

# Supporting Information

## A QM/MM Study of the Bergman Reaction of Dynemicin A in the Minor Groove of DNA

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## 1. Computational details

*Gas phase calculations.* The Bergman cyclization in both model and naturally occurring compounds is well described by density functional theory (DFT). DFT accounts for long-range correlation effects through the local exchange potential and, in an unspecified way, through the inherent self-interaction error.<sup>1-4</sup> The use of broken-symmetry unrestricted DFT (BS-UDFT) specifically incorporates long-range correlation into the calculation and thus the choice of the functional is dictated by the need to avoid double counting of the correlation effects.<sup>5,6</sup> The B3LYP functional<sup>7-11</sup> in connection with either the 3-21G<sup>12</sup> or 6-31G(d,p)<sup>13</sup> basis set satisfies this requirement and therefore these combinations were used for the optimization of all structures in the gas phase. The calculations in the gas phase were performed in Gaussian03.<sup>14</sup>

*System preparation, classical simulations.* There is no experimentally determined structure for dynemicin A complexed with DNA. Therefore, we constructed a duplex 10-mer B-DNA sequence with the same primary sequence – d(CTACTACTGG) – that has previously been used.<sup>15</sup> All fully classical calculations (including: hydration of the system, molecular mechanics (MM) minimizations and MD simulations) were carried out with the CHARMM program.<sup>16-18</sup> The DNA segment was minimized and the gas-phase optimized anionic dynemicin ligand (**2**) was docked into the minor groove of the segment using AutoDock<sup>19,20</sup> in the same manner as previously described for the neutral ligand.<sup>15</sup>

The system was neutralized by the addition of counterions ( $\text{Mg}^{2+}/\text{Na}^+$ ) around the **2**-DNA complex. One  $\text{Mg}^{2+}$  ion was placed at each end of the duplex, to satisfy the -2 charge of the two terminal residues.  $\text{Na}^+$  ions were then positioned opposite to each of the  $\text{PO}_4^-$  groups along the backbone (18 in total). Finally, the  $\text{Na}^+$  ion closest to the  $\text{COO}^-$

group of the ligand was converted into a  $\text{Mg}^{2+}$  ion, yielding a total system charge of 0. The neutral system thus obtained was solvated by a sphere of TIP3P waters<sup>21</sup> of radius 30 Å, centered about the geometric center of the bound ligand. All waters that overlapped<sup>22</sup> with the complex were deleted.

The system was minimized for 500 Steepest Descent steps, followed by 10,000 ABNR (Adopted Basis Newton Raphson) steps,<sup>17</sup> and subsequently equilibrated for 1ns at 310K using Langevin Dynamics with a 1 fs time-step. The SHAKE algorithm<sup>23</sup> was employed for all bonds involving hydrogen. A stochastic boundary potential<sup>24</sup> was used to maintain the structure of the water sphere during the MD simulation. In both the CHARMM minimization and subsequent equilibration the structure of **2** was kept fixed. The standard CHARMM force field<sup>25-28</sup> does not contain parameter or topology data for the dynamicin A ligand; therefore, the MM parameters and topology of dynamicin A were automatically generated with QUANTA98 using the CHARMM forcefield.<sup>29</sup> While these parameters are not strictly compatible with the academic version of CHARMM, the restraining of the ligand geometry ensures that only the van der Waals parameters and partial charges are important during the MM minimization and equilibration. To this end, the parameterization of the ligand was considered sufficient.

The RMSD for the DNA atoms of the final structure relative to the minimized DNA is 1.3 Å. In addition we observed that the H-bond network between the DNA bases was conserved, with only minor deviations for the terminal base pairs. The counterions remained associated to the charge groups to which they were assigned and the development of solvent shells around the charged species was evident. The reproduction of these qualitative features as well as the acceptable RMSD for the DNA structure

indicate that the final system is well equilibrated and therefore provides a reasonable starting point for the QM/MM optimization.

*QM/MM setup.* The QM region is electronically coupled to the MM region by including the point charges from the environment into the one electron QM Hamiltonian (see the main text for the definition of QM and MM regions). In an attempt to increase the efficiency of the calculations, a cutoff of 15 Å was initially applied for the inclusion of point charges. However, this approach led to a fluctuating number of point charges entering the Hamiltonian, which resulted in inconsistent absolute energies between the stationary points. Thus, for all results reported no QM/MM cutoff was applied.

The QM/MM geometry optimizations were performed with HDLC\_OPT – a linear scaling, microiterative algorithm that employs a set of hybrid delocalized coordinates – as implemented in CHEMSHELL.<sup>30-32</sup> The residues defined for this optimization procedure were taken as the standard residues defined for CHARMM (e.g. nucleic acids, waters, dynemicin).

In QM/MM optimizations of complex large systems, it is important to ensure the connectivity between the reactants, products, and the associated transition states (TS). To achieve this goal, we first located **TS(2-3)** and performed a frequency calculation on the QM region of the optimized structure, at the QM/MM level. The presence of a single imaginary frequency corresponding to the reaction coordinate confirmed that the correct TS had been found. The ligand was then shifted slightly towards the reactant (product) side, and the structure was then optimized to attain **2 (3)**. Starting from the optimized structure of **3-T**, the geometry was altered towards that of **TS(3-4)** and optimized to locate **TS(3-4)** whose correct nature was again verified by a frequency calculation.

Finally, the geometry was modified slightly towards **4** and then fully optimized to obtain the structure of the product. Using this procedure, we have established that a contiguous path connects the minima and transition states during the reactions studied.

*Approximating the singlet biradical.* The optimization of the singlet biradical was extremely time-consuming, due primarily to the necessity of using a quadratically convergent SCF routine. Therefore, we tested the validity of using the optimized structure of the triplet (**3-T**) to calculate the energy of the singlet (**3-S**).

**Table S1.** Comparison of geometries and relative energies for **3-S** and **3-T**.<sup>a</sup>

|            | $\Delta E$ | C23-C24 | C24-C25 | C25-C26 | C26-C27 | C27-C28 | C28-C23 |
|------------|------------|---------|---------|---------|---------|---------|---------|
| <b>3-S</b> | 0.0 (3.3)  | 1.377   | 1.373   | 1.429   | 1.373   | 1.376   | 1.430   |
| <b>3-T</b> | 2.8 (0.4)  | 1.383   | 1.381   | 1.414   | 1.381   | 1.385   | 1.414   |

<sup>a</sup>Energies in kcal/mol relative to the energy of the optimized singlet biradical; bond lengths in Å (see Scheme 1 of the paper for atom numbering). Structures optimized at the B3LYP/3-21G level. Values in parentheses correspond to a single point calculation of the structure using the alternative electronic state.

