New Cellulose-Supported Reagent: a Sustainable Approach to Guanidines

Andrea Porcheddu,* Giampaolo Giacomelli, Alessandra Chighine, and Simonetta

Masala

Dipartimento di Chimica, Università degli Studi di Sassari, Via Vienna 2, I-07100 Sassari, Italy

Supporting Information

General Methods. All reagents and solvents were obtained from Aldrich, Avocado or Acros Organics. Perloza AV-100, particle size 100-250 mm was obtained from Iontosorb (Czech Republic); loading of aniline bead cellulose is about 0.5 mmol/g. Commercial bead cellulose is supplied as pre-swollen (water) gel. Cellulose beads were agitated using standard laboratory shakers or roller mixers. All reagents and solvents employed were reagent grade materials purified by standard methods and distilled before use. All conventional reactions were run under dry nitrogen in flamedried glassware using standard techniques unless otherwise stated. Thin-layer chromatography (TLC) analysis was performed with Merck Kieselgel 60 F254 plates and visualized using UV light, KMnO₄ staining and the Sakaguchi reagent (spray of a 0.1% solution of 8-hydroxyquinoline in acetone followed by a spray of a mixture of

^{*} To whom correspondence should be addressed. E-mail: anpo@uniss.it

0.2 mL of bromine in 100 mL of 0.5 N NaOH). Melting points were determined in open capillary tubes on a Buchi apparatus and are uncorrected. The experiments were performed using using a flask equipped with a reflux condenser mounted outside the apparatus. FT-IR spectra were recorded as KBr pellets on Bruker Equinox 55 spectrometer (value in cm⁻¹). Low resolution electronspray mass spectrometry was performed using VG Quattro (quadrupole-hexapole-quadrupole, QHQ) mass spectrometer system (Fisons Instruments, VG Analytical; Manchester, UK). Analytical HPLC was performed using a reversed phase Nucleosil C18 (5µm) column (250 x 2 mm) (Grom, Herremberg, Germany). Solvents used for HPLC elution were (A) H₂O containing 0.1% TFA, (B) 95% acetonitrile in H₂O containing 0.1% TFA. The column was eluted with a linear gradient of solvent A going to 21% solvent B in A over the course of 5 min at a flow rate of 1.0 mL/min (220 nm detection). H NMR (300 MHz) and proton-decoupled ¹³C NMR (75.4 MHz) Fourier transform spectra were obtained with Varian VXR-300 spectrometer in CDCl₃ using CDCl₃ solutions and TMS as an internal standard. Chemical shifts are expressed in (δ) ppm values, and coupling constants are expressed in Hertz (Hz). The following abbreviations are used: s = singlet, d = doublet, m = multiplet and brs = broad singlet. Elemental analyses were performed on a Perkin-Elmer 420 B analyzer.

1H-Pyrazole-1-carboxamidine Hydrochloride (**1**). To pyrazole (8.17 g, 0.12 mol) and cyanamide (5.05 g, 0.12 mol) in *p*-dioxane (120 mL) was added 4 N HC1 (31 mL) in *p*-dioxane. The mixture was gently refluxed with stirring for 2 h under nitrogen. During the course of the reaction the product crystallizes. After cooling to room temperature, anhydrous ether (30 mL) was added and the mixture allowed to stand for 30 min. The white crystalline product was collected by filtration, washed with anhydrous ether, and dried to constant weight in vacuo to yield 15.8 g (90%): mp 164-165 °C (lit.¹ mp 165-66 °C); ¹H NMR (DMSO-d₆) δ (*ppm*): 9.77 (brs, 4H), 9.06 (d, 1 H), 8.12 (s, 1 H), 6.85 (t, 1 H). 13 C NMR (DMSO-d₆) δ (*ppm*): 153.2, 146.1, 132.2, 112.4. (ESI + ve ion): 111.1 [(M+H)⁺].

