

Supporting Information

1,3-Dithianes as Acyl Anion Equivalents in Pd-Catalyzed Asymmetric Allylic Substitution

Kun Yao,[†] Delong Liu,^{*,†} Qianjia Yuan,[‡] Tsuneo Imamoto,[‡] Yangang Liu,[†] and Wanbin Zhang^{*,†,‡}

[†]School of Pharmacy and [‡]School of Chemistry and Chemical Engineering, Shanghai Jiao Tong University, 800 Dongchuan Road, Shanghai 200240, P. R. China

Fax: (+)-86-21-5474-3265; Phone: (+)-86-21-5474-3265; e-mail: wanbin@sjtu.edu.cn

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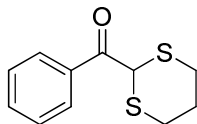
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1. General Information

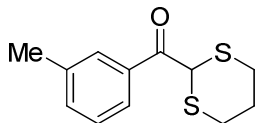
All the reactions were monitored by TLC (thin layer chromatography) using UV light to visualize the course of reaction. Anhydrous THF, DME, Et₂O, 1,4-dioxane, and toluene were prepared by distillation over sodium-benzophenone prior to use. ¹H, ¹⁹F, and ¹³C NMR spectra were obtained using a Varian MERCURY plus-400 spectrometer with TMS as an internal standard. HRMS was performed on a Bruker solariX FTICR Mass Spectrometer at the Instrumental Analysis Center of Shanghai Jiao Tong University. Melting points were measured with SGW X-4 micro melting point apparatus. 2-Aryl-1,3-dithianes **2** were prepared according to literature procedures.^[1] All commercially available reagents were used as received.

2. Preparation of 2-Aroyl-1,3-dithianes **4**

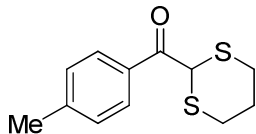
General procedure: To a solution of 1,3-dithiane (601 mg, 5 mmol) in THF (10 mL) at -40 °C was added *n*-BuLi (5 mmol, 2.5 M in hexane), and the solution was stirred for 1 h. This mixture was added dropwise to a solution of ethyl benzoates (6 mmol) in THF (10 mL) at -40 °C. The solution was slowly warmed to RT and stirred overnight. The reaction was quenched with water and diluted with EtOAc (90 mL). The resulting mixture was washed with 1M HCl, H₂O, and brine, dried over Na₂SO₄, and then concentrated in vacuum. The residue was purified by flash column chromatography to give the corresponding 2-aryl-1,3-dithianes.



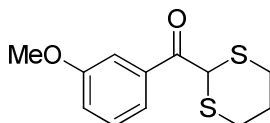
(1,3-Dithian-2-yl)(phenyl)methanone (4a):^[2] White solid (448 mg, 40%). mp: 92–93 °C; ¹H NMR (400 MHz, CDCl₃): δ 7.98–7.92 (m, 2H), 7.61–7.55 (m, 1H), 7.50–7.45 (m, 2H), 5.15 (s, 1H), 3.45–3.35 (m, 2H), 2.73–2.65 (m, 2H), 2.25–2.00 (m, 2H).



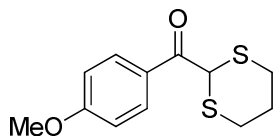
(1,3-Dithian-2-yl)(*m*-tolyl)methanone (4b): Yellow solid (404 mg, 34%). mp: 42–43 °C; ¹H NMR (400 MHz, CDCl₃): δ 7.80–7.66 (m, 2H), 7.40–7.26 (m, 2H), 5.14 (s, 1H), 3.43–3.31 (m, 2H), 2.74–2.60 (m, 2H), 2.42 (s, 3H), 2.22–1.98 (m, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 193.1, 138.8, 134.9, 134.3, 129.4, 128.8, 126.1, 42.9, 26.7, 25.4, 21.6; IR: 2920, 1674, 1275, 656; HRMS (APCI) [M+H]⁺ calcd for C₁₂H₁₅OS₂ 239.0564; found 239.0569.



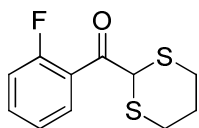
(1,3-Dithian-2-yl)(*p*-tolyl)methanone (4c):^[3] Yellow solid (381 mg, 32%). mp: 123–125 °C; ¹H NMR (400 MHz, CDCl₃): δ 7.85 (d, *J* = 8.4 Hz, 2H), 7.26 (d, *J* = 7.6 Hz, 2H), 5.15 (s, 1H), 3.38 (t, *J* = 12.0 Hz, 2H), 2.68 (ddd, *J* = 9.2, 6.0, 3.2 Hz, 2H), 2.41 (s, 3H), 2.20–2.00 (m, 2H).



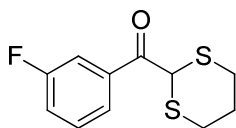
(1,3-Dithian-2-yl)(3-methoxyphenyl)methanone (4d): White solid (520 mg, 41%). mp: 68–69 °C; ^1H NMR (400 MHz, CDCl_3): δ 7.51–7.45 (m, 2H), 7.40–7.30 (m, 1H), 7.14–7.04 (m, 1H), 5.12 (s, 1H), 3.84 (s, 3H), 3.42–3.32 (m, 2H), 2.66 (ddd, $J = 8.8, 5.6, 2.8$ Hz, 2H), 2.18–1.92 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3): δ 192.7, 160.2, 136.2, 129.9, 121.3, 120.0, 113.4, 55.7, 42.8, 26.7, 25.4; IR: 1636, 649; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{12}\text{H}_{15}\text{O}_2\text{S}_2$ 255.0513; found 255.0513.



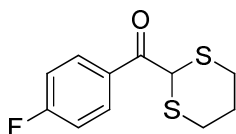
(1,3-Dithian-2-yl)(4-methoxyphenyl)methanone (4e): White solid (493 mg, 38%). mp: 56–57 °C; ^1H NMR (400 MHz, CDCl_3): δ 7.92 (d, $J = 8.8$ Hz, 2H), 6.92 (d, $J = 8.8$ Hz, 2H), 5.15 (s, 1H), 3.85 (s, 3H), 3.38–3.28 (m, 2H), 2.72–2.64 (m, 2H), 2.20–1.98 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3): δ 191.7, 163.9, 131.3, 127.4, 114.1, 55.7, 43.5, 27.2, 25.5; IR: 2920, 1666, 1262, 1170, 596; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{12}\text{H}_{15}\text{O}_2\text{S}_2$ 255.0513; found 255.0513.



(1,3-Dithian-2-yl)(2-fluorophenyl)methanone (4f): White solid (423 mg, 35%). mp: 63–64 °C; ^1H NMR (400 MHz, CDCl_3): δ 7.96–7.86 (m, 1H), 7.55–7.46 (m, 1H), 7.30–7.18 (m, 1H), 7.13–7.04 (m, 1H), 5.00 (s, 1H), 3.28 (t, $J = 12.4$ Hz, 2H), 2.61–2.50 (m, 2H), 2.25–1.90 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3): δ 189.8, 160.9 (d, $J_{\text{C-F}} = 252.3$ Hz), 134.8 (d, $J_{\text{C-F}} = 9.1$ Hz), 131.6 (d, $J_{\text{C-F}} = 2.0$ Hz), 124.7 (d, d, $J_{\text{C-F}} = 3.2$ Hz), 123.7 (d, $J_{\text{C-F}} = 12.6$ Hz), 116.6 (d, $J_{\text{C-F}} = 24.1$ Hz), 45.6 (d, $J_{\text{C-F}} = 8.3$ Hz), 25.7, 25.1; ^{19}F NMR (376 MHz, CDCl_3): δ -108.5; IR: 2923, 1677, 1608, 1265, 743, 628; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{11}\text{H}_{12}\text{OFS}_2$ 243.0314; found 243.0313.

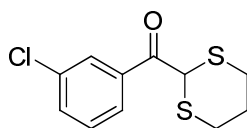


(1,3-Dithian-2-yl)(3-fluorophenyl)methanone (4g): White solid (412 mg, 34%). mp: 76–78 °C; ^1H NMR (400 MHz, CDCl_3): δ 7.71–7.58 (m, 2H), 7.49–7.35 (m, 1H), 7.28–7.20 (m, 1H), 5.05 (s, 1H), 3.33 (t, $J = 12.0$ Hz, 2H), 2.71–2.59 (m, 2H), 2.20–1.90 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3): δ 191.2, 162.8 (d, $J_{\text{C-F}} = 246.9$ Hz), 136.7 (d, $J_{\text{C-F}} = 6.3$ Hz), 130.3 (d, $J_{\text{C-F}} = 7.7$ Hz), 124.3 (d, $J_{\text{C-F}} = 3.0$ Hz), 120.4 (d, $J_{\text{C-F}} = 21.4$ Hz), 115.6 (d, $J_{\text{C-F}} = 22.5$ Hz), 42.4, 26.3, 25.0; ^{19}F NMR (376 MHz, CDCl_3): δ -111.4; IR: 1654, 673, 496; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{11}\text{H}_{12}\text{OFS}_2$ 243.0314; found 243.0313.

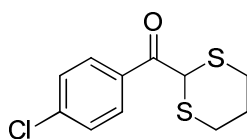


(1,3-Dithian-2-yl)(4-fluorophenyl)methanone (4h): White solid (497 mg, 41%). mp: 88–90 °C; ^1H NMR (400 MHz, CDCl_3): δ 7.94 (t, $J = 7.2$ Hz, 2H), 7.10 (t, $J = 8.8$ Hz, 2H), 5.09 (s, 1H), 3.31 (t, $J = 12.8$ Hz, 2H), 2.75–2.60 (m, 2H), 2.20–1.95 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3): δ 191.1,

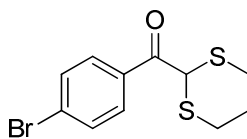
165.7 (d, $J_{\text{C-F}} = 254.4$ Hz), 131.4 (d, $J_{\text{C-F}} = 9.3$ Hz), 130.8 (d, $J_{\text{C-F}} = 2.9$ Hz), 115.9 (d, $J_{\text{C-F}} = 21.8$ Hz), 42.8, 26.6, 25.0; ^{19}F NMR (376 MHz, CDCl_3): δ -114.4; IR: 2904, 1642, 596; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{11}\text{H}_{12}\text{OFS}_2$ 243.0314; found 243.0318.



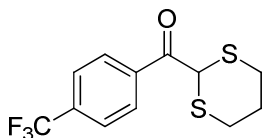
(3-Chlorophenyl)(1,3-dithian-2-yl)methanone (4i): Yellow solid (388 mg, 30%). mp: 88–90 °C; ^1H NMR (400 MHz, CDCl_3): δ 7.91 (t, $J = 2.0$ Hz, 1H), 7.82–7.73 (m, 1H), 7.55–7.50 (m, 1H), 7.40 (t, $J = 7.9$ Hz, 1H), 5.06 (s, 1H), 3.39–3.30 (m, 2H), 2.67 (ddd, $J = 8.8, 5.6, 2.8$ Hz, 2H), 2.22–1.94 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3): δ 191.4, 136.6, 135.3, 133.4, 130.2, 129.0, 126.9, 42.7, 26.6, 25.2; IR: 1637, 649; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{11}\text{H}_{12}\text{OS}_2\text{Cl}$ 259.0018; found 259.0027.



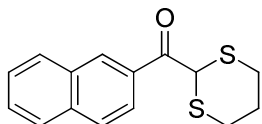
(4-Chlorophenyl)(1,3-dithian-2-yl)methanone (4j): Yellow solid (453 mg, 35%). mp: 97–98 °C; ^1H NMR (400 MHz, CDCl_3): δ 7.94–7.81 (m, 2H), 7.47–7.36 (m, 2H), 5.08 (s, 1H), 3.38–3.28 (m, 2H), 2.67 (ddd, $J = 8.8, 6.0, 2.8$ Hz, 2H), 2.19–1.94 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3): δ 191.6, 140.0, 133.1, 130.4, 129.2, 42.9, 26.8, 25.3; IR: 1642, 610, 413; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{11}\text{H}_{12}\text{OS}_2\text{Cl}$ 259.0018; found 259.0024.



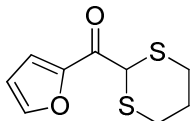
(4-Bromophenyl)(1,3-dithian-2-yl)methanone (4k): White solid (521 mg, 34%). mp: 98–99 °C; ^1H NMR (400 MHz, CDCl_3): δ 7.83–7.75 (m, 2H), 7.62–7.55 (m, 2H), 5.07 (s, 1H), 3.49–3.17 (m, 2H), 2.75–2.60 (m, 2H), 2.20–1.95 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3): δ 191.7, 133.6, 132.2, 130.5, 128.7, 42.9, 26.7, 25.3; IR: 1654, 497; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{11}\text{H}_{12}\text{OS}_2\text{Br}$ 302.9513; found 302.9521.



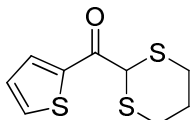
(1,3-Dithian-2-yl)(4-(trifluoromethyl)phenyl)methanone (4l): White solid (585 mg, 40%). mp: 70–72 °C; ^1H NMR (400 MHz, CDCl_3): δ 8.02 (d, $J = 7.6$ Hz, 2H), 7.70 (d, $J = 7.6$ Hz, 2H), 5.08 (s, 1H), 3.38–3.28 (m, 2H), 2.73–2.56 (m, 2H), 2.22–1.94 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3): δ 191.2, 137.5, 134.5 (d, $J_{\text{C-F}} = 32.7$ Hz), 129.0, 126.2 (q, $J = 271.7$ Hz), 125.7 (q, $J_{\text{C-F}} = 3.6$ Hz), 42.5, 26.2, 24.9; ^{19}F NMR (376 MHz, CDCl_3): δ -63.2; IR: 2925, 1677, 1271, 1067, 668, 648; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{12}\text{H}_{12}\text{OS}_2\text{F}_3$ 293.0282; found 293.0290.



(1,3-Dithian-2-yl)(naphthalen-2-yl)methanone (4m): Yellow solid (384 mg, 28%). mp: 80–82 °C; ¹H NMR (400 MHz, CDCl₃): δ 8.46 (s, 1H), 8.05–7.75 (m, 4H), 7.62–7.50 (m, 2H), 5.32 (s, 1H), 3.48–3.36 (m, 2H), 2.80–2.65 (m, 2H), 2.26–2.02 (m, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 192.9, 135.9, 132.7, 132.1, 130.5, 129.9, 128.9, 128.8, 127.9, 127.1, 124.6, 43.1, 26.9, 25.4; IR: 1692, 1384, 610, 408; HRMS (APCI) [M+H]⁺ calcd for C₁₅H₁₅OS₂ 275.0564; found 275.0572.



(1,3-Dithian-2-yl)(furan-2-yl)methanone (4n): White solid (396 mg, 37%). mp: 141–142 °C; ¹H NMR (400 MHz, CDCl₃): δ 7.58–7.55 (m, 1H), 7.28–7.24 (m, 1H), 6.55 (dd, *J* = 3.6, 1.6 Hz, 1H), 4.89 (s, 1H), 3.50–3.36 (m, 2H), 2.69–2.46 (m, 2H), 2.24–1.96 (m, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 182.2, 150.3, 146.7, 118.5, 112.9, 40.9, 26.1, 25.3; IR: 1657, 595; HRMS (APCI) [M+H]⁺ calcd for C₉H₁₁O₂S₂ 215.0200; found 215.0206.



(1,3-Dithian-2-yl)(thiophen-2-yl)methanone (4o): White solid (357 mg, 31%). mp: 138–139 °C; ¹H NMR (400 MHz, CDCl₃): δ 7.74–7.62 (m, 2H), 7.15–7.10 (m, 1H), 4.91 (s, 1H), 3.49–3.39 (m, 2H), 2.65 (ddd, *J* = 8.4, 5.6, 2.8 Hz, 2H), 2.22–2.00 (m, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 186.4, 141.5, 134.6, 132.8, 128.4, 43.4, 26.7, 25.3; IR: 2922, 2850, 1640, 1384, 419; HRMS (APCI) [M+H]⁺ calcd for C₉H₁₁OS₃ 230.9972; found 230.9977.

3. Optimization of the Reaction Conditions^[a]

Table S1 Screening the effect of ligand, additive, and base on the reaction

Entry	Ligand	Additive	Yield (%) ^{[b][4]}	Ee (%) ^[c]	
1	L1	-	36	25	
2	L2	-	32	20	
3	L3	-	NP	-	
4	L4	-	27	63	
5	L5	-	33	59	
6	L6	-	43	73	
7	L6	LiCl	48	75	
8	L6	LiBr	46	79	
9	L6	LiI	47	85	
10	L6	CaI ₂	NP	-	
11	BenzP*	-	41	78	
12	BenzP*	LiF	42	68	
13	BenzP*	LiCl	Trace	72	
14	BenzP*	LiBr	34	90	

15	BenzP*	LiI	52	92
16	QuinoxP*	LiI	41	90
17 ^[d]	BenzP*	LiI	NP	-
18 ^[e]	BenzP*	LiI	NP	-
19 ^[f]	BenzP*	LiI	NP	-

[a] Unless otherwise noted, the reaction of **1a** (0.1 mmol) with **2a** (0.2 mmol) was carried out using a catalytic system consisting of $[\text{Pd}(\eta^3\text{-C}_3\text{H}_5\text{Cl})_2]$ (2.5 mol%) and ligand (5.5 mol%) in the presence of LiHMDS (0.15 mmol) and an additive (0.1 mmol) in THF (3 mL) under a nitrogen atmosphere at RT for 12 h. [b] Isolated yield. [c] Determined by HPLC using a chiral Daicel OD-H column. [d] NaHMDS was used as a base. [e] KHMDS was used as a base. [f] LDA was used as a base.

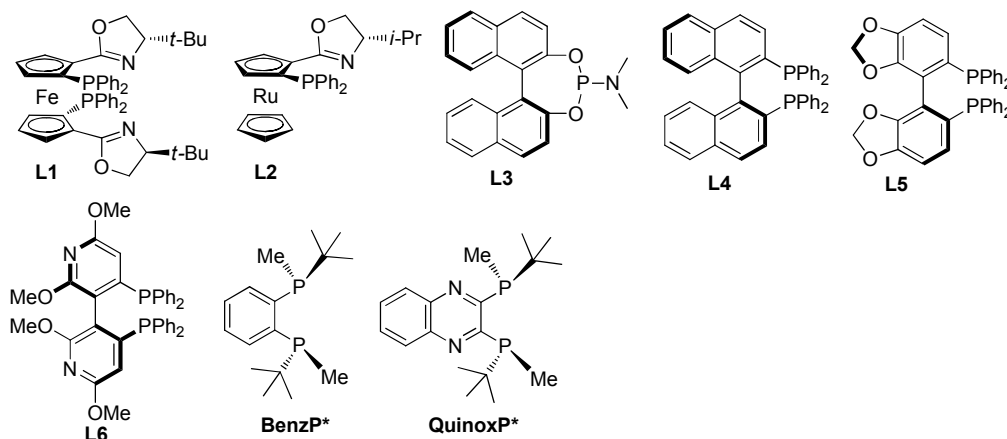


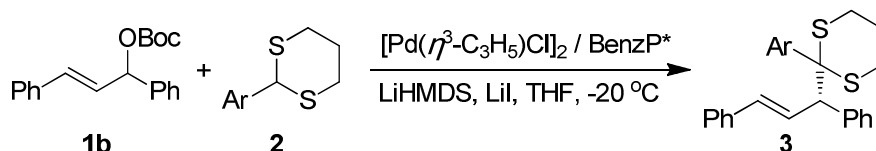
Table S2 Screening the effect of ligand and base on the reaction

Reaction scheme: **1b** + **4a** $\xrightarrow[\text{Base, THF}]{[\text{Pd}(\eta^3\text{-C}_3\text{H}_5\text{Cl})_2] / \text{Ligand}}$ **5a**. The reaction involves the coupling of **1b** and **4a** to form **5a** using a palladium catalyst and various ligands and bases.

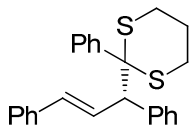
Entry	Ligand	Base (equiv)	Yield (%) ^[b]	Ee (%) ^[c]
1	L4	<i>t</i> BuOK (2)	67	27
2	L7	<i>t</i> BuOK (2)	trace	-
3	BenzP*	<i>t</i> BuOK (2)	51	93
4	BenzP*	DBU (2)	trace	92
5	BenzP*	LiHMDS (2)	NP	-
6	BenzP*	<i>t</i> BuONa (2)	91	95
7	BenzP*	<i>t</i> BuONa (2.5)	33	95
8	BenzP*	<i>t</i> BuONa (1.5)	93	95
9	BenzP*	NaH (2)	NP	-
10^[d]	BenzP*	NaH (1.5)	96	95

[a] Unless otherwise noted, the reaction of **1b** (0.15 mmol) with **4a** (0.1 mmol) was carried out using a catalytic system consisting of $[\text{Pd}(\eta^3\text{-C}_3\text{H}_5\text{Cl})_2]$ (2.5 mol%) and ligand (5.5 mol%) in the presence of a base in THF (4 mL) under a nitrogen atmosphere at RT for 12 h. [b] Isolated yield. [c] Determined by HPLC using a chiral Daicel IC-3 column. [d] 15-Crown-5 (0.12 mmol) was used.

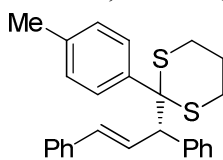
4. Pd-Catalyzed Allylic Substitution with 2-Aryl-1,3-dithiane Nucleophiles



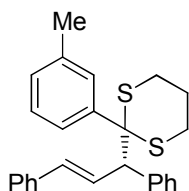
To an oven-dried glassware were added $[\text{Pd}(\eta^3\text{-C}_3\text{H}_5)\text{Cl}]_2$ (0.9 mg, 2.5 mol%), BenzP* (1.6 mg, 5.5 mol%), and anhydrous THF (1.0 mL). To another dry glassware were added **2** (0.2 mmol), ultra-dry LiI (0.1 mmol, 13.3 mg), and anhydrous THF (1.0 mL). LiHMDS (1.5 – 5 equiv) was then added to the second glassware. The mixture was stirred at RT under a nitrogen atmosphere for 1 h. Then a solution of allylic carbonate **1b** (31 mg, 0.10 mmol) in anhydrous THF (1.0 mL) was added to the first glassware. The solution in the first glassware was added dropwise to another one. The reaction mixture was stirred at $-20\text{ }^\circ\text{C}$ for 12 h, quenched with H_2O , and extracted with EtOAc (5 mL $\times$ 3). The combined organic layer was washed with brine, dried over anhydrous Na_2SO_4 , and then concentrated in vacuum. The residue was then purified by preparative TLC (cyclohexane:anisole = 10:1) to give the desired product.



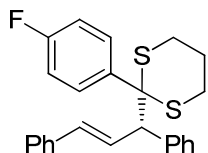
(S)-2-(1,3-Diphenylallyl)-2-phenyl-1,3-dithiane (3a): Using 1.5 equiv of LiHMDS to obtain a white solid (36.9 mg, 95%). mp: $102\text{--}104\text{ }^\circ\text{C}$; $[\alpha]_{\text{D}}^{20} = -1.5$ (c 0.21, CHCl_3); ^1H NMR (400 MHz, CDCl_3): δ 7.78 (d, $J = 7.2$ Hz, 2H), 7.45–7.08 (m, 11H), 6.91–6.85 (m, 2H), 6.63–6.49 (m, 2H), 4.06 (d, $J = 8.0$ Hz, 1H), 2.71–2.51 (m, 4H), 1.97–1.80 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3): δ 138.4, 138.3, 137.2, 133.2, 131.0, 130.0, 128.5, 128.1, 127.7, 127.5, 127.4, 127.2, 127.1, 126.5, 64.0, 61.7, 27.7, 27.6, 24.8; IR: 2917, 2850, 1631, 1383, 1265, 738; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{25}\text{H}_{25}\text{S}_2$ 389.1398; found 389.1407; Daicel Chiralpak OD-H column, n -hexane/ i -PrOH = 98/2, 1 mL/min, 254 nm: $t_{\text{major}} = 6.6$ min, $t_{\text{minor}} = 8.2$ min.



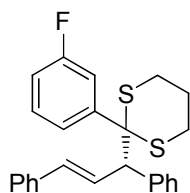
(S)-2-(1,3-Diphenylallyl)-2-(*p*-tolyl)-1,3-dithiane (3b): Using 4 equiv of LiHMDS to obtain yellow oil (30.2 mg, 75%). $[\alpha]_{\text{D}}^{20} = -4.4$ (c 0.16, CHCl_3); ^1H NMR (400 MHz, CDCl_3): δ 7.59 (d, $J = 8.0$ Hz, 2H), 7.39–7.06 (m, 10H), 6.85 (d, $J = 6.8$ Hz, 2H), 6.49 (d, $J = 4.6$ Hz, 2H), 4.01 (t, $J = 4.4$ Hz, 1H), 2.68–2.51 (m, 4H), 2.38 (s, 3H), 1.90–1.80 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3): δ 138.3, 137.2, 136.8, 135.1, 133.0, 131.0, 130.00, 128.7, 128.4, 127.8, 127.3, 127.1, 126.5, 63.7, 61.6, 27.6, 27.5, 24.8, 21.0; IR: 2920, 2849, 1633, 1385, 418; Daicel Chiralpak OD-H column, n -hexane/ i -PrOH = 98/2, 1 mL/min, 254 nm: $t_{\text{major}} = 5.3$ min, $t_{\text{minor}} = 6.7$ min.



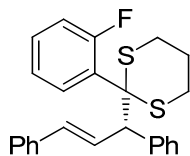
(S)-2-(1,3-Diphenylallyl)-2-(*m*-tolyl)-1,3-dithiane (3c): Using 5 equiv of LiHMDS to obtain yellow oil (31.0 mg, 77%); $[\alpha]_D^{20} = +1.0$ (*c* 0.40, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.60–7.44 (m, 2H), 7.38–7.03 (m, 10H), 6.81 (d, *J* = 7.3 Hz, 2H), 6.50–6.46 (m, 2H), 3.97 (d, *J* = 7.2 Hz, 1H), 2.70–2.50 (m, 4H), 2.30 (s, 3H), 1.90–1.80 (m, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 138.3, 138.1, 137.6, 137.2, 133.0, 131.6, 130.0, 128.4, 128.0, 127.9, 127.8, 127.7, 127.3, 127.2, 127.1, 126.4, 63.9, 61.5, 27.6, 27.5, 24.8, 21.6; IR: 2920, 2856, 1641, 1384, 629; HRMS (APCI) $[M+H]^+$ calcd for C₂₆H₂₇S₂ 403.1554; found 403.1513; Daicel Chiralpak OD-H column, *n*-hexane/*i*-PrOH = 98/2, 1 mL/min, 254 nm: *t*_{major} = 5.0 min, *t*_{minor} = 6.2 min.



(S)-2-(1,3-Diphenylallyl)-2-(4-fluorophenyl)-1,3-dithiane (3d): Using 5 equiv of LiHMDS to obtain a white solid (33.2 mg, 82%). mp: 134–135 °C; $[\alpha]_D^{20} = -5.3$ (*c* 0.68, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.68 (dd, *J* = 8.4, 5.6 Hz, 2H), 7.39–7.06 (m, 8H), 6.99 (t, *J* = 8.8 Hz, 2H), 6.83 (d, *J* = 7.6 Hz, 2H), 6.48 (d, *J* = 4.8 Hz, 2H), 4.02 – 3.92 (t, *J* = 4.4 Hz, 1H), 2.70–2.50 (m, 4H), 2.00–1.78 (m, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 161.9 (d, *J*_{C-F} = 246.0 Hz), 138.0, 137.0, 134.2 (d, *J*_{C-F} = 2.0 Hz), 133.3, 132.8 (d, *J*_{C-F} = 8.0 Hz), 129.9, 128.5, 127.5, 127.4, 127.2, 126.4, 114.8, 114.6, 63.2, 61.6, 27.5, 27.4, 24.7; ¹⁹F NMR (376 MHz, CDCl₃): δ -115.95; IR: 2919, 2849, 1645, 1384, 678, 433; HRMS (APCI) $[M+H]^+$ calcd for C₂₅H₂₄S₂F 407.1303; found 407.1316; Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 99/1, 0.7 mL/min, 254 nm: *t*_{major} = 6.2 min, *t*_{minor} = 8.3 min.

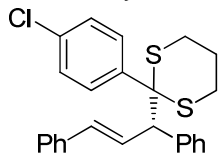


(S)-2-(1,3-Diphenylallyl)-2-(3-fluorophenyl)-1,3-dithiane (3e): Using 3 equiv of LiHMDS to obtain a white solid (39.4 mg, 97%). mp: 40–42 °C; $[\alpha]_D^{20} = +0.9$ (*c* 0.15, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.54–7.44 (m, 2H), 7.38–7.09 (m, 9H), 6.97 (td, *J* = 8.0, 2.4 Hz, 1H), 6.86–6.81 (m, 2H), 6.59–6.42 (m, 2H), 3.96 (d, *J* = 8.6 Hz, 1H), 2.66–2.51 (m, 4H), 1.93–1.78 (m, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 163.0 (d, *J*_{C-F} = 243.4 Hz), 142.0 (d, *J*_{C-F} = 6.7 Hz), 138.0, 137.0, 133.4, 129.9, 129.3 (d, *J*_{C-F} = 8.0 Hz), 128.5, 127.5, 127.4, 127.3, 127.2, 126.6 (d, *J*_{C-F} = 2.7 Hz), 126.5, 117.9 (d, *J*_{C-F} = 23.7 Hz), 114.1 (d, *J*_{C-F} = 21.1 Hz), 63.4 (d, *J*_{C-F} = 1.5 Hz), 61.7, 27.8, 27.5, 24.6; ¹⁹F NMR (376 MHz, CDCl₃): δ -113.4; IR: 2920, 2850, 1633, 1384, 1265, 963, 869, 698, 419; HRMS (APCI) $[M+H]^+$ calcd for C₂₅H₂₄S₂F 407.1303; found 407.1288; Daicel Chiralpak OD-H column, *n*-hexane/*i*-PrOH = 98/2, 1 mL/min, 254 nm: *t*_{major} = 5.9 min, *t*_{minor} = 7.1 min.

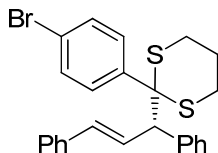


(S)-2-(1,3-Diphenylallyl)-2-(2-fluorophenyl)-1,3-dithiane (3f): Using 1.5 equiv of LiHMDS to obtain a white solid (24.9 mg, 61%). mp: 116–117 °C; $[\alpha]_D^{20} = +14.9$ (*c* 0.50, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.61 (t, *J* = 8.0 Hz, 1H), 7.40–6.81 (m, 14H), 6.40 (d, *J* = 15.6 Hz, 1H), 4.38

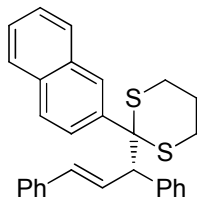
(d, $J = 8.8$ Hz, 1H), 2.85–2.55 (m, 4H), 1.94–1.78 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3): δ 161.0 (d, $J_{\text{C-F}} = 250.0$ Hz), 138.9, 137.3, 133.7, 132.9, 129.7, 129.5 (d, $J_{\text{C-F}} = 8.8$ Hz), 128.5, 128.4, 128.1 (d, $J_{\text{C-F}} = 7.1$ Hz), 127.4, 127.2, 127.0, 126.5, 123.3 (d, $J_{\text{C-F}} = 3.6$ Hz), 117.2 (d, $J_{\text{C-F}} = 24.3$ Hz), 61.6 (d, $J_{\text{C-F}} = 6.3$ Hz), 58.4 (d, $J_{\text{C-F}} = 1.9$ Hz), 28.1, 27.9, 24.4; ^{19}F NMR (376 MHz, CDCl_3): δ –101.1; IR: 2920, 2849, 1633, 1384, 697, 963, 419; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{25}\text{H}_{24}\text{S}_2\text{F}$ 407.1303; found 407.1293; Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 99/1, 0.7 mL/min, 254 nm: $t_{\text{major}} = 10.0$ min, $t_{\text{minor}} = 9.6$ min.



(S)-2-(4-Chlorophenyl)-2-(1,3-diphenylallyl)-1,3-dithiane (3g): Using 4 equiv of LiHMDS to obtain a white solid (41.5 mg, 98%). mp: 138–140 °C; $[\alpha]_{\text{D}}^{20} = -17.5$ (*c* 0.19, CHCl_3); ^1H NMR (400 MHz, CDCl_3): δ 7.66 (d, $J = 8.8$ Hz, 2H), 7.39–7.11 (m, 10H), 6.89–6.82 (m, 2H), 6.55–6.43 (m, 2H), 3.99 (d, $J = 8.0$ Hz, 1H), 2.66–2.37 (m, 4H), 1.92–1.78 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3): δ 137.9, 137.2, 137.0, 133.5, 133.2, 132.5, 129.9, 128.5, 128.1, 127.5, 127.4, 127.3, 127.1, 126.5, 63.3, 61.5, 27.6, 27.5, 24.7; IR: 2921, 2851, 1637, 1481, 1452, 1384, 1093, 617, 419; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{25}\text{H}_{24}\text{S}_2\text{Cl}$ 423.1008; found 423.0987; Daicel Chiralpak OD-H column, *n*-hexane/*i*-PrOH = 98/2, 1 mL/min, 254 nm: $t_{\text{major}} = 6.2$ min, $t_{\text{minor}} = 8.8$ min.