In Table S1, we compare the bond lengths of the optimized triplet and singlet biradical warheads, along with the relative energies of the structures. In addition, the destabilization resulting from using the geometry of one optimized electronic structure to calculate the other electronic state is also given. The most accurate singlet-triplet (S-T) splitting results from the energy difference of the structures ( $\Delta E(S-T) = 2.8$  kcal/mol) when optimized in the correct electronic states (i.e. **3-S** and **3-T**). However, the bond distances within the *p*-benzyne warhead are similar in the singlet and triplet optimized structures as has been discussed in the literature. There is a slight elongation ( $< 0.01$  Å) of

the bonds involving the radical carbon atoms (C24 and C27) for the **3-T** structure. Conversely, the remote C-C bonds are slightly shorter ( $\approx 0.015$  Å). These geometric changes are small, as reflected in the energy difference between the optimized singlet and the singlet at the optimized triplet geometry ( $\Delta E = 0.4$  kcal/mol). The alternative case of using the triplet at the optimized singlet geometry causes an increase in the S-T splitting by a similar amount ( $\Delta\Delta E = 0.5$  kcal/mol) due to the further destabilization of the triplet through using a non-optimized geometry. These minor energy changes indicate that the use of the optimized triplet geometry in determining the S-T splitting is a justified approximation. As such, during the QM/MM optimization of the Bergman cyclization reaction the singlet biradical system was calculated at the optimized triplet geometry.

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## 2. Gas phase calculations with larger basis sets

For the purpose of checking whether a larger basis set or the inclusion of diffuse functions would significantly affect the results of the anionic species we carried out B3LYP calculations with the 6-31G(d,p) and 6-31+G(d,p) basis sets. Results are summarized and compared with B3LYP/3-21G energies in Table S2.

**Table S2.** Effect of basis set size on the energy profile for anionic dynemicin A.<sup>a</sup>

|                | $\Delta E$ |            |             | $\mu^b$ |            |             |
|----------------|------------|------------|-------------|---------|------------|-------------|
|                | 3-21G      | 6-31G(d,p) | 6-31+G(d,p) | 3-21G   | 6-31G(d,p) | 6-31+G(d,p) |
| <b>2</b>       | 0.0        | 0.0        | 0.0         | 15.0    | 16.6       | 17.1        |
| <b>TS(2-3)</b> | 20.4       | 20.4       | 20.5        | 14.7    | 16.6       | 17.1        |
| <b>3-S</b>     | -0.8       | -3.2       | -1.9        | 15.0    | 16.6       | 17.2        |
| <b>3-T</b>     | 2.0        | -1.1       | 0.3         | 15.0    | 16.6       | 17.2        |
| <b>TS(3-4)</b> | 22.1       | 24.5       | 24.3        | 14.8    | 16.4       | 17.0        |
| <b>4</b>       | -9.1       | -6.7       | -7.0        | 14.9    | 16.5       | 17.1        |

<sup>a</sup>Energies in kcal/mol, dipole moments in Debye. The 6-31+G(d,p) calculations were performed as single point calculations on the 6-31G(d,p) optimized structures.

<sup>b</sup>The origin for the dipole moments is taken as the center of nuclear charge.

The calculated activation barriers are largely unchanged when using the larger basis sets. However, the exothermicity of the singlet formation is predicted to be slightly larger with the 6-31G(d,p) basis set ( $\Delta\Delta E = -2.3$  kcal/mol). The second TS, leading to the acyclic enediyne isomer **4** is calculated to be higher by a corresponding amount and the formation of the product is less exothermic. The addition of diffuse functions to the larger basis set has only a minor effect on the relative energies and generally tends to bring the

results into closer agreement with the 3-21G results. As all of the differences in the relative energies are in the order of 2-3 kcal/mol, we consider this as a validation of the usefulness of the 3-21G basis set, in conjunction with the B3LYP functional for studying the Bergman cyclization of the anionic form of dynemicin A.

### 3. Analysis of interaction energies

In this section the influence of the environment on the reaction sequence **2** → **4** is discussed by a decomposition of calculated interaction energies into individual components.

*3.1 Bulk environment interactions.* In Table S3 the contributions to the relative MM energy differences, between the optimized stationary points, is given with respect to the three segments – water, DNA and counterions – which comprise the environment. In addition, the MM component of the interaction energy between the ligand (QM) and each of these segments is listed. This MM component represents only the van der Waals interactions between dynemicin A and the environment, as the electrostatic interaction energy is contained within the QM energy term and will be considered subsequently.

**Table S3.** Variations in the MM interaction energies.<sup>a</sup>

|              | $\Delta E$                |       |      |      | $\Delta E$                |       |      |      |
|--------------|---------------------------|-------|------|------|---------------------------|-------|------|------|
|              | Ligand                    | Water | DNA  | Ions | Ligand                    | Water | DNA  | Ions |
|              | <b>2</b> → <b>TS(2-3)</b> |       |      |      | <b>TS(2-3)</b> → <b>3</b> |       |      |      |
| <b>Water</b> | -0.2                      | 2.3   |      |      | -0.7                      | -2.7  |      |      |
| <b>DNA</b>   | -0.1                      | -2.7  | 0.4  |      | -0.3                      | 1.3   | 0.5  |      |
| <b>Ions</b>  | 0.0                       | 0.8   | 0.5  | 0.6  | 0.0                       | -0.7  | -1.5 | 0.3  |
|              | <b>3</b> → <b>TS(3-4)</b> |       |      |      | <b>TS(3-4)</b> → <b>4</b> |       |      |      |
| <b>Water</b> | 0.2                       | -3.2  |      |      | 1.9                       | 1.0   |      |      |
| <b>DNA</b>   | -0.2                      | 3.1   | 0.6  |      | -0.5                      | -0.1  | 1.6  |      |
| <b>Ions</b>  | 0.0                       | -1.1  | -0.1 | 0.3  | 0.0                       | 0.6   | -1.3 | -0.1 |

<sup>a</sup>Energies are reported in kcal/mol. Structures optimized at the B3LYP/3-21G//CHARMM level of theory.

The data presented in Table S3 shows that the interaction of the ligand with the environment is relatively consistent (variations of less than 1 kcal/mol) in all cases, except for **4**. In the reaction step moving from **TS(3-4)** to **4** there is a change in the van der Waals interactions between the ligand and the water, which leads to a 1.9 kcal/mol destabilization. However, this destabilization is partially offset by more favorable interactions between the ligand and the DNA ( $\Delta E = -0.5$  kcal/mol).

Differences in the solvent sphere account for the largest variations in the MM energies. However, changes in the solvent sphere are generally compensated such that the system as a whole remains stable. For example, in the formation of **TS(2-3)**, the re-orientation of water molecules leads to less favorable water-water interactions ( $\Delta E = 2.3$  kcal/mol), while the water-DNA interaction becomes more stabilizing ( $\Delta E = -2.7$  kcal/mol). The acyclic enediyne **4** is an exception since the change in the water configuration results in a net destabilization of the system by 3.4 kcal/mol (relative to **TS(3-4)**).