Colorimetric assay:

β-Naphthol test. A few beads of the cellulose were successively washed with DMF and MeOH. To a small portion (5 mg) of cellulose in MeOH (1 mL) at 0° C, 0.2 mL water-solution of NaNO₂ (2%) and 0.2 mL water-solution of HCl (2.5 mol/l) were gradually added. After 1 min the activated support was separated and rinsed with H₂O (4x3 mL). To a filtered off support 0.3 mL aqueous solution of β-naphthol (1% in NaOH 1N) was added and the mixture was shaken for 30 sec and the beads rinsed

(1) Bredereck, H.; Effenberger, F.; Hajek, M. Chem. Ber. 1965, 98, 3178.

several times with a phosphate buffer (0.1 mol/l), H₂O and MeOH. If free primary aromatic amino groups are present on the solid support the beads turn red. It is recommended that for each test a blank experiment be run in parallel.

Guanidines synthesis

Cellulos

$$NH_2$$
 NH_2
 NH

Preparation of solid supported reagent 8

Aniline Cellulose **6** (2 g, 1 mmol) was added to a chilled solution of NaNO₂ (0.21 g, 3.0 mmol) in H_2O (20 ml). The mixture was shaken at 0°C while 37% aq. HCl (2.6 mL, 3.1 mol) was added in portions over 10 min. Then the slurry was shaken at 0°C for a further 3 h, filtered and washed with chilled water (5 x 20 ml). The resulting cellulose-bound diazonium salt **7** was transferred into water (10 ml) at 0°C. Then an ice water solution (15 ml) of 1*H*-pyrazole-1-carboxamidine hydrochloride **3** (0.59 g, 4 mmol) and NaHCO₃ (0.42 g 5 mmol) was added in portions at 0°C over 10 min. The reaction mixture was shaken at 0°C for 3 h and for 12h at room temperature. After

filtration, the cellulose was washed with H₂O (20 mL) and stirred in EtOH (20 mL) for 1 h, filtered again and washed with EtOH (20 mL), H₂O (20 mL) and EtOH (20 mL). The cellulose 8 was then swollen in EtOH (15 mL). A few beads of cellulose were dried in high vacum for 1 d to yield pale brown beads of 8. Functionalization of all cellulose-bound amines was verified by the β-naftol test.² The efficiency of the transformation according to nitrogen elemental analysis proved to be over 99%. IR (KBr): 3228, 3097, 1675 (s), 1600, 1518, 1441, 1379, 1340(s), 1240, 1153 (s), 828

General procedure to prepare guanidines 10a-l

The cellulose 8 previously obtained was suspended in EtOH (10 ml/g resin) and treated with the amine (5 equiv) and heated at reflux for 12 h. The cellulose beads were filtered and then washed sequentially with EtOH, NMP, 2% aq. NaHSO₄, H₂O, EtOH (3 x 15 ml/g of cellulose), and dried in vacuum. The corresponding guanidine was cleaved from the polymer by treatment with 1N HCl in H₂O (15 ml) under mechanical agitation for 3 h. Subsequently; the cellulose was filtered and washed with H₂O (2 x 10 ml) and EtOH (15 ml). The cleavage step was repeated three times. The cellulose was then washed finally with H₂O (2 x 15 ml) and the combined filtrates were concentrated under vacuum to afford the desired guanidine 10 in yields up to 98%.

(2) Gaggini, F.; Porcheddu, A.; Reginato, G.; Rodriquez, M.; Taddei, M. J. Comb. Chem. 2004; 6, 805-810.

$$\begin{array}{c} {}^{\ominus}\text{Cl} \, {}^{\oplus}\text{NH}_2 \\ {}^{N} \, {}^{N}\text{NH}_2 \end{array}$$

n-Butylguanidine hydrochloride **10a.** 98% yield. Colourless oil. ¹H NMR (DMSO- d_6) δ 7.71 (brs, 1H), 7.15 (brs, 4H), 3.07 (m, J = 6.6, 2H), 1.46 (m, 2H), 1.34 (m, 2H), 0.86 (t, J = 7.2 Hz, 3H). ¹³C NMR (DMSO- d_6): δ (ppm) 158.4, 37.9, 33.4, 20.1, 14.3. HPLC purity 99%. (ESI + ve ion). Calcd for C₅H₁₄N₃: 116.1 [(M+H)⁺]. Found: 116.1. Anal. Calcd for C₅H₁₄ClN₃: C, 39.60; H, 9.31; N, 27.71. Found: C, 39.51; H, 9.26; N, 27.79.

