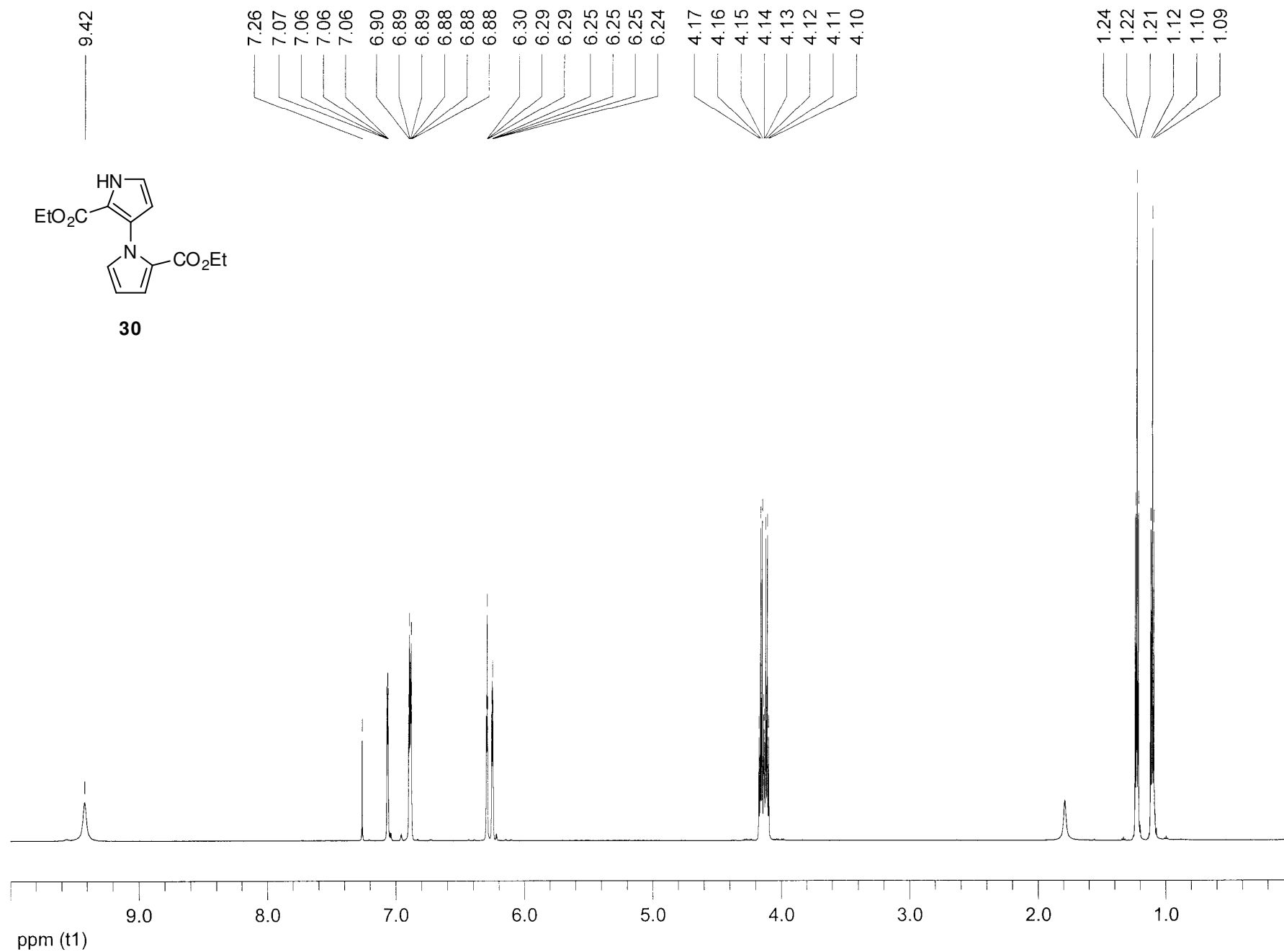
^a 75 MHz. ^b 600 MHz.

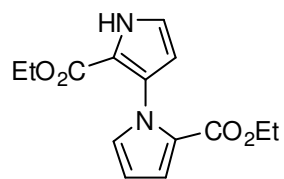
¹H, ¹³C, and HMBC NMR Spectral Data for **31** (CDCl₃)

position	δ _c ^a	δ _H , mult. (J, Hz) ^b	HMBC
1		10.45, br s	2,4
2	126.8		
3	120.5 ^c		
4	102.2		
5	106.0 ^c		
6	158.2		
7	61.3	4.13, m	6,8
8	13.8	1.08, t (7.2)	7
2'	125.9		
3'	120.0	7.17, s	2',5',6'
4'	100.5 ^c		
5'	114.3		
6'	158.6		
7'	60.7	4.20, q (7.2)	6',8'
8'	14.1	1.26, t (7.2)	7'