(S)-2-(4-Bromophenyl)-2-(1,3-diphenylallyl)-1,3-dithiane (3h): Using 3 equiv of LiHMDS to obtain a white solid (45.8 mg, 98%). mp: 145–146 °C; $[\alpha]_{\text{D}}^{20} = -10.4$ (*c* 0.28, CHCl_3); ^1H NMR (400 MHz, CDCl_3): δ 7.61 (d, $J = 8.4$ Hz, 2H), 7.46 (d, $J = 8.8$ Hz, 2H), 7.40–7.10 (m, 8H), 6.87 (d, $J = 6.8$ Hz, 2H), 6.59–6.40 (m, 2H), 4.00 (d, $J = 7.6$ Hz, 1H), 2.74–2.50 (m, 4H), 1.98–1.75 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3): δ 137.9, 137.8, 137.0, 133.5, 133.0, 131.1, 129.9, 128.5, 127.5, 127.3, 127.1, 126.5, 121.6, 63.3, 61.4, 27.6, 27.5, 24.7; IR: 2921, 2850, 1640, 1384, 1093, 619, 419; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{25}\text{H}_{24}\text{S}_2\text{Br}$ 467.0503; found 467.0490; Daicel Chiralpak OD-H column, *n*-hexane/*i*-PrOH = 98/2, 1 mL/min, 254 nm: $t_{\text{major}} = 6.1$ min, $t_{\text{minor}} = 8.7$ min.



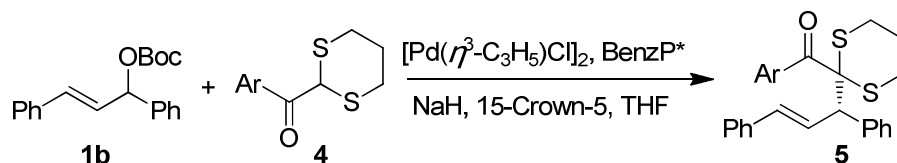
(S)-2-(1,3-Diphenylallyl)-2-(naphthalen-2-yl)-1,3-dithiane (3i): Using 3 equiv of LiHMDS to obtain yellow oil (43.1 mg, 98%). $[\alpha]_{\text{D}}^{20} = -26.3$ (*c* 0.20, CHCl_3); ^1H NMR (400 MHz, CDCl_3): δ 8.21 (s, 1H), 7.96–7.76 (m, 4H), 7.51 (dt, $J = 14.0, 6.8$ Hz, 2H), 7.44–7.02 (m, 8H), 6.85 (d, $J = 7.4$ Hz, 2H), 6.59–6.51 (m, 2H), 4.14 (d, $J = 3.7$ Hz, 1H), 2.70–2.58 (m, 4H), 2.00–1.75 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3): δ 138.2, 137.1, 135.9, 133.2, 133.1, 132.3, 130.9, 130.0, 128.5, 128.4, 127.7, 127.6, 127.5, 127.4, 127.3, 127.2, 126.5, 126.4, 126.0, 64.0, 61.6, 27.7, 27.6, 24.8; IR: 2919, 2849, 1633, 1384, 742, 696, 419; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{29}\text{H}_{27}\text{S}_2$ 439.1554;



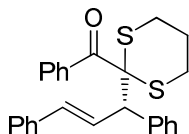
The chemical structure shows a 1,3-dithiane ring substituted at the 4-position with a thien-2-yl group. The 2-position of the thiane ring is substituted with a 1,3-bis(phenyl)prop-1-en-1-yl group. The stereochemistry at the 2-position is indicated by a dashed bond to the phenyl group and a solid bond to the propenyl chain.

Cc1ccc(cc1)/C=C/[C@H](c2ccc(C)cc2)S3CCCCC3S4CCCCC4S4

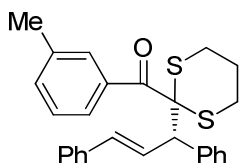
5. Pd-Catalyzed Allylic Substitution with 2-Aroyl-1,3-dithiane Nucleophiles



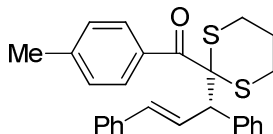
S10



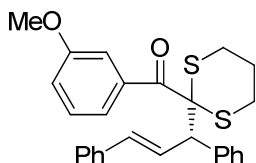
(S)-(2-(1,3-Diphenylallyl)-1,3-dithian-2-yl)(phenyl)methanone (5a): White solid (40.1 mg, 96%). mp: 125–126 °C; $[\alpha]_D^{20} = +63.4$ (*c* 0.70, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 8.08–7.98 (m, 2H), 7.53–7.45 (m, 1H), 7.45–7.08 (m, 12H), 6.79 (dd, *J* = 16.0, 8.8 Hz, 1H), 6.11 (d, *J* = 15.6 Hz, 1H), 4.41 (d, *J* = 8.5 Hz, 1H), 3.32–3.12 (m, 2H), 2.76–2.66 (m, 2H), 2.08–1.98 (m, 1H), 1.86–1.76 (m, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 196.8, 138.5, 138.4, 137.1, 133.5, 131.7, 130.2, 130.1, 128.7, 128.2, 128.1, 127.8, 127.7, 127.6, 126.8, 66.1, 56.5, 28.4, 24.3; IR: 1642, 1384, 497; HRMS (APCI) $[M+H]^+$ calcd for C₂₆H₂₅OS₂ 417.1347; found 417.1342; Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 98/2, 1 mL/min, 254 nm: *t*_{major} = 10.1 min, *t*_{minor} = 11.1 min.



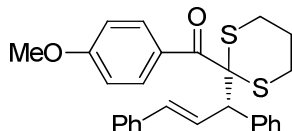
(S)-(2-(1,3-Diphenylallyl)-1,3-dithian-2-yl)(*m*-tolyl)methanone (5b): Yellow oil (40.5 mg, 94%). $[\alpha]_D^{20} = +70.7$ (*c* 0.20, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.89 (d, *J* = 7.6 Hz, 1H), 7.68 (s, 1H), 7.40–7.10 (m, 12H), 6.79 (dd, *J* = 15.6, 8.4 Hz, 1H), 6.10 (d, *J* = 16.0 Hz, 1H), 4.40 (d, *J* = 8.4 Hz, 1H), 3.32–3.08 (m, 2H), 2.80–2.59 (m, 2H), 2.34 (s, 3H), 2.10–1.95 (m, 1H), 1.90–1.71 (m, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 197.2, 138.6, 138.5, 137.9, 137.1, 133.5, 132.5, 130.8, 130.2, 128.7, 128.1, 127.9, 127.8, 127.7, 127.6, 127.1, 126.8, 66.2, 56.5, 28.4, 24.3, 21.7; IR: 2923, 2847 1691, 1658, 1383, 496, 434; HRMS (APCI) $[M+H]^+$ calcd for C₂₇H₂₇OS₂ 431.1503; found 431.1514; Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 98/2, 1 mL/min, 254 nm: *t*_{major} = 9.7 min, *t*_{minor} = 10.8 min.



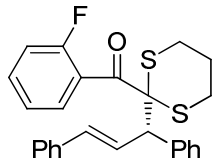
(S)-(2-(1,3-Diphenylallyl)-1,3-dithian-2-yl)(*p*-tolyl)methanone (5c): White solid (41.8 mg, 97%). mp: 136–138 °C; $[\alpha]_D^{20} = +79.0$ (*c* 0.18, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.98 (d, *J* = 8.4 Hz, 2H), 7.43–7.12 (m, 12H), 6.79 (dd, *J* = 15.6, 8.4 Hz, 1H), 6.14 (d, *J* = 15.6 Hz, 1H), 4.46 (d, *J* = 8.4 Hz, 1H), 3.29–3.06 (m, 2H), 2.79–2.60 (m, 2H), 2.41 (s, 3H), 2.06–1.97 (m, 1H), 1.87–1.77 (m, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 196.2, 148.0, 142.5, 138.4, 137.1, 135.5, 133.4, 130.4, 130.1, 128.8, 128.7, 128.1, 127.7, 127.6, 126.8, 66.6, 56.5, 28.4, 24.3, 21.7; IR: 2920, 2849 1637, 1471, 1384, 1117, 518, 446; HRMS (APCI) $[M+H]^+$ calcd for C₂₇H₂₇OS₂ 431.1503; found 431.1520; Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 97/3, 0.5 mL/min, 254 nm: *t*_{major} = 17.8 min, *t*_{minor} = 18.6 min.



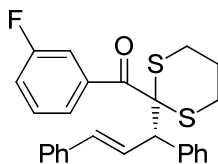
(S)-(2-(1,3-Diphenylallyl)-1,3-dithian-2-yl)(3-methoxyphenyl)methanone (5d): Yellow oil (42.4 mg, 95%). $[\alpha]_D^{20} = +60.1$ (c 0.27, CHCl_3); ^1H NMR (400 MHz, CDCl_3): δ 7.73–7.65 (m, 1H), 7.43–7.36 (m, 1H), 7.35–7.10 (m, 11H), 7.03 (ddd, $J = 8.4, 2.8, 0.8$ Hz, 1H), 6.77 (dd, $J = 15.6, 8.4$ Hz, 1H), 6.09 (d, $J = 15.6$ Hz, 1H), 4.40 (d, $J = 8.4$ Hz, 1H), 3.73 (s, 3H), 3.32–3.10 (m, 2H), 2.76–2.64 (m, 2H), 2.08–1.98 (m, 1H), 1.90–1.75 (m, 1H); ^{13}C NMR (100 MHz, CDCl_3): δ 196.9, 159.2, 139.8, 138.5, 137.0, 133.5, 130.2, 129.1, 128.7, 128.1, 127.8, 127.6, 126.8, 122.4, 118.2, 114.9, 66.1, 56.5, 55.5, 28.4, 24.2; IR: 2920, 2849 1677, 1584, 1272, 955, 404; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{27}\text{H}_{27}\text{O}_2\text{S}_2$ 447.1452; found 447.1456; Daicel Chiralpak IE column, n -hexane/ i -PrOH = 95/5, 1 mL/min, 254 nm: $t_{\text{major}} = 17.0$ min, $t_{\text{minor}} = 11.5$ min.



(S)-(2-(1,3-Diphenylallyl)-1,3-dithian-2-yl)(4-methoxyphenyl)methanone (5e): Yellow oil (43.8 mg, 98%). $[\alpha]_D^{20} = +69.4$ (c 0.30, CHCl_3); ^1H NMR (400 MHz, CDCl_3): δ 8.22–8.07 (m, 2H), 7.51–7.10 (m, 10H), 6.94–6.73 (m, 3H), 6.18 (d, $J = 16.0$ Hz, 1H), 4.48 (d, $J = 8.4$ Hz, 1H), 3.87 (s, 3H), 3.29–3.01 (m, 2H), 2.79–2.60 (m, 2H), 2.10–1.92 (m, 1H), 1.90–1.79 (m, 1H); ^{13}C NMR (100 MHz, CDCl_3): δ 194.8, 162.6, 138.4, 137.1, 133.4, 132.7, 130.0, 128.7, 128.1, 128.0, 127.8, 127.7, 127.6, 126.8, 113.3, 66.9, 56.7, 55.6, 28.5, 24.4; IR: 2919, 2848 1665, 1598, 1420, 1260, 742, 696; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{27}\text{H}_{27}\text{O}_2\text{S}_2$ 447.1452; found 447.1462; Daicel Chiralpak IE column, n -hexane/ i -PrOH = 95/5, 1 mL/min, 254 nm: $t_{\text{major}} = 25.3$ min, $t_{\text{minor}} = 17.0$ min.

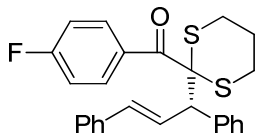


(S)-(2-(1,3-Diphenylallyl)-1,3-dithian-2-yl)(2-fluorophenyl)methanone (5f): White solid (26.1 mg, 66%). mp: 114–115 °C; $[\alpha]_D^{20} = +37.0$ (c 0.38, CHCl_3); ^1H NMR (400 MHz, CDCl_3): δ 7.81 (t, $J = 6.4$ Hz, 1H), 7.63–6.88 (m, 13H), 6.76 (dd, $J = 15.6, 8.0$ Hz, 1H), 6.15 (d, $J = 15.6$ Hz, 1H), 4.18 (d, $J = 8.0$ Hz, 1H), 3.32–2.86 (m, 2H), 2.75–2.52 (m, 2H), 2.09–1.93 (m, 1H), 1.87–1.67 (m, 1H); ^{13}C NMR (100 MHz, CDCl_3): δ 195.2, 159.8 (d, $J_{\text{C-F}} = 252.8$ Hz), 138.2, 136.8, 133.4, 131.9 (d, $J_{\text{C-F}} = 8.5$ Hz), 130.2, 129.5, 128.4, 128.0, 127.5, 127.4, 126.5, 123.2, 122.9 (d, $J_{\text{C-F}} = 3.9$ Hz), 116.4 (d, $J_{\text{C-F}} = 21.9$ Hz), 64.8, 55.9, 27.9, 27.7, 23.7; ^{19}F NMR (376 MHz, CDCl_3): δ -110.8; IR: 2920, 2848 1644, 1384, 543, 418, 404; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{26}\text{H}_{24}\text{OS}_2\text{F}$ 435.1253; found 435.1241; Daicel Chiralpak IC-3 column, n -hexane/ i -PrOH = 97/3, 1 mL/min, 254 nm: $t_{\text{major}} = 11.3$ min, $t_{\text{minor}} = 12.5$ min.

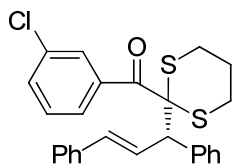


(S)-(2-(1,3-Diphenylallyl)-1,3-dithian-2-yl)(3-fluorophenyl)methanone (5g): Using $t\text{BuONa}$ (0.15 mmol) and 15-crown-5 (0.12 mmol) as base to obtain yellow oil (42.1 mg, 97%). $[\alpha]_D^{20} =$

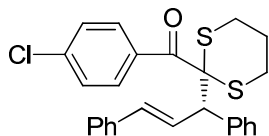
+28.5 (*c* 0.57, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.76–7.63 (m, 2H), 7.40–7.04 (m, 12H), 6.75 (dd, *J* = 16.0, 8.4 Hz, 1H), 6.10 (d, *J* = 15.6 Hz, 1H), 4.29 (d, *J* = 8.4 Hz, 1H), 3.34–3.16 (m, 1H), 3.16–3.02 (m, 1H), 2.76–2.59 (m, 2H), 2.16–1.89 (m, 1H), 1.89–1.72 (m, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 195.3, 161.7 (d, *J*_{C-F} = 245.7 Hz), 140.2, 140.1, 137.9, 136.6, 133.3, 129.8, 129.5 (d, *J*_{C-F} = 7.8 Hz), 128.5, 128.0, 127.6 (d, *J*_{C-F} = 9.0 Hz), 127.2, 126.5, 125.7 (d, *J*_{C-F} = 3.0 Hz), 118.5 (d, *J*_{C-F} = 21.1 Hz), 116.9 (d, *J*_{C-F} = 23.7 Hz), 65.7, 56.3, 28.2, 23.9; ¹⁹F NMR (376 MHz, CDCl₃): δ -50.0; IR: 2920, 2849 1645, 1507, 1488, 1384, 634, 442, 418; HRMS (APCI) [M+H]⁺ calcd for C₂₆H₂₄OS₂F 435.1253; found 435.1254; Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 98/2, 0.3 mL/min, 254 nm: *t*_{major} = 21.6 min, *t*_{minor} = 22.4 min.



(S)-(2-(1,3-Diphenylallyl)-1,3-dithian-2-yl)(4-fluorophenyl)methanone (5h): Using *t*BuONa (0.15 mmol) and 15-crown-5 (0.12 mmol) as base to obtain yellow solid (43.1 mg, 99%); mp: 146–147 °C; [α]_D²⁰ = +71.0 (*c* 0.3, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 8.12–7.93 (m, 2H), 7.44–7.10 (m, 10H), 7.00 (t, *J* = 8.8 Hz, 2H), 6.76 (dd, *J* = 16.0, 8.4 Hz, 1H), 6.11 (d, *J* = 16.0 Hz, 1H), 4.30 (d, *J* = 8.4 Hz, 1H), 3.24 (t, *J* = 13.2 Hz, 1H), 3.09 (t, *J* = 12.8 Hz, 1H), 2.83–2.56 (m, 2H), 2.10–1.93 (m, 1H), 1.92–1.72 (m, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 195.1, 164.6 (d, *J*_{C-F} = 252.2 Hz), 138.0, 136.7, 134.3 (d, *J*_{C-F} = 3.3 Hz), 133.2, 132.5 (d, *J*_{C-F} = 8.6 Hz), 129.7, 128.5, 128.0, 127.6, 127.5, 127.3, 126.5, 114.9 (d, *J*_{C-F} = 21.4 Hz), 66.0, 56.5, 28.2, 23.9; ¹⁹F NMR (376 MHz, CDCl₃): δ -107.1; IR: 2920, 2848, 1645, 1555, 1417, 442, 419; HRMS (APCI) [M+H]⁺ calcd for C₂₆H₂₄OS₂F 435.1253; found 435.1266; Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 98/2, 1 mL/min, 254 nm: *t*_{major} = 7.3 min, *t*_{minor} = 7.8 min.

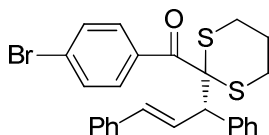


(S)-(3-Chlorophenyl)(2-(1,3-diphenylallyl)-1,3-dithian-2-yl)methanone (5i): Yellow oil (42.9 mg, 95%). [α]_D²⁰ = +27.3 (*c* 0.25, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.92 (s, 1H), 7.81 (d, *J* = 8.0 Hz, 1H), 7.44 (d, *J* = 7.9 Hz, 1H), 7.38–7.16 (m, 11H), 6.76 (dd, *J* = 16.0, 8.4 Hz, 1H), 6.11 (d, *J* = 16.0 Hz, 1H), 4.27 (d, *J* = 8.4 Hz, 1H), 3.34–3.17 (m, 1H), 3.17–3.03 (m, 1H), 2.82–2.59 (m, 2H), 2.17–1.98 (m, 1H), 1.98–1.76 (m, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 195.7, 140.2, 138.2, 136.9, 134.0, 133.6, 131.6, 130.1, 130.0, 129.4, 128.7, 128.3, 128.1, 127.8, 127.7, 127.5, 126.8, 66.0, 56.6, 28.4, 24.1; IR: 2914, 2844, 1677, 1658, 1650 1641, 1631, 1383, 531, 464, 401; HRMS (APCI) [M+H]⁺ calcd for C₂₆H₂₄OS₂Cl 451.0957; found 451.0958; Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 98/2, 0.8 mL/min, 254 nm: *t*_{major} = 8.2 min, *t*_{minor} = 8.6 min.

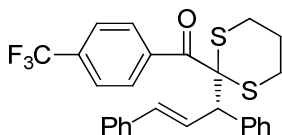


(S)-(4-Chlorophenyl)(2-(1,3-diphenylallyl)-1,3-dithian-2-yl)methanone (5j): White solid (43.3 mg, 96%). mp: 128–130 °C; [α]_D²⁰ = +50.3 (*c* 0.59, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.96 (d,

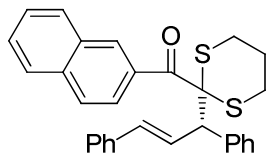
$J = 8.8$ Hz, 2H), 7.49–7.03 (m, 12H), 6.78 (dd, $J = 16.0$, 8.4 Hz, 1H), 6.14 (d, $J = 16.0$ Hz, 1H), 4.31 (d, $J = 8.4$ Hz, 1H), 3.33–3.17 (m, 1H), 3.17–3.02 (m, 1H), 2.81–2.61 (m, 2H), 2.15–1.90 (m, 1H), 1.90–1.67 (m, 1H); ^{13}C NMR (100 MHz, CDCl_3): δ 195.7, 138.2, 138.1, 136.9, 136.8, 133.6, 131.6, 130.0, 128.7, 128.4, 128.3, 127.9, 127.8, 127.5, 126.8, 66.2, 56.7, 28.4, 24.2; IR: 1658, 1650 1641, 1631, 649, 496; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{26}\text{H}_{24}\text{OS}_2\text{Cl}$ 451.0957; found 451.0960; Daicel Chiralpak IE column, n -hexane/ i -PrOH = 98/2, 1 mL/min, 254 nm: $t_{\text{major}} = 13.8$ min, $t_{\text{minor}} = 10.7$ min.



(S)-(4-Bromophenyl)(2-(1,3-diphenylallyl)-1,3-dithian-2-yl)methanone (5k): White solid (44.6 mg, 90%). mp: 143–145 °C; $[\alpha]_{\text{D}}^{20} = +43.0$ (c 0.46, CHCl_3); ^1H NMR (400 MHz, CDCl_3): δ 7.92–7.84 (m, 2H), 7.54–7.44 (m, 2H), 7.37–7.08 (m, 10H), 6.77 (dd, $J = 15.6$, 8.4 Hz, 1H), 6.13 (d, $J = 16.0$ Hz, 1H), 4.30 (d, $J = 8.4$ Hz, 1H), 3.31–3.17 (m, 1H), 3.17–3.02 (m, 1H), 2.79–2.55 (m, 2H), 2.15–1.94 (m, 1H), 1.94–1.76 (m, 1H); ^{13}C NMR (100 MHz, CDCl_3): δ 195.9, 138.2, 137.2, 136.9, 133.6, 131.7, 131.3, 130.0, 128.7, 128.3, 127.9, 127.8, 127.5, 126.8, 126.6, 66.2, 56.7, 28.4, 24.2; IR: 2922, 2853, 1643, 1384 744, 526, 518, 501; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{26}\text{H}_{24}\text{OS}_2\text{Br}$ 495.0452; found 495.0459; Daicel Chiralpak IE column, n -hexane/ i -PrOH = 98/2, 1 mL/min, 254 nm: $t_{\text{major}} = 14.5$ min, $t_{\text{minor}} = 11.3$ min.

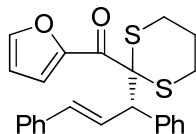


(S)-(2-(1,3-Diphenylallyl)-1,3-dithian-2-yl)(4-(trifluoromethyl)phenyl)methanone (5l): Yellow oil (44.6 mg, 92%). $[\alpha]_{\text{D}}^{20} = +30.2$ (c 0.28, CHCl_3); ^1H NMR (400 MHz, CDCl_3): δ 7.98 (d, $J = 8.0$ Hz, 2H), 7.55 (d, $J = 8.4$ Hz, 2H), 7.35–6.96 (m, 10H), 6.74 (dd, $J = 15.6$, 8.0 Hz, 1H), 6.08 (d, $J = 16.0$ Hz, 1H), 4.20 (d, $J = 8.4$ Hz, 1H), 3.34–3.19 (m, 1H), 3.18–3.04 (m, 1H), 2.81–2.61 (m, 2H), 2.16–1.94 (m, 1H), 1.94–1.75 (m, 1H); ^{13}C NMR (100 MHz, CDCl_3): δ 196.4, 141.9, 138.2, 136.8, 133.7, 133.0 (q, $J_{\text{C-F}} = 32.6$ Hz), 130.2, 130.0, 128.7, 128.3, 127.9, 127.8, 127.3, 126.8, 126.6 (q, $J_{\text{C-F}} = 271.0$ Hz), 125.0 (d, $J_{\text{C-F}} = 3.3$ Hz), 65.6, 56.6, 28.4, 24.0; ^{19}F NMR (376 MHz, CDCl_3): δ -63.0; IR: 2922, 2850, 1639, 1507, 1471, 1384, 739, 704, 517, 457, 419; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{27}\text{H}_{24}\text{OS}_2\text{F}_3$ 485.1221; found 485.1218; Daicel Chiralpak IC-3 column, n -hexane/ i -PrOH = 99/1, 0.6 mL/min, 254 nm: $t_{\text{major}} = 9.4$ min, $t_{\text{minor}} = 9.9$ min.

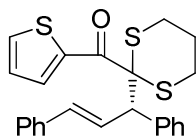


(S)-(2-(1,3-Diphenylallyl)-1,3-dithian-2-yl)(naphthalen-2-yl)methanone (5m): Yellow oil (45.7 mg, 98%). $[\alpha]_{\text{D}}^{20} = +65.8$ (c 0.66, CHCl_3); ^1H NMR (400 MHz, CDCl_3): δ 8.74 (s, 1H), 8.01–7.48 (m, 7H), 7.38–7.11 (m, 9H), 6.82 (dd, $J = 15.6$, 8.4 Hz, 1H), 6.11 (d, $J = 15.6$ Hz, 1H), 4.51 (d, $J = 8.8$ Hz, 1H), 3.38–3.09 (m, 2H), 2.90–2.57 (m, 2H), 2.12–1.98 (m, 1H), 1.96–1.78 (m, 1H); ^{13}C NMR (100 MHz, CDCl_3): δ 196.9, 138.4, 137.0, 135.7, 134.8, 133.6, 132.4, 131.0, 130.2, 129.8, 128.7, 128.3, 128.2, 127.9, 127.8, 127.7, 126.8, 66.7, 56.7, 28.5, 24.3; IR: 2920, 2847, 1650, 1641,

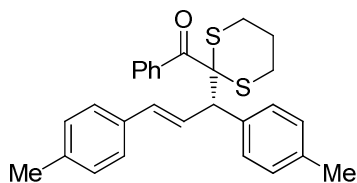
1631, 1384, 610, 408; HRMS (APCI) $[M+H]^+$ calcd for $C_{30}H_{27}OS_2$ 467.1503; found 467.1516; Daicel Chiralpak IE column, *n*-hexane/*i*-PrOH = 95/5, 1 mL/min, 254 nm: t_{major} = 17.2 min, t_{minor} = 12.4 min.



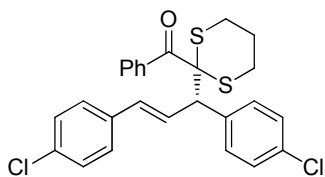
(S)-(2-(1,3-Diphenylallyl)-1,3-dithian-2-yl)(furan-2-yl)methanone (5n): Yellow oil (40.2 mg, 99%). $[\alpha]_D^{20}$ = +38.7 (*c* 0.46, $CHCl_3$); 1H NMR (400 MHz, $CDCl_3$): δ 7.73–7.13 (m, 12H), 6.84 (dd, J = 16.0, 8.8 Hz, 1H), 6.50 (dd, J = 3.2, 1.6 Hz, 1H), 6.32 (d, J = 16.0 Hz, 1H), 4.71 (d, J = 8.4 Hz, 1H), 3.22–3.03 (m, 1H), 3.00–2.80 (m, 1H), 2.74–2.54 (m, 2H), 2.01–1.87 (m, 1H), 1.87–1.72 (m, 1H); ^{13}C NMR (100 MHz, $CDCl_3$): δ 182.3, 151.1, 145.4, 138.1, 136.9, 133.1, 129.4, 128.5, 128.0, 127.7, 127.6, 127.5, 126.6, 120.5, 112.0, 66.4, 55.3, 28.2, 28.0, 24.0; IR: 2920, 2847, 1631, 1384, 496; HRMS (APCI) $[M+H]^+$ calcd for $C_{24}H_{23}O_2S_2$ 407.1139; found 407.1149; Daicel Chiralpak IE column, *n*-hexane/*i*-PrOH = 95/5, 1 mL/min, 254 nm: t_{major} = 24.8 min, t_{minor} = 16.2 min.



(S)-(2-(1,3-Diphenylallyl)-1,3-dithian-2-yl)(thiophen-2-yl)methanone (5o): White solid (41.8 mg, 99%). mp: 108–110 °C; $[\alpha]_D^{20}$ = +45.7 (*c* 0.28, $CHCl_3$); 1H NMR (400 MHz, $CDCl_3$): δ 8.20–8.07 (m, 1H), 7.67–7.53 (m, 1H), 7.44–7.12 (m, 10H), 7.13–7.09 (m, 1H), 6.80 (dd, J = 15.6, 8.8 Hz, 1H), 6.28 (d, J = 15.6 Hz, 1H), 4.49 (d, J = 8.4 Hz, 1H), 3.19–2.82 (m, 2H), 2.81–2.48 (m, 2H), 2.05–1.91 (m, 1H), 1.90–1.74 (m, 1H); ^{13}C NMR (100 MHz, $CDCl_3$): δ 187.9, 140.3, 138.0, 137.0, 135.6, 133.7, 133.6, 129.9, 128.7, 128.2, 127.9, 127.8, 127.6, 127.4, 126.8, 68.4, 57.1, 28.5, 28.4, 24.3; IR: 2922, 2850, 1640, 1384 457, 441, 409; HRMS (APCI) $[M+H]^+$ calcd for $C_{24}H_{23}OS_3$ 423.0911; found 423.0927; Daicel Chiralpak IE column, *n*-hexane/*i*-PrOH = 95/5, 1 mL/min, 254 nm: t_{major} = 22.8 min, t_{minor} = 13.6 min.

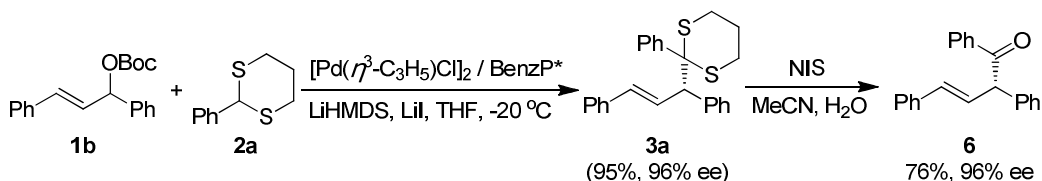


(S)-(2-(1,3-Di-*p*-tolylallyl)-1,3-dithian-2-yl)(phenyl)methanone (5p): Yellow oli (40.9 mg, 92%). $[\alpha]_D^{20}$ = +17.4 (*c* 0.46, $CHCl_3$); 1H NMR (400 MHz, $CDCl_3$): δ 8.05 (d, J = 7.2 Hz, 2H), 7.51–7.43 (m, 1H), 7.41–7.31 (m, 2H), 7.18 (d, J = 8.0 Hz, 2H), 7.11 (d, J = 8.0 Hz, 2H), 7.05 (d, J = 7.6 Hz, 2H), 7.01 (d, J = 8.0 Hz, 2H), 6.69 (dd, J = 15.6, 8.4 Hz, 1H), 6.03 (d, J = 16.0 Hz, 1H), 4.37 (d, J = 8.4 Hz, 1H), 3.36–3.02 (m, 2H), 2.79–2.54 (m, 2H), 2.30 (s, 3H), 2.28 (s, 3H), 2.05–1.92 (m, 1H), 1.86–1.71 (m, 1H); ^{13}C NMR (100 MHz, $CDCl_3$): δ 196.6, 138.2, 137.3, 137.1, 135.2, 134.0, 133.1, 131.6, 130.1, 129.7, 129.1, 128.6, 127.9, 126.4, 126.2, 66.2, 55.7, 28.1, 24.0, 21.2, 21.1; IR: 3011, 2797, 1867, 1640, 1338, 457, 441, 419; HRMS (APCI) $[M+H]^+$ calcd for $C_{28}H_{29}OS_2$ 445.1660; found 445.1664; Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 98/2, 1 mL/min, 254 nm: t_{major} = 8.8 min, t_{minor} = 10.9 min.



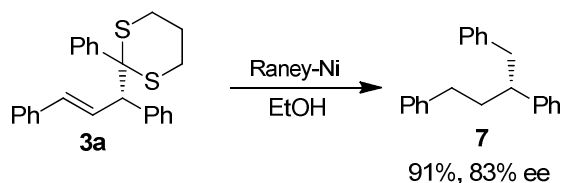
(S)-(2-(1,3-Di(4-chlorophenyl)allyl)-1,3-dithian-2-yl)(phenyl)methanone (5q): Yellow oil (46.6 mg, 96%). $[\alpha]_D^{20} = +15.3$ (*c* 0.20, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.98 (d, *J* = 7.6 Hz, 2H), 7.51 – 7.44 (m, 1H), 7.38 – 7.31 (m, 2H), 7.23 – 7.09 (m, 8H), 6.68 (dd, *J* = 15.6, 8.4 Hz, 1H), 5.97 (d, *J* = 16.0 Hz, 1H), 4.34 (d, *J* = 8.4 Hz, 1H), 3.33 – 3.09 (m, 2H), 2.79 – 2.61 (m, 2H), 2.06 – 1.97 (m, 1H), 1.89 – 1.70 (m, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 196.1, 138.0, 136.4, 135.1, 133.4, 133.3, 132.2, 131.8, 131.2, 129.9, 128.6, 128.1, 128.0, 127.7, 127.6, 65.4, 55.5, 28.1, 23.9; IR: 3566, 2922, 2850, 1868, 1637, 1385 456, 419; HRMS (APCI) $[M+H]^+$ calcd for C₂₆H₂₃OS₂Cl₂ 485.0567; found 485.0577; Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 98/2, 1 mL/min, 254 nm: *t*_{major} = 8.1 min, *t*_{minor} = 8.7 min.