$$\begin{array}{c} H \stackrel{Cl}{\underset{NH_2}{\smile}} \oplus \\ NH_2 \end{array}$$

tert-Butylguanidine hydrochloride 10b. 91% yield. ¹H NMR (DMSO- d_6): δ (ppm) 7.48 (s, 1H), 7.00 (brs, 4H), 1.29 (s, 9H). ¹³C NMR (DMSO- d_6): δ (ppm) 156.3, 51.5, 29.2. (ESI + ve ion): Calcd for $C_5H_{14}N_3$: 116.1 [(M+H)⁺]. Found: 116.1. HPLC purity 99%. Anal. Calcd for $C_5H_{14}ClN_3$: C, 39.60; H, 9.31; N, 27.71. Found: C, 39.68; H, 9.36; N, 27.75.

$$\begin{array}{c} H^{Cl} \oplus \\ N \longrightarrow NH_2 \end{array}$$

Cyclohexylguanidine Hydrochloride 10c. 95% yield. white crystalline solid: 225-226 °C (litt.³ 224-226). ¹H NMR (DMSO-*d*6): δ (ppm) 7.55 (d, J = 8.3 Hz, 1H), 6.63–7.41 (brs, 4H), 3.45–3.30 (m, 1H), 1.74–1.89 (m, 2H), 1.60–1.71 (m, 2H), 1.50–1.58

(3) Braun, C. E. J. Am. Chem. Soc. 1933, 55, 1280-1284.

(m, 1H), 1.09–1.37 (m, 5H), 13 C NMR ((DMSO- d_6): 156.0, 49.1, 32.2, 24.8, 23.9. (ESI + ve ion). Calcd for $C_7H_{16}N_3$: 142.1 [(M+H)⁺]. Found: 142.3 [(M+H)⁺]. HPLC purity 98%. Anal. Calcd for $C_7H_{16}CIN_3$: C, 47.32; H, 9.08; N, 23.65. Found: C, 47.24; H, 9.17; N, 23.62.

$$\begin{array}{c} \text{Cl} & \oplus \\ \text{NH}_2 \\ \hline & \text{N} \\ \text{H} \end{array}$$

Benzylguanidine hydrochloride 10d. 98% yield. White crystalline solid, mp 173-174°C. ¹H NMR (DMSO- d_6): δ (ppm) 8.15 (brs, 1H), 6.84–7.62 (m, 9H), 4.35 (brs, 2H). ¹³C NMR (DMSO- d_6): δ (ppm) 162.8, 141.7, 121.9, 118.0, 114.0, 110.3. (ESI + ve ion). Calcd for $C_8H_{12}N_3$: 150.1 [(M+H)⁺]. Found: 150.2. HPLC purity 99%. Anal. Calcd for $C_8H_{12}ClN_3$: C, 51.76; H, 6.51; N, 22.63. Found: C, 51.81; H, 6.44; N, 22.66.

$$\begin{array}{c} Cl \stackrel{\oplus}{N}H_2 \\ MeOOC \stackrel{N}{N} NH_2 \end{array}$$

Methyl 2-(amidinoamino)acetate hydrochloride 10e. 65% yield. White hygroscopic foam. 1 H NMR (DMSO- d_{6}): δ (ppm) 7.45 (s, 1H), 7.02 (brs, 4H), 3.58 (s, 3H), 3.21 (s, 2H). 13 C NMR (DMSO- d_{6}): δ (ppm) 172.3, 159.2, 52.1, 40.2. (ESI + ve ion). Calcd for $C_{4}H_{10}N_{3}O_{2}$: 132.1 [(M+H)⁺]. Found: 132.3. HPLC purity 97%. Anal. Calcd for $C_{4}H_{10}ClN_{3}O_{2}$: C, 28.67; H, 6.01; N, 25.07. Found: C, 28.71; H, 6.13; N, 25.02.