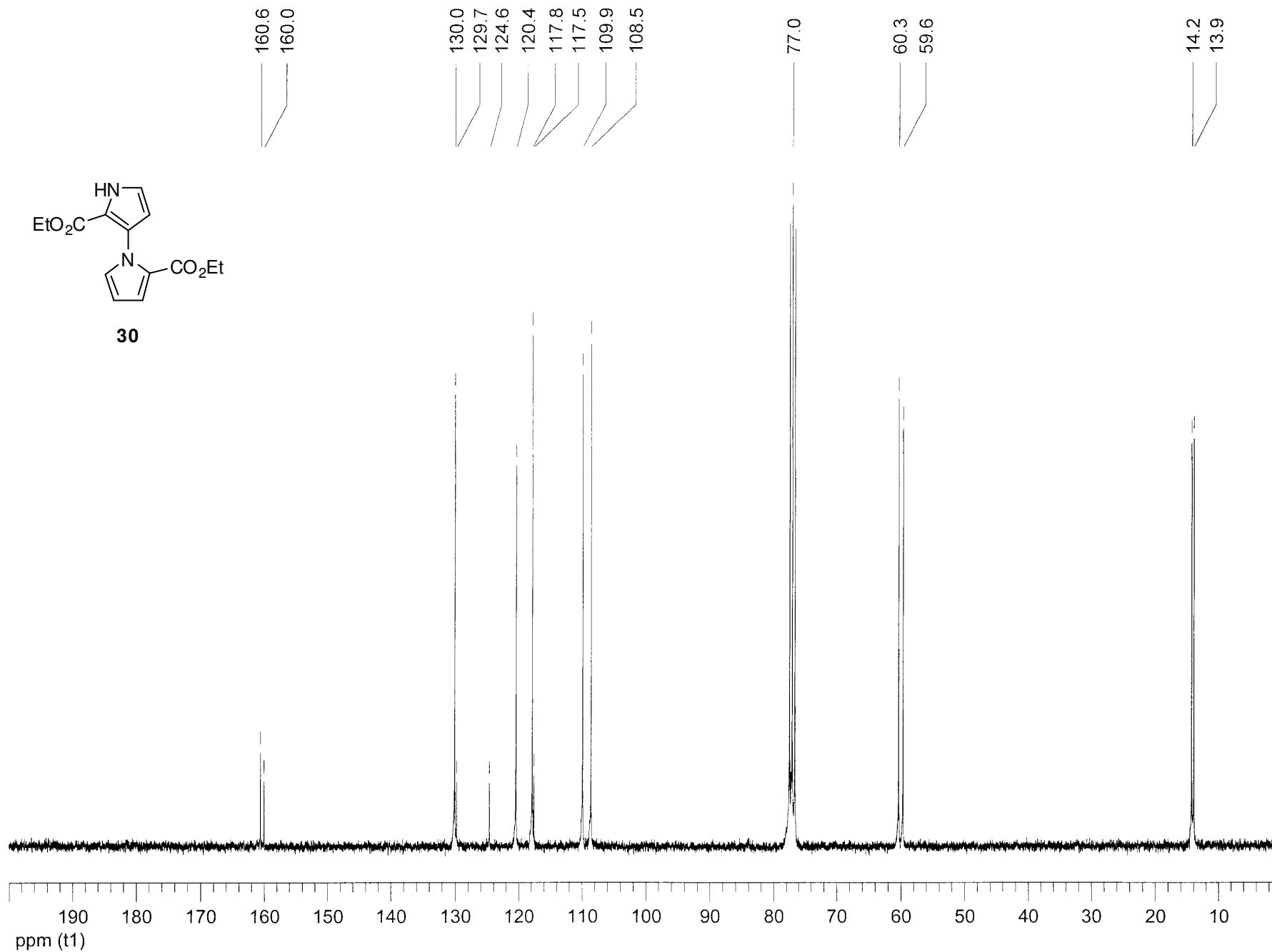
^a 75 MHz. ^b 600 MHz. ^c Carbon assignments were based on ¹³C data for **30**.