*3.2 Electrostatic interactions between selected residues and the ligand.* The electrostatic effect of individual residues on the reaction profiles can be assessed by removing the partial charges assigned to a residue in the MM region and then recalculating the QM/MM energy. The resulting change in the QM energy is a measure of the electrostatic influence of the particular residue on the QM region. The changes caused by the charge deletion influence only the QM component of the reaction energy (column 5 of Table 2), as the van der Waals interactions, and thus the MM component, are unaffected by this procedure.

Table S4 lists the total electrostatic interaction energies of the ligand with strands 1 and 2 of the DNA and with the counterions, as obtained from the charge deletion analysis, and their variation during the reaction. Detailed individual interaction energies are given in Tables S5 and S6 of this summary. It is obvious from Table S4 that the total electrostatic interaction of the anionic ligand is strongly repulsive with DNA and strongly attractive with the counterions, as expected from the formal charges. Proceeding along the reaction path, these electrostatic interactions become progressively less favorable overall (by 1.0-3.9 kcal/mol), mostly due to the increased repulsive interactions with DNA strand 1, which are only partially compensated by the other interactions. One should emphasize however, that these variations remain minor compared with the total interaction energies.

**Table S4.** Electrostatic interactions between the proximal non-water residues with **2** ( $\Delta E$ ) and their variation along the reaction path ( $\Delta\Delta E$ , relative to **2**).<sup>a</sup>

|             | $\Delta E$ | $\Delta\Delta E$ |          |                |          |
|-------------|------------|------------------|----------|----------------|----------|
|             | <b>2</b>   | <b>TS(2-3)</b>   | <b>3</b> | <b>TS(3-4)</b> | <b>4</b> |
| Strand 1    | 141.1      | 3.8              | 2.3      | 3.8            | 3.3      |
| Strand 2    | 158.2      | -1.6             | 0.6      | 0.6            | -0.6     |
| Counterions | -196.2     | -1.2             | -1.2     | -1.6           | 1.2      |
| Sum         | 103.1      | 1.0              | 1.7      | 2.8            | 3.9      |

<sup>a</sup>The individual effects from the proximal DNA bases considered were summed into two groups based on the strand. The interactions of the individual counterions considered have been summed together. For a complete list of the individual interactions see section 3.3. Relative energies are reported in kcal/mol.

It should be pointed out that the individual electrostatic interaction energies between the ligand and the environmental residues do not sum up to the differences  $E(\text{QM}) - E(\text{QM}(\text{gas})//\text{QM}/\text{MM})$  from Table 2 of the main paper, for two reasons: First we considered only a selected set of residues and neglected the more remote ones. Second, the charge deletion analysis assumes that the individual interactions are additive, which does not hold strictly because of cooperative effects. The purpose of the present analysis is not to reproduce  $E(\text{QM}) - E(\text{QM}(\text{gas})//\text{QM}/\text{MM})$ , but to check for specific environmental effects on the reaction profile. We find as our main result that there are no pronounced qualitative effects of this kind.

*3.3 DNA and counterions energy decomposition.* The counterions and DNA residues within 10 Å of the geometric center of **TS(2-3)** are listed in Tables S5 and S6, respectively, which also provide their electrostatic interaction energies with the ligand in the reactant and their variation during the reaction. Consistent with the anionic nature of the ligand, each of the counter-cations has a stabilizing interaction, while the anionic DNA residues are all strongly destabilizing (17-43 kcal/mol, see Table S6).

**Table S5.** Electrostatic interaction energies of the surrounding counterions with **2** ( $\Delta E$ ) and their variation along the reaction path ( $\Delta\Delta E$ ).<sup>a</sup>

| Residue | $\Delta E / R$ | $\Delta\Delta E / R$ |            |                |           |
|---------|----------------|----------------------|------------|----------------|-----------|
|         | <b>2</b>       | <b>TS(2-3)</b>       | <b>3</b>   | <b>TS(3-4)</b> | <b>4</b>  |
| Na6     | -27.7/11.17    | -1.0/11.07           | 0.0/11.08  | -0.6/11.05     | 0.1/11.02 |
| Na7     | -24.9/12.23    | -1.3/12.19           | -0.5/12.16 | -0.9/12.17     | 0.3/13.14 |
| Na8     | -26.1/14.44    | 0.2/14.47            | -0.4/14.46 | 0.1/14.47      | 0.0/14.58 |
| Na16    | -37.5/9.21     | 0.4/9.28             | -0.5/9.25  | -0.2/9.25      | 0.3/9.38  |
| Mg17    | -80.0/8.00     | 0.5/8.01             | 0.2/8.03   | 0.0/8.01       | 0.5/8.04  |

<sup>a</sup> Energies in kcal/mol. R is the distance between the ion and the C30 atom of the ligand in Å (see Scheme 1 of the paper for atom numbering).

Going from **2** to **TS(2-3)** the electrostatic interactions with the proximal counterions have a mixed effect. Two sodium ions are moderately stabilizing, relative to **2** ( $\Delta\Delta E(\text{Na6,Na7}) = -2.3$  kcal/mol); however, the remaining three cations have a combined destabilizing effect of 1.1 kcal/mol. For the intermediate (**3**) the interactions with the counterions are again more favorable relative to **2** ( $\Delta\Delta E = -1.2$  kcal/mol), with only Mg17 being slightly destabilizing. The electrostatic stabilization offered by the counterions is maintained in the formation of **TS(3-4)** ( $\Delta\Delta E = -1.6$  kcal/mol), but is lost in the final product ( $\Delta\Delta E = 0.6$  kcal/mol).

The Mg17 ion clearly has the largest effect on the QM energy; however, the position of this ion is essentially constant and thus the relative energy differences between systems are small. The Na8 counterion is actually the closest in terms of the minimum distance to any ligand atom, although it is further from the C30 atom.

For the direct interaction of the counterion with the negative charge on the carboxylate group of the ligand, one expects a correlation between the changes in the distance

between the two atoms and the changes in the relative energies. This is seen for Mg17, which is closest to C30 atom: as the counterion distance to C30 increases, the relative stabilization caused by the counterion decreases. However, for the sodium counterions that are more remote, there is generally no such clear correlation (except for specific cases, see e.g. Na16, **3** vs. **4**, Table S3). One should keep in mind, of course, that the counterions also polarize the electronic structure of the ligand and that this polarization may vary in a non-linear fashion because it depends both on the distance and the screening between the ligand and the counterion.

In an overall assessment, the contributions of the individual counterions to the changes in the reaction energetics are always quite small, especially when compared with the much larger total interaction energy.