6. Large Scale Synthesis of 3a and the Derivatization of 3a and 5a

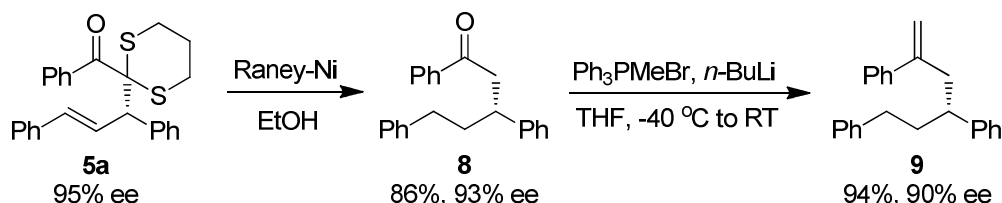


To an oven-dried glassware were added $[Pd(\eta^3-C_3H_5)Cl]_2$ (36 mg, 2.5 mol%), BenzP* (64 mg, 5.5 mol%), and anhydrous THF (20.0 mL). To another dry glassware were added **2a** (8 mmol, 1.57 g), ultra-dry LiI (4 mmol, 532 mg), and anhydrous THF (20 mL). Then LiHMDS (1M in THF, 6 mL) was added to the second glassware. The mixture was stirred at RT under a nitrogen atmosphere for 1 h. Then a solution of allylic carbonate **1b** (4 mmol, 1.24 g) in anhydrous THF (20 mL) was added to the first glassware. The solution in the first glassware was added to another one. The reaction mixture was stirred at $-20\text{ }^\circ\text{C}$ for 12 h, quenched with H₂O, and extracted with EtOAc (80 mL $\times$ 3). The combined organic layer was washed with brine, dried over anhydrous Na₂SO₄, and then concentrated in vacuum. The residue was purified by preparative HPLC (*n*-hexane/*i*-PrOH = 10:1) to give the product **3a** as a white solid (1.48 g, 95%, 96% ee).

To a solution of **3a** (77.6 mg, 0.2 mmol) in MeCN (9.0 mL) and water (0.5 mL) at $-20\text{ }^\circ\text{C}$ was added NIS (112.5 mg, 0.5 mmol), and the mixture was stirred and warmed to RT. The reaction was monitored by TLC until full consumption of **3a**. The reaction mixture was diluted with DCM, washed with brine, dried over anhydrous Na₂SO₄, and concentrated in vacuum. The residue was purified by flash column chromatography to give the compound **6** as a white solid (45.3 mg, 76%, 96% ee, the column chromatography purification should be quickly carried out to avoid the racemization of **6**). mp: 43–45 $^\circ\text{C}$; $[\alpha]_D^{20} = +24.4$ (*c* 0.10, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 8.01 (d, *J* = 7.2 Hz, 2H), 7.65–7.07 (m, 13H), 6.76 (dd, *J* = 15.6, 8.0 Hz, 1H), 6.46 (d, *J* = 15.6 Hz, 1H), 5.45 (d, *J* = 8.0 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 198.8, 138.9, 137.0, 136.6, 133.4, 132.3, 129.3, 129.1, 128.9, 128.8, 128.7, 128.6, 127.8, 127.5, 126.7, 57.5; IR: 2921, 2850, 1644, 1384 696, 517, 456; HRMS (APCI) $[M+H]^+$ calcd for C₂₂H₁₉O 299.1436; found 299.1449; Daicel Chiralpak AD-H column, *n*-hexane/*i*-PrOH = 98/2, 1 mL/min, 254 nm: *t*_{major} = 24.8 min, *t*_{minor} = 28.4 min.



To a solution of **3a** (38.8 mg, 0.1 mmol) in EtOH (15 mL) was added Raney-Ni (water slurry, 2 mL). The reaction was monitored by TLC until full consumption of **3a**. The reaction mixture was filtrated, dried over anhydrous Na₂SO₄, and concentrated in vacuum. The residue was purified by flash column chromatography to give the compound **7** as yellow oil (26.1 mg, 91%, 83% ee). [α]_D²⁰ = -29.9 (*c* 0.10, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.33–7.08 (m, 11H), 7.05 (d, *J* = 7.6 Hz, 2H), 6.99 (d, *J* = 7.6 Hz, 2H), 2.99–2.76 (m, 3H), 2.55–2.32 (m, 2H), 2.10–1.85 (m, 2H).^[5] Daicel Chiralpak OD-H column, *n*-hexane, 1 mL/min, 220 nm: *t*_{major} = 46.8 min, *t*_{minor} = 30.7 min.



To a solution of **5a** (83.1 mg, 0.2 mmol) in EtOH (8.0 mL) was added Raney-Ni (water slurry, 6 mL), and the mixture was stirred at RT for 2 h. The reaction mixture was filtrated, dried over anhydrous Na₂SO₄, and concentrated in vacuum. The residue was purified by flash column chromatography to give the compound **8** as a white solid (54.1 mg, 86%, 93% ee).^[6] [α]_D²⁰ = -5.0 (*c* 0.41, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.85 (d, *J* = 7.2 Hz, 2H), 7.55–6.98 (m, 13H), 3.45–3.15 (m, 3H), 2.52–2.41 (m, 2H), 2.15–2.03 (m, 1H), 2.03–1.85 (m, 1H); Daicel Chiralpak OD-H column, *n*-hexane/*i*-PrOH = 98/2, 1 mL/min, 254 nm: *t*_{major} = 12.3 min, *t*_{minor} = 16.3 min.

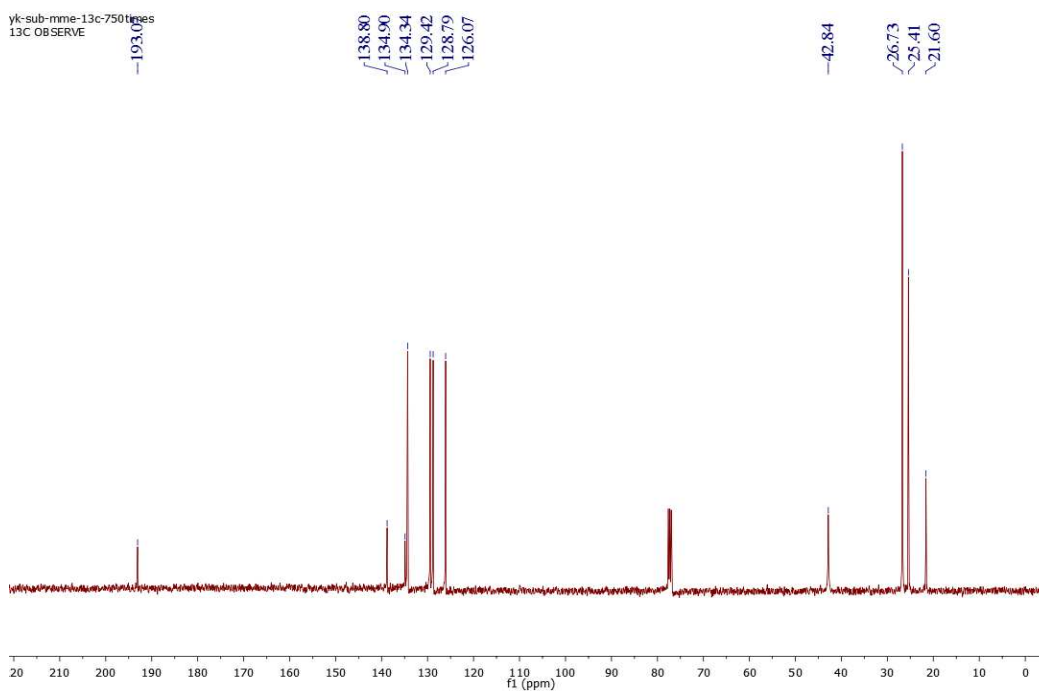
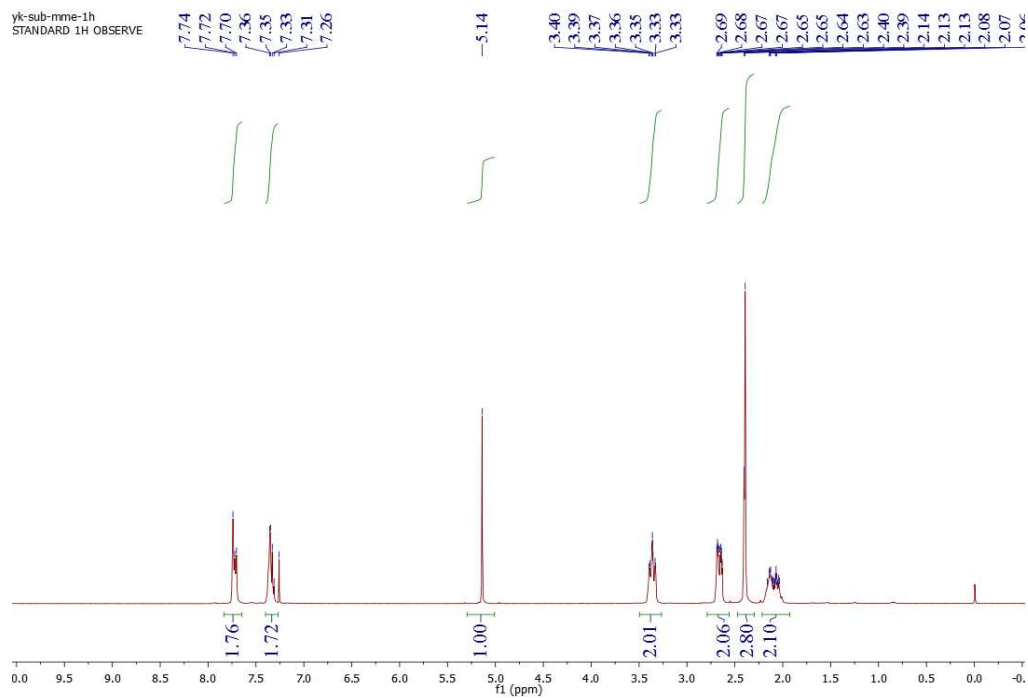
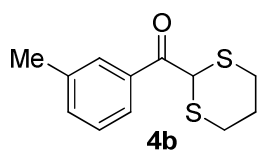
To a solution of Ph₃PMeBr (107.1 mg, 0.3 mmol) in anhydrous THF (2 mL) was added *n*-BuLi (0.25 mmol, 2.5 M in hexane) dropwise at -40 °C, and the mixture was stirred for 1 h. Then a solution of **8** (31.4 mg, 0.1 mmol) in anhydrous THF (2.0 mL) was added dropwise to the above solution. The reaction mixture was warmed to RT and stirred overnight, filtrated through celite, and concentrated in vacuum. The residue was purified by flash column chromatography to give the compound **9** as colorless oil (29.2 mg, 94%, 90% ee).^[7] [α]_D²⁰ = -2.0 (*c* 0.30, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.34–7.00 (m, 15H), 5.15 (d, *J* = 1.6 Hz, 1H), 4.89 (d, *J* = 1.2 Hz, 1H), 2.88–2.75 (m, 2H), 2.73–2.63 (m, 1H), 2.46–2.28 (m, 2H), 2.10–1.98 (m, 1H), 1.94–1.81 (m, 1H); Daicel Chiralpak OD-H column, *n*-hexane, 1 mL/min, 254 nm: *t*_{major} = 44.0 min, *t*_{minor} = 32.0 min.

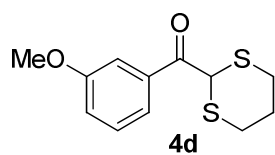
7. Reference

- [1] Samajdar, S.; Basu, M. K.; Becker, F. F.; Banik, B. K. *Tetrahedron Lett.* **2001**, 42, 4425.
- [2] Page, P. C. B.; Wilkes, R. D.; Namwindwa, E. S.; Witty, M. J. *Tetrahedron* **1996**, 52, 2125.
- [3] Balasubramaniam, S.; Kommidi, H.; Aidhen, I. S. *Tetrahedron Lett.* **2011**, 52, 2683.
- [4] A byproduct was also formed and identified as hexa-1,5-diene-1,3,4,6-tetrayltetraabenzene. ¹H NMR (400 MHz, CDCl₃): δ 7.38–7.05 (m, 20H), 6.53 (dd, *J* = 15.6, 5.6 Hz, 1H), 6.42 (d, *J* = 16 Hz, 1H), 6.33 (dd, *J* = 15.6, 5.2 Hz, 1H), 6.22 (d, *J* = 16 Hz, 1H), 3.94–3.83 (m, 2H). The spectrum is consistent with the published data, see: Shirakawa, E.; Hironaka, K.; Otsuka, H.; Hayashi, T. *Chem. Commun.* 2006, 3927.

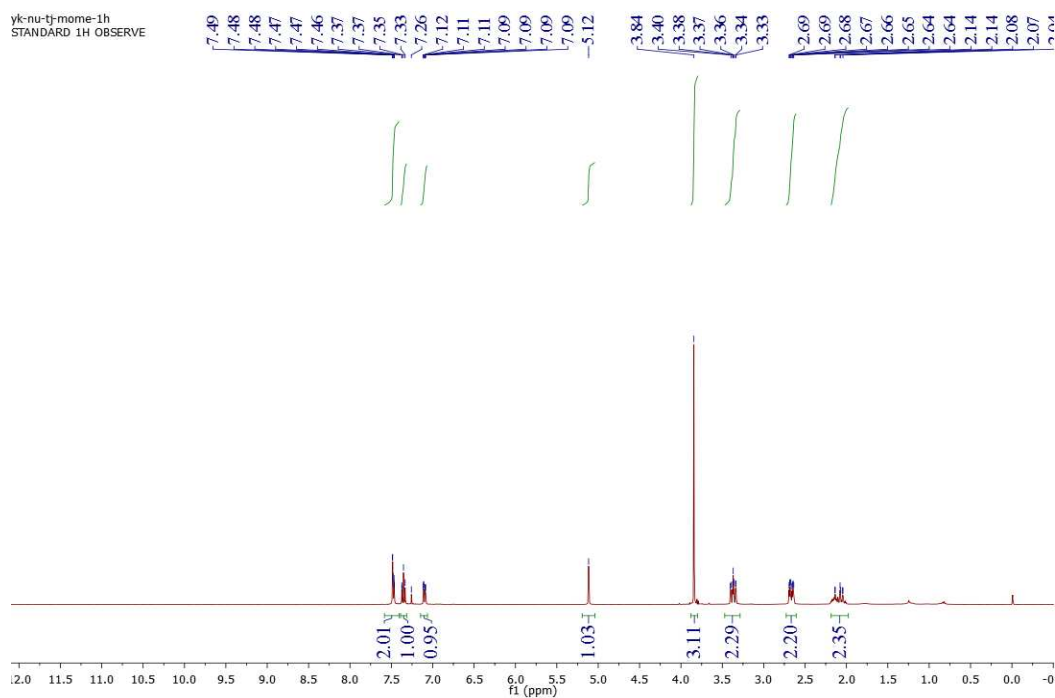
- [5] Prementine, G. S.; Tirrell, D. A. *Macromolecules* **1989**, *22*, 52.
- [6] Tottleben, M. J.; Curran, D. P.; Wipf, P. *J. Org. Chem.* **1992**, *57*, 1740.
- [7] Ellis, G. A.; T. Wyche, P.; Fry, C. G.; Braun, D. R.; Bugni, T. S. *Mar. Drugs* **2014**, *12*, 1013.

8. NMR Spectra

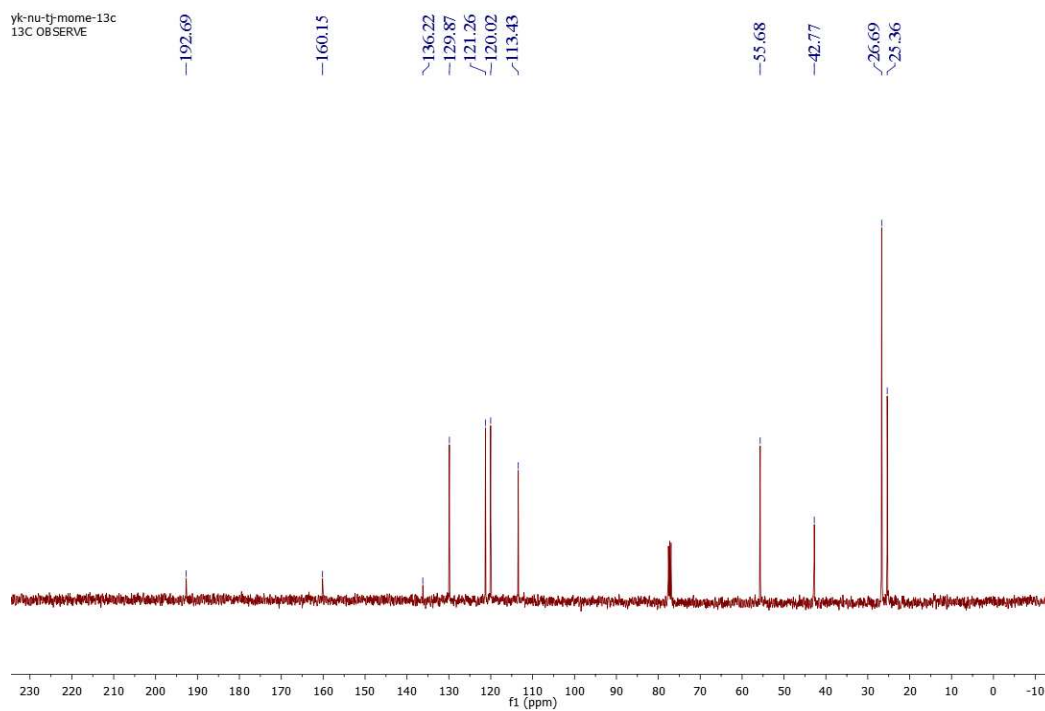


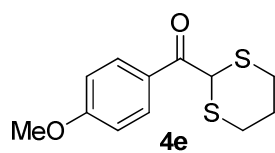


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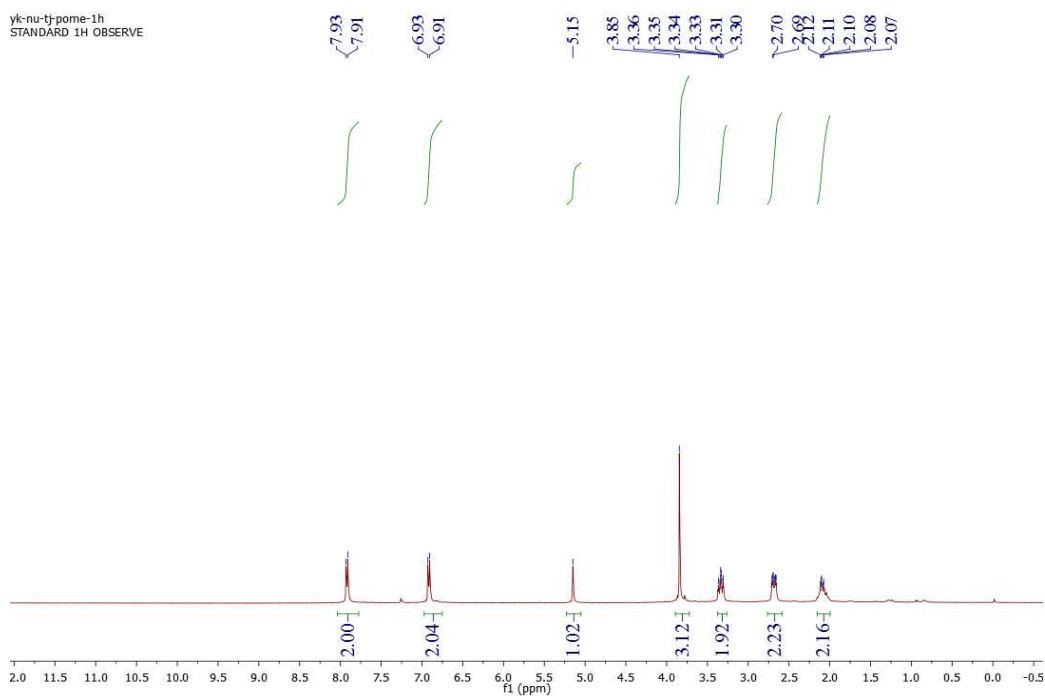


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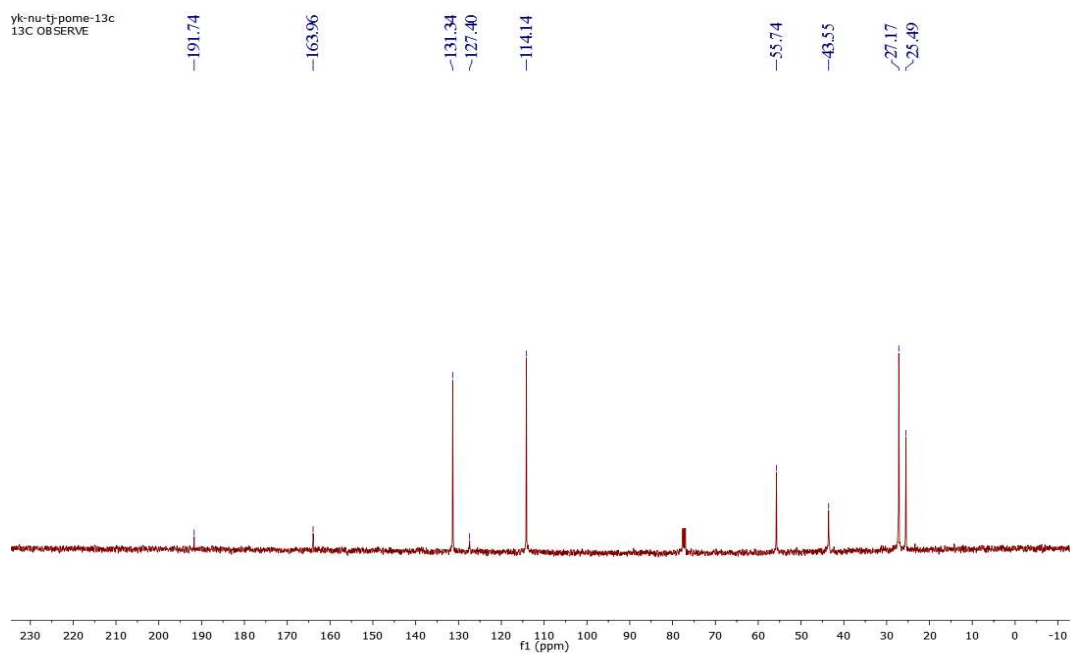


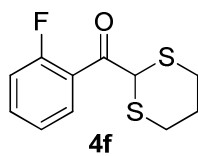


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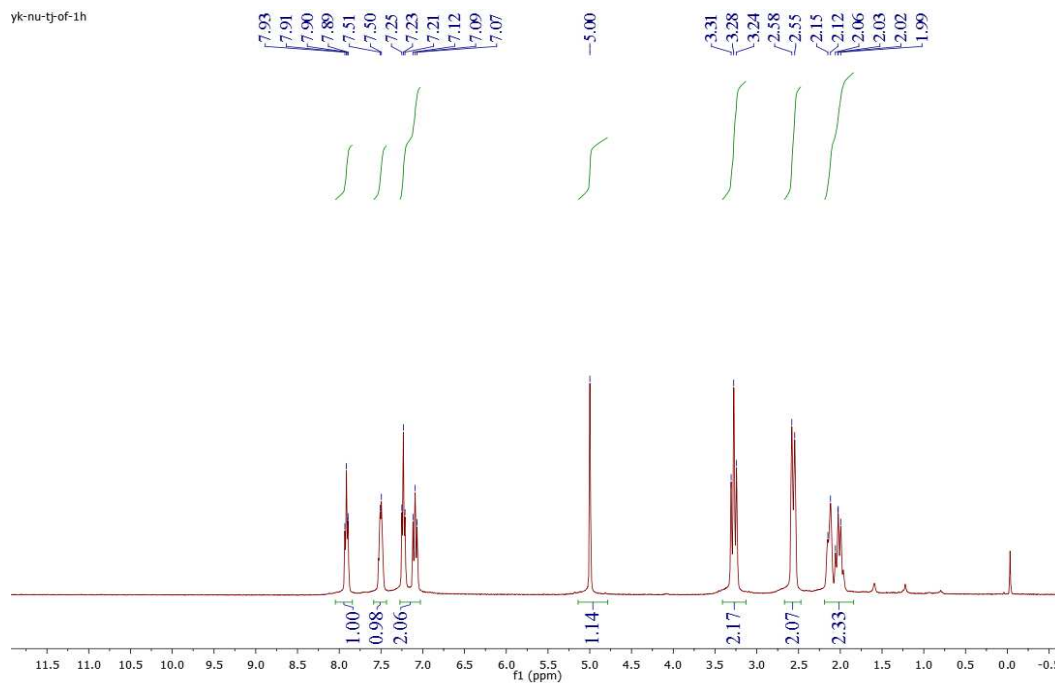


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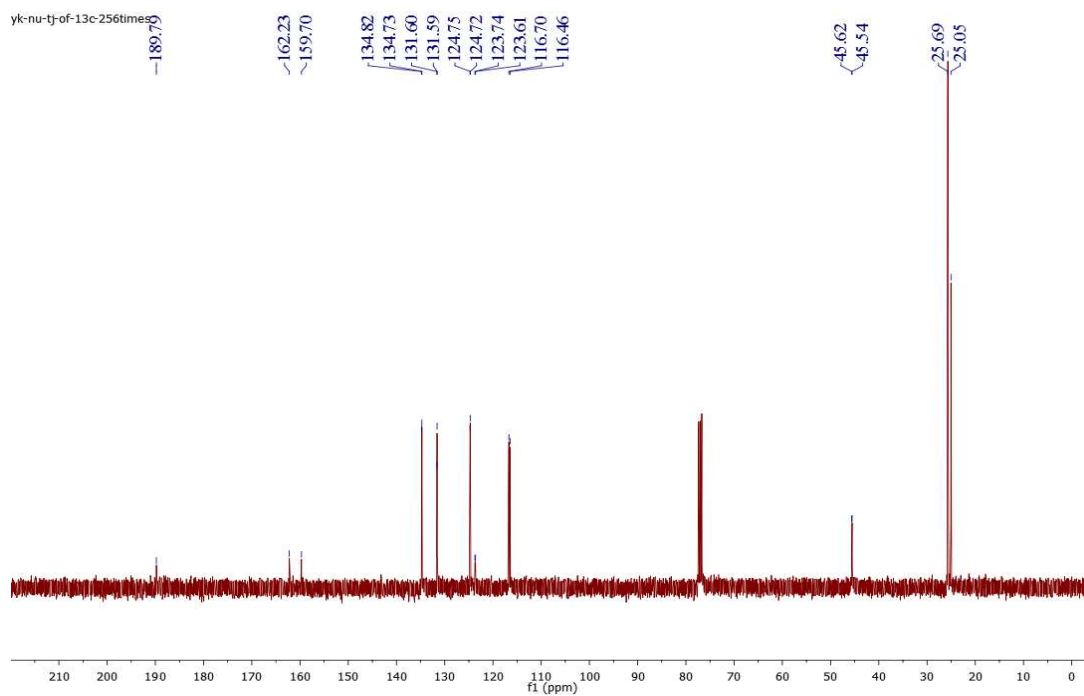


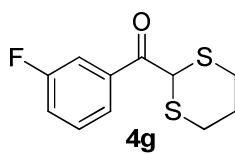


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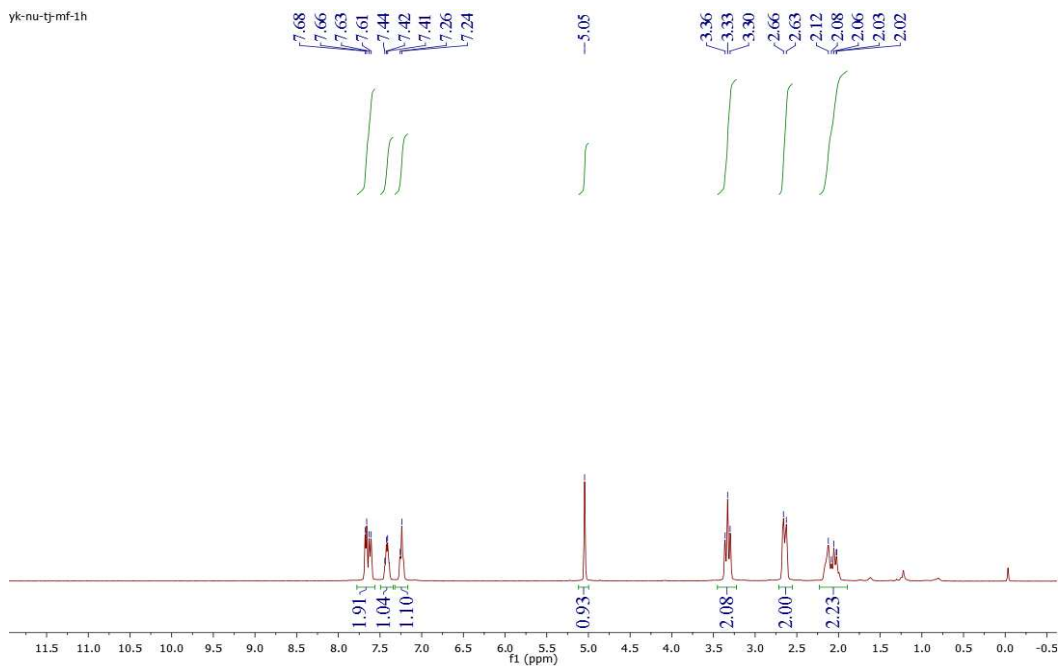


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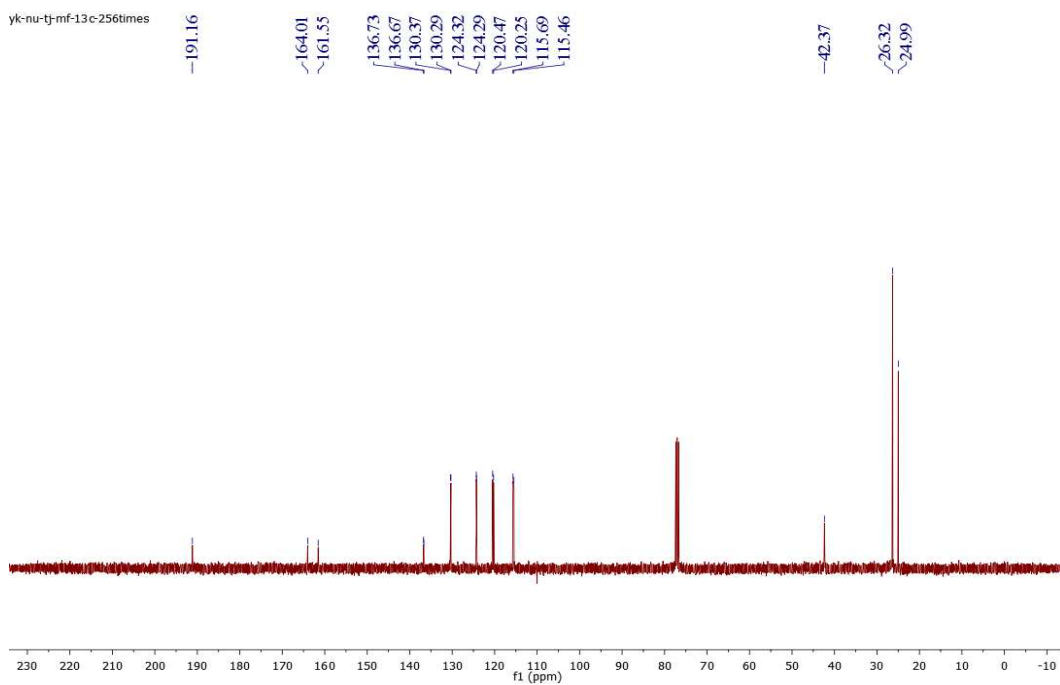


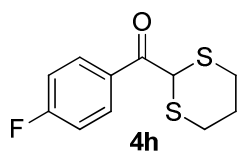


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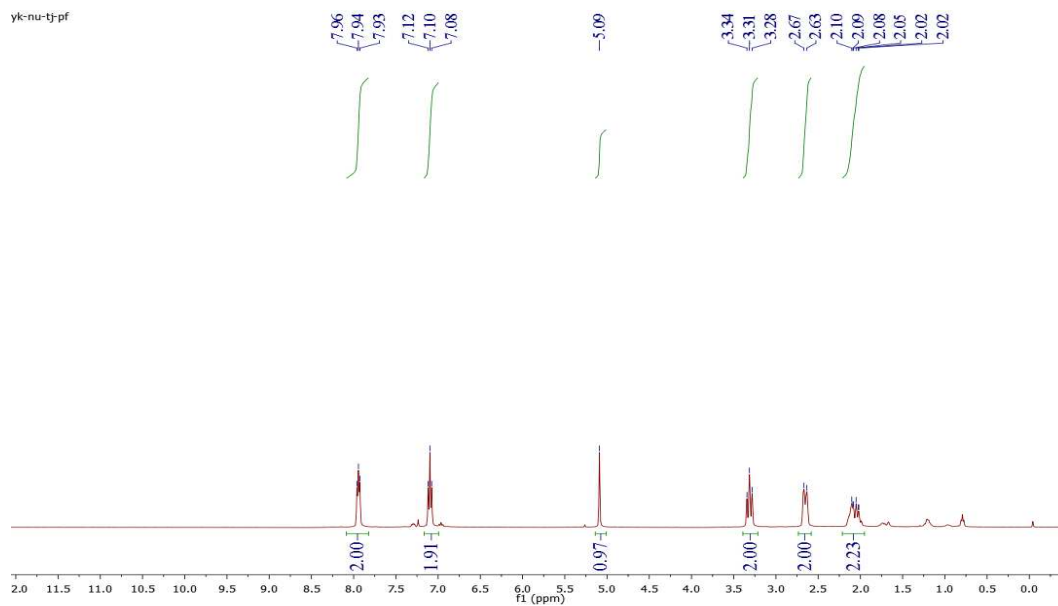