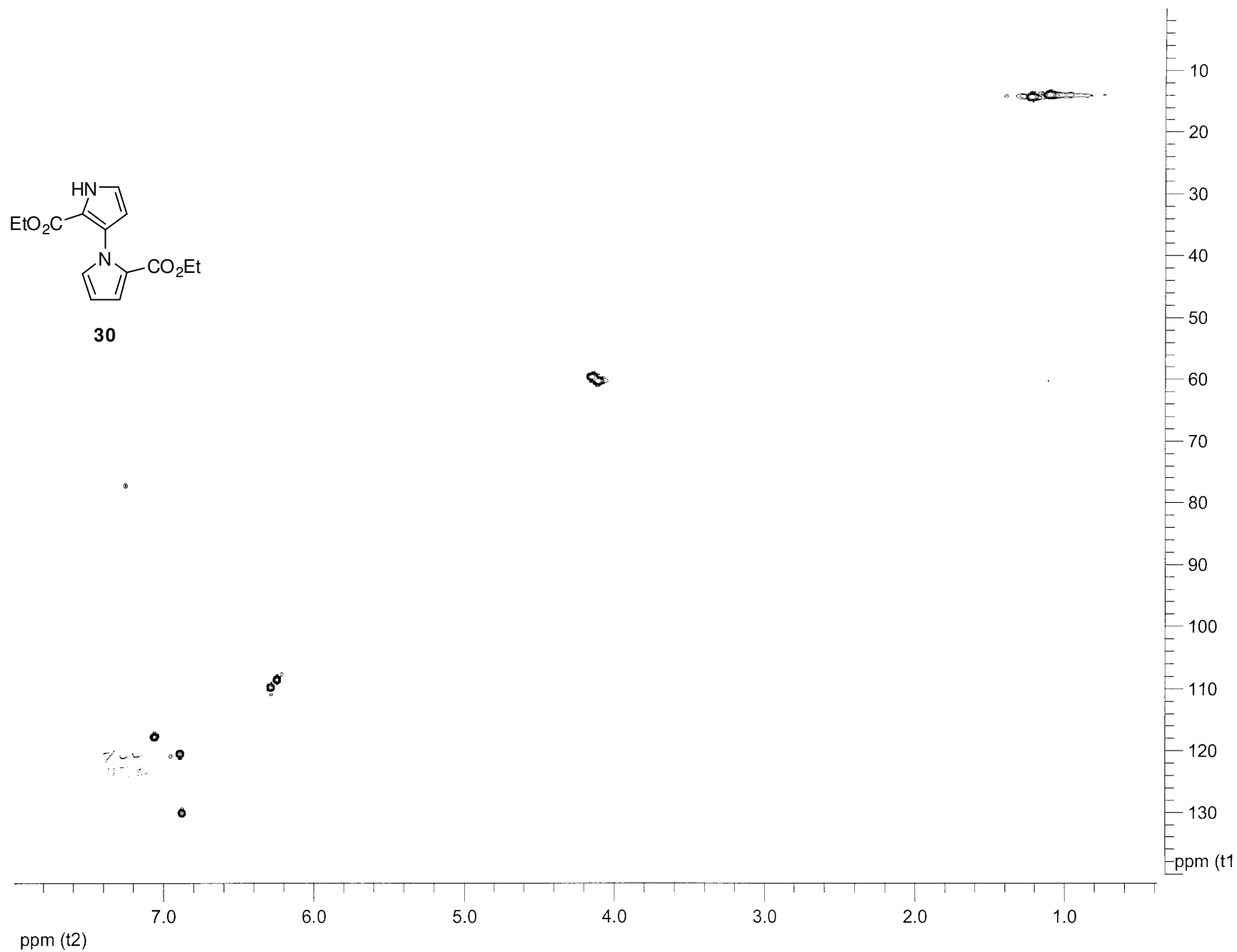
¹H NMR (600 MHz, CDCl₃) of **30**

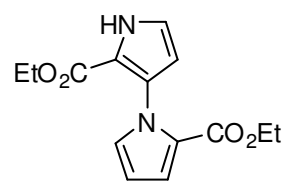




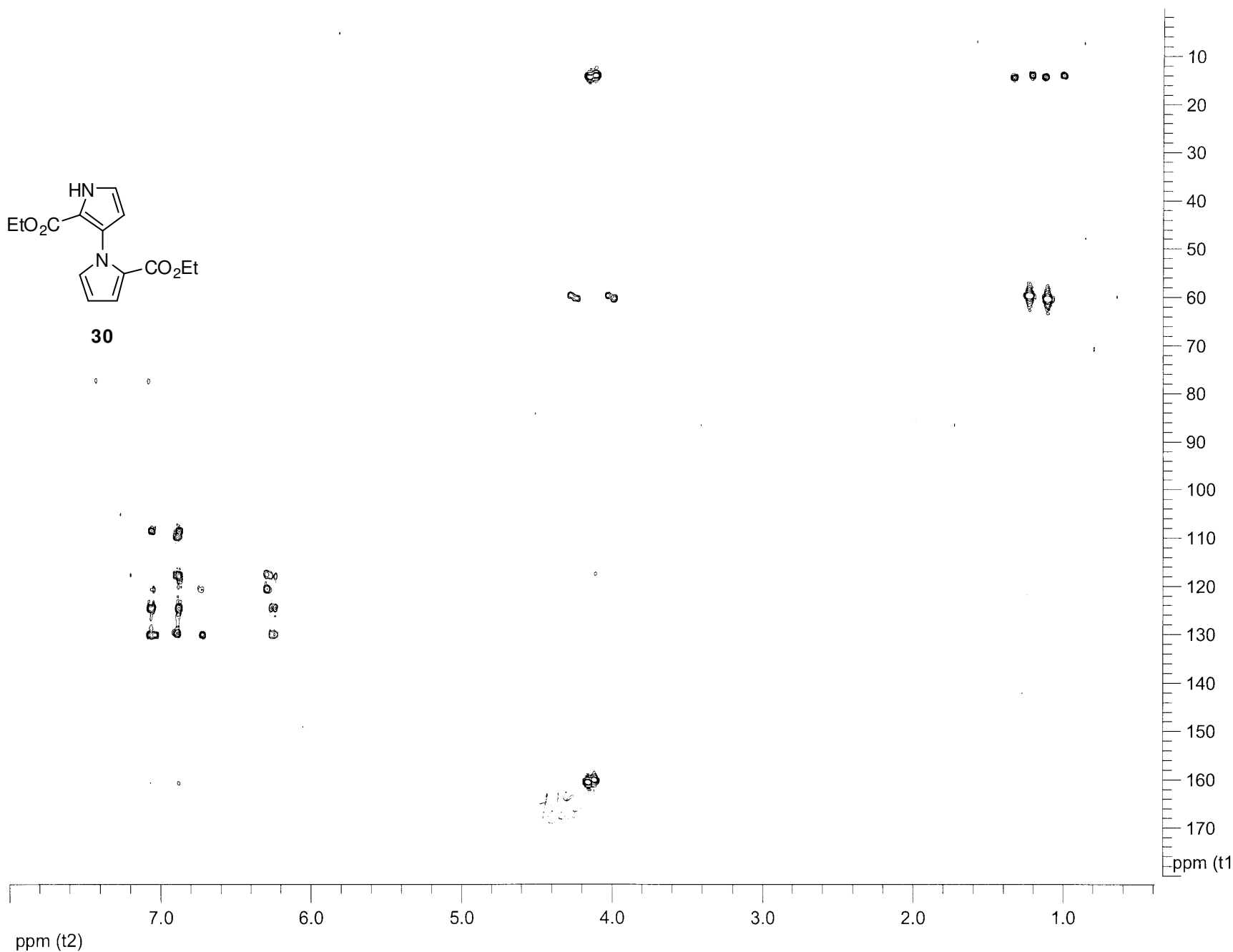