**Table S6.** Electrostatic interaction energies of the surrounding DNA bases with **2** ( $\Delta E$ ) and their variation along the reaction path ( $\Delta\Delta E$ ). Energies in kcal/mol.

| Residue  | $\Delta E$ | $\Delta\Delta E$ |          |                |          |
|----------|------------|------------------|----------|----------------|----------|
|          | <b>2</b>   | <b>TS(2-3)</b>   | <b>3</b> | <b>TS(3-4)</b> | <b>4</b> |
| 1:THY5   | 24.5       | 0.1              | 0.0      | 0.2            | 0.2      |
| 1:ADE6   | 34.4       | 0.7              | 0.1      | 1.0            | 0.4      |
| 1:CYT7   | 34.0       | 2.0              | 0.8      | 1.9            | 1.3      |
| 1:THY8   | 26.3       | 1.1              | 1.2      | 0.7            | 1.2      |
| 1:GUA9   | 21.9       | -0.1             | 0.2      | 0.0            | 0.2      |
| 1: Total |            | 3.8              | 2.2      | 3.8            | 3.3      |
| 2:GUA4   | 17.7       | -0.1             | 0.0      | 0.0            | -0.1     |
| 2:THY5   | 26.7       | -0.4             | 0.2      | 0.0            | -0.3     |
| 2:ADE6   | 37.3       | -0.6             | 0.0      | -0.1           | -0.5     |
| 2:GUA7   | 43.8       | -0.4             | 0.5      | 0.6            | 0.2      |
| 2:THY8   | 32.7       | -0.1             | -0.1     | 0.1            | 0.1      |
| 2: Total |            | -1.6             | 0.6      | 0.6            | -0.6     |

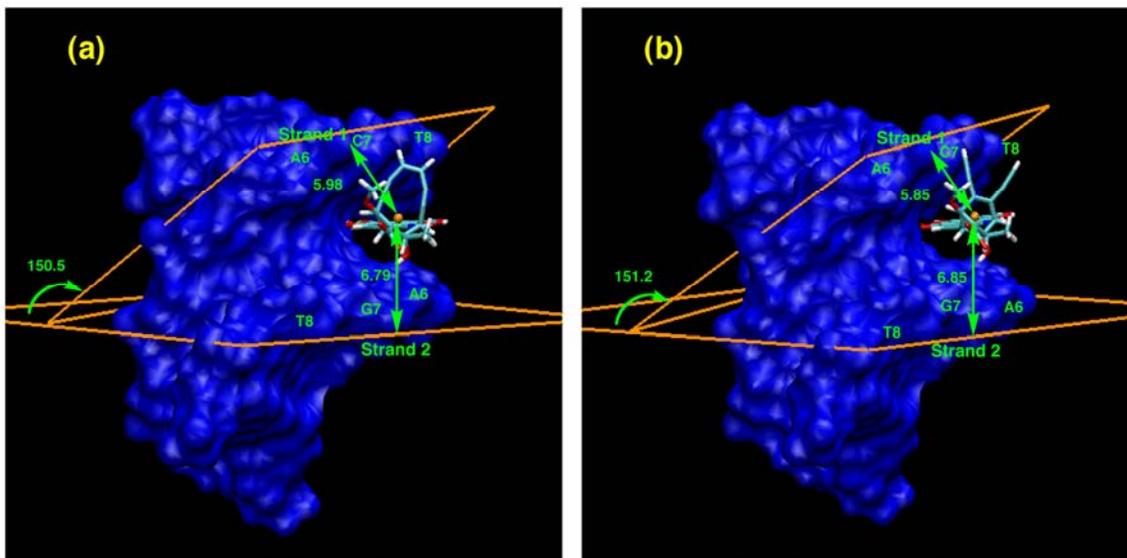
We now consider DNA-ligand interactions (Table S6). Following the reaction path from **2** to **TS(2-3)** leads to less favorable interactions ( $\Delta\Delta E = 3.8$  kcal/mol) between the ligand and strand 1 of the DNA (Table S4). However, these are partially offset by the more favorable interaction of strand 2 with the ligand ( $\Delta\Delta E = -1.7$  kcal/mol). The comparison of electrostatic interactions in **3** and **2** reveals that the interactions with the first DNA strand are also destabilizing ( $\Delta\Delta E = 2.2$  kcal/mol). Moreover, the stabilization of the radical intermediate state resulting from interactions is not as prevalent as in the

case of **TS(2-3)**, with the second DNA strand also causing an increase in the relative energy by 0.6 kcal/mol.

Summing the individual interactions of the residues listed in Table S5 and Table S6 for the formation of **TS(3-4)** and **4**, relative to **2**, leads to a destabilization of 0.8 and 3.9 kcal/mol, respectively. The larger destabilization of **4** arises predominantly from less favorable interactions of the ligand with strand 1 of the DNA helix. These changes can be understood by the change in the ligand position that results from the open enediyne structure.

In Figure S1, two planes are constructed to describe the minor groove of DNA. The bottom plane (Figure S1) is defined by the three P atoms of residues THY8, GUA7 and ADE6, which form part of the second strand and lie underneath the ligand. The upper plane is defined by the three P atoms of residues CYT7, THY8 and GUA9, which form part of the first strand, located above the ligand. The angle between the two planes, as indicated in Figure S1, is 150.5°. The formation of **4** results in a slight increase of this angle to 151.2° due to the interaction of the ligand with the backbone atoms of the upper strand. The formation of the more open head unit, present in **4**, results in the wedging of the head unit between two PO<sub>4</sub><sup>-</sup> groups and brings the methyl group of the methoxy substituent close to the PO<sub>4</sub><sup>-</sup> group (C7) of the backbone.

**Figure S1.** Change in the position of the ligand [(a) **2**; (b) **4**] relative to the DNA strands. Distances in Å, angles in °.



The change in the distance between the geometric center (orange ball in Figure S1) of the ligand and the upper and lower planes indicates the upward movement of the ligand. The distance to the upper plane decreases from 5.98 Å to 5.85 Å ( $\Delta R = -0.13$  Å) during formation of **3**, whereas the distance to the lower plane increases from 6.79 Å to 6.85 Å ( $\Delta R = 0.06$  Å). Clearly, the open structure of **4** results in a tightly wedged complex, with the ligand being drawn into the DNA minor groove.

The C atoms of the anthraquinone unit of the ligand define an additional plane, which can be used to determine the angle between the ligand and the lower plane of DNA. The angle between this plane (not shown) and the bottom plane is  $3.8^\circ$ , indicating that **1** is essentially parallel to the backbone atoms of the DNA second strand. For the product (**3**) this orientation is essentially conserved with a minor decrease in the angle between the bottom plane and the anthraquinone plane to  $3.2^\circ$ .