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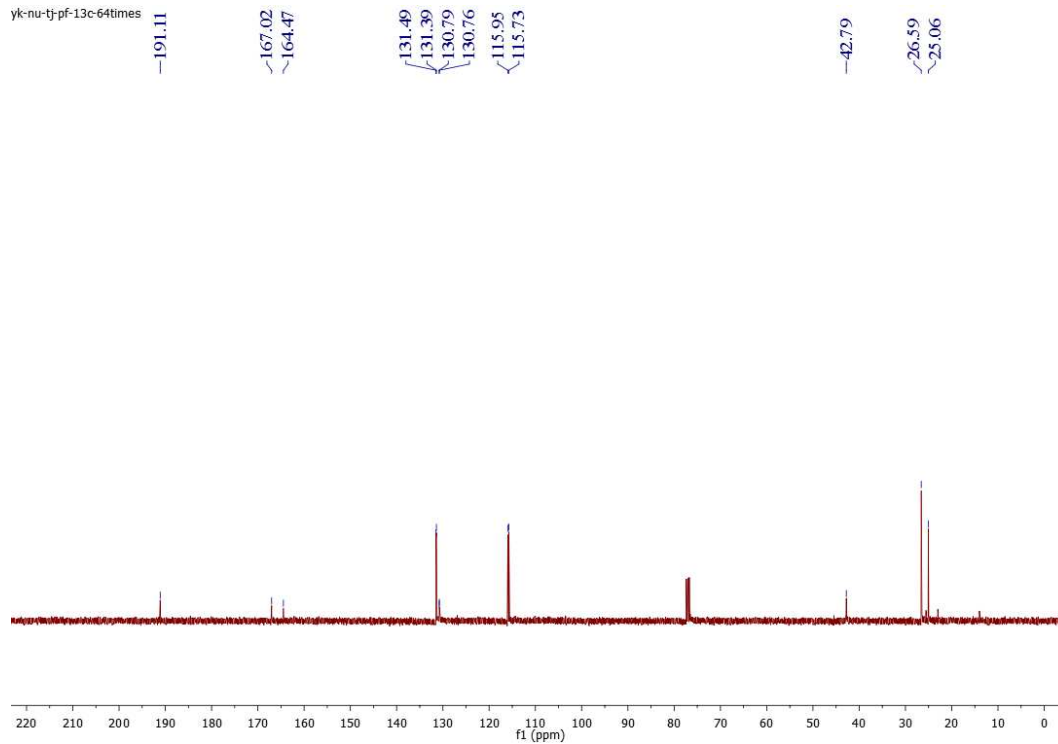


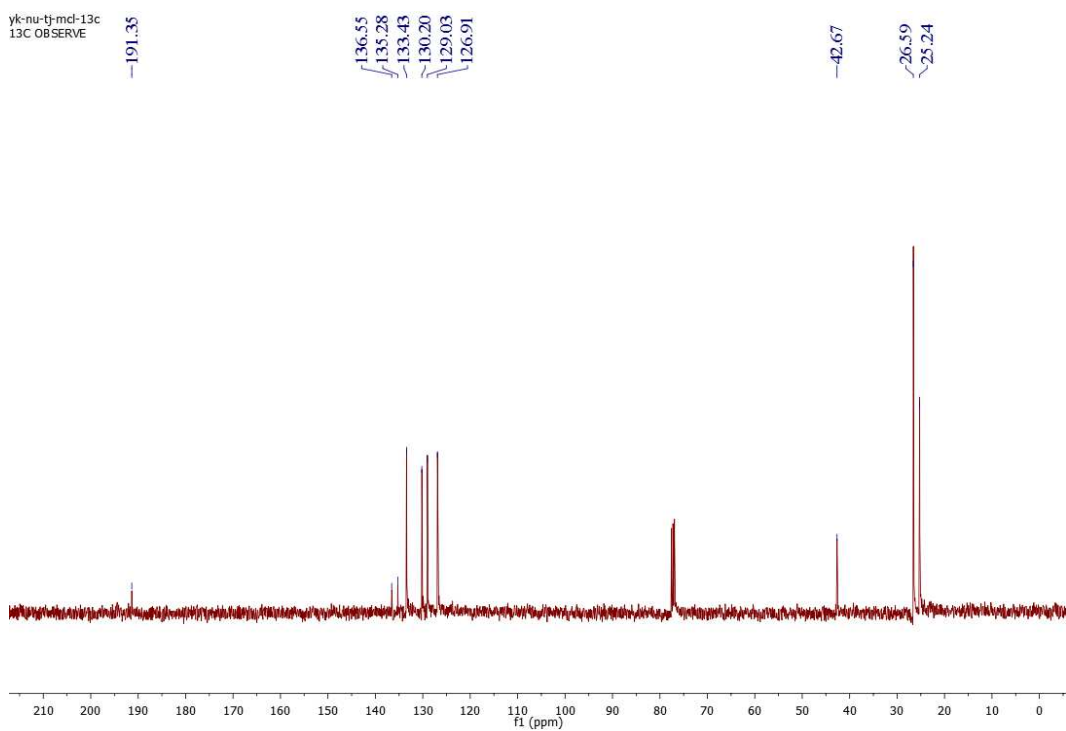
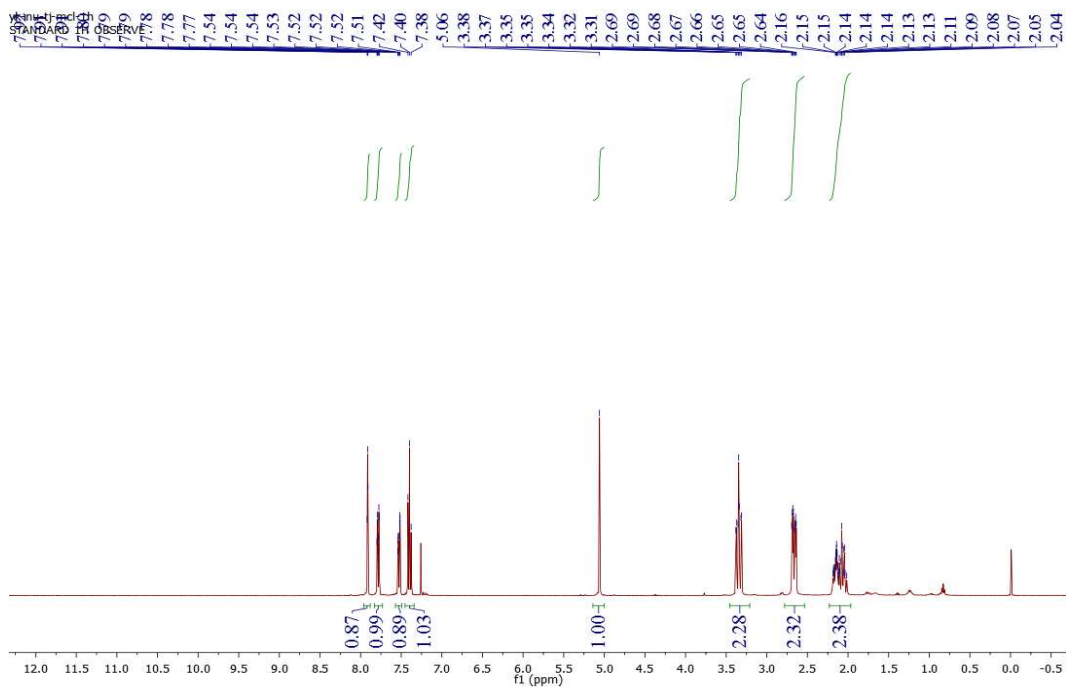
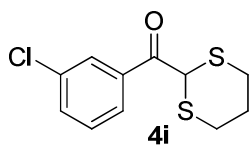


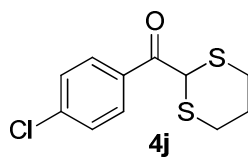
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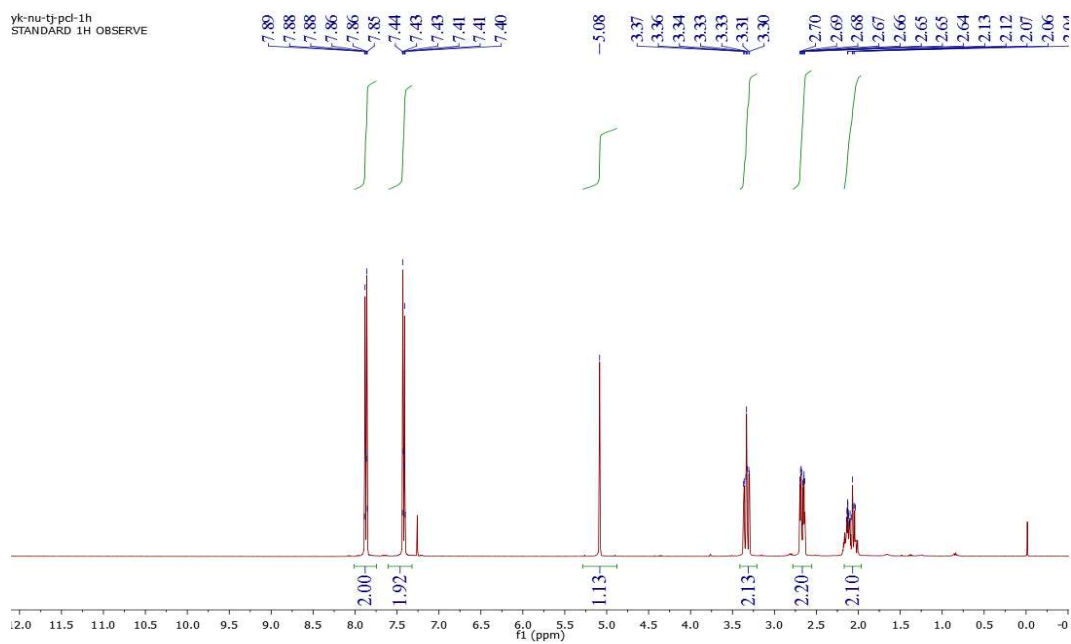
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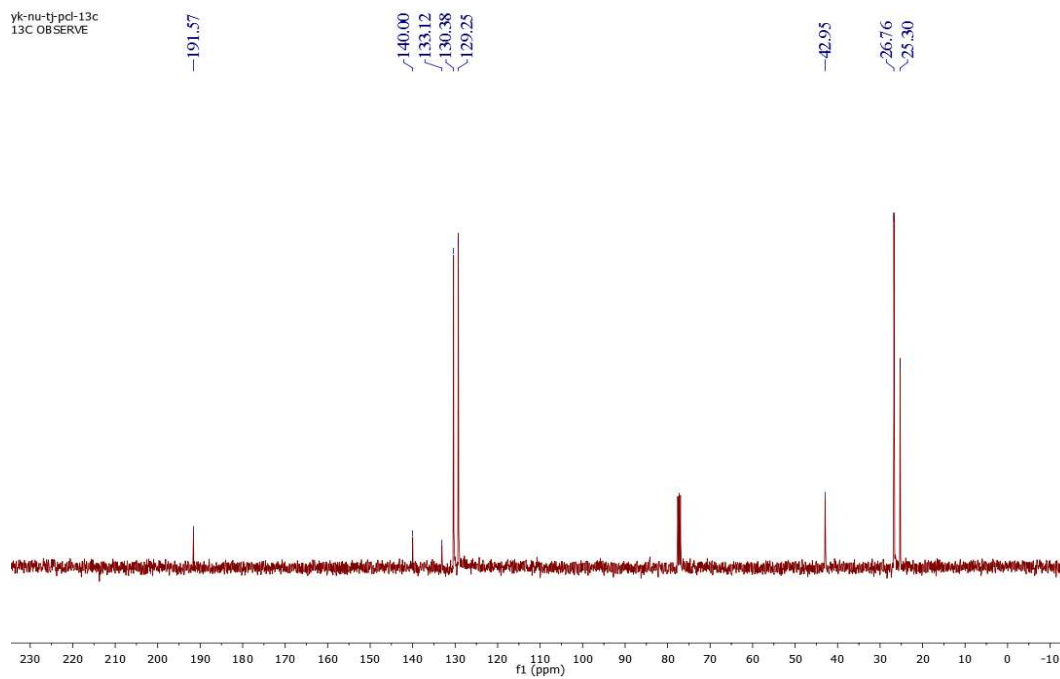


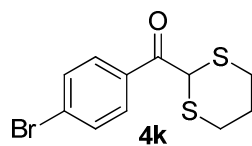


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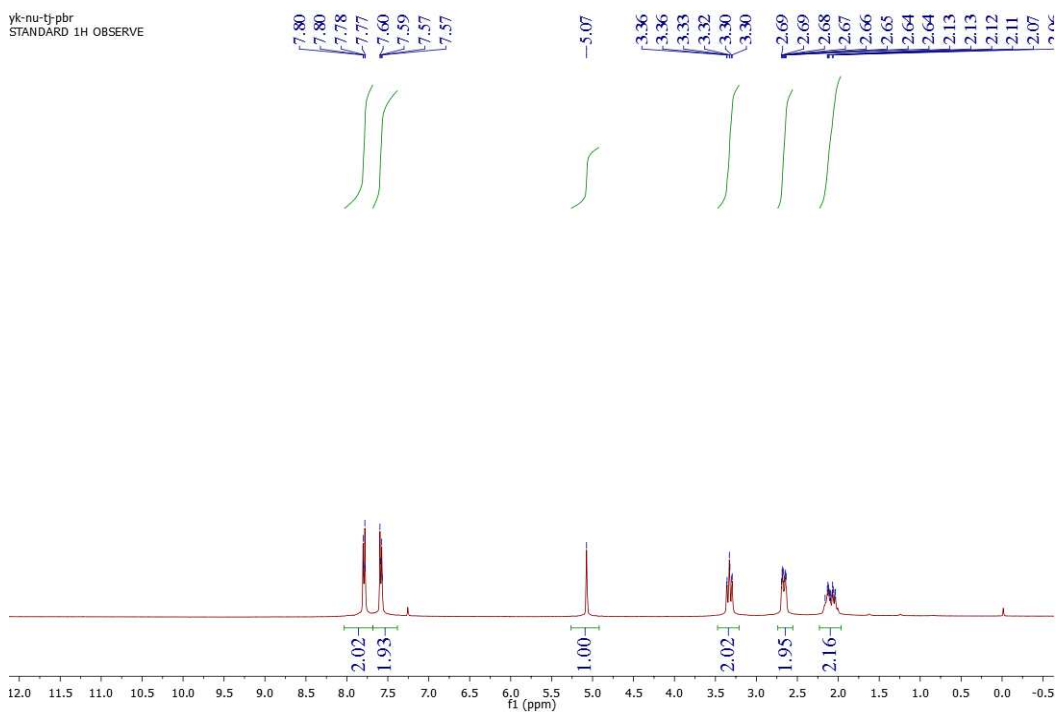


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13C OBSERVE

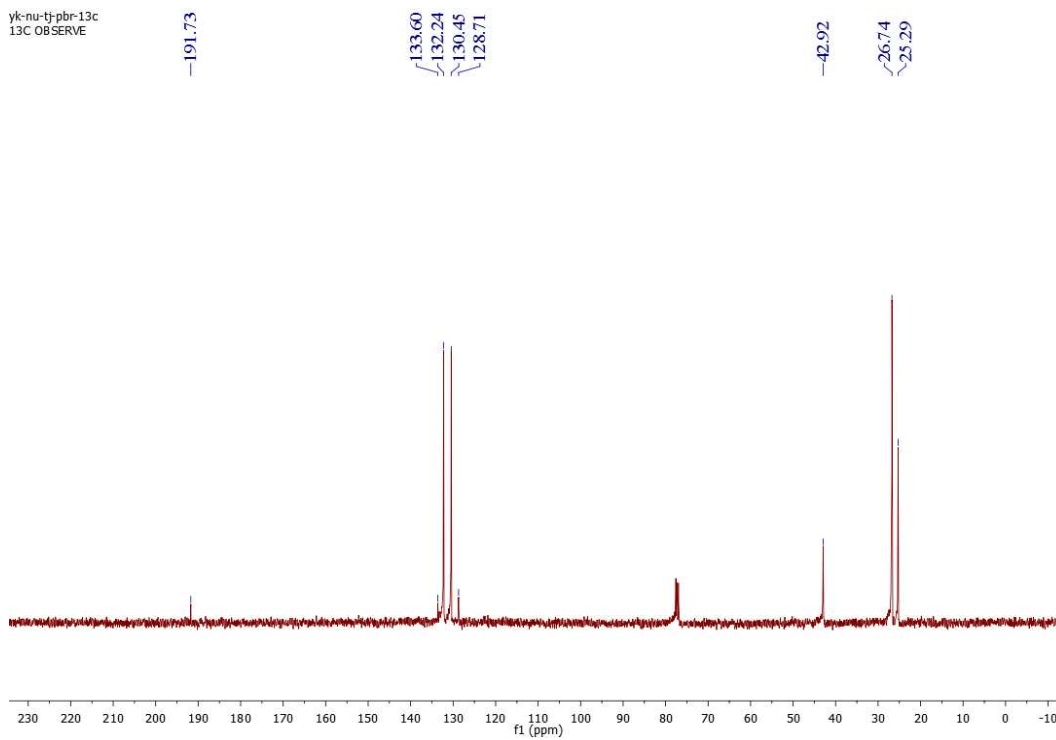


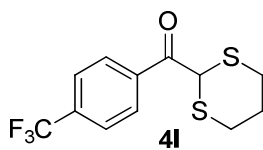


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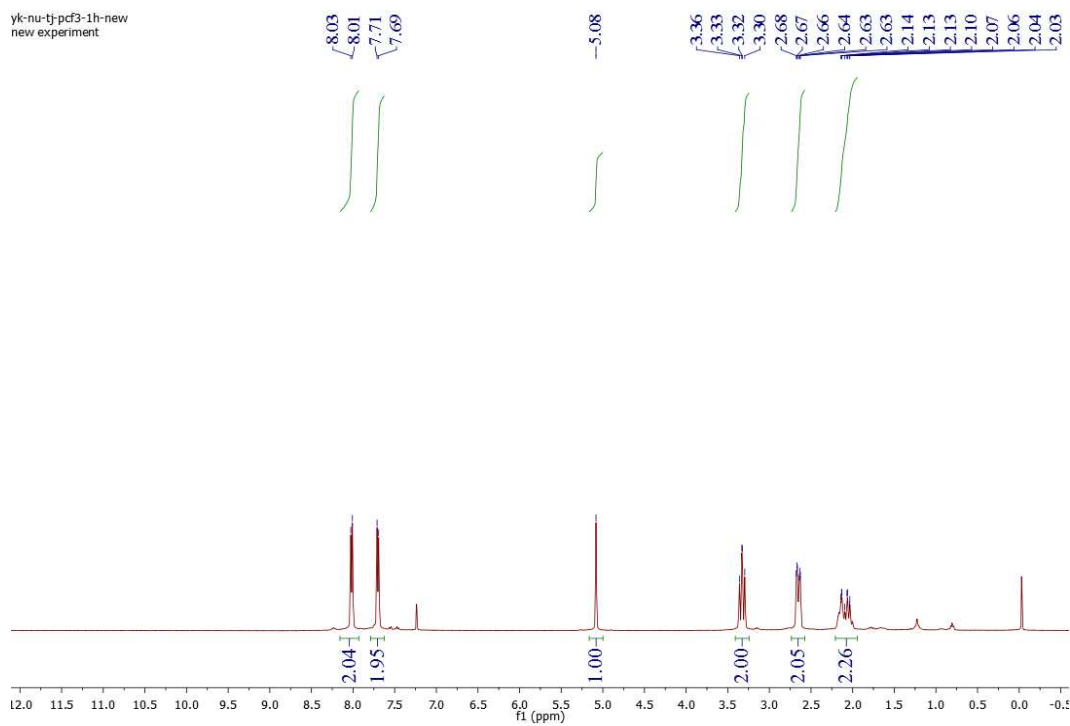


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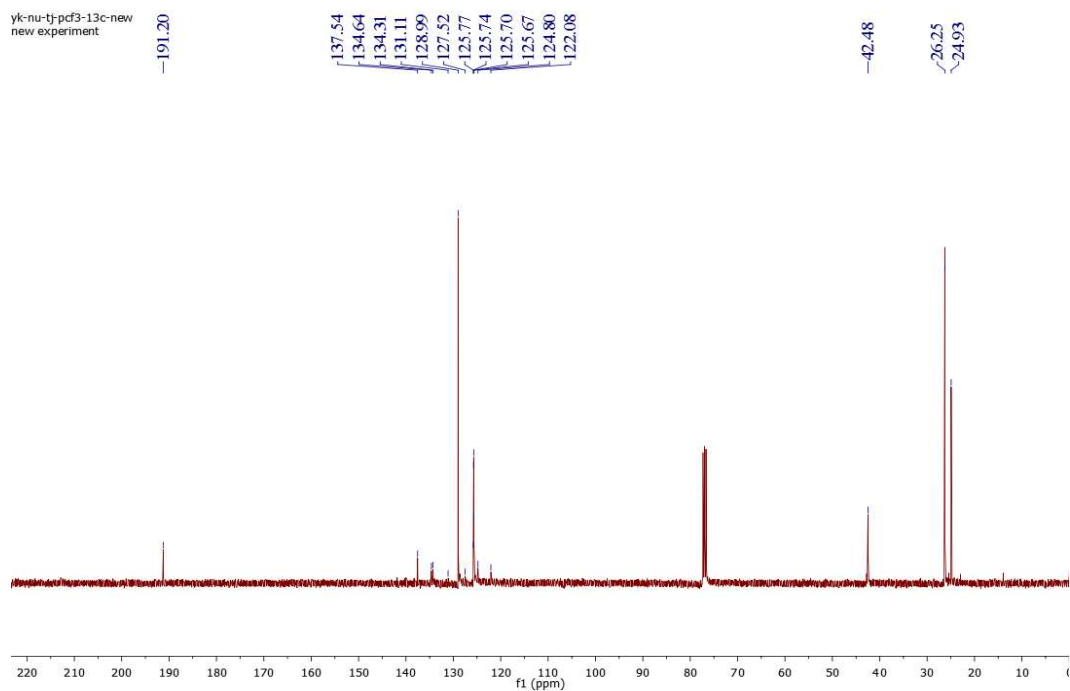


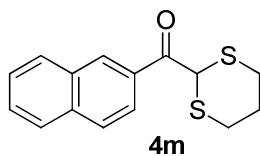


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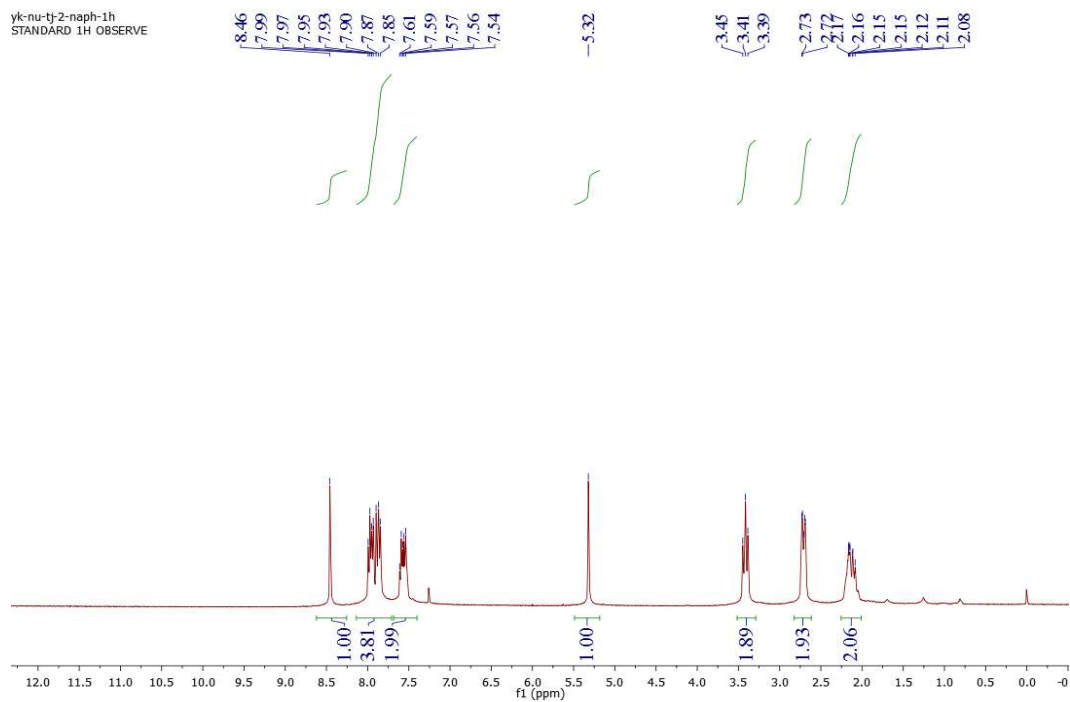


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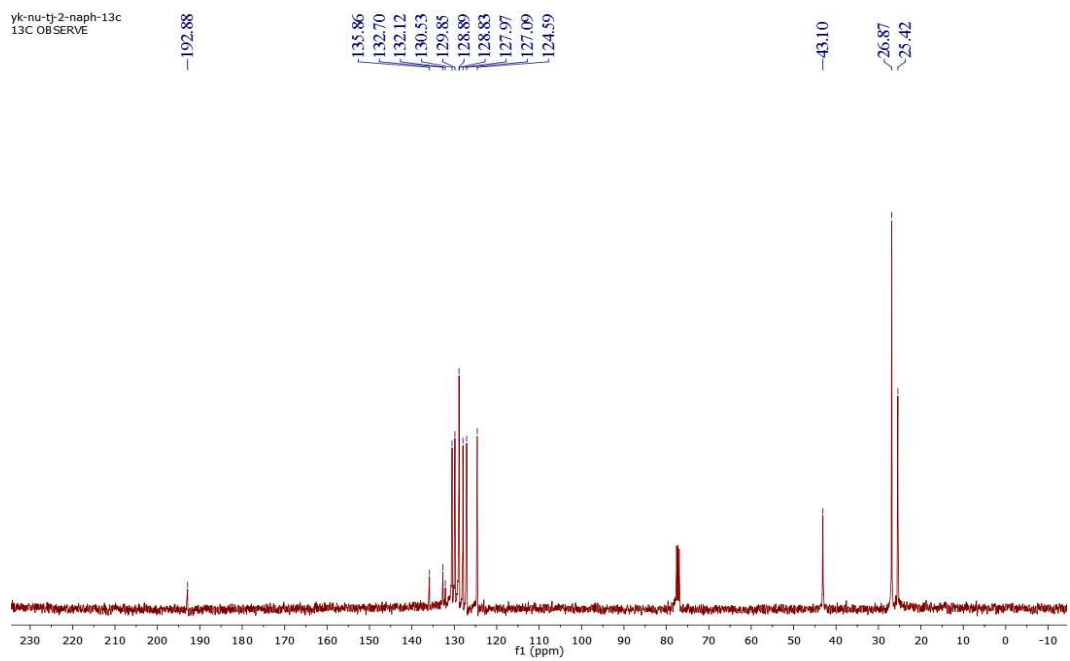


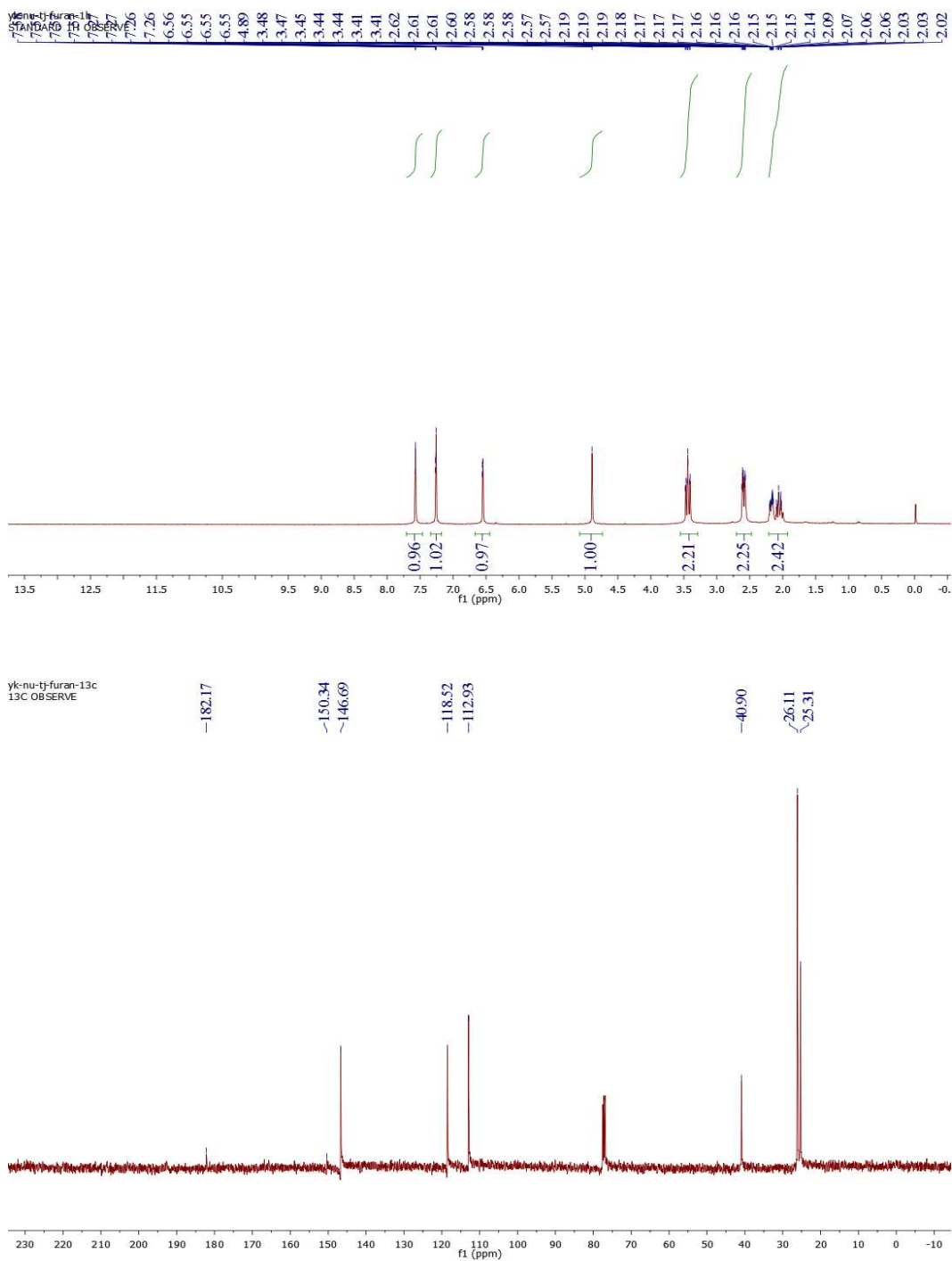


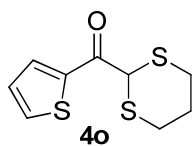
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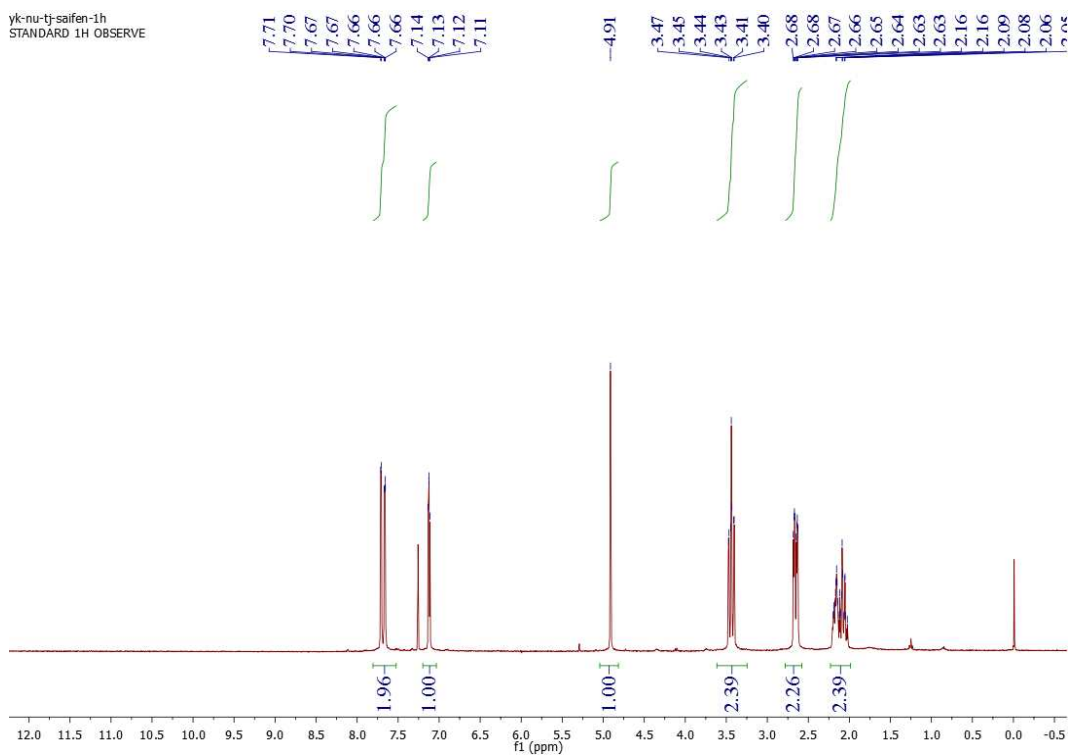
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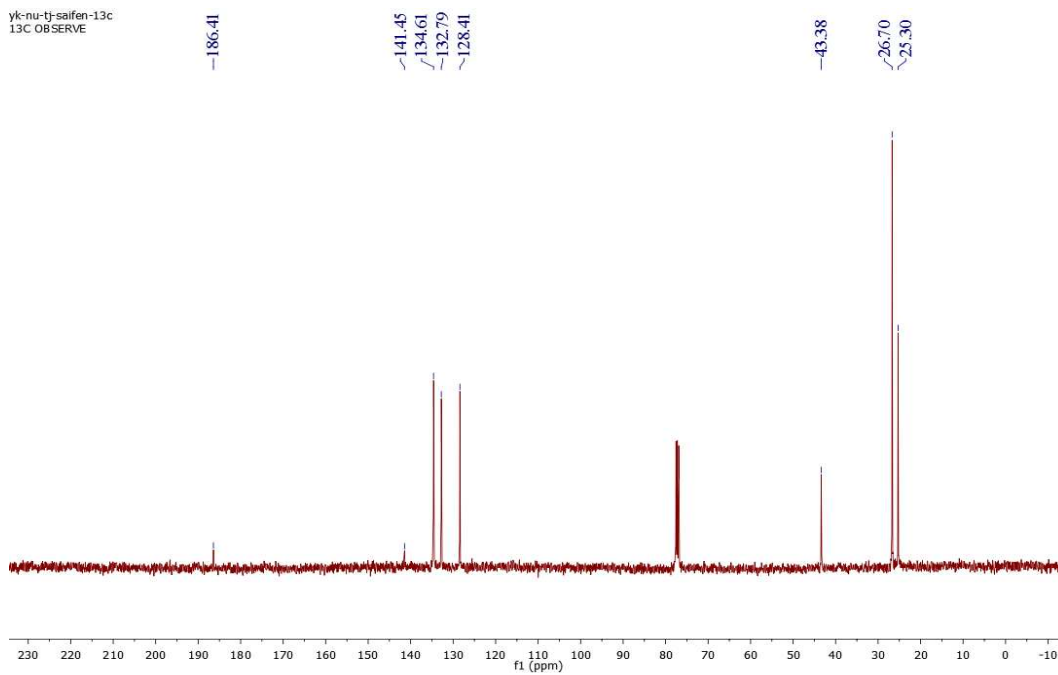