30







30

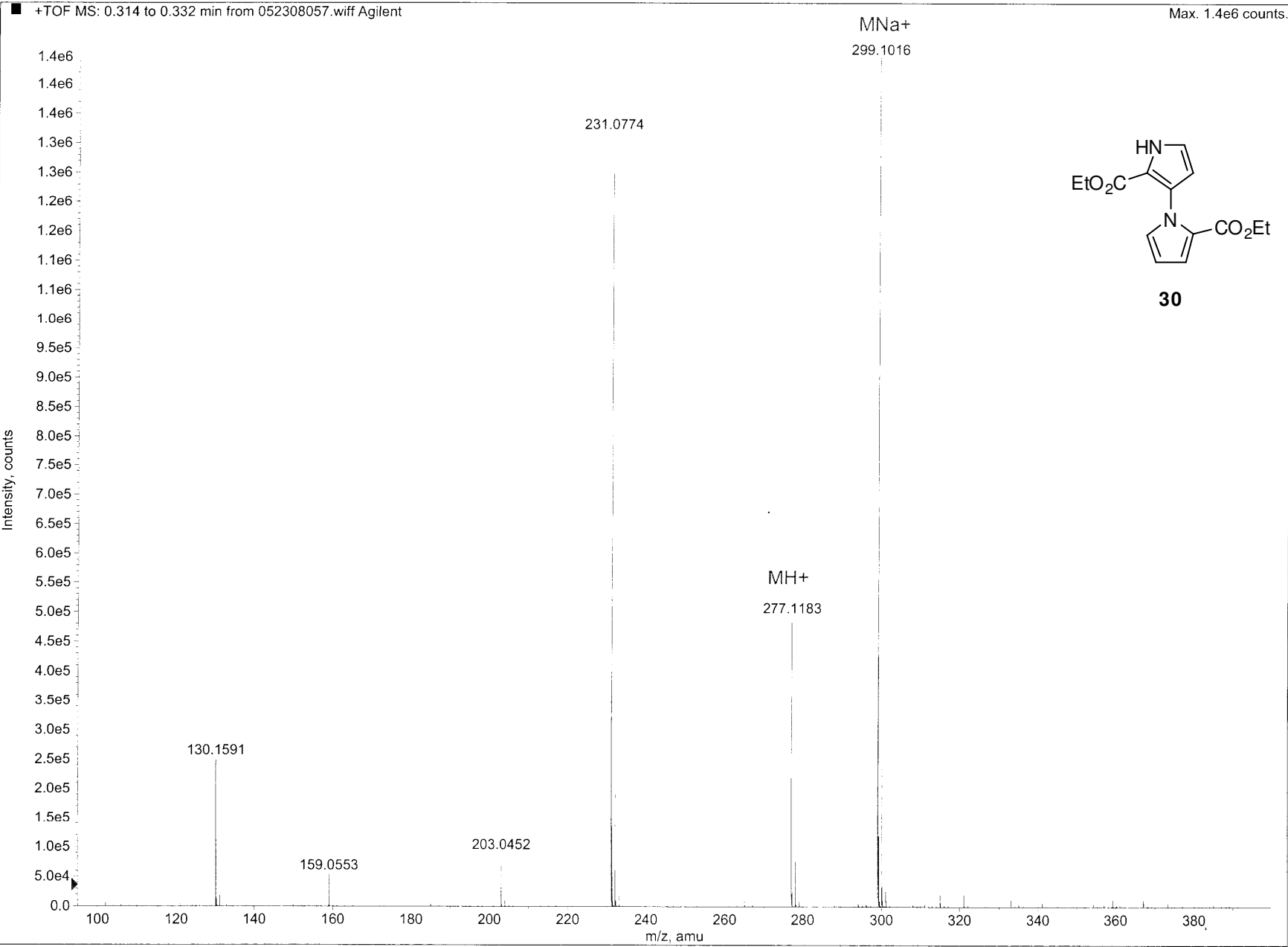


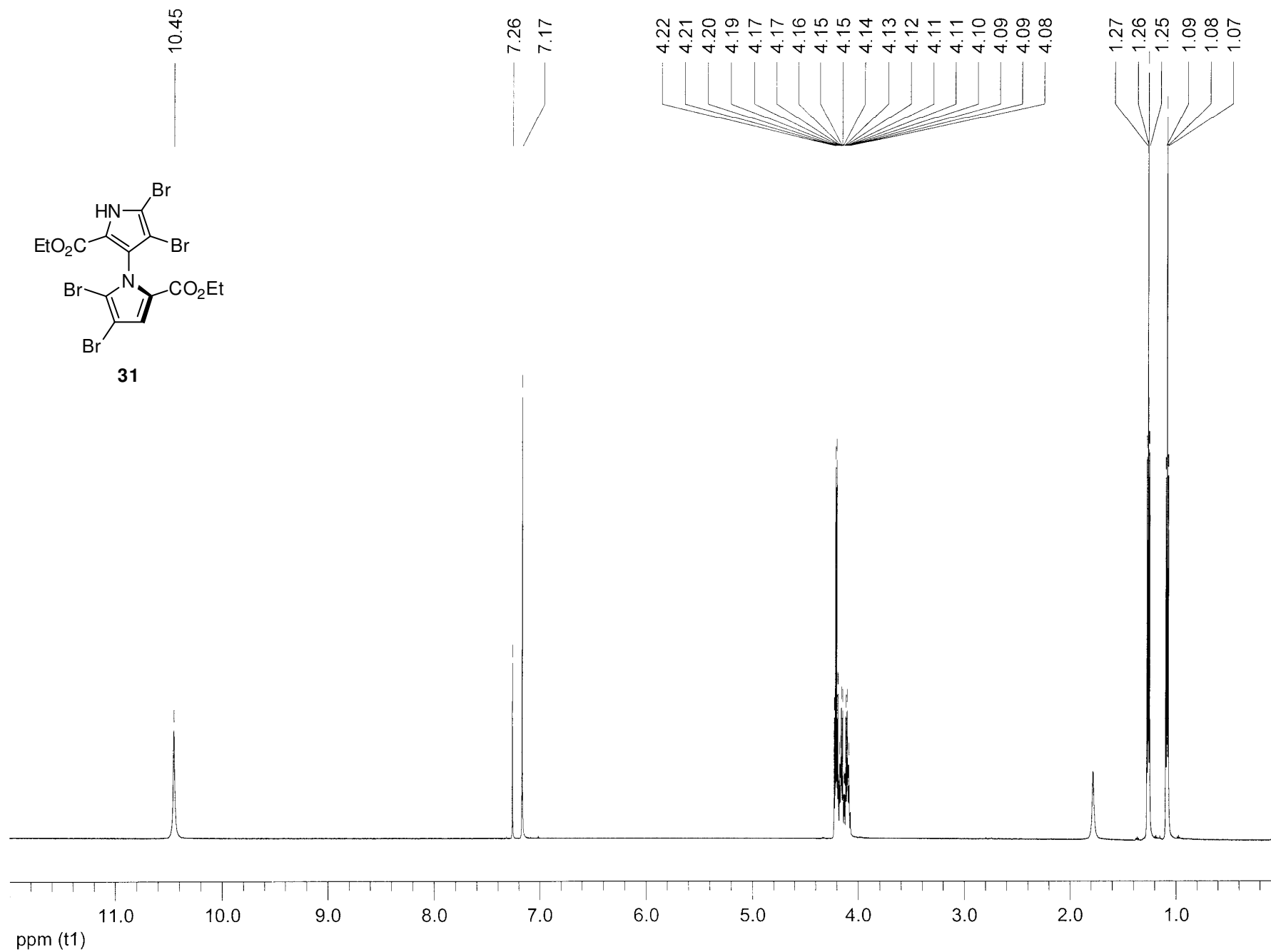
Polarity/Scan Type: Positive

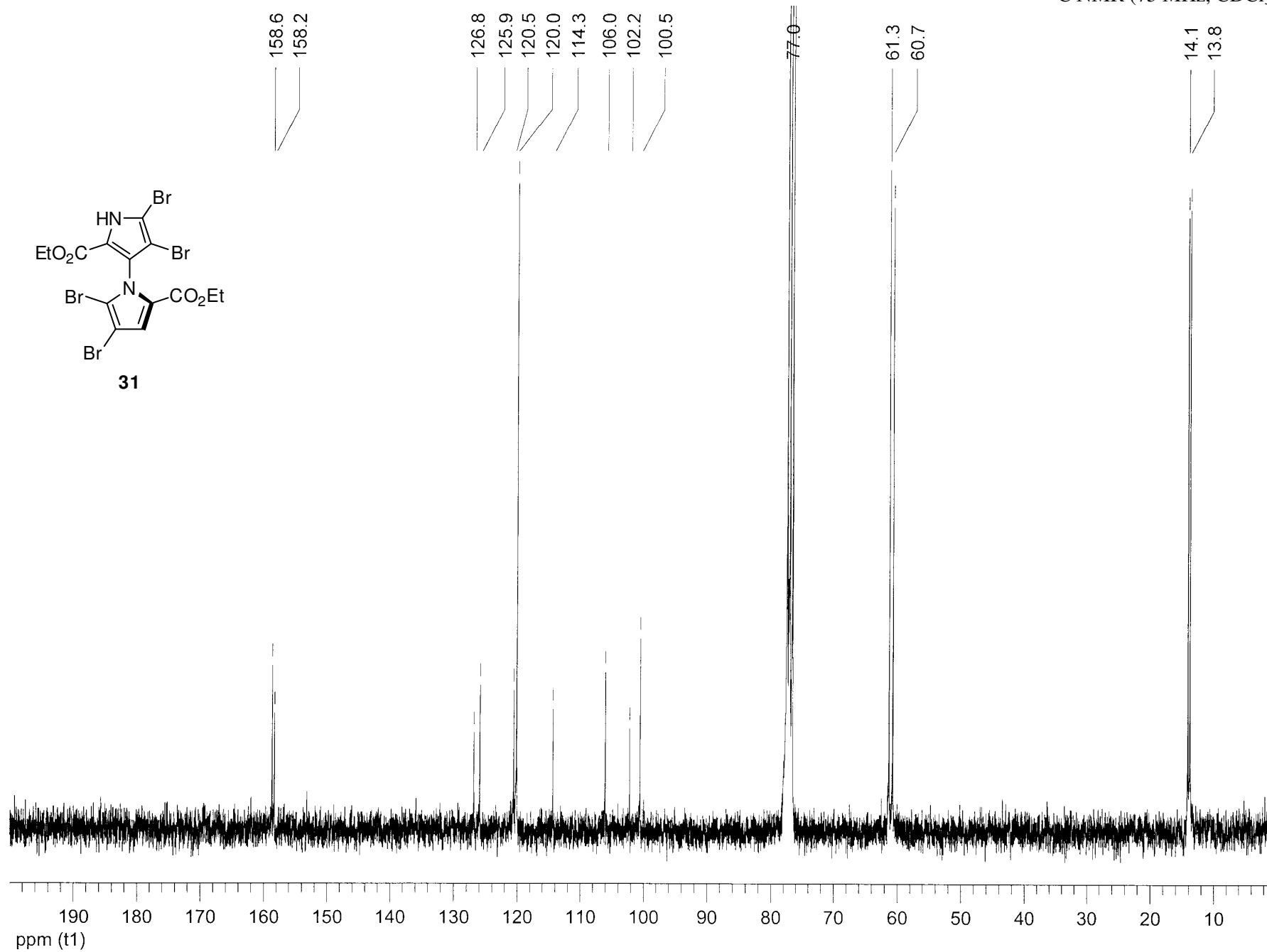