*3.4 Water energy decomposition.* The charge deletion analysis was also applied to the electrostatic interactions between the ligand and the water molecules in the primary

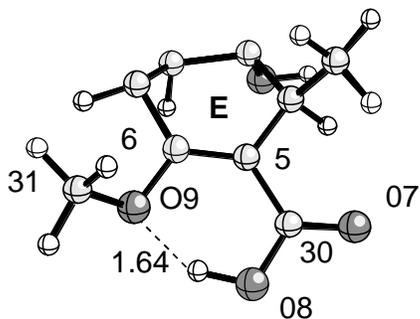
solvent shell (i.e., within 4 Å of the ligand). Table S7 addresses these water molecules, which form H-bonds with the ligand at the specified oxygen acceptor atoms. It lists the variations along the reaction path (relative to **2**) of the associated H-bond energies ( $\Delta\Delta E$  as obtained from charge deletion analysis, see main paper) and H-bond lengths ( $\Delta R$ ).

**Table S7.** The effect of specific H-bonds between the ligand and neighboring water molecules (see text).  $\Delta\Delta E$  in kcal/mol,  $\Delta R$  in Å, both relative to **2**. For atom numbering, see Scheme 1 in the paper.

|           | $\Delta\Delta E / \Delta R$ |            |                |            |
|-----------|-----------------------------|------------|----------------|------------|
| O(acc)    | <b>TS(2-3)</b>              | <b>3</b>   | <b>TS(3-4)</b> | <b>4</b>   |
| <b>O1</b> | -0.2/-0.03                  | -0.5/-0.06 | -0.5/-0.07     | -0.3/-0.05 |
| <b>O4</b> | -0.1/0.0                    | -0.2/0.00  | -0.1/0.00      | 0.0/0.00   |
| <b>O5</b> | -0.2/-0.01                  | -0.4/-0.02 | -0.3/-0.03     | -1.4/-0.09 |
| <b>O7</b> | -0.2/0.0                    | -0.2/0.00  | -0.2/0.00      | -0.2/-0.01 |
| <b>O7</b> | 0.2/0.02                    | 0.0/0.01   | 0.0/0.00       | 0.3/0.02   |
| <b>O8</b> | 0.2/0.01                    | 0.4/0.01   | 0.7/0.02       | 0.7/0.02   |
| <b>O8</b> | -0.2/-0.01                  | -0.1/-0.01 | 0.0/-0.01      | 0.0/-0.02  |
| <b>O9</b> | 0.2/0.01                    | 0.9/0.03   | 0.6/0.02       | -0.8/0.02  |

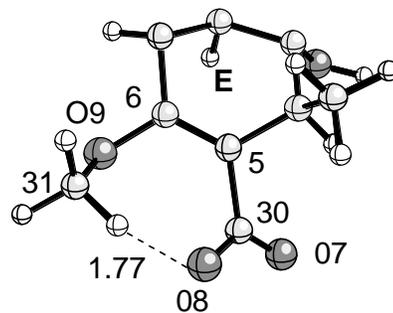
When  $\Delta R$  is significant ( $>0.01$  Å), the changes  $\Delta\Delta E$  correlate well with  $\Delta R$ . However, generally speaking, the energy changes along the reaction path are rather small and tend to compensate each other. For example, after summing the individual changes, the total effect for **TS(2-3)** is a decrease in the activation barrier by 0.3 kcal/mol.

As pointed out in the main text, H-bonding does not change in the rearrangement of **2** to **4**. This is documented in Figure S2, which compares H-bonding for neutral and anionic dynemicin structures along the reaction path.

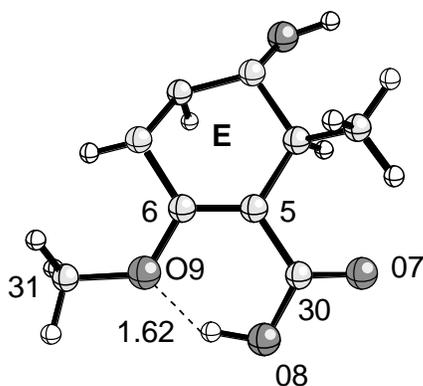


$31-O9-6-5 = 133.8$   
 $O9-6-5-30 = -2.7$   
 $6-5-30-07 = -176.1$   
 $6-5-30-08 = 4.1$   
 $5-30-08-H = 0.1$

2

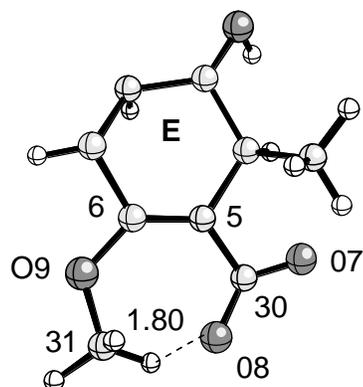


$31-O9-6-5 = 71.6$   
 $O9-6-5-30 = -2.7$   
 $6-5-30-07 = 132.3$   
 $6-5-30-08 = -47.5$   
 $5-30-08-H = 25.8$

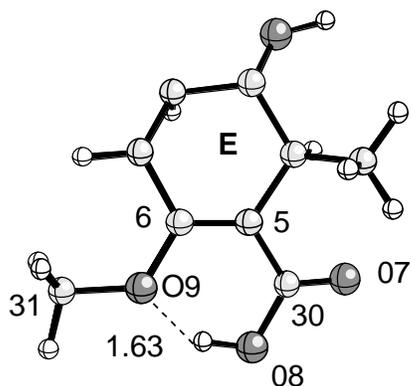


$31-O9-6-5 = -171.5$   
 $O9-6-5-30 = 8.8$   
 $6-5-30-07 = 170.6$   
 $6-5-30-08 = -8.6$   
 $5-30-08-H = 1.1$

TS(2-3)