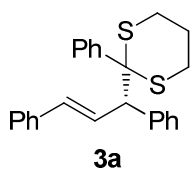


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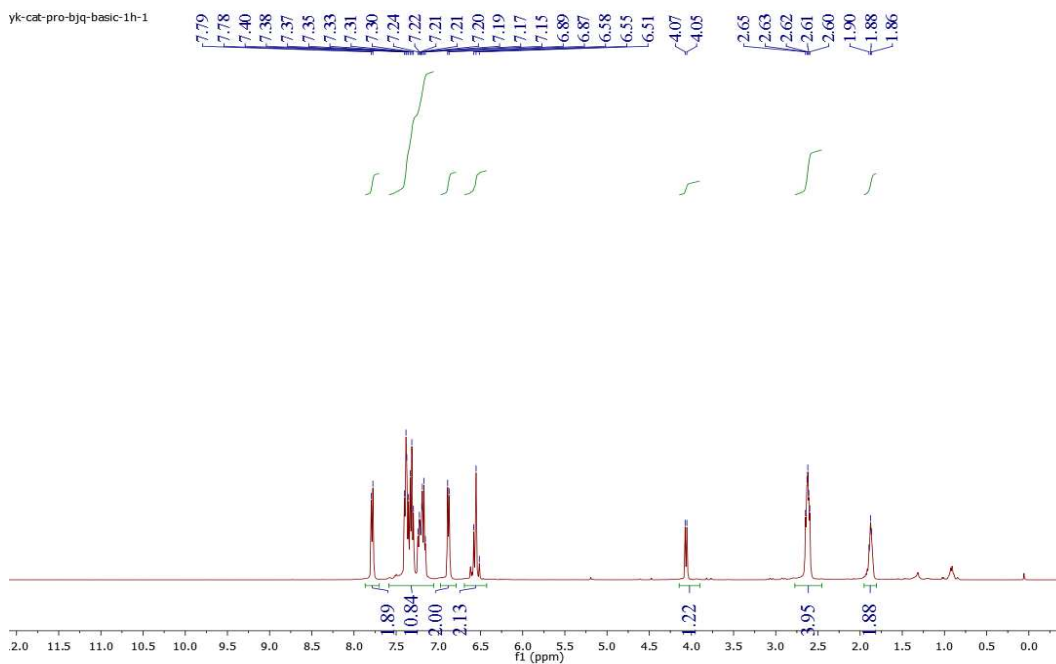


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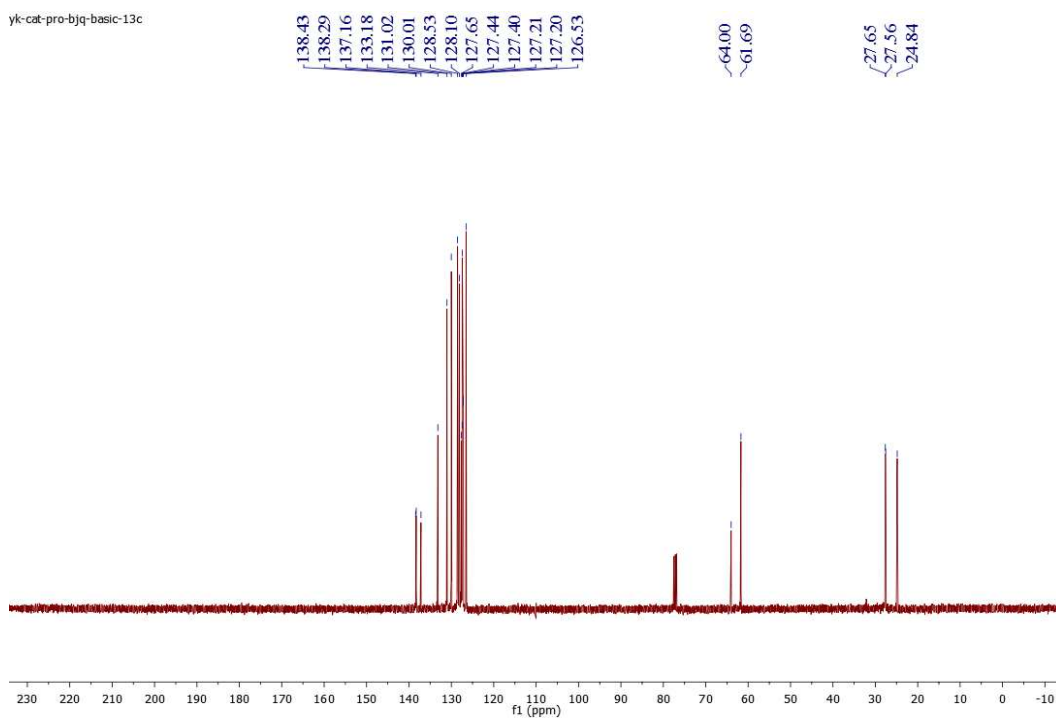


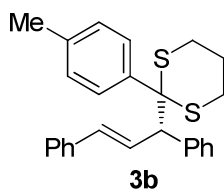


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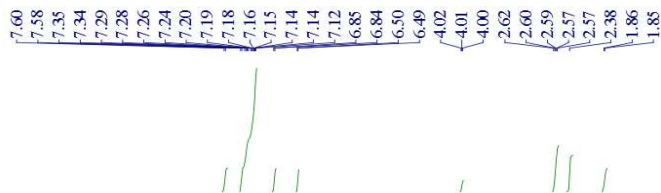


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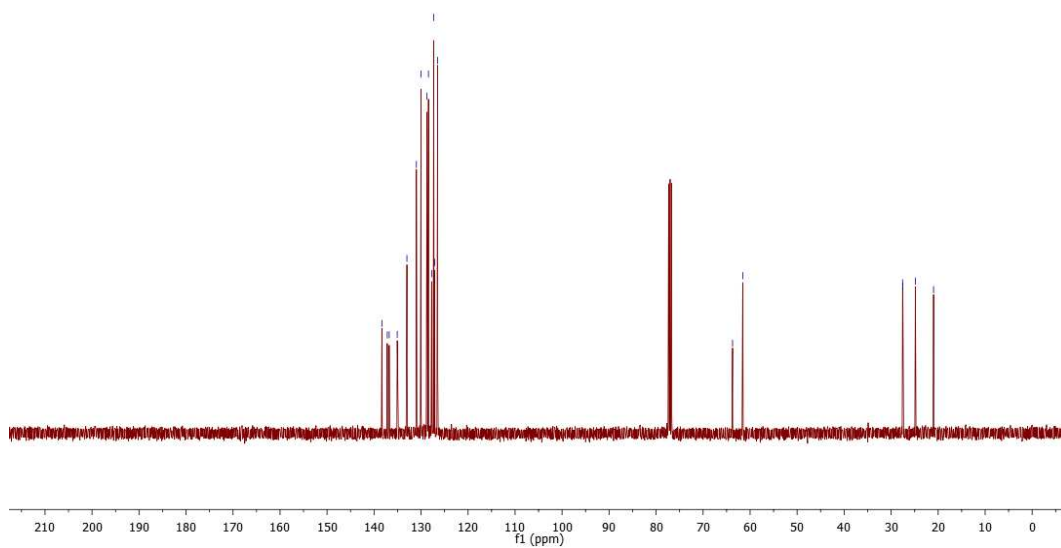


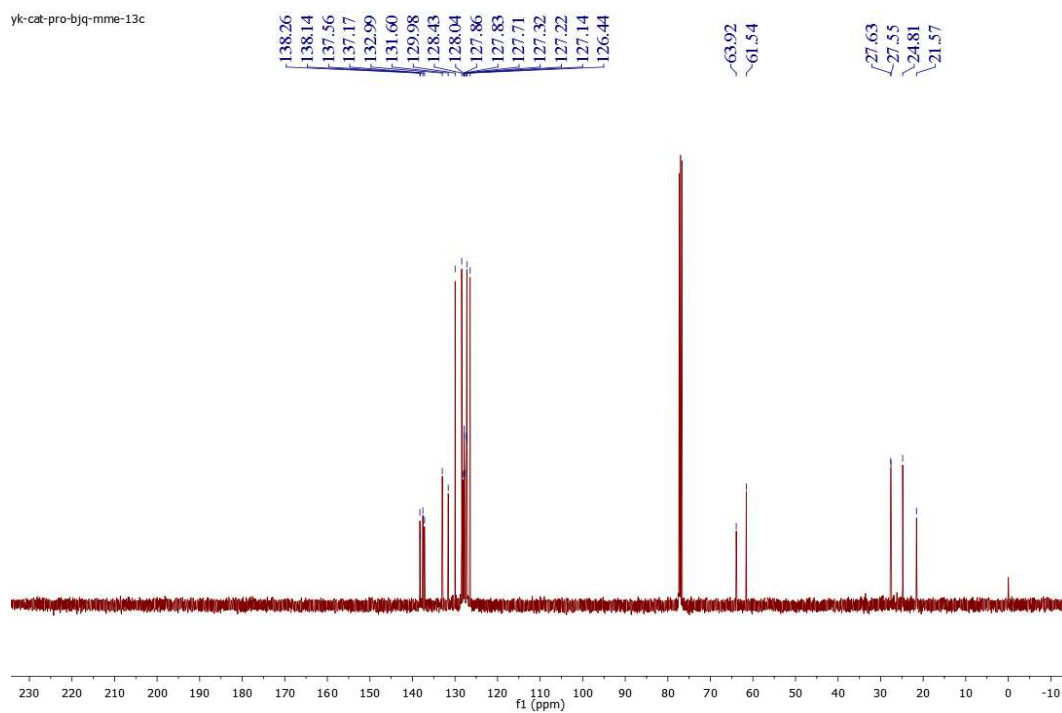
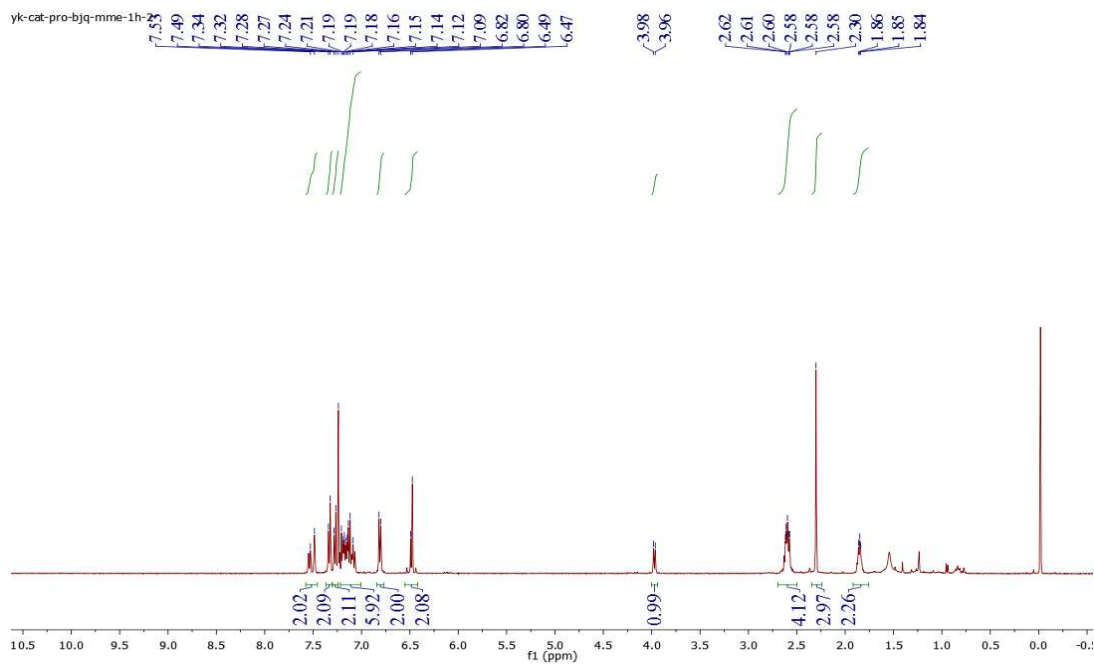
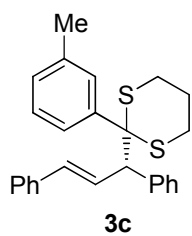


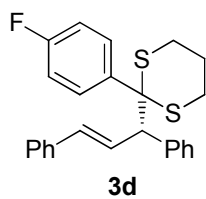
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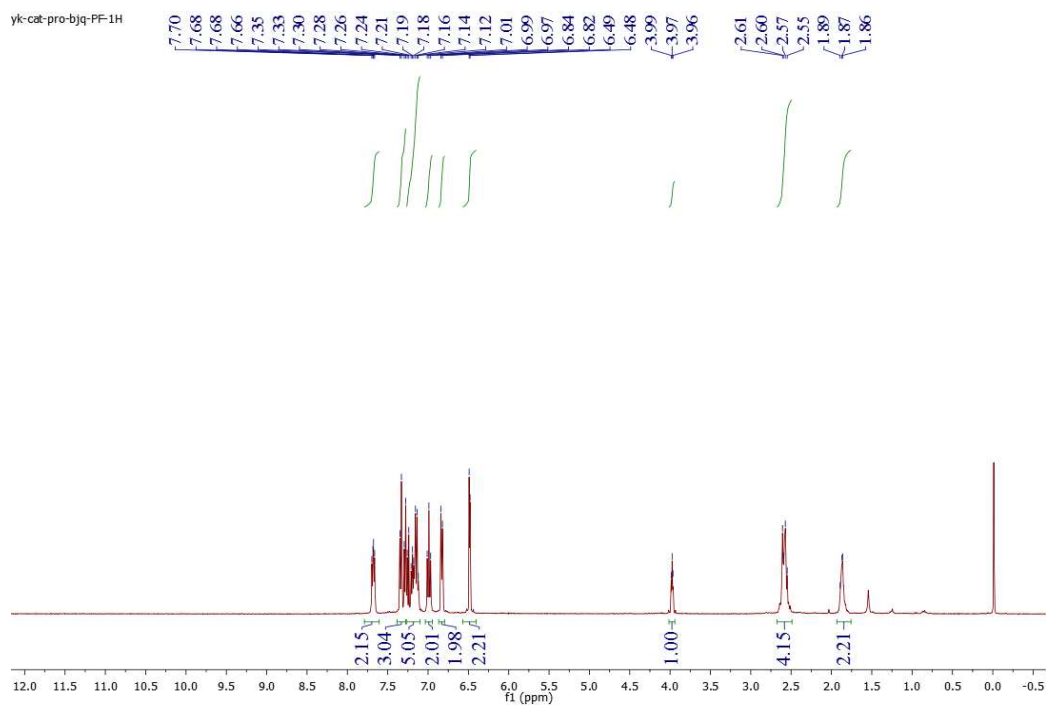
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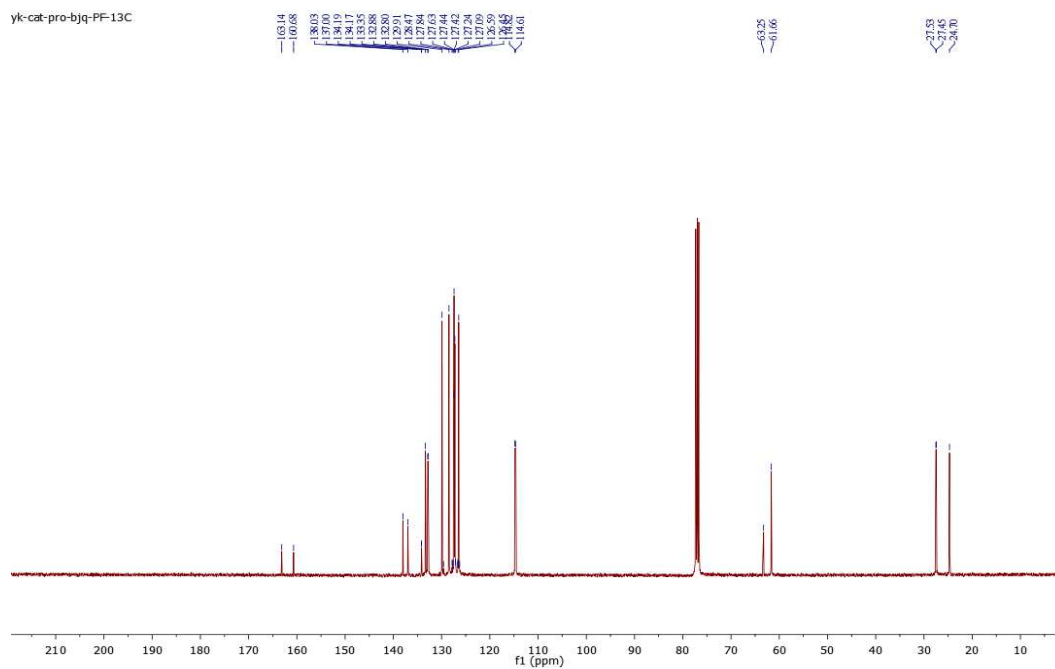


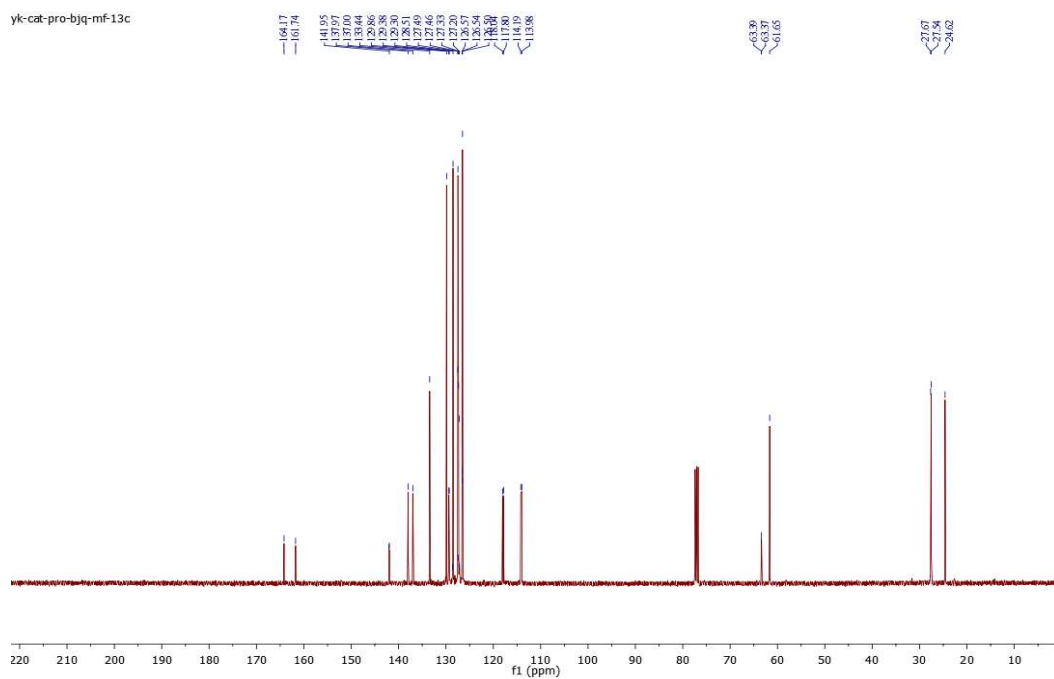
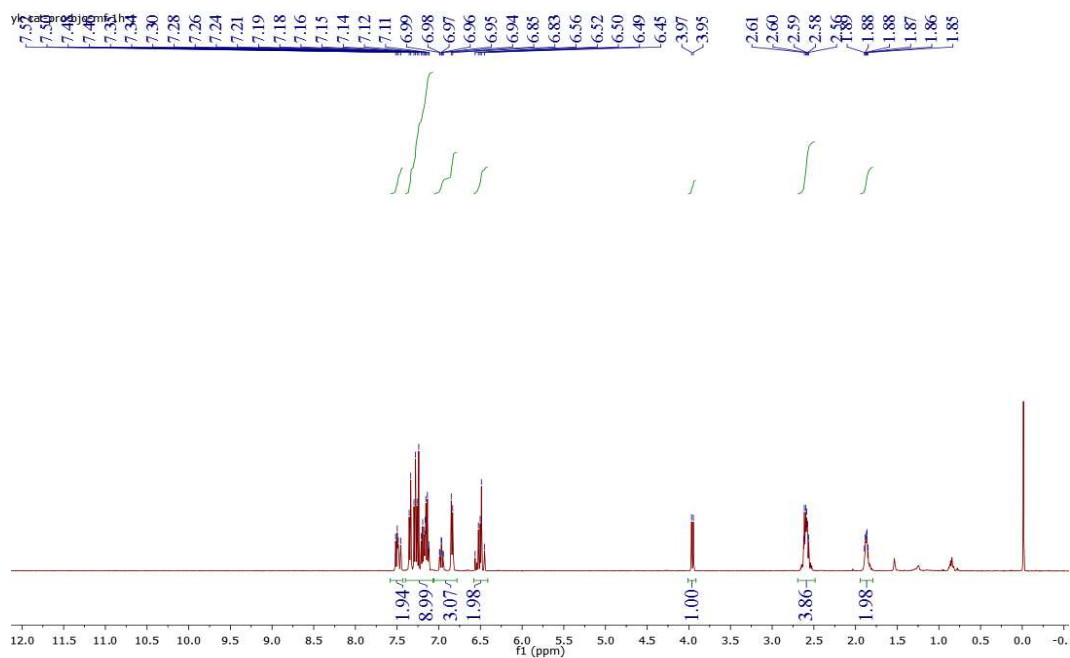
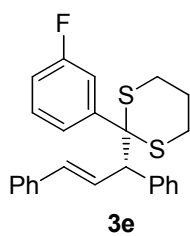


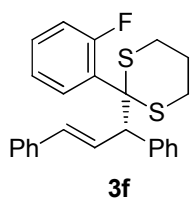
yk-cat-pro-bjq-PF-1H



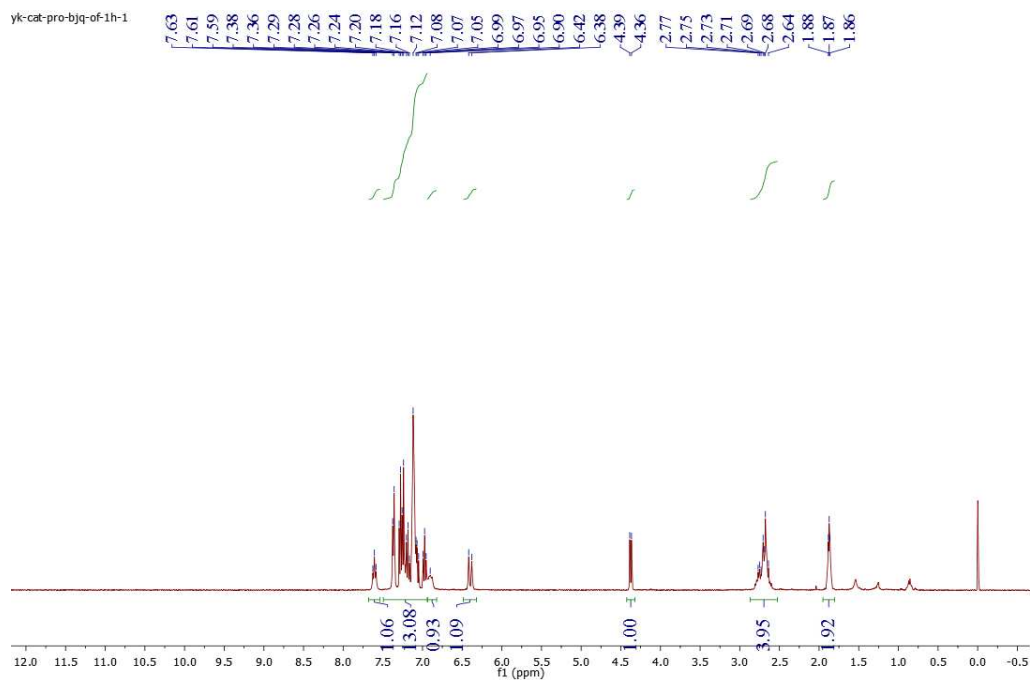
yk-cat-pro-bjq-PF-13C



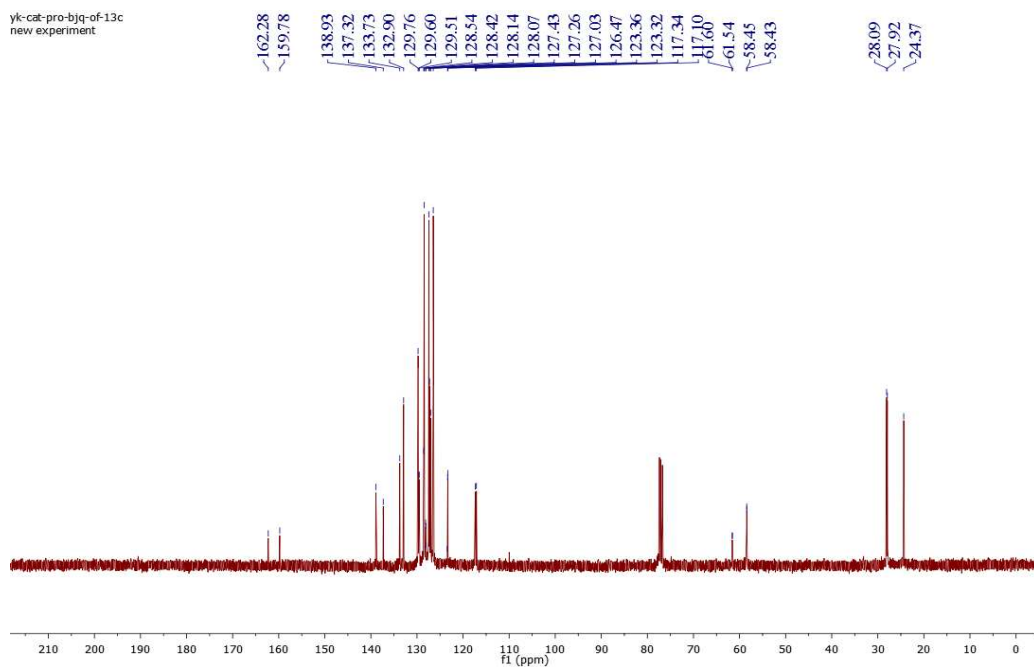


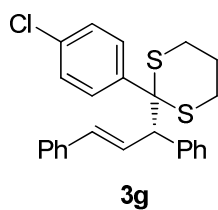


yk-cat-pro-bjq-of-1h-1

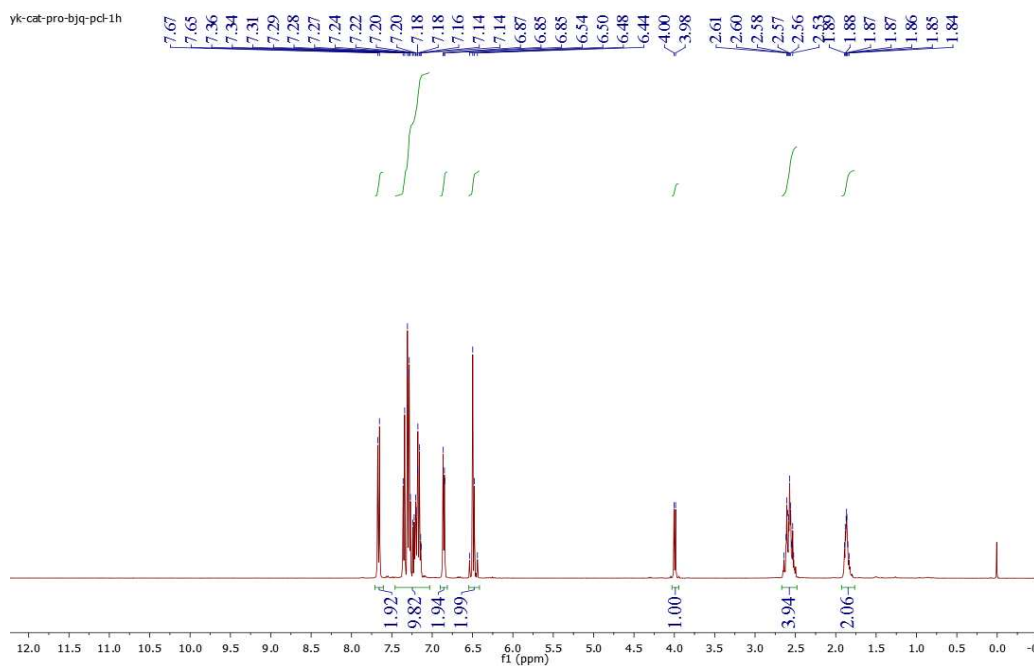


yk-cat-pro-bjq-of-13c
new experiment

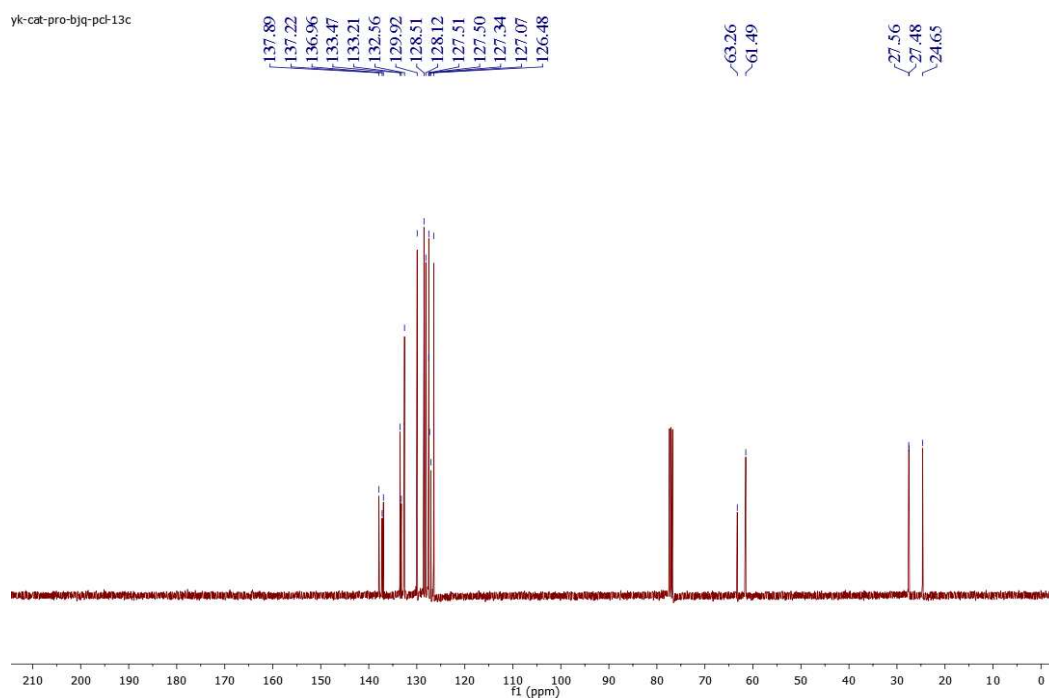


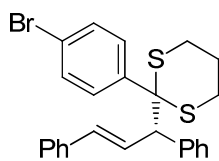


yk-cat-pro-bjq-pcl-1h



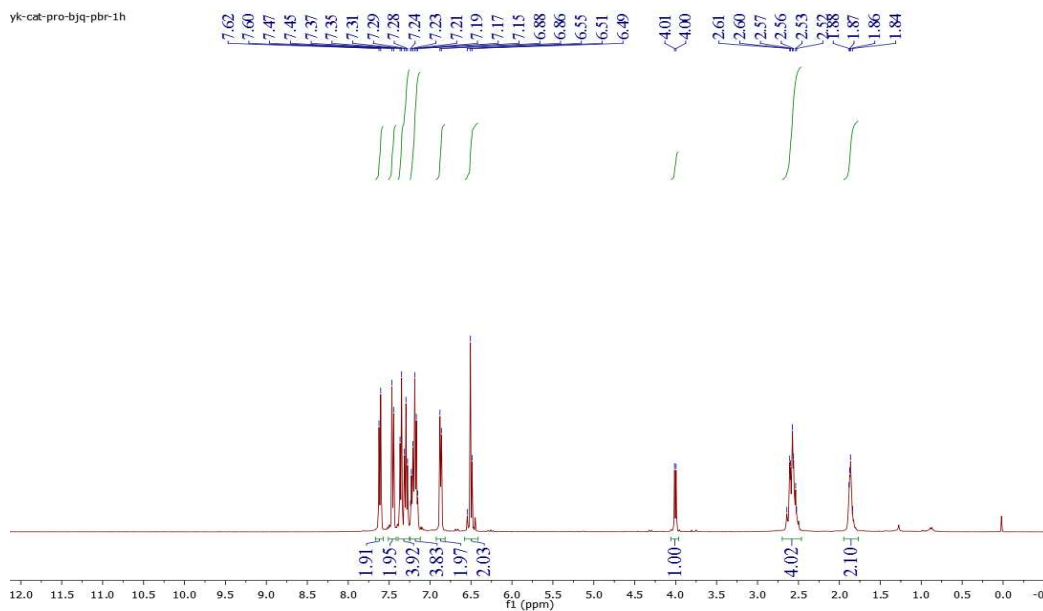
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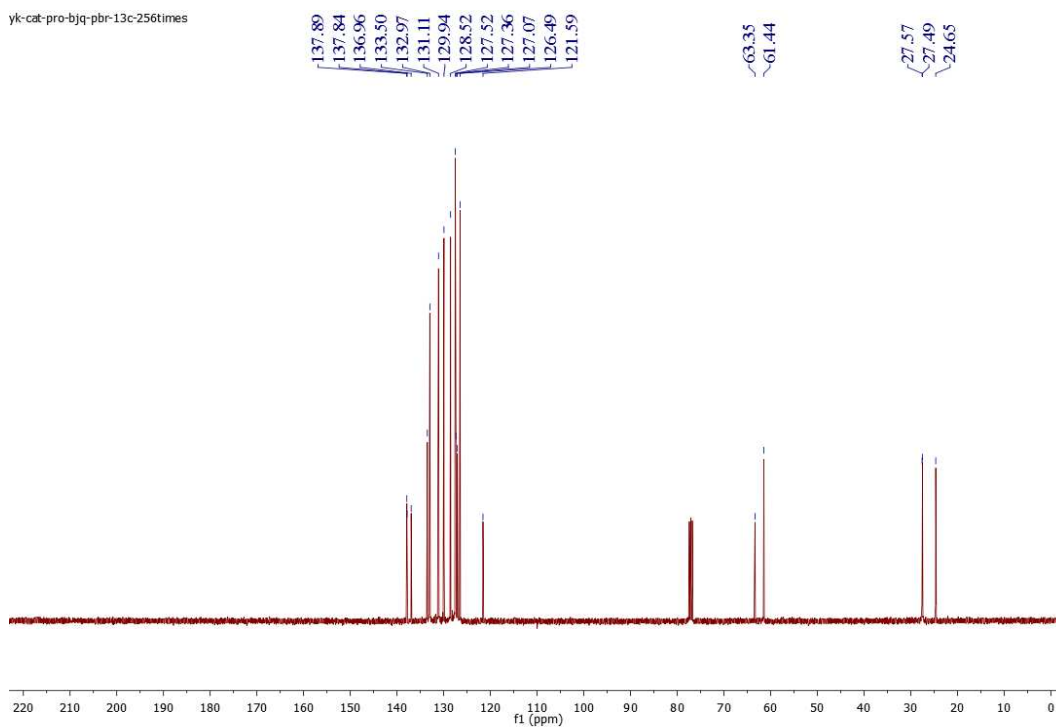