Sample Name: flch4384

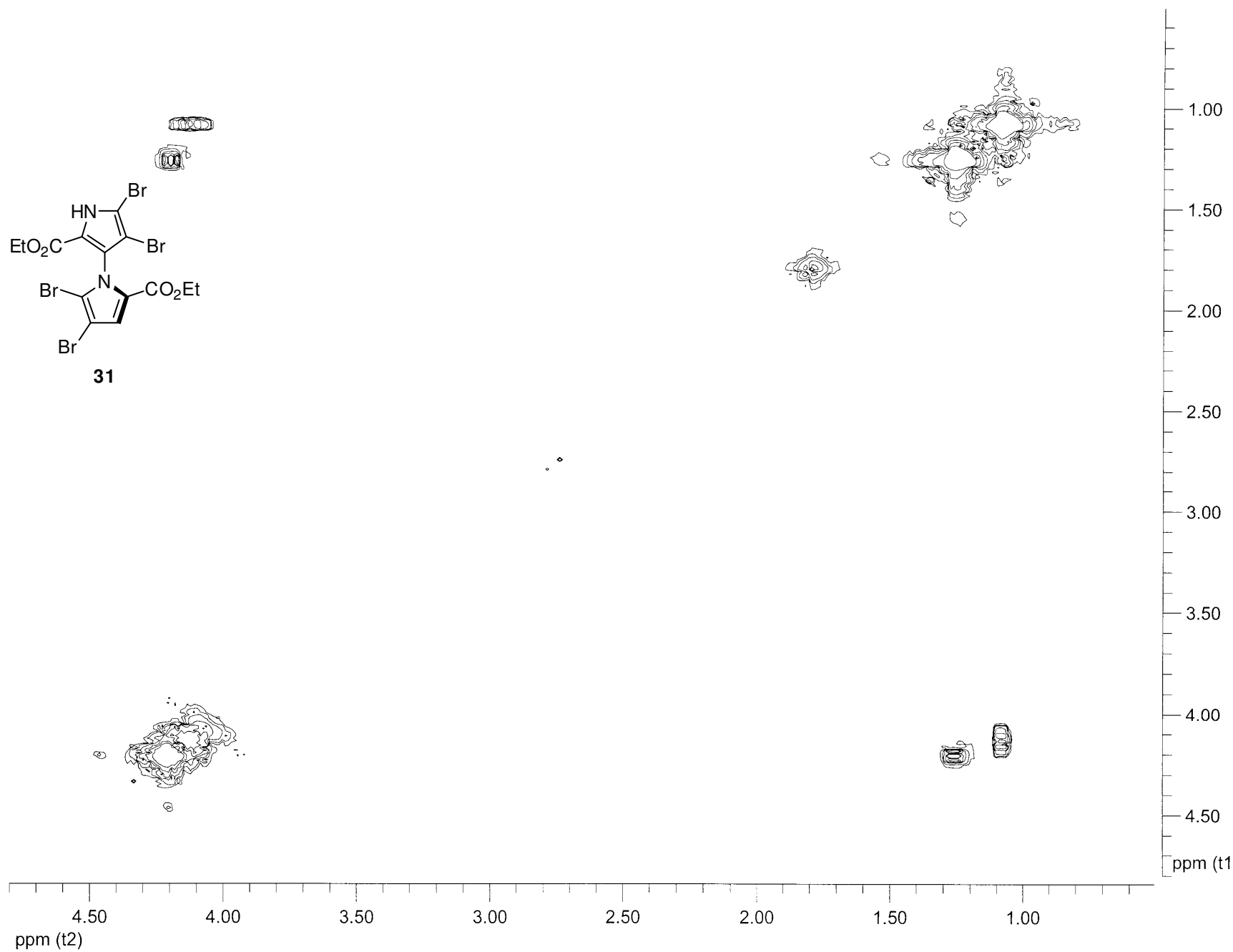
Acq. Date: Friday, May 23, 2008

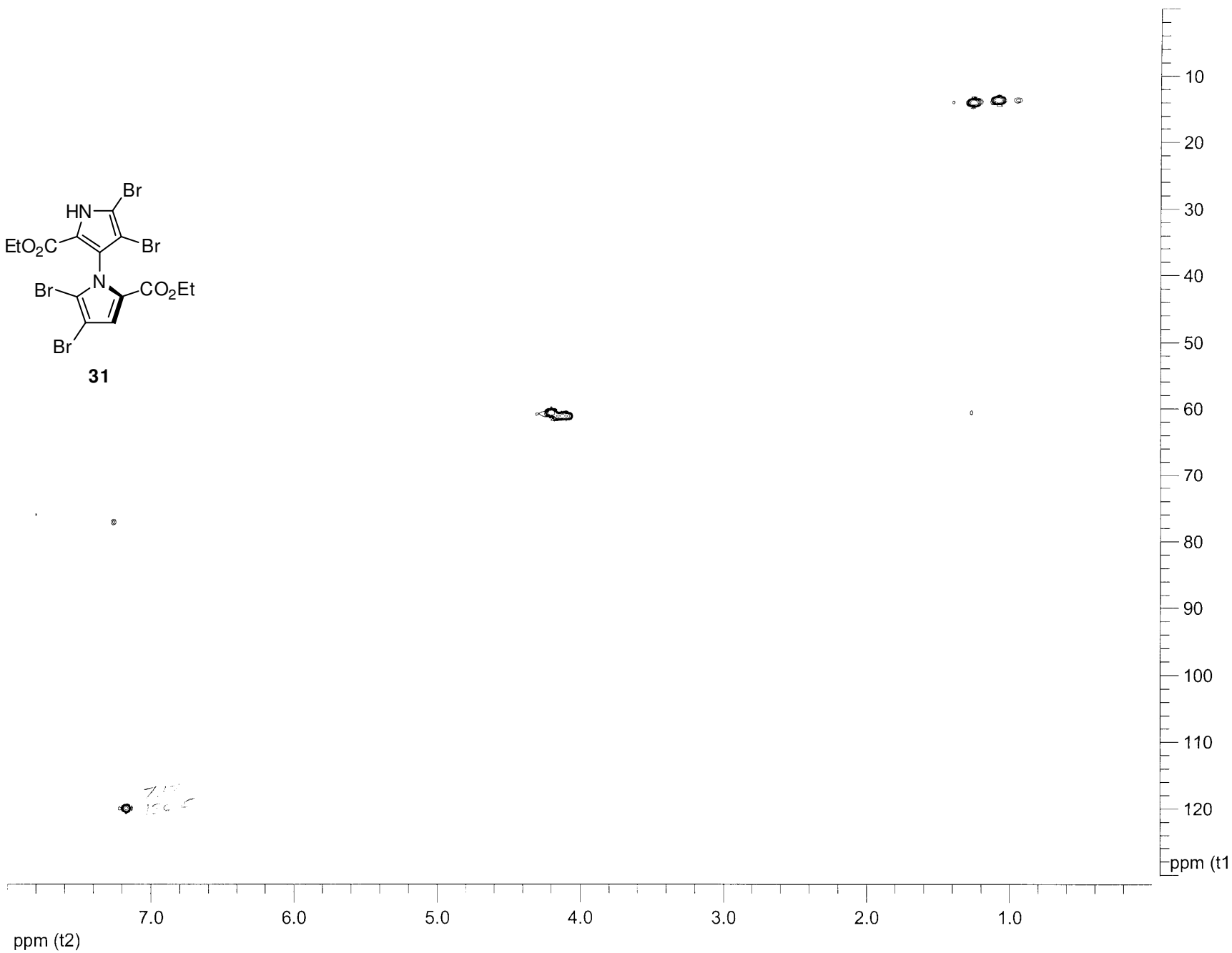