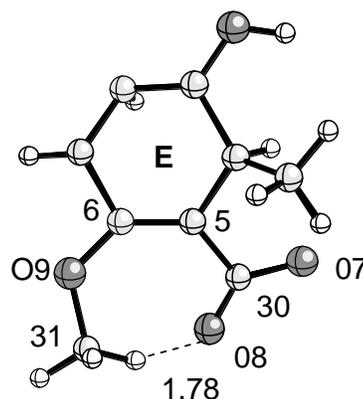


$31-O9-6-5 = 39.7$   
 $O9-6-5-30 = 14.6$   
 $6-5-30-07 = 167.9$   
 $6-5-30-08 = -9.7$   
 $5-30-08-H = -27.8$

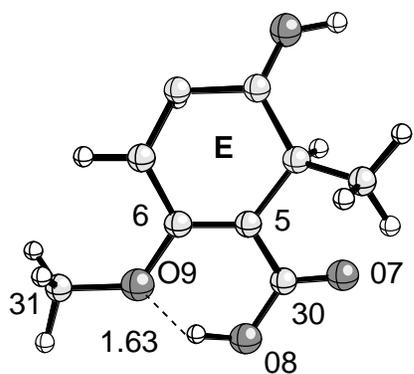


$31-O9-6-5 = -172.2$   
 $O9-6-5-30 = 7.5$   
 $6-5-30-07 = 172.6$   
 $6-5-30-08 = -6.8$   
 $5-30-08-H = 0.7$

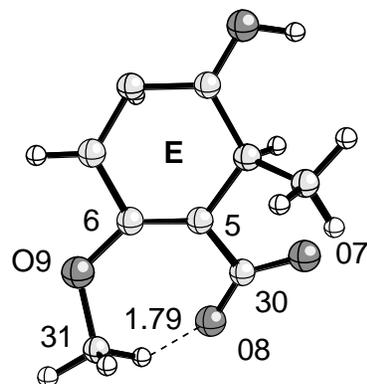
3



$31-O9-6-5 = 30.7$   
 $O9-6-5-30 = 17.4$   
 $6-5-30-07 = 174.9$   
 $6-5-30-08 = -2.4$   
 $5-30-08-H = -33.9$

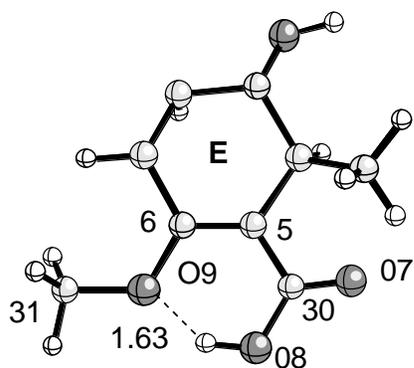


TS(3-4)

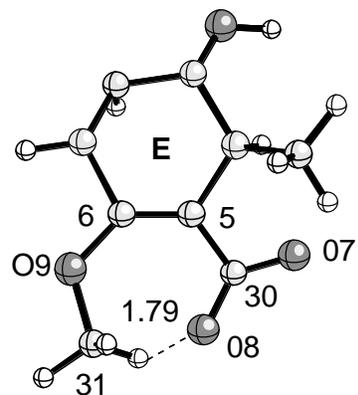


$$\begin{aligned} 31-O9-6-5 &= -178.2 \\ O9-6-5-30 &= 5.9 \\ 6-5-30-O7 &= 173.9 \\ 6-5-30-O8 &= -5.5 \\ 5-30-O8-H &= 0.7 \end{aligned}$$

$$\begin{aligned} 31-O9-6-5 &= 31.8 \\ O9-6-5-30 &= 15.8 \\ 6-5-30-O7 &= 175.6 \\ 6-5-30-O8 &= -2.1 \\ 5-30-O8-H &= -33.3 \end{aligned}$$



4



$$\begin{aligned} 31-O9-6-5 &= -161.7 \\ O9-6-5-30 &= 7.2 \\ 6-5-30-O7 &= 173.0 \\ 6-5-30-O8 &= -6.5 \\ 5-30-O8-H &= 0.3 \end{aligned}$$

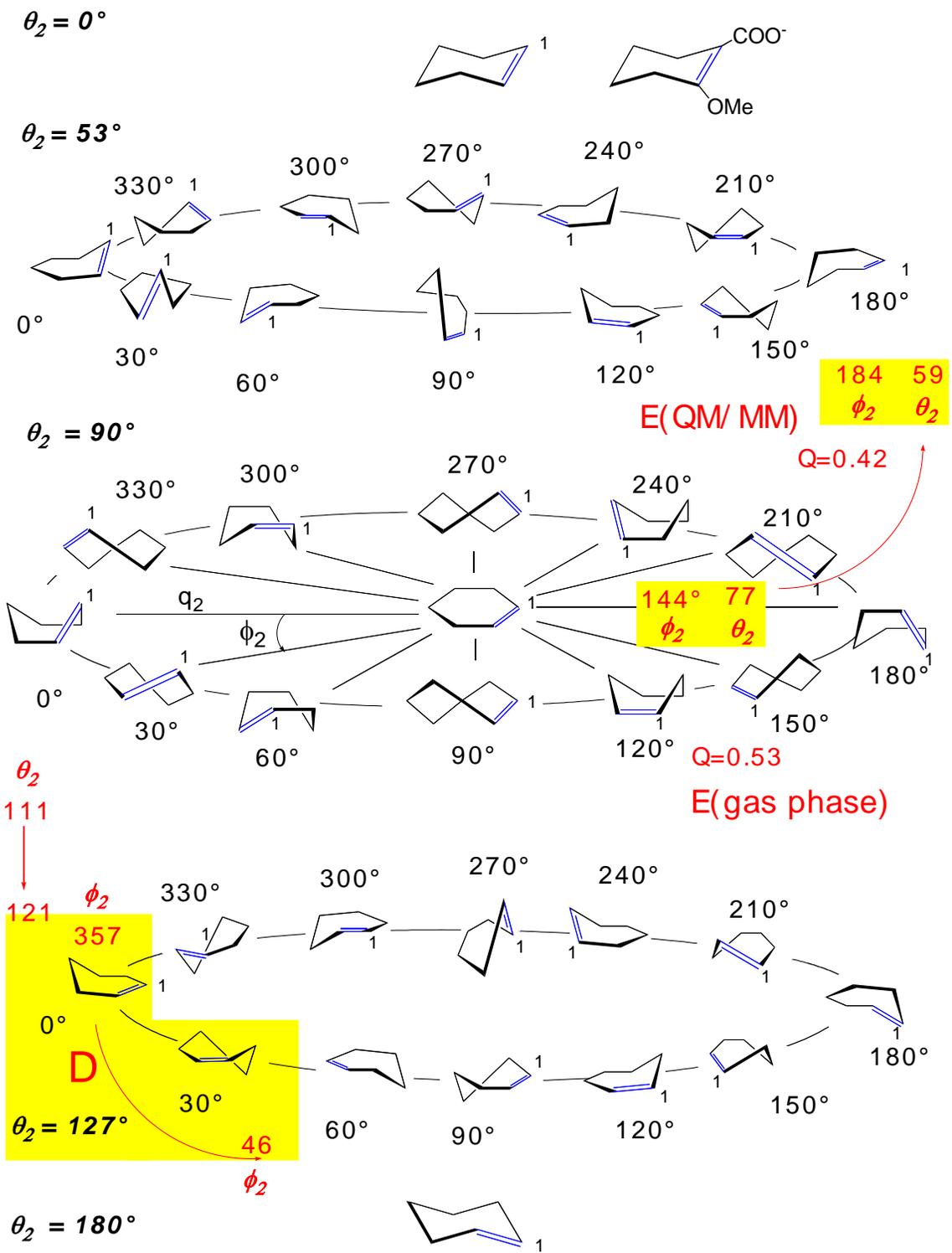
$$\begin{aligned} 31-O9-6-5 &= 34.9 \\ O9-6-5-30 &= 13.4 \\ 6-5-30-O7 &= 175.5 \\ 6-5-30-O8 &= -2.3 \\ 5-30-O8-H &= -32.8 \end{aligned}$$

**Figure S2.** Calculated geometrical parameters for the H-bonds involving the carboxyl and methoxy groups located at ring E.