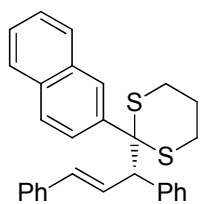
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yk-cat-pro-bjq-pbr-1h

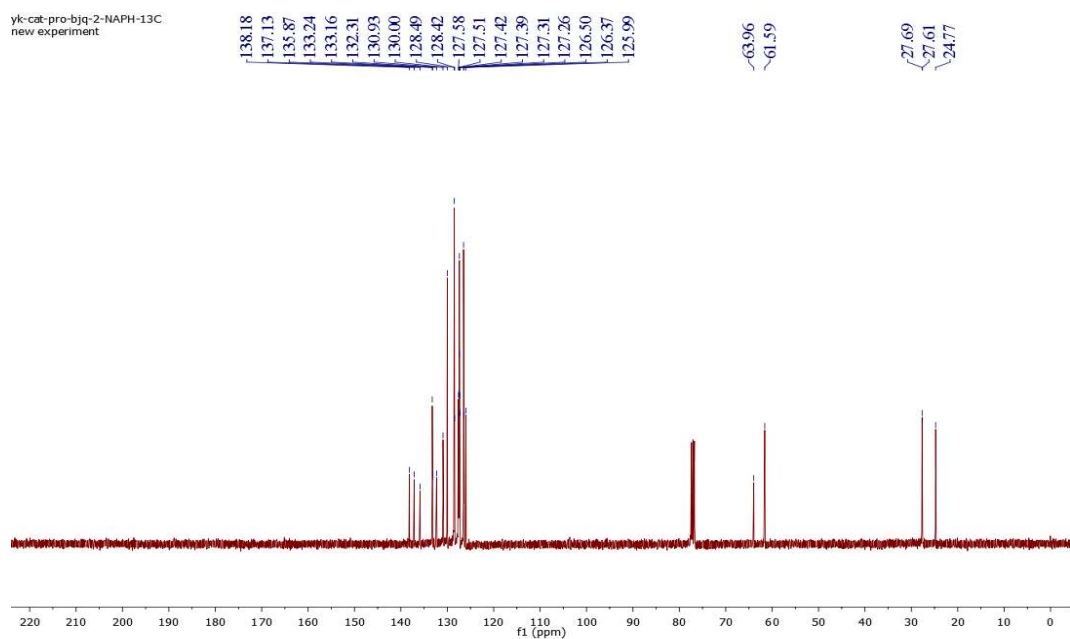
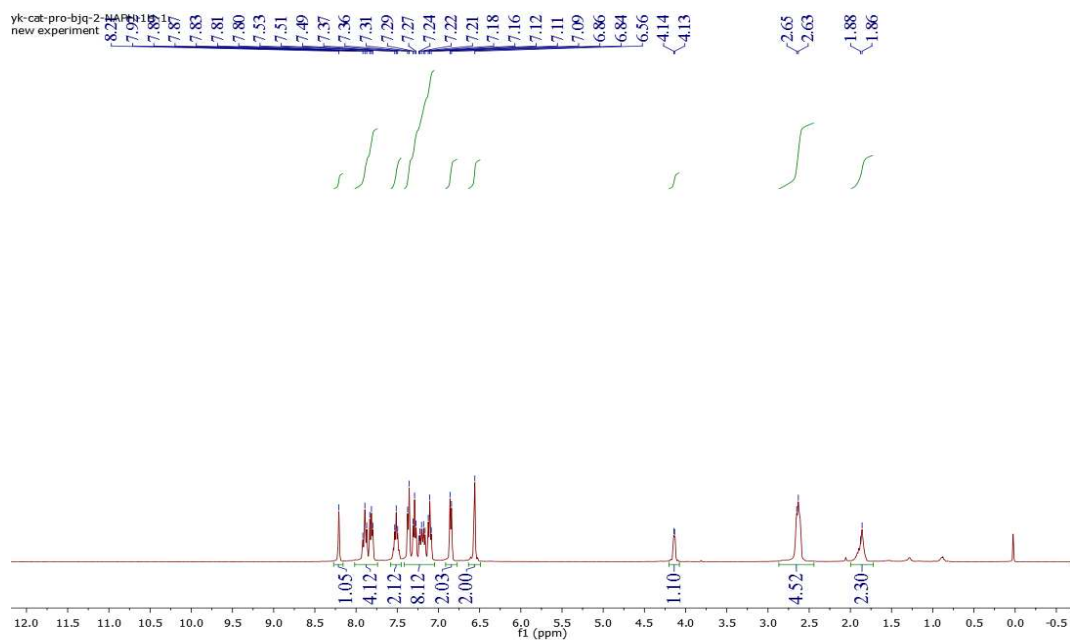


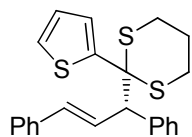
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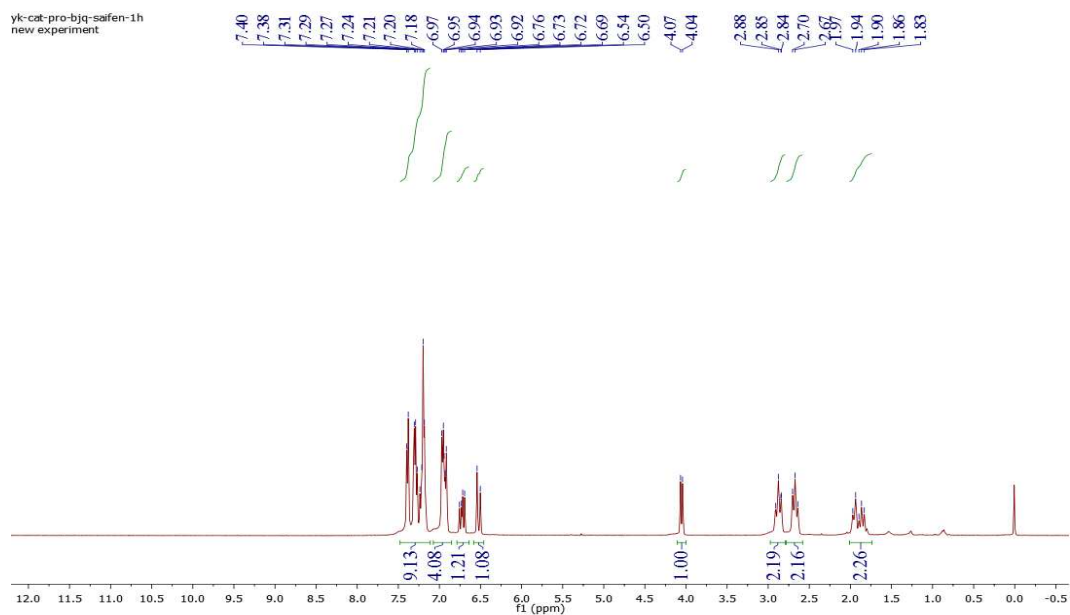
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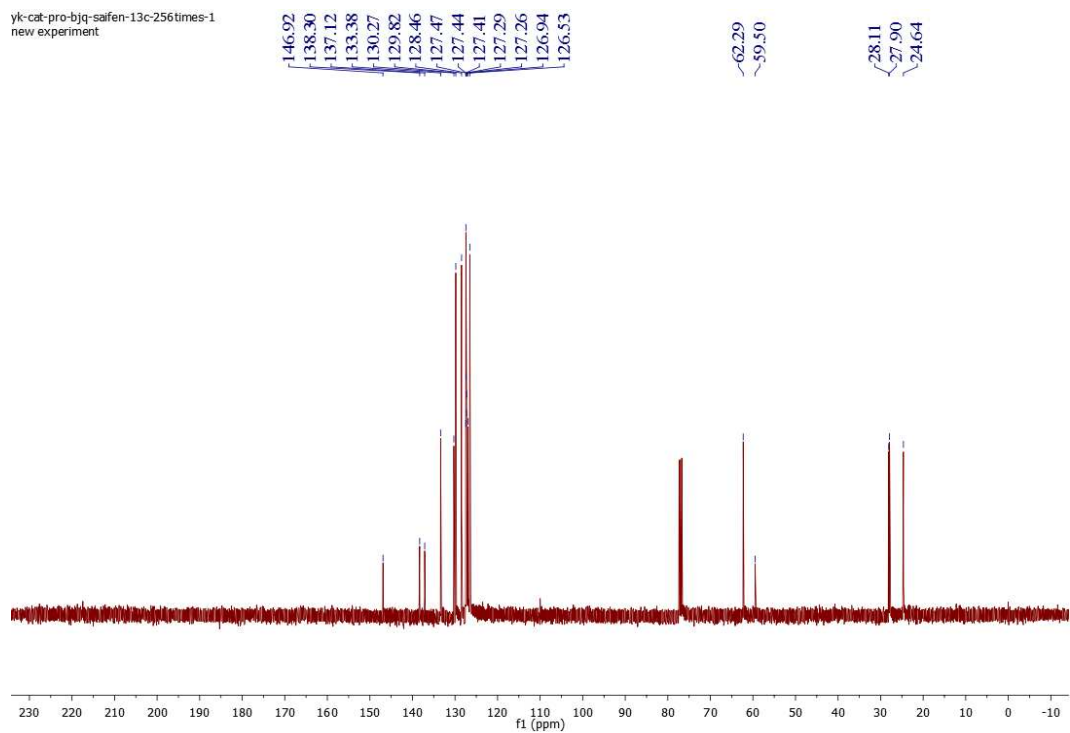


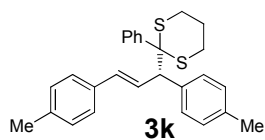
3j

yk-cat-pro-bjq-saifen-1h
new experiment

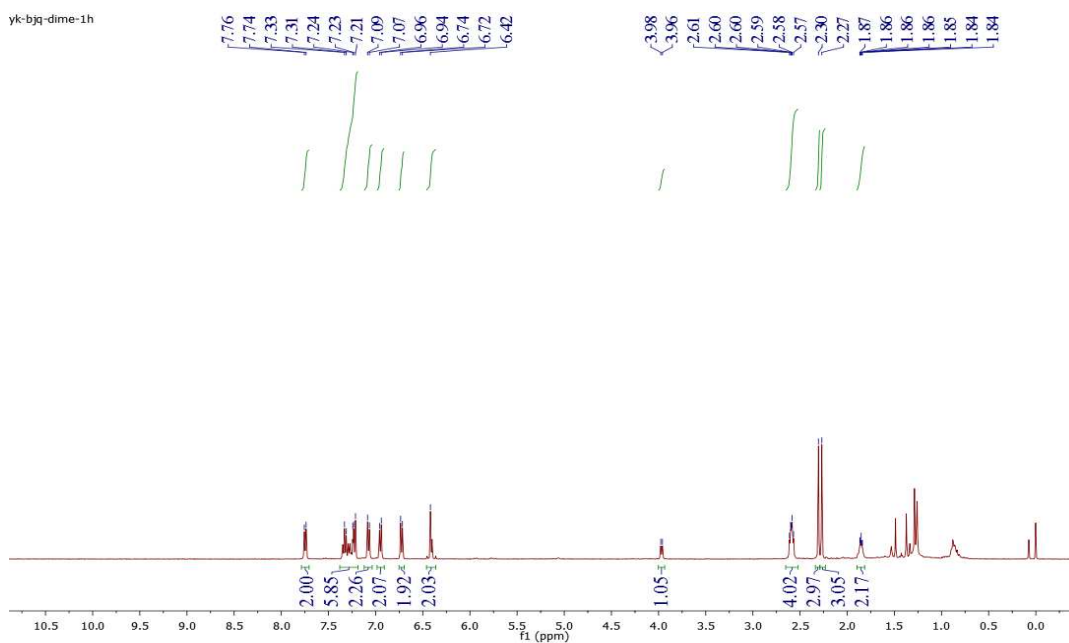


yk-cat-pro-bjq-saifen-13c-256times-1
new experiment

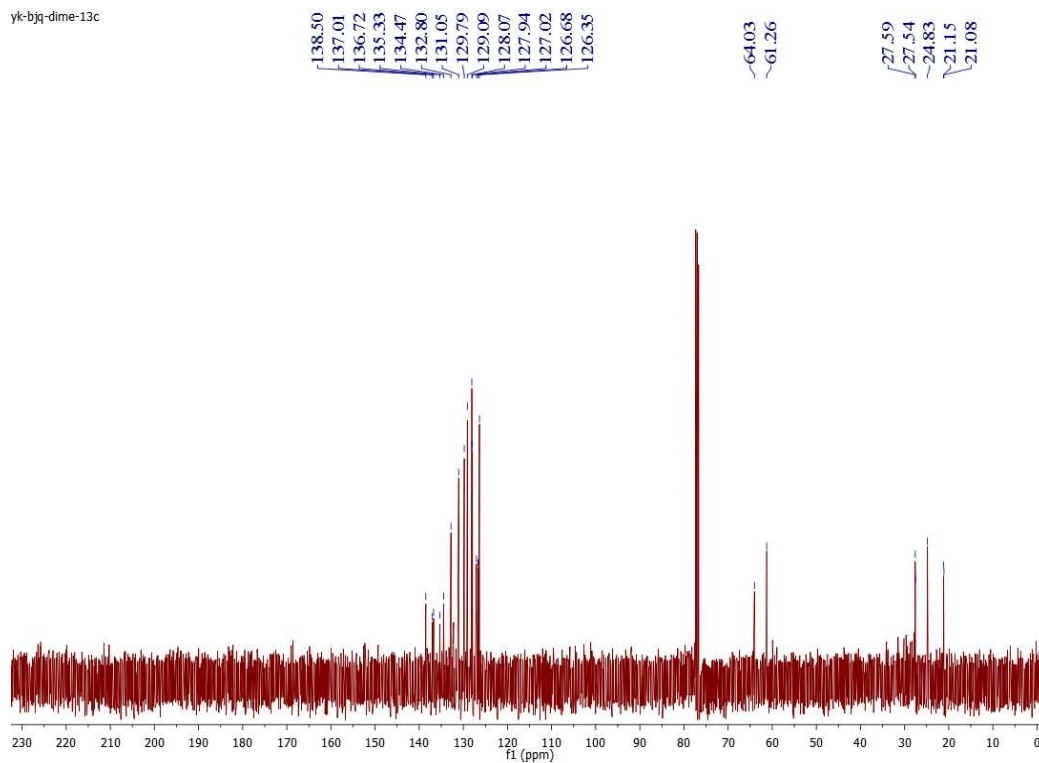


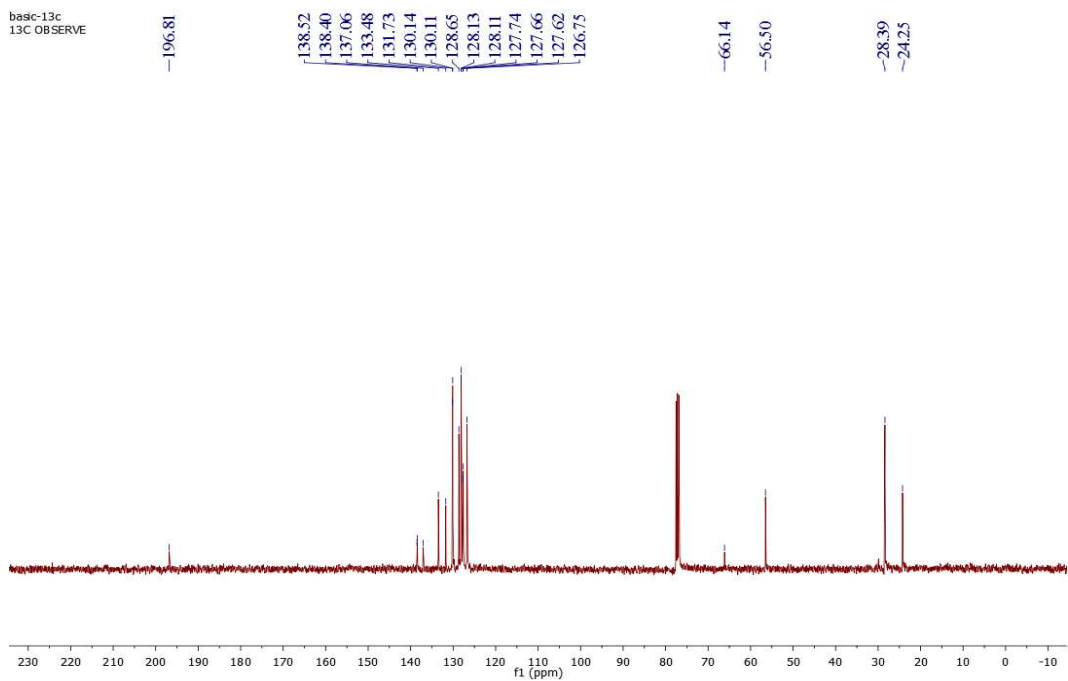
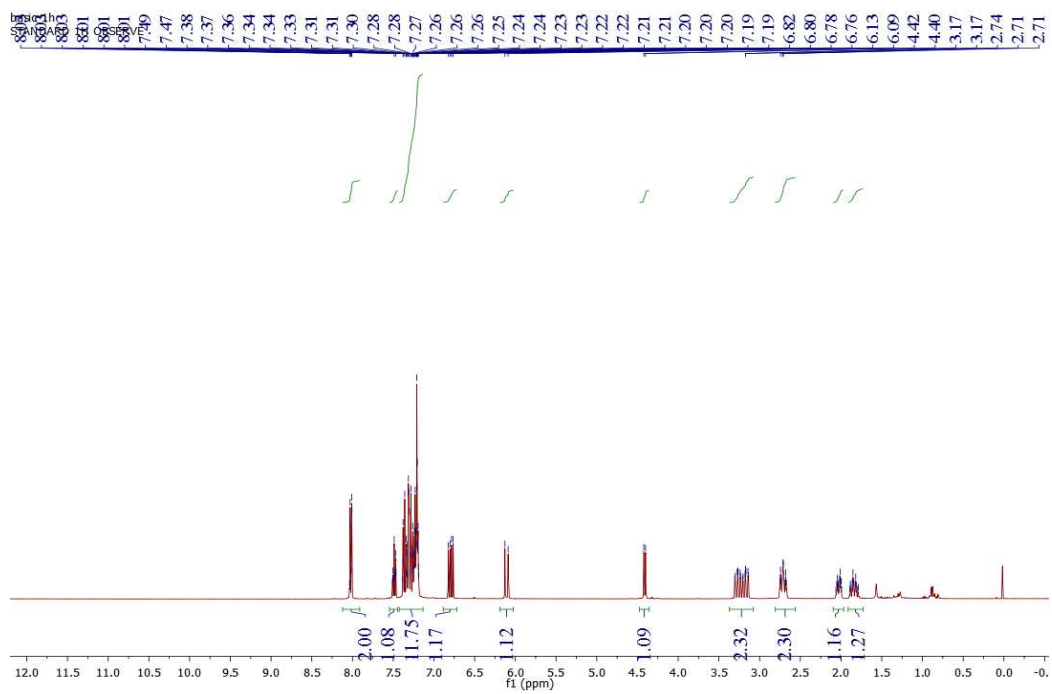
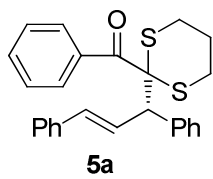


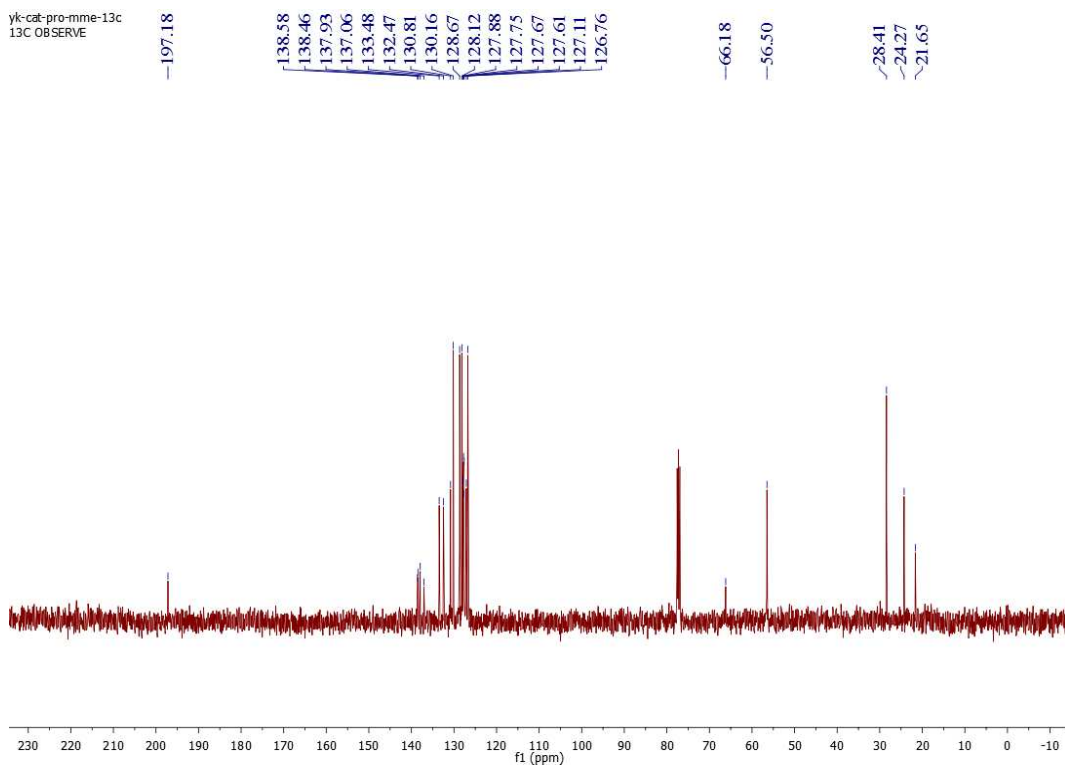
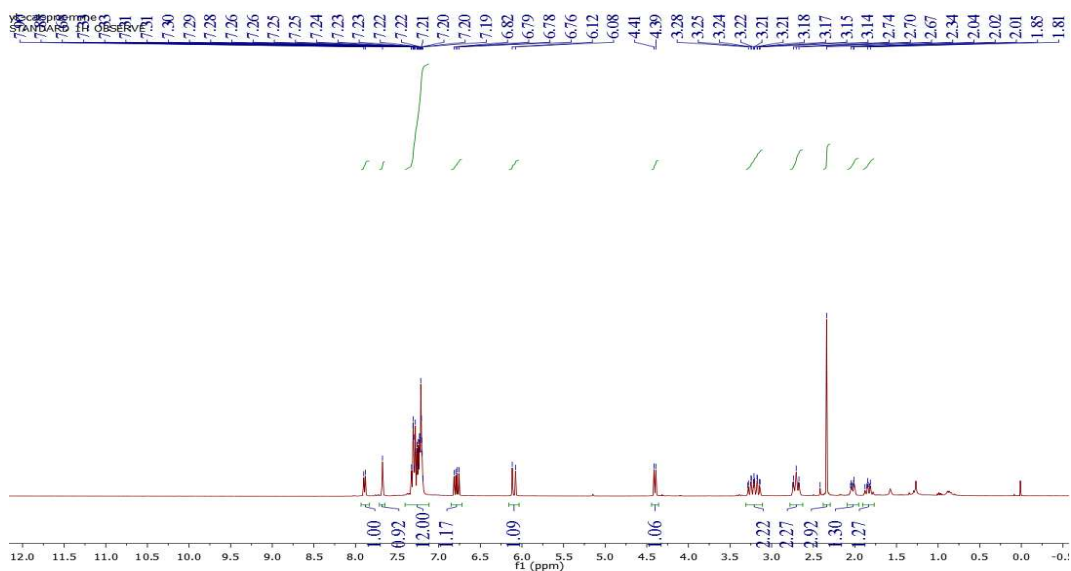
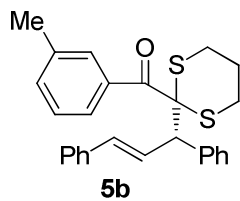
yk-bjq-dime-1h

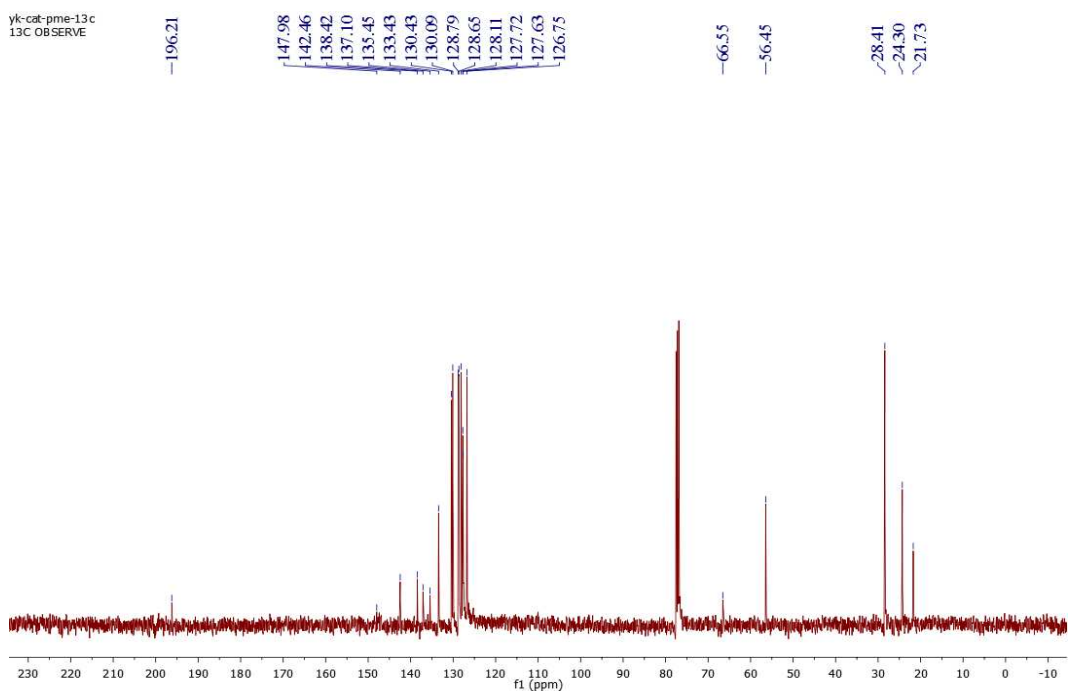
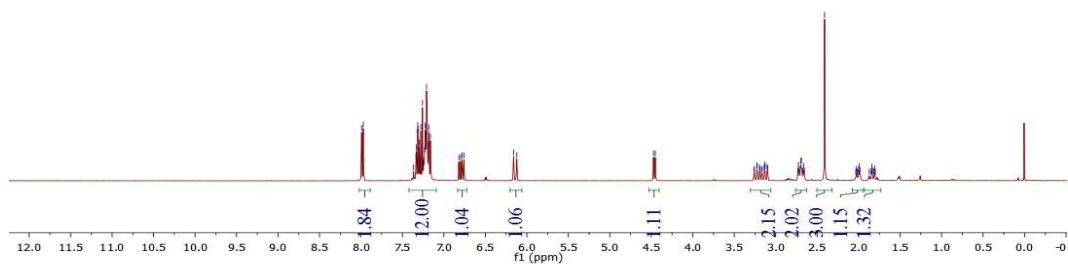
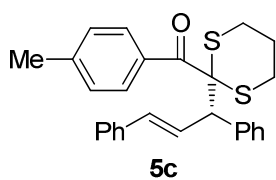


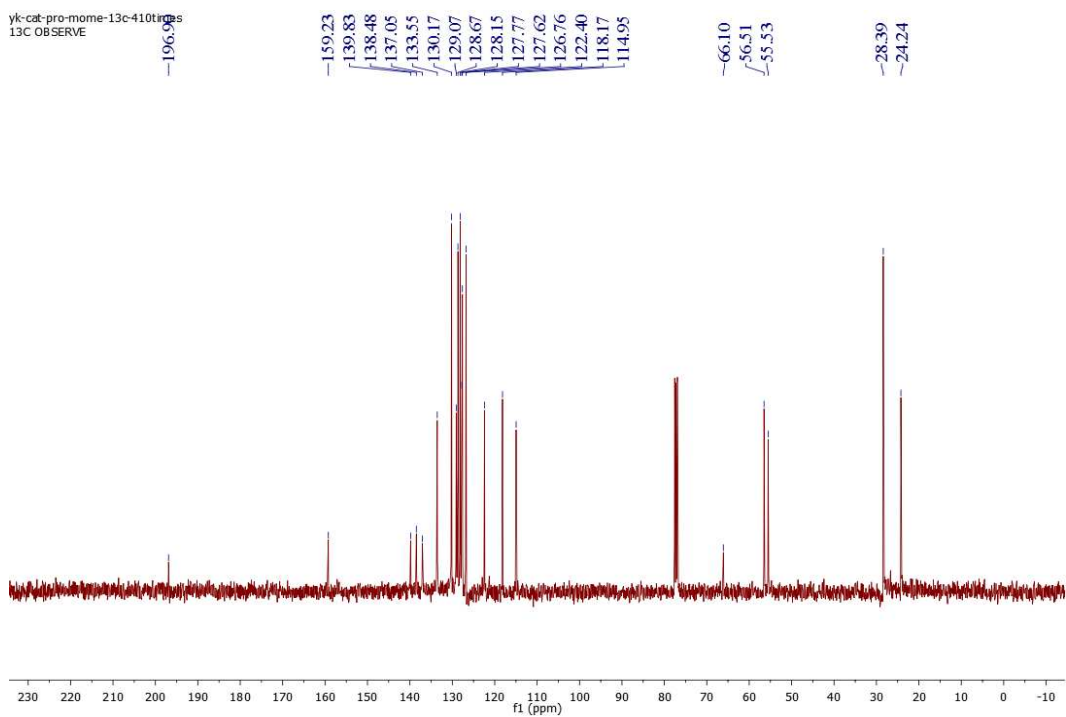
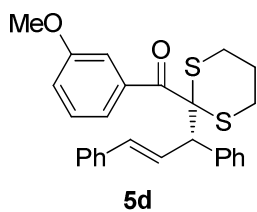
yk-bjq-dime-13c