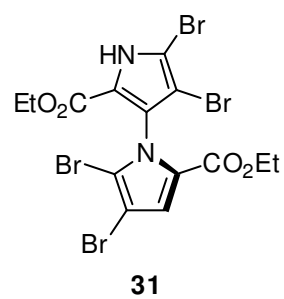
Acq. File: 052308057.wiff

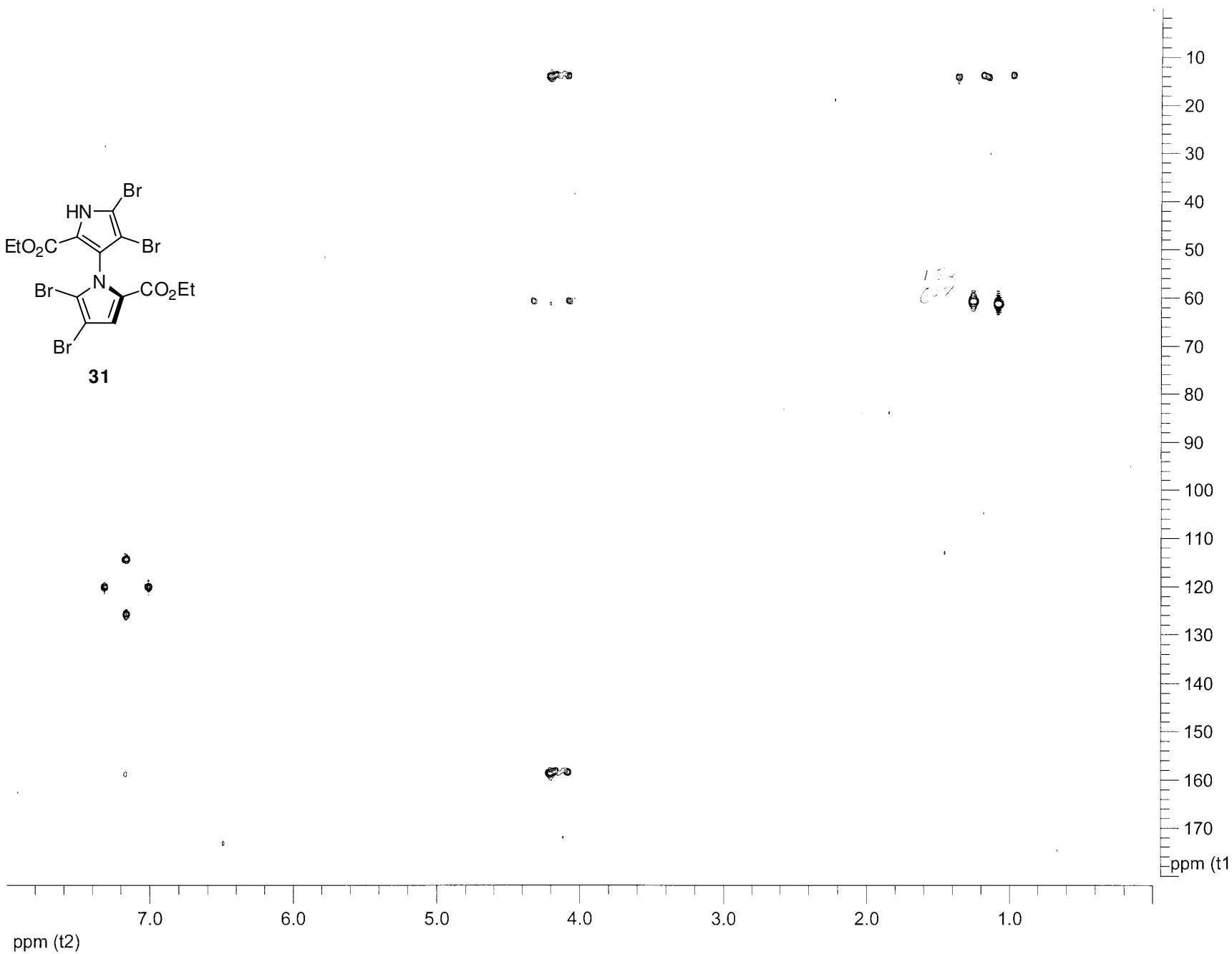
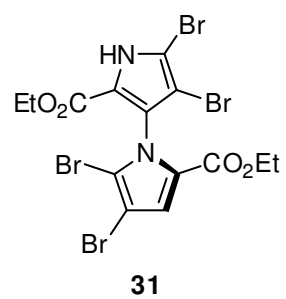