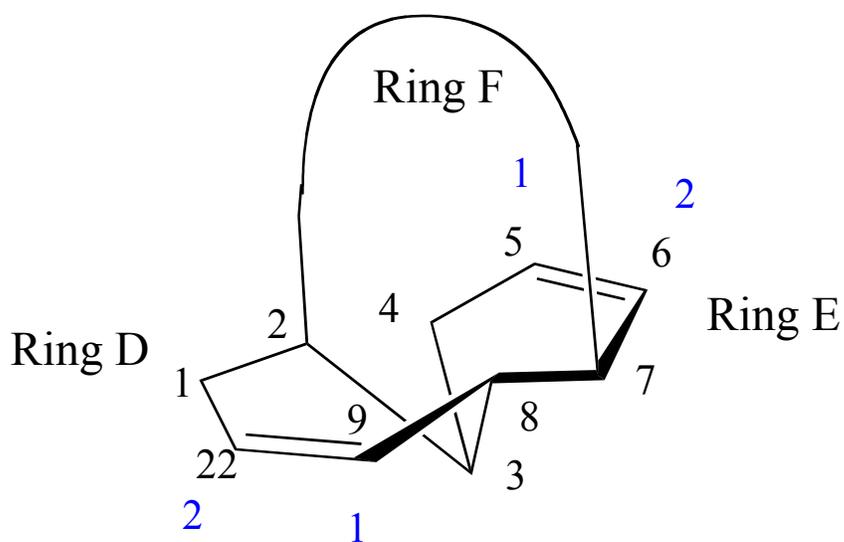
#### **4. Conformational analysis of rings E and D in dynemicin**

Ring F of dynemicin (**2**) is deformed by steric interactions with the back of the minor groove upon an edge-wise insertion into the minor groove between strands 1 and 2, which implies a change of the puckering of rings E and D. In the main paper it is discussed that in the course of the Bergman cyclization ring E converts from a twist-boat form to a half-boat form with lower degree of puckering to give the triple bonds a chance of approaching each other. This is indicated in Figure S3, which schematically describes the deformation of ring E on the conformational globe of a cyclohexene ring when **2** is transferred from the gas phase into the minor groove.

In Figure S4, the numbering system used in Figure S3 is explained by relating atom numbers of Scheme 1 (see main text) to atom numbers used in Figure S3 (clockwise counting for each ring) and presenting the conformations adopted by rings E and D in the minor groove.



**Figure S3.** Conformational globe of cyclohexene spanned by puckering amplitudes  $q_2$  and  $q_3$  and pseudorotational phase angle  $\phi_2$  (cylindrical coordinate system). Alternatively the spherical coordinate system including the total puckering amplitude  $Q$ , the pseudorotational phase angle  $\phi_2$ , and the angle  $\theta_2$  can be used. Cyclohexene is the appropriate model for rings E and D of dynemicin **2**. The deformation of these rings when **2** is transferred from the gas phase into the minor groove is indicated.



**Figure S4.** Schematic representation of rings D, E, and F in the conformations adopted in the minor groove. Atom C5 corresponds to 1 in Figure S3 for ring E, atom C9 to 1 in Figure S3 for ring D.

## 5. CHARMM topology file for dynemicin A.

```
RESI DYNA      -1.000
ATOM N1  NP   -0.400
ATOM C1  CT    0.050
ATOM C2  CT    0.250
ATOM C3  CT   -0.050
ATOM C4  CUA1  0.000
ATOM C5  C    0.150
ATOM C6  CT   -0.050
ATOM C7  CT   -0.050
ATOM C21 C6R   0.050
ATOM C22 CUY1  0.000
ATOM C23 CUY1  0.000
ATOM C24 CUA1 -0.100
ATOM C25 CUA1 -0.100
ATOM C26 CUY1  0.000
ATOM C27 CUY1  0.000
ATOM C28 CT   -0.150
ATOM C29 C    0.200
ATOM C30 CT    0.030
ATOM O1  OT   -0.650
ATOM O7  OC   -0.600
ATOM O8  OC   -0.600
ATOM O9  OE   -0.330
ATOM H4  H    0.250
ATOM H5  HA    0.050
ATOM H6  HA    0.050
ATOM H7  HA    0.050
ATOM H11 HA    0.100
ATOM H12 HA    0.100
ATOM H13 HA    0.050
ATOM H14 HA    0.050
ATOM H15 HA    0.050
ATOM H16 HA    0.050
ATOM H17 HA    0.050
ATOM H18 HA    0.050
ATOM H19 HA    0.050
ATOM H20 HO    0.400
GROUP !Anthraquinone moiety
ATOM C8  C6R   0.000
ATOM C9  C6R  -0.130
ATOM C10 C6R   0.250
ATOM C11 C6R   0.000
```

|          |     |        |
|----------|-----|--------|
| ATOM C12 | C   | 0.550  |
| ATOM C13 | C6R | 0.000  |
| ATOM C14 | C6R | 0.250  |
| ATOM C15 | C6R | -0.130 |
| ATOM C16 | C6R | -0.130 |
| ATOM C17 | C6R | 0.250  |
| ATOM C18 | C6R | 0.000  |
| ATOM C19 | C   | 0.550  |
| ATOM C20 | C6R | 0.000  |
| ATOM O2  | OT  | -0.650 |
| ATOM O3  | O   | -0.550 |
| ATOM O4  | OT  | -0.650 |
| ATOM O5  | OT  | -0.650 |
| ATOM O6  | O   | -0.550 |
| ATOM H1  | HO  | 0.400  |
| ATOM H2  | HO  | 0.400  |
| ATOM H3  | HO  | 0.400  |
| ATOM H8  | HA  | 0.130  |
| ATOM H9  | HA  | 0.130  |
| ATOM H10 | HA  | 0.130  |
| BOND N1  | C1  |        |
| BOND N1  | C21 |        |
| BOND N1  | H4  |        |
| BOND C1  | C2  |        |
| BOND C1  | C22 |        |
| BOND C1  | H5  |        |
| BOND C2  | C3  |        |
| BOND C2  | C7  |        |
| BOND C2  | O1  |        |
| BOND C3  | C28 |        |
| BOND C3  | H6  |        |
| BOND C3  | C4  |        |
| BOND C4  | C5  |        |
| BOND C4  | C29 |        |
| BOND C5  | C6  |        |
| BOND C5  | O9  |        |
| BOND C6  | C7  |        |
| BOND C6  | C27 |        |
| BOND C6  | H7  |        |
| BOND C7  | C8  |        |
| BOND C7  | H19 |        |
| BOND C8  | C9  |        |
| BOND C8  | C21 |        |
| BOND C9  | C10 |        |
| BOND C9  | H8  |        |
| BOND C10 | C11 |        |