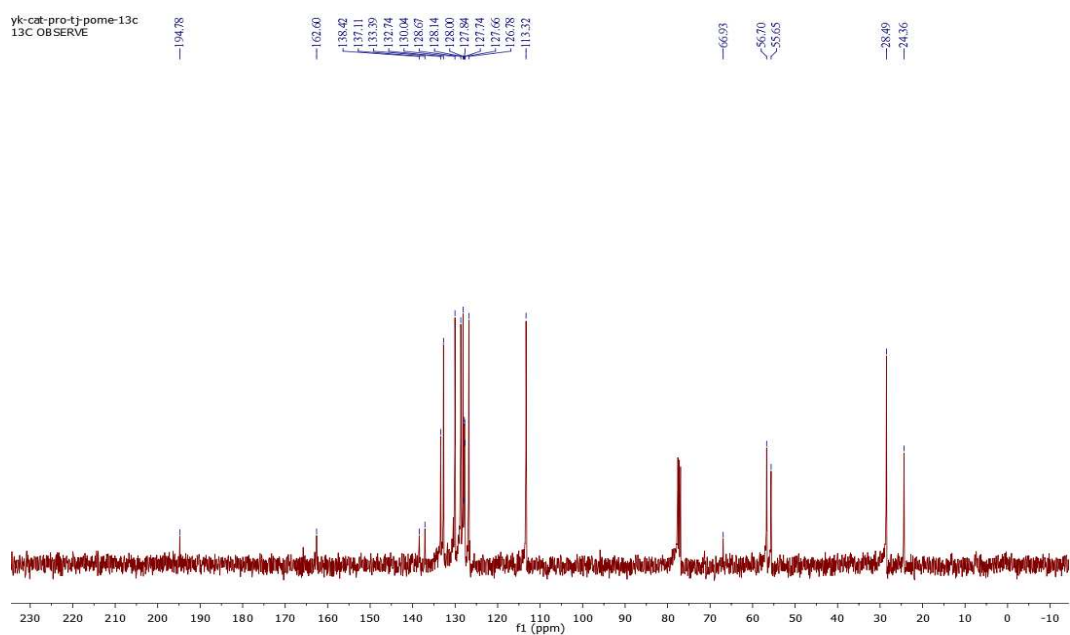
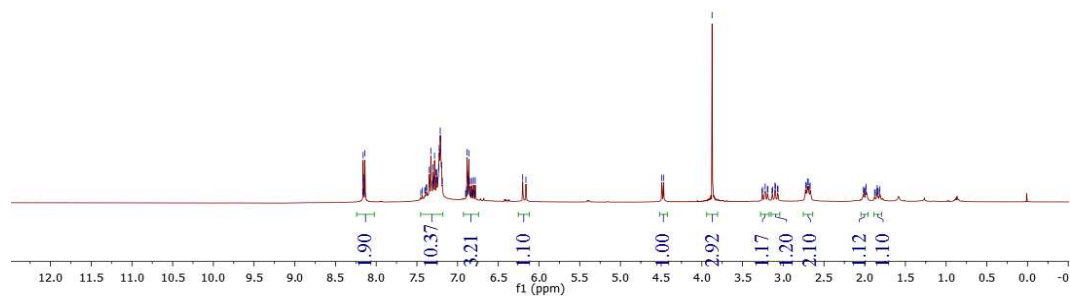
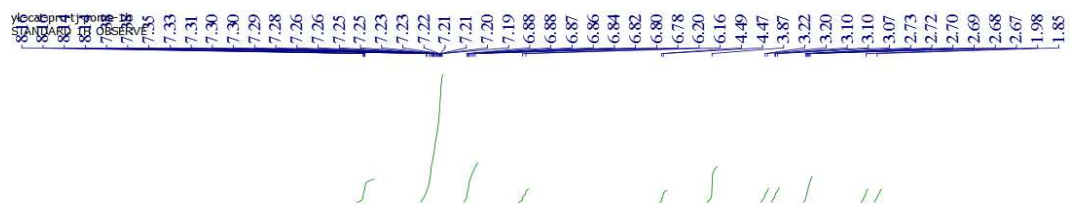
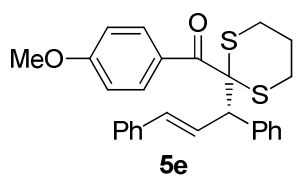


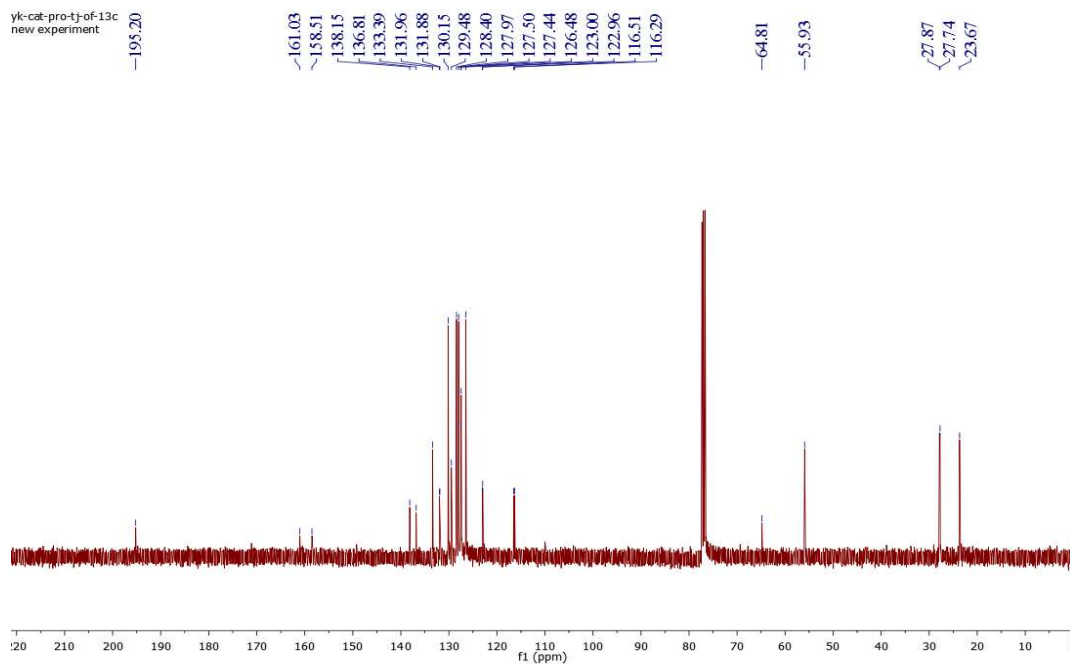
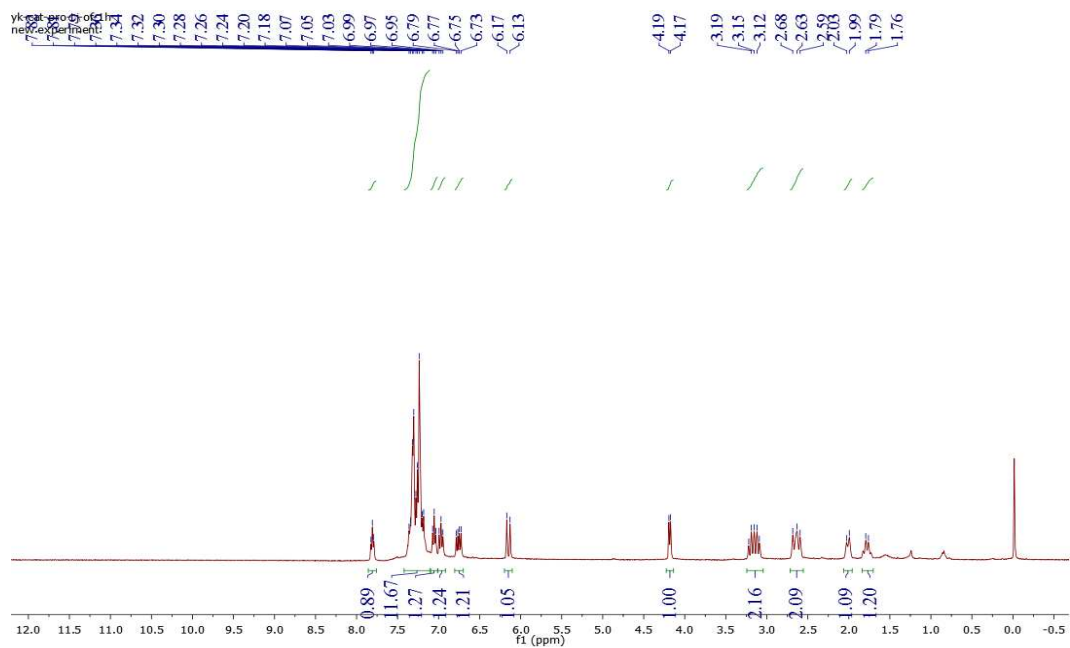
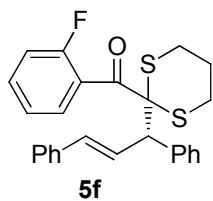


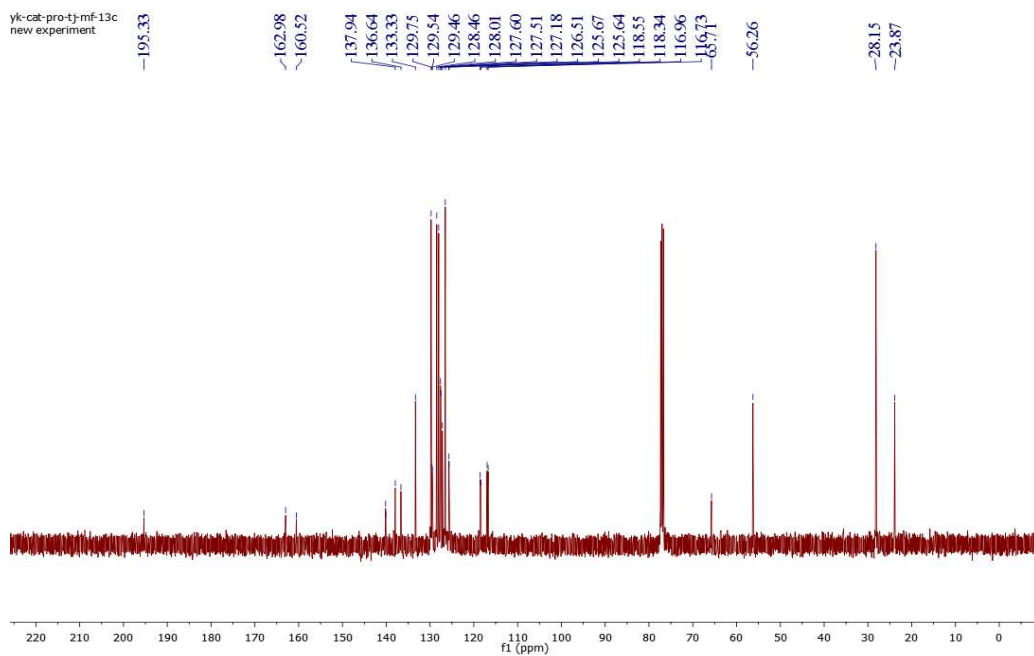
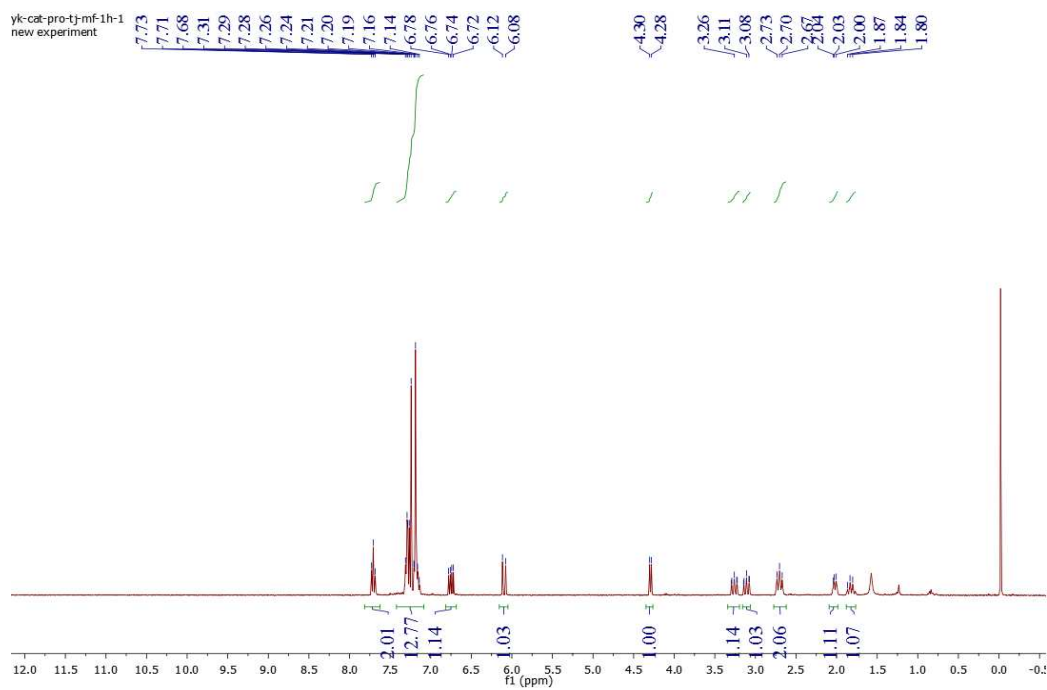
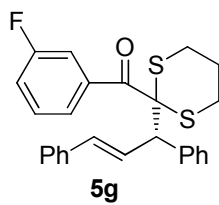


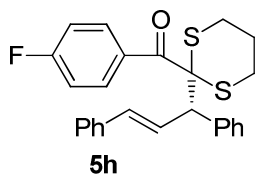




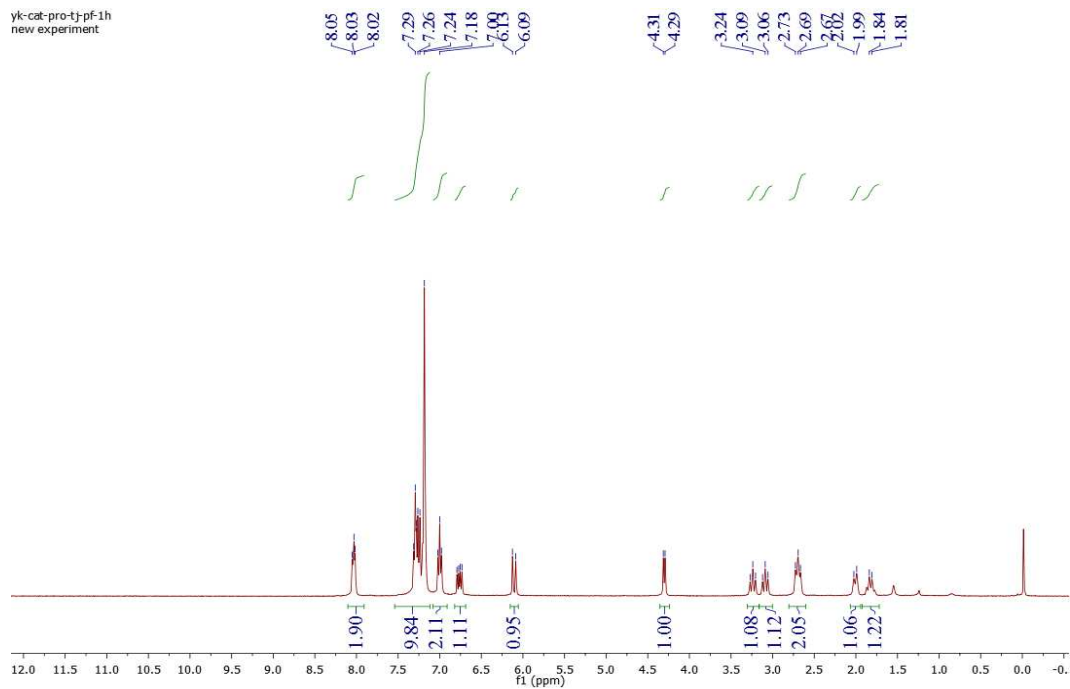




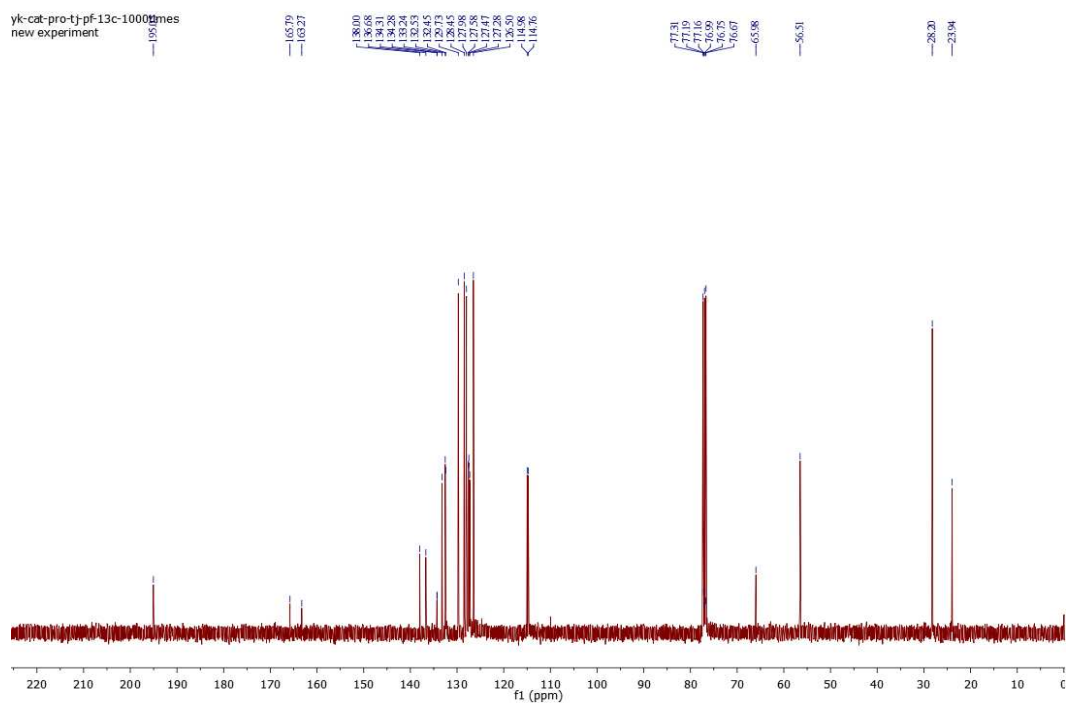


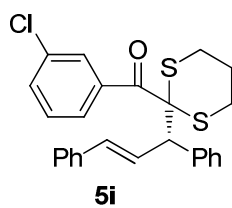


yk-cat-pro-tj-pf-1h
new experiment

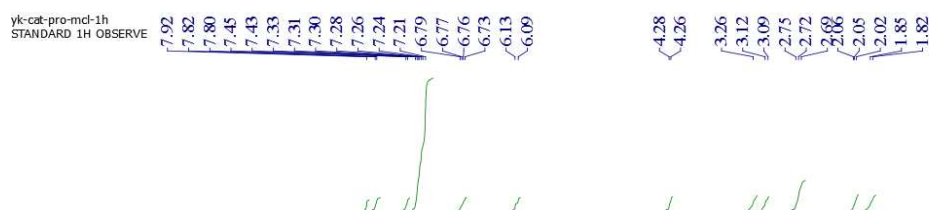


yk-cat-pro-tj-pf-13c-1000times
new experiment

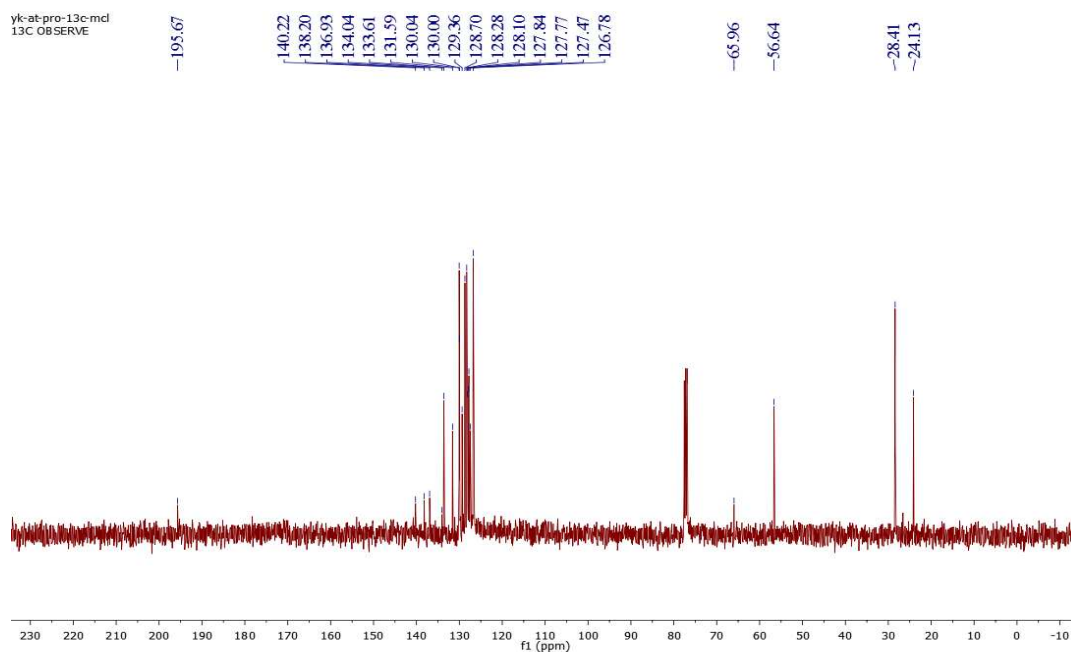


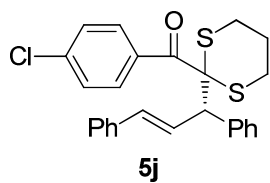


yk-cat-pro-mcl-1h
STANDARD 1H OBSERVE

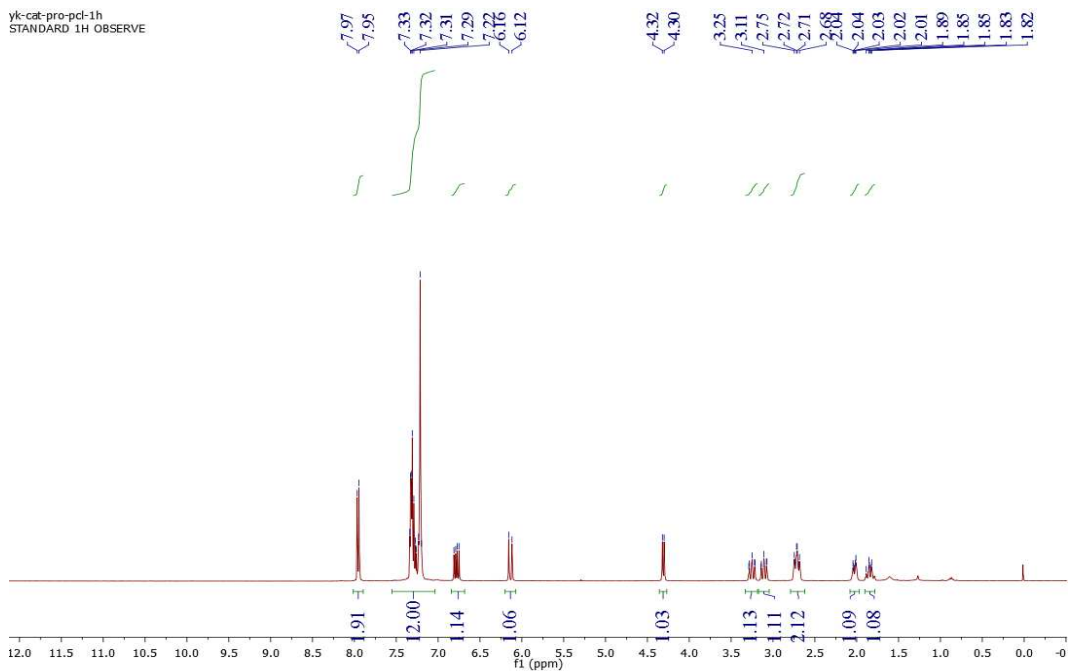


yk-at-pro-13c-mcl
13C OBSERVE

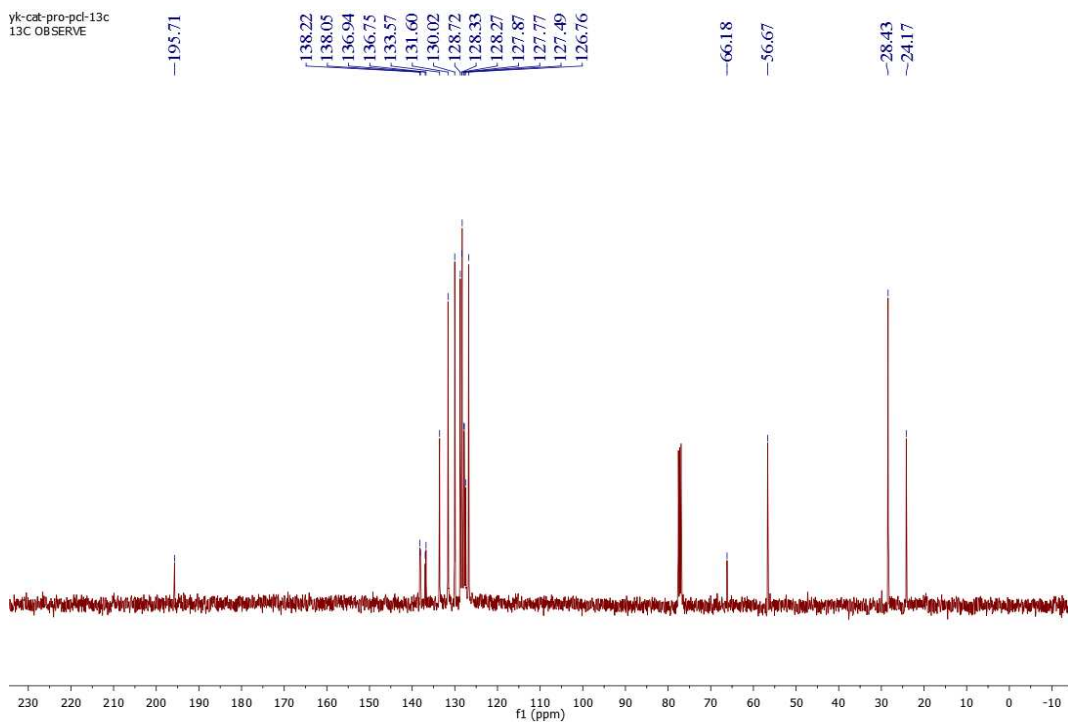




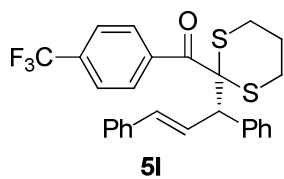
yk-cat-pro-pd-1h
STANDARD 1H OBSERVE



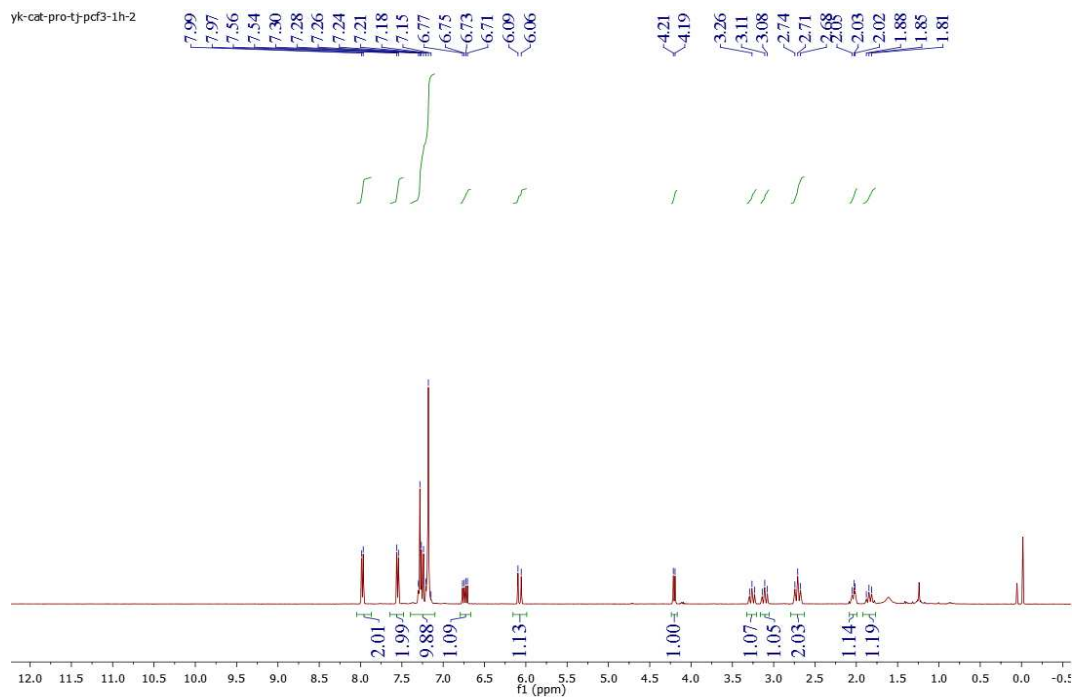
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13C OBSERVE