$$\begin{array}{c} H \stackrel{Cl^{\ominus}_{\oplus}}{\longrightarrow} NH_2 \end{array}$$

Phenylguanidine hydrochloride 10f. 71% yield. ¹H NMR (DMSO-d₆): 7.24 (d,

J=7.4 Hz, 2H), 7.29 (t, J=7.4 Hz, 1H), 7.45 (t, J=7.9 Hz, 2H), 7.61 (brs, 4H), 10.07 (brs, 1H). ¹³C NMR ((DMSO- d_6): δ (ppm) 158.2, 142.6, 129.2, 128.8, 118.3, 115.7. (ESI + ve ion). Calcd for $C_7H_{10}N_3$: 135.1 [(M+H)⁺]. Found: 135.2 [(M+H)⁺]. HPLC purity 97%. Anal. Calcd for $C_7H_{10}ClN_3$: C, 48.99; H, 5.87; N, 20.66. Found: C, 48.90; H, 5.79; N, 20.67.

$$\begin{array}{c|c} \text{MeO} & \stackrel{\ominus \oplus}{\underset{\text{N}}{\bigcap}} \text{NH}_2 \\ & & \text{NH}_2 \\ \end{array}$$

(4-Methoxyphenyl)guanidine hydrochloride 10g. 78% yield. white crystalline solid: mp 143-144 °C (lit.⁴ mp 144–146 °C). ¹H NMR (DMSO- d_6): δ (ppm) 8.37 (brs, 1H), 7.36 (broad s, 4H), 7.18 (d, J = 8.9 Hz, 2H), 7.00 (d, J = 8.9 Hz, 2H), 3.77 (s, 3H). ¹³C NMR (DMSO- d_6): δ (ppm) 158.5, 150.1, 136.9, 117.9, 115.6, 61.1. (ESI + ve ion). Calcd for $C_8H_{12}N_3$: 166.1 [(M+H)⁺]. Found: 166.3. HPLC purity 98%. Anal. Calcd for $C_8H_{12}C1N_3O$: C, 47.65; H, 6.00; N, 20.84. Found: C, 47.59; H, 5.97; N, 20.90.

Piperidine-1-carboxamidine hydrochloride 10i. 93% yield. White solid, mp 186-87 °C, (lit. mp 187-188 °C); ¹H NMR (DMSO- d_6): δ (ppm): 5.2-4.3 (br s, 3H), 3.31 (t, J=5.25 Hz, 4H), 1.54-1.37 (m, 6H). ¹³C NMR (DMSO- d_6): δ (ppm): 158.8, 45.9, 25.1, 24.0. (ESI + ve ion). Calcd for C₆H₁₄N₃: 128.1 [(M+H)⁺]. Found: 127.9. HPLC

(4) Bernatowicz, M. S.; Wu, Y.; Matsueda, G. R. *J. Org.Chem.* **1992**, *57*, 2497-2502. (5) Bannard, R. A. B.; Casselman, A. A.; Cockburn, W. F.; Brown, G. M. *Can. J. Chem.* **1958**, *36*, 1541.

purity 98%. Anal. Calcd for $C_6H_{14}ClN_3$: C, 44.04; H, 8.62; N, 25.68. Found: C, 44.12; H, 8.70; N, 25.57.

Diisopropylguanidine hydrochloride 10l. 91% yield. White hygroscopic foam. 1 H NMR (DMSO-d₆): δ (ppm) 7.03 (br s, 4H), 3.93 (h, J = 6.8 Hz, 2H), 1.24 (d, J = 6.8 Hz, 12H). 13 C NMR (DMSO-d₆): δ (ppm): 158.2, 43.4, 21.8. (ESI + ve ion). Calcd for $C_{7}H_{8}N_{3}$: 143.1 [(M+H)⁺]. Found: 143.2. [(M+H)⁺]. HPLC purity 97%. Anal. Calcd for $C_{7}H_{8}CIN_{3}$: C, 46.79; H, 10.10; N, 23.38. Found: C, 46.85; H, 10.01; N, 23.27.