Polarity/Scan Type: Positive

Sample Name: flch4385

Acq. Date: Friday, May 23, 2008

Acq. File: 052308058.wiff

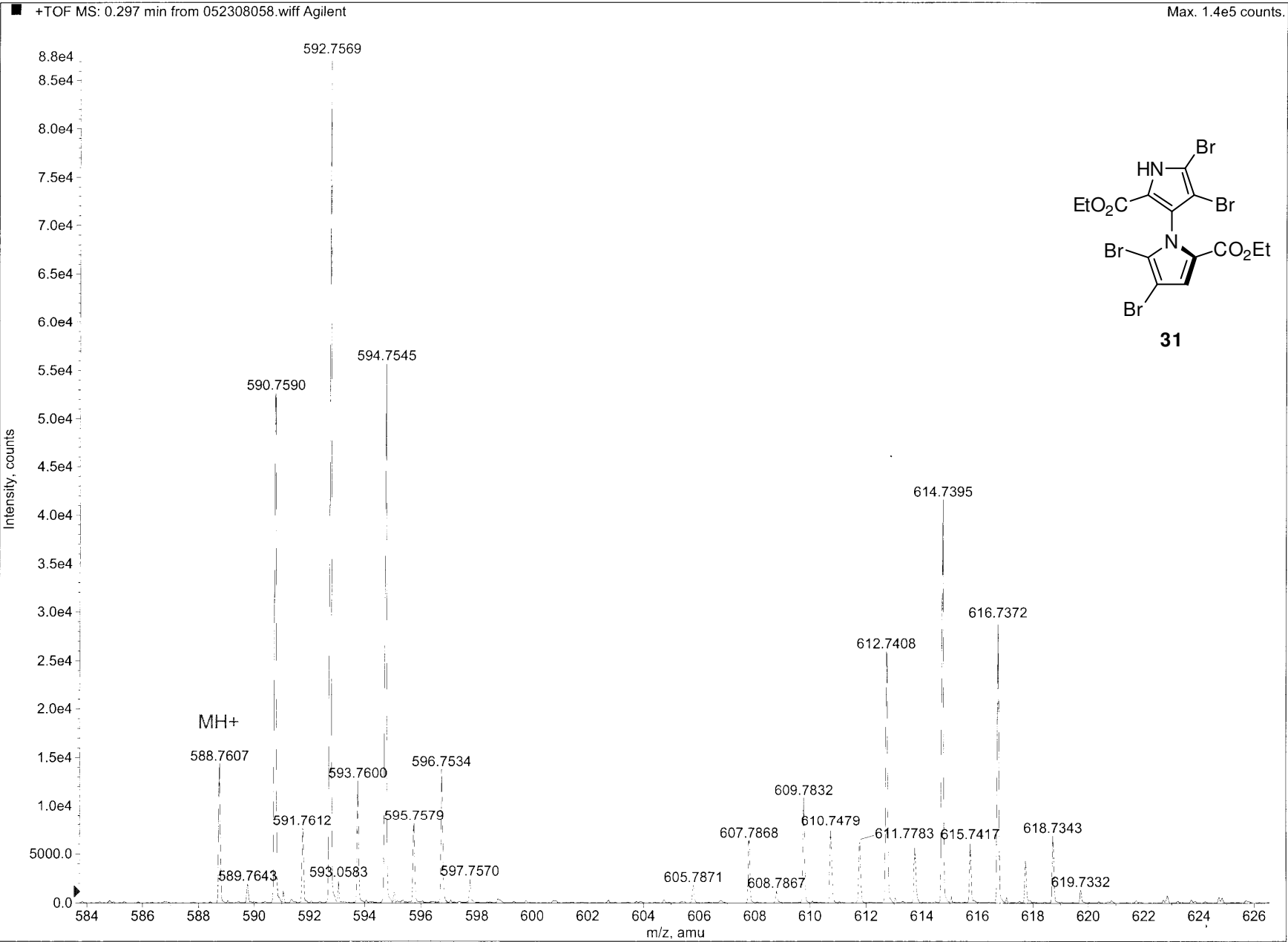


Figure S1. ORTEP of marinopyrrole F (**6**)