BOND C10 O2  
BOND C11 C12  
BOND C11 C20  
BOND C12 C13  
BOND C12 O3  
BOND C13 C14  
BOND C13 C18  
BOND C14 C15  
BOND C14 O4  
BOND C15 C16  
BOND C15 H9  
BOND C16 C17  
BOND C16 H10  
BOND C17 C18  
BOND C17 O5  
BOND C18 C19  
BOND C19 C20  
BOND C19 O6  
BOND C20 C21  
BOND C22 C23  
BOND C23 C24  
BOND C24 C25  
BOND C24 H11  
BOND C25 C26  
BOND C25 H12  
BOND C26 C27  
BOND C28 H13  
BOND C28 H14  
BOND C28 H15  
BOND C29 O7  
BOND C29 O8  
BOND C30 O9  
BOND C30 H16  
BOND C30 H17  
BOND C30 H18  
BOND H1 O2  
BOND H2 O4  
BOND H3 O5  
BOND O1 H20

## 6. List of active atoms for the optimized structures.

In this section we list the numbers for the active atoms during the QM/MM optimization of each structure discussed in the paper. The atom numbers are consistent with those in the analogous structure contained in the PDB file. Throughout the course of the reaction the active atom list remained the same.

### Active atom list for RRR – 2853 atoms.

|     |     |     |     |     |     |     |     |     |     |     |     |     |     |     |     |     |    |    |
|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|----|----|
| 1   | 2   | 3   | 4   | 5   | 6   | 7   | 8   | 9   | 10  | 11  | 12  | 13  | 14  | 15  | 16  | 17  | 18 | 19 |
| 20  | 21  | 22  | 23  | 24  | 25  | 26  | 27  | 28  | 29  | 30  | 31  | 32  | 33  | 34  | 35  | 36  | 37 |    |
| 38  | 39  | 40  | 41  | 42  | 43  | 44  | 45  | 46  | 47  | 48  | 49  | 50  | 51  | 52  | 53  | 54  | 55 |    |
| 56  | 57  | 58  | 59  | 60  | 61  | 62  | 63  | 64  | 65  | 66  | 67  | 68  | 69  | 70  | 71  | 72  | 73 |    |
| 74  | 75  | 76  | 77  | 78  | 79  | 80  | 81  | 82  | 83  | 84  | 85  | 86  | 87  | 88  | 89  | 90  | 91 |    |
| 92  | 93  | 94  | 95  | 96  | 97  | 98  | 99  | 100 | 101 | 102 | 103 | 104 | 105 | 106 | 107 | 108 |    |    |
| 109 | 110 | 111 | 112 | 113 | 114 | 115 | 116 | 117 | 118 | 119 | 120 | 121 | 122 | 123 | 124 |     |    |    |
| 125 | 126 | 127 | 128 | 129 | 130 | 131 | 132 | 133 | 134 | 135 | 136 | 137 | 138 | 139 | 140 |     |    |    |
| 141 | 142 | 143 | 144 | 145 | 146 | 147 | 148 | 149 | 150 | 151 | 152 | 153 | 154 | 155 | 156 |     |    |    |
| 157 | 158 | 159 | 160 | 161 | 162 | 163 | 164 | 165 | 166 | 167 | 168 | 169 | 170 | 171 | 172 |     |    |    |
| 173 | 174 | 175 | 176 | 177 | 178 | 179 | 180 | 181 | 182 | 183 | 184 | 185 | 186 | 187 | 188 |     |    |    |
| 189 | 190 | 191 | 192 | 193 | 194 | 195 | 196 | 197 | 198 | 199 | 200 | 201 | 202 | 203 | 204 |     |    |    |
| 205 | 206 | 207 | 208 | 209 | 210 | 211 | 212 | 213 | 214 | 215 | 216 | 217 | 218 | 219 | 220 |     |    |    |
| 221 | 222 | 223 | 224 | 225 | 226 | 227 | 228 | 229 | 230 | 231 | 232 | 233 | 234 | 235 | 236 |     |    |    |
| 237 | 238 | 239 | 240 | 241 | 242 | 243 | 244 | 245 | 246 | 247 | 248 | 249 | 250 | 251 | 252 |     |    |    |
| 253 | 254 | 255 | 256 | 257 | 258 | 259 | 260 | 261 | 262 | 263 | 264 | 265 | 266 | 267 | 268 |     |    |    |
| 269 | 270 | 271 | 272 | 273 | 274 | 275 | 276 | 277 | 278 | 279 | 280 | 281 | 282 | 283 | 284 |     |    |    |
| 285 | 286 | 287 | 288 | 289 | 290 | 291 | 292 | 293 | 294 | 295 | 296 | 297 | 298 | 299 | 300 |     |    |    |
| 301 | 302 | 303 | 304 | 305 | 306 | 307 | 308 | 309 | 310 | 311 | 312 | 313 | 314 | 315 | 316 |     |    |    |
| 317 | 318 | 319 | 320 | 321 | 322 | 323 | 324 | 325 | 326 | 327 | 328 | 329 | 330 | 331 | 332 |     |    |    |
| 333 | 334 | 335 | 336 | 337 | 338 | 339 | 340 | 341 | 342 | 343 | 344 | 345 | 346 | 347 | 348 |     |    |    |
| 349 | 350 | 351 | 352 | 353 | 354 | 355 | 356 | 357 | 358 | 359 | 360 | 361 | 362 | 363 | 364 |     |    |    |
| 365 | 366 | 367 | 368 | 369 | 370 | 371 | 372 | 373 | 374 | 375 | 376 | 377 | 378 | 379 | 380 |     |    |    |
| 381 | 382 | 383 | 384 | 385 | 386 | 387 | 388 | 389 | 390 | 391 | 392 | 393 | 394 | 395 | 396 |     |    |    |
| 397 | 398 | 399 | 400 | 401 | 402 | 403 | 404 | 405 | 406 | 407 | 408 | 409 | 410 | 411 | 412 |     |    |    |
| 413 | 414 | 415 | 416 | 417 | 418 | 419 | 420 | 421 | 422 | 423 | 424 | 425 | 426 | 427 | 428 |     |    |    |
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| 445 | 446 | 447 | 448 | 449 | 450 | 451 | 452 | 453 | 454 | 455 | 456 | 457 | 458 | 459 | 460 |     |    |    |
| 461 | 462 | 463 | 464 | 465 | 466 | 467 | 468 | 469 | 470 | 471 | 472 | 473 | 474 | 475 | 476 |     |    |    |
| 477 | 478 | 479 | 480 | 481 | 482 | 483 | 484 | 485 | 486 | 487 | 488 | 489 | 490 | 491 | 492 |     |    |    |
| 493 | 494 | 495 | 496 | 497 | 498 | 499 | 500 | 501 | 502 | 503 | 504 | 505 | 506 | 507 | 508 |     |    |    |
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10463 10464

## **7. PDB coordinates for QM/MM optimized structures.**

The PDB coordinates (in a separate PDB file) correspond to the final QM/MM optimized geometry of each structure discussed in the paper.