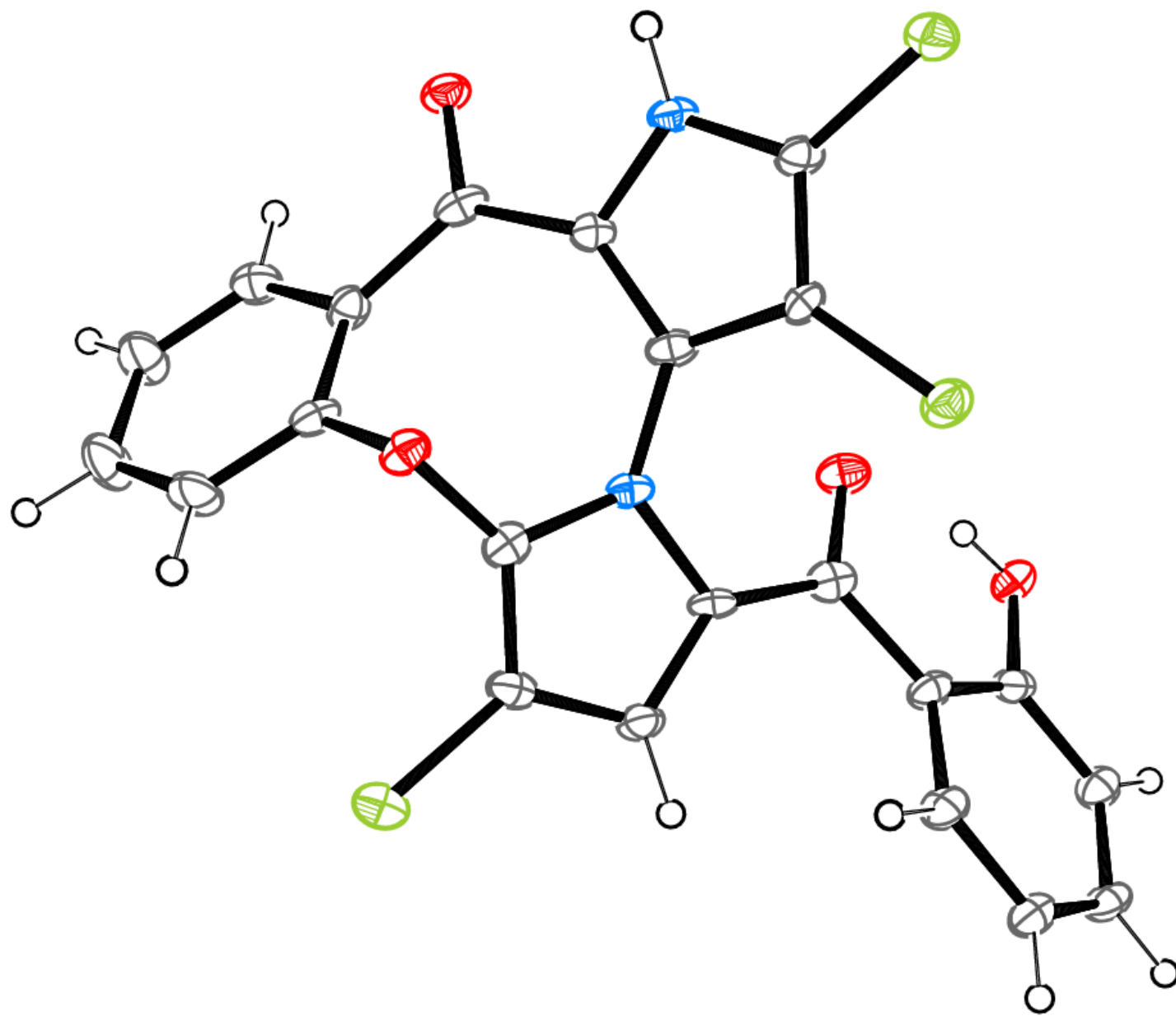


Figure S2. ORTEP of bromophenol **19**

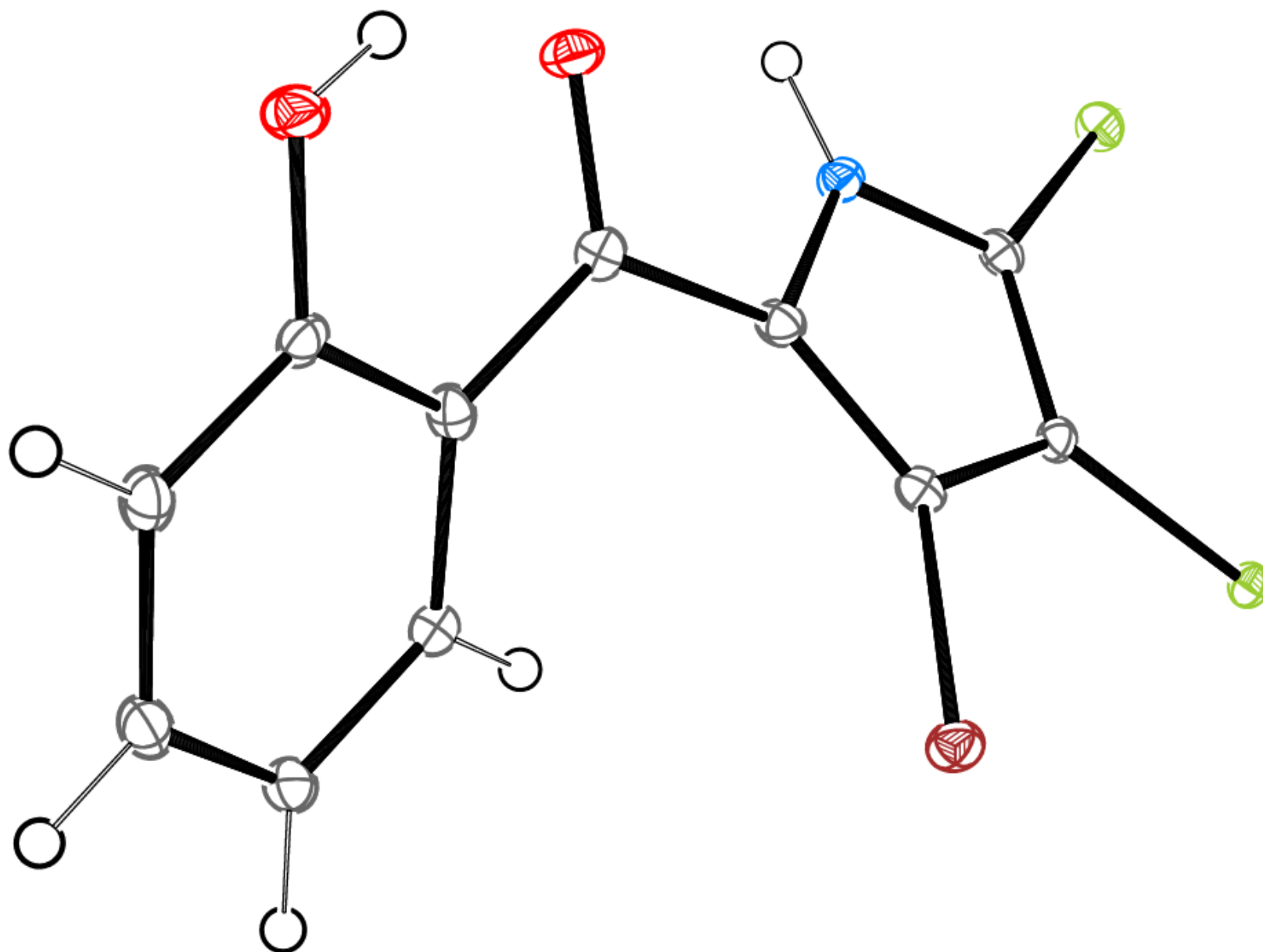


Figure S3. ORTEP of bipyrrole **31**