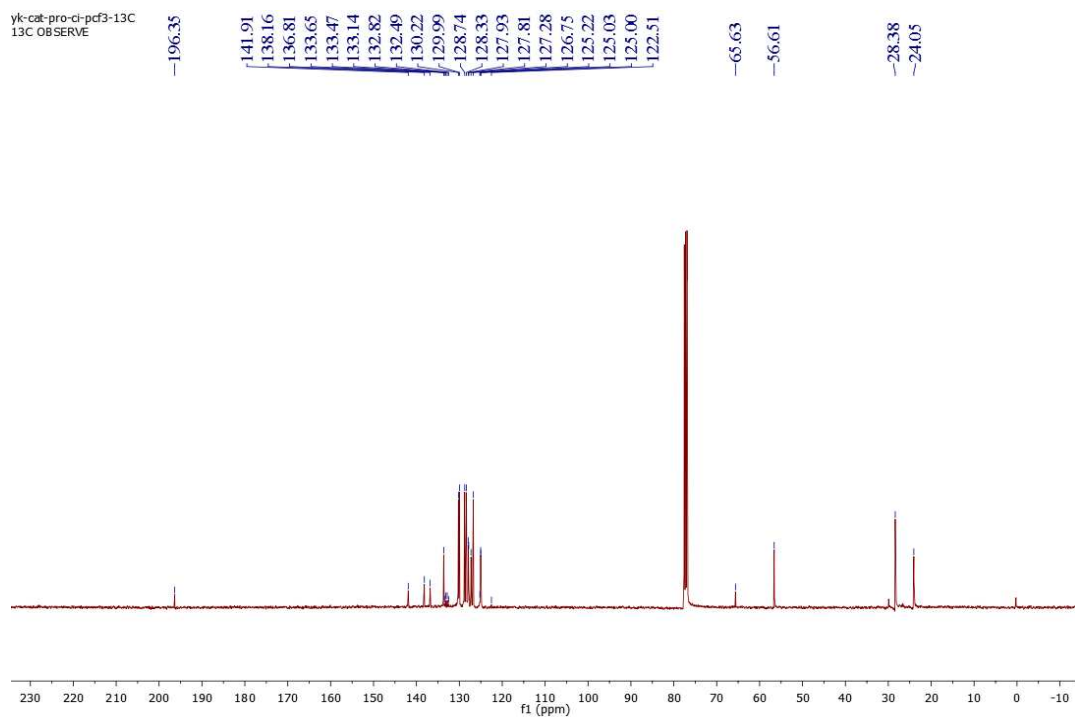


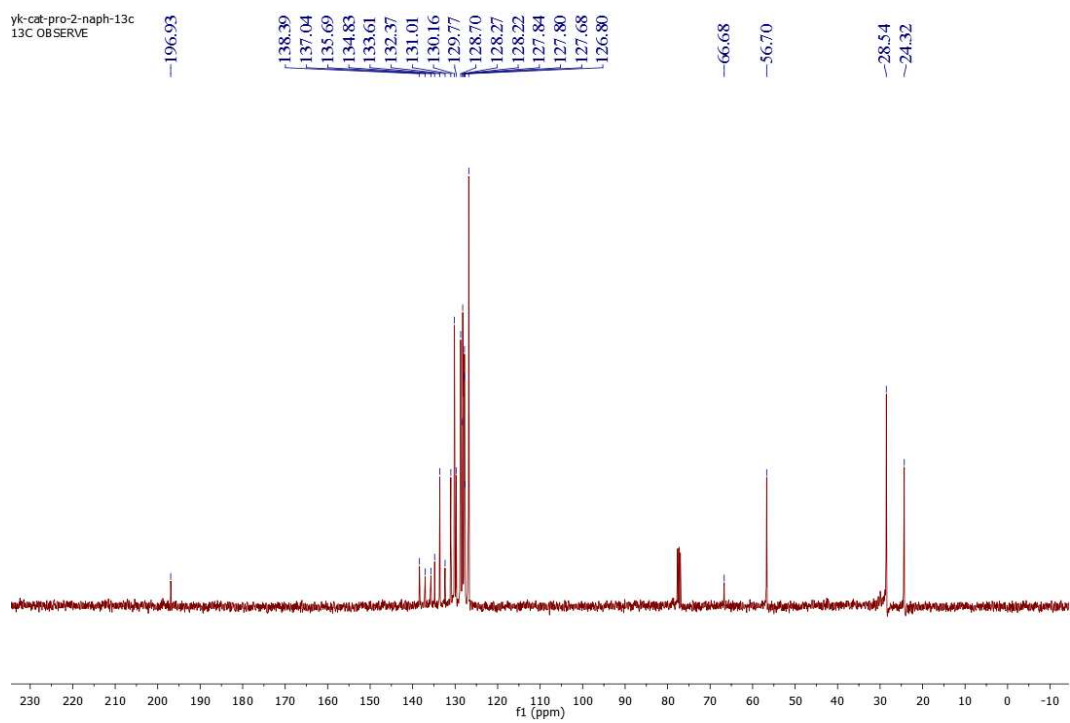
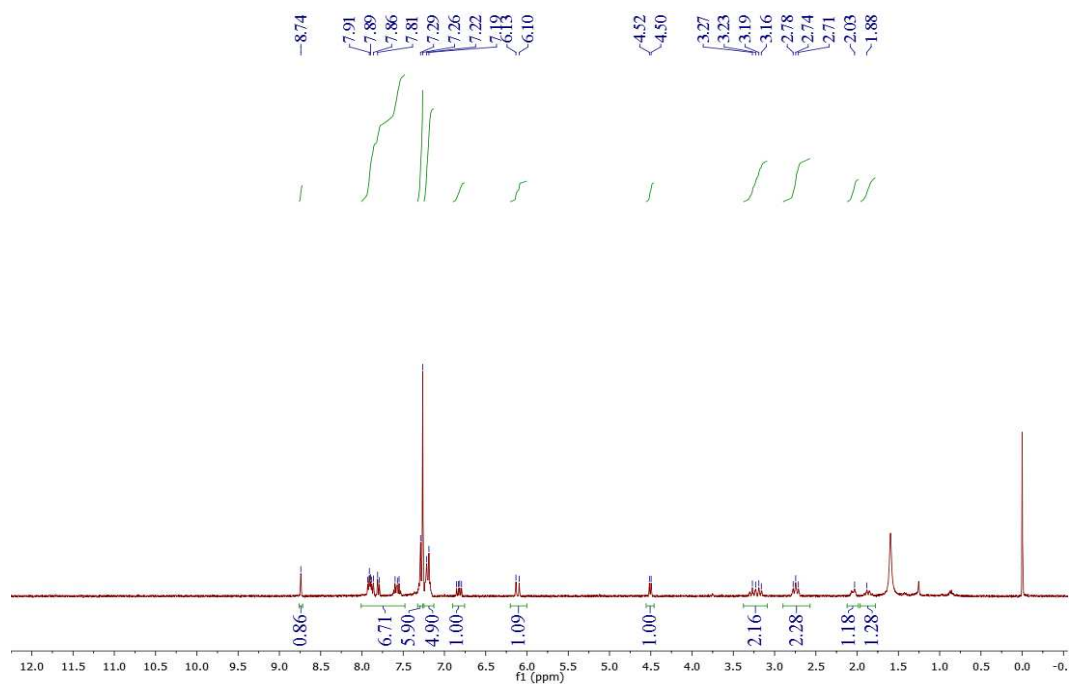
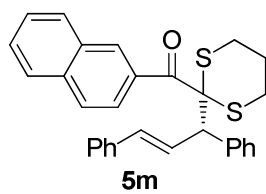


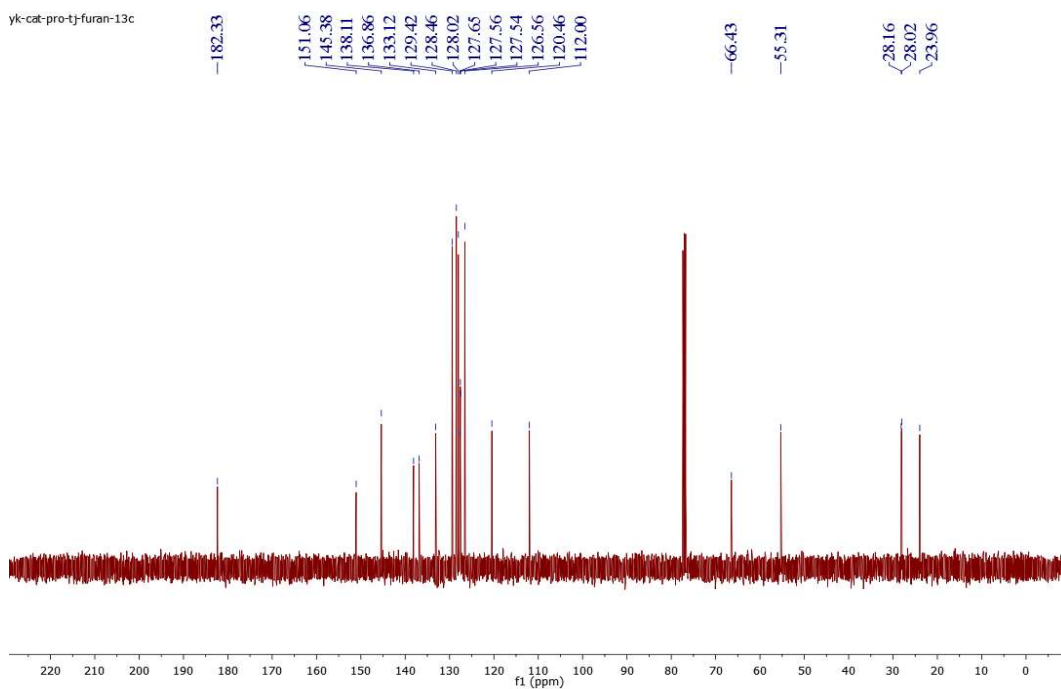
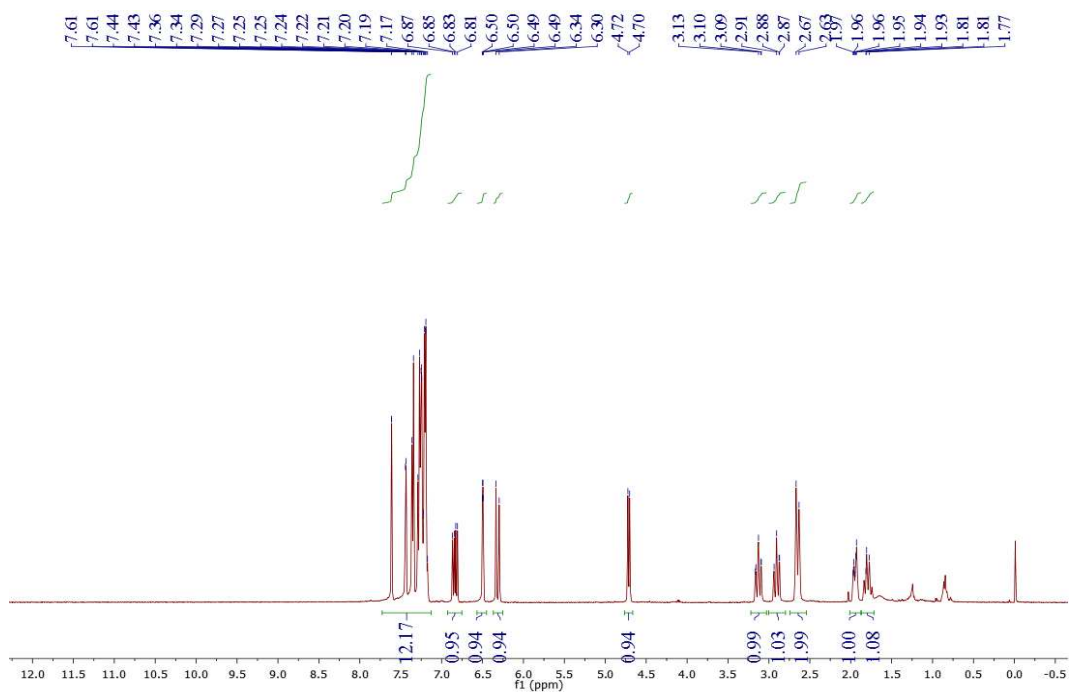
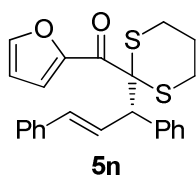
yk-cat-pro-tj-pcf3-1h-2

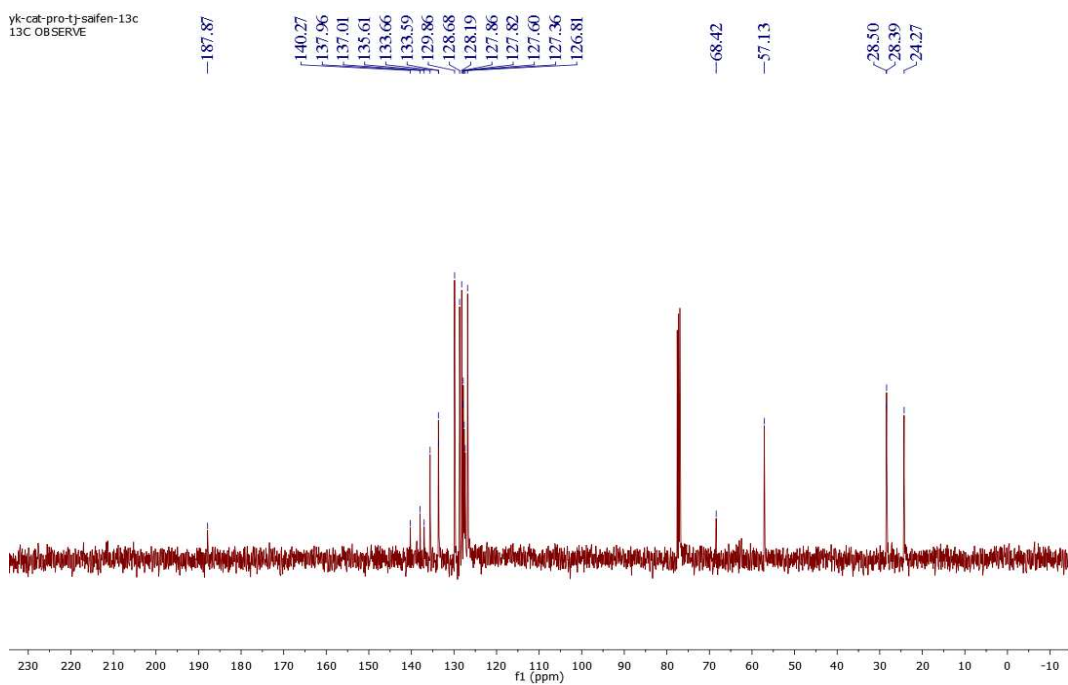
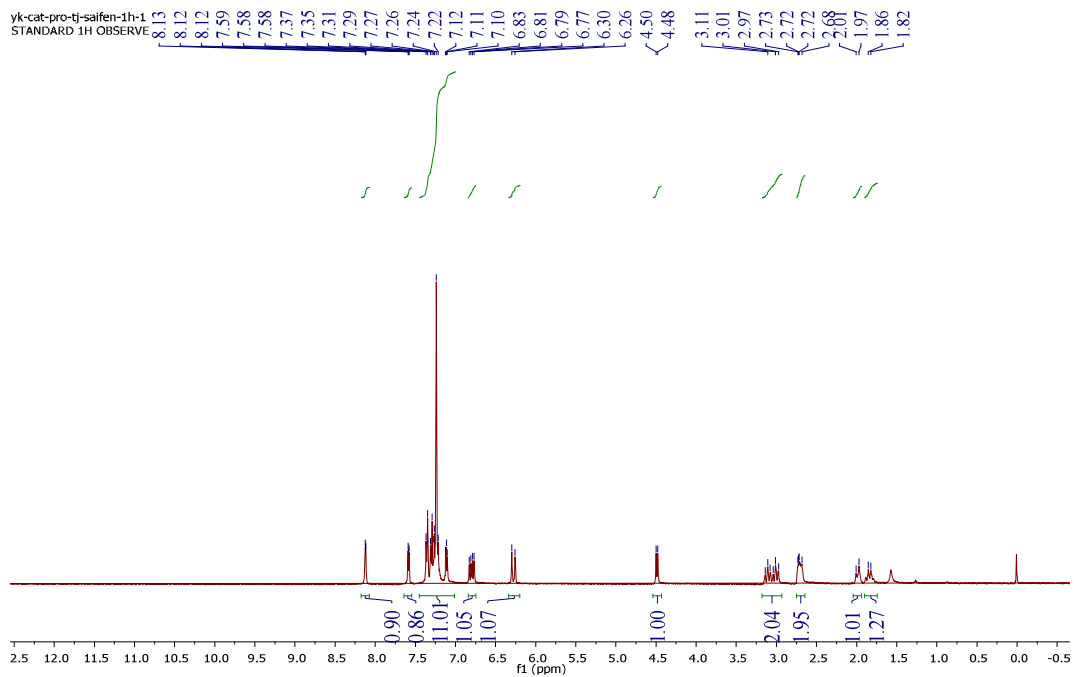
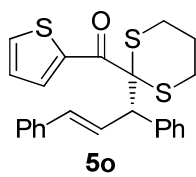


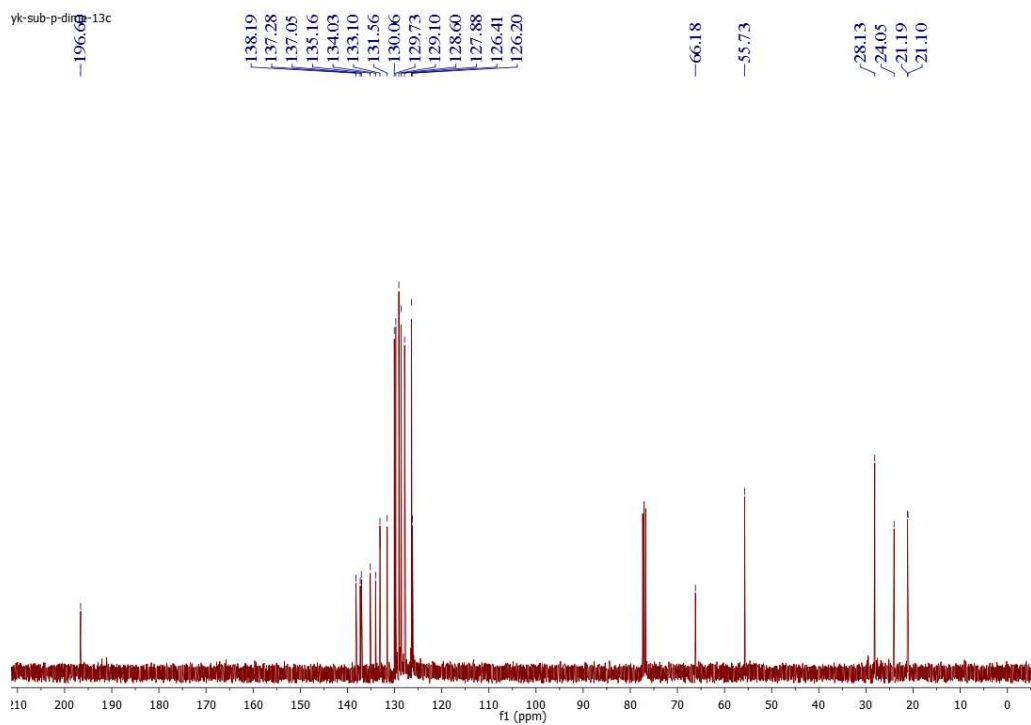
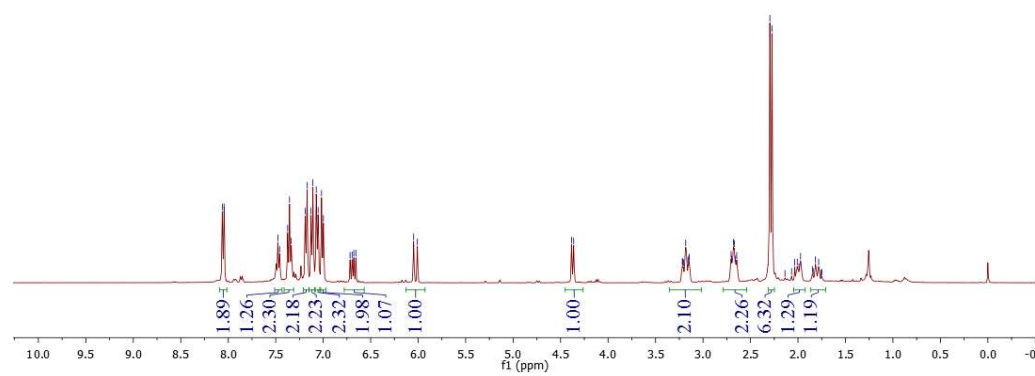
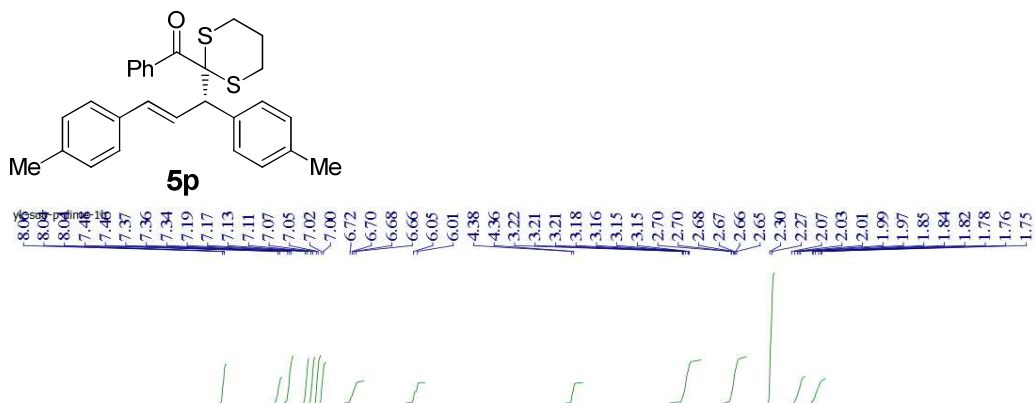
yk-cat-pro-ci-pcf3-13C
13C OBSERVE

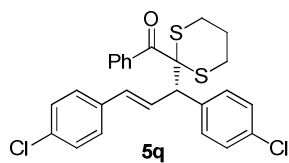




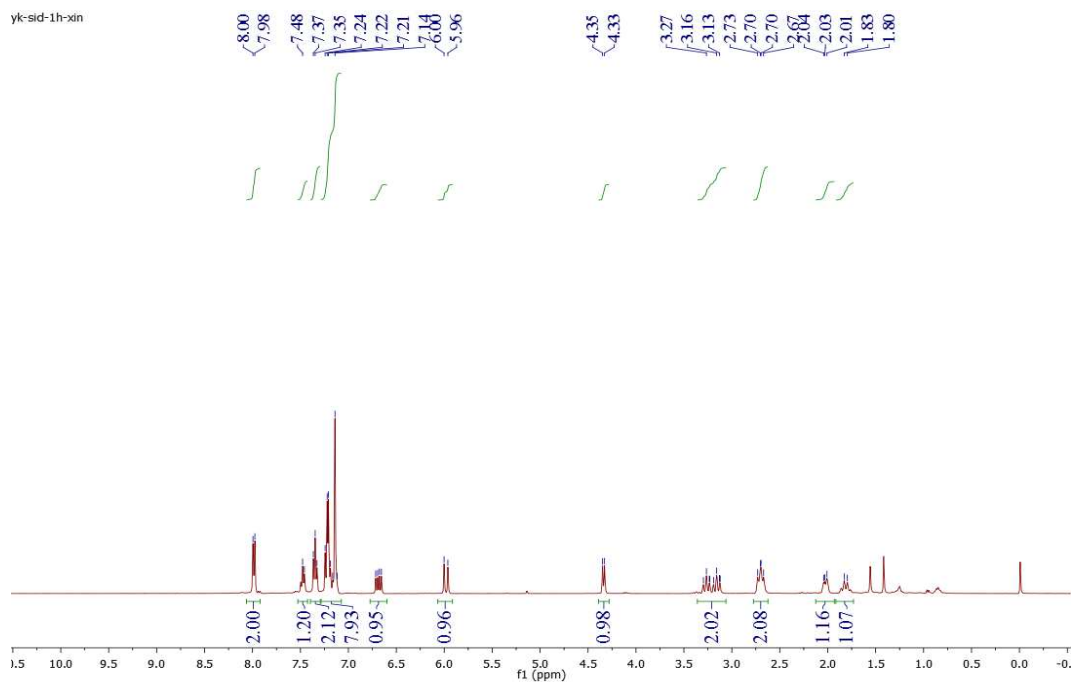




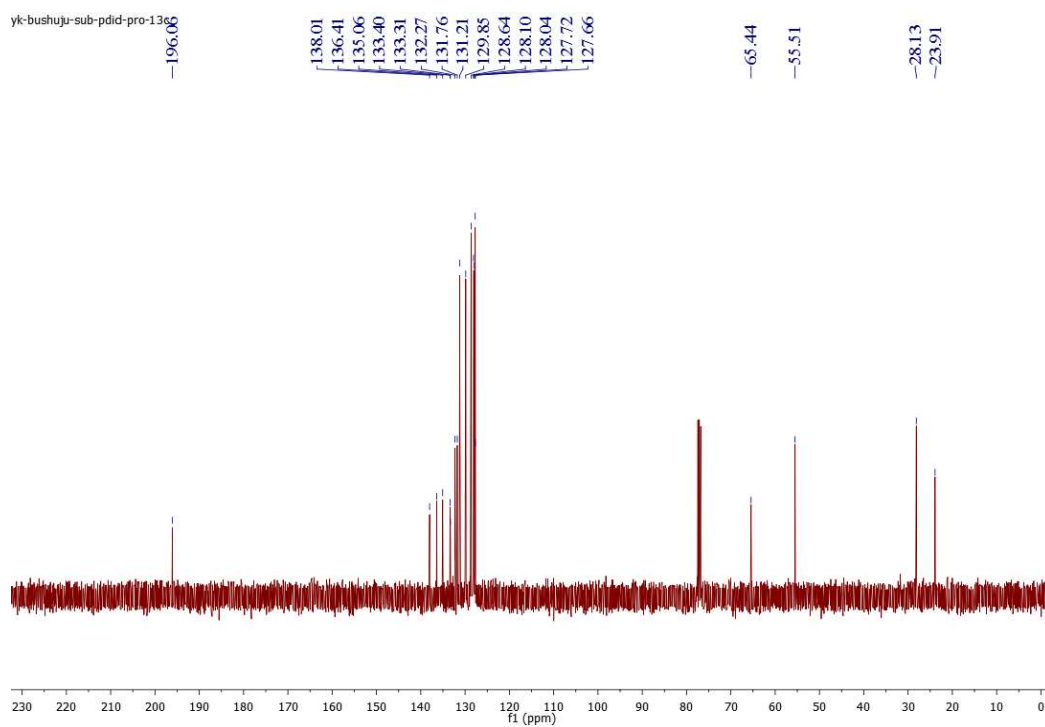


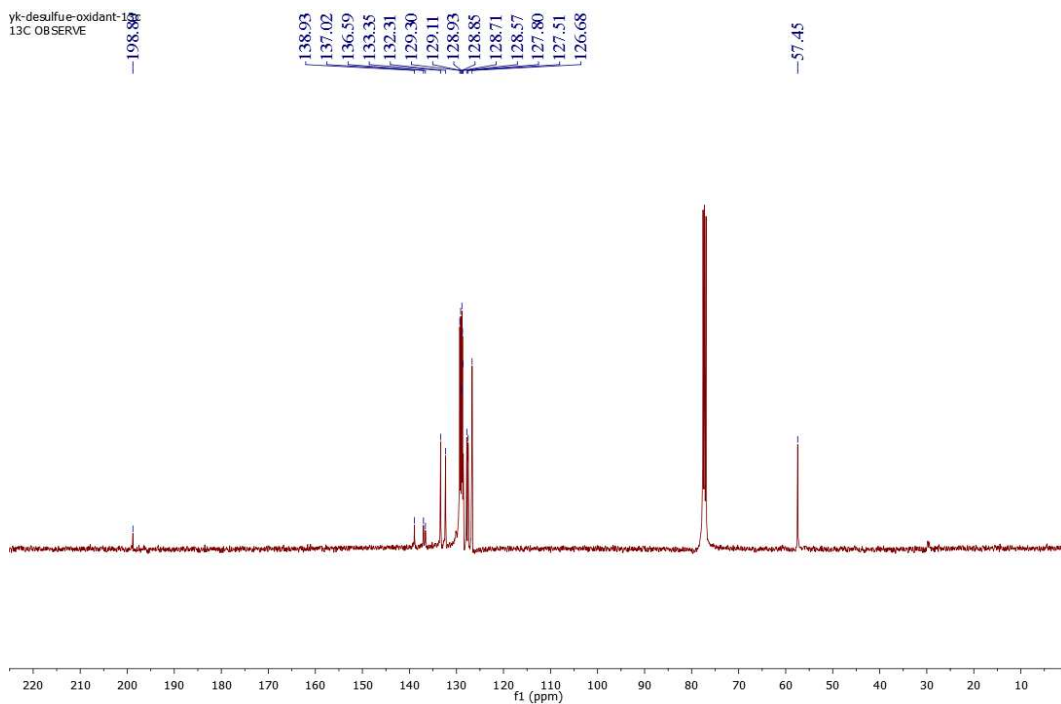
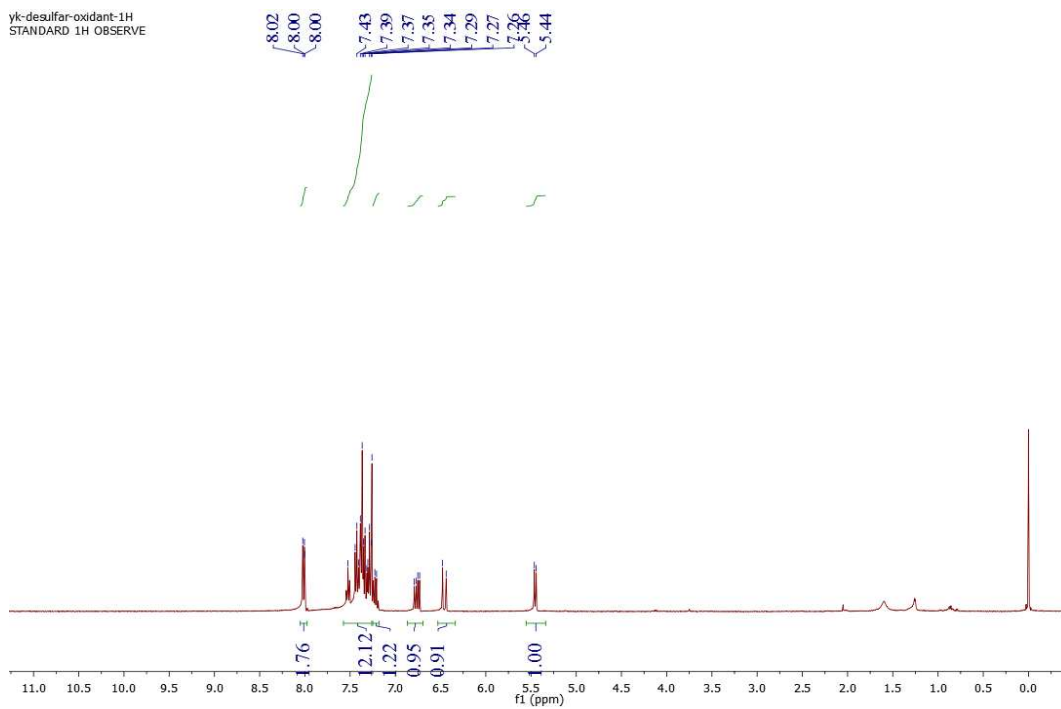
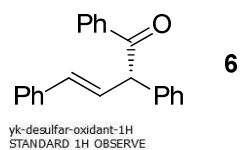


yk-sid-1h-xin

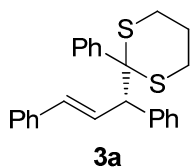


yk-bushuju-sub-pdid-pro-13



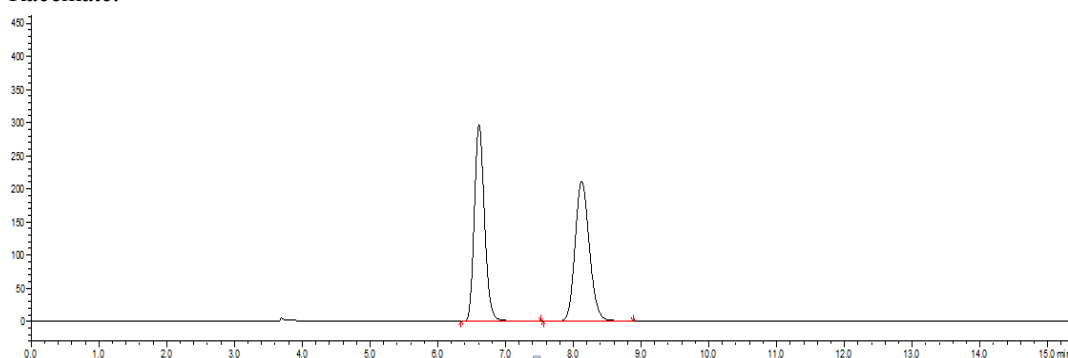


9. HPLC Data



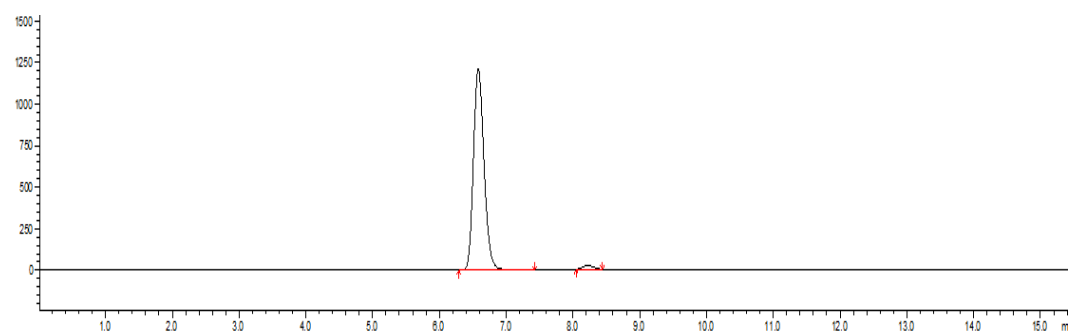
96% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak OD-H column, *n*-hexane/*i*-PrOH = 98/2, 1 mL/min, 254 nm.

Racemate:

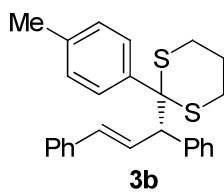


	Retention Time (min)	Area (%)
Peak 1	6.608	50.031
Peak 2	8.123	49.969

Chiral:

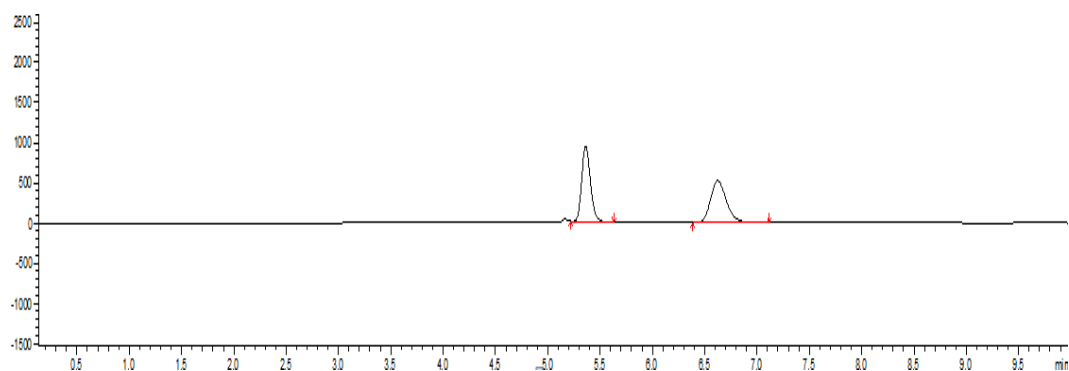


	Retention Time (min)	Area (%)
Peak 1	6.582	97.935
Peak 2	8.225	2.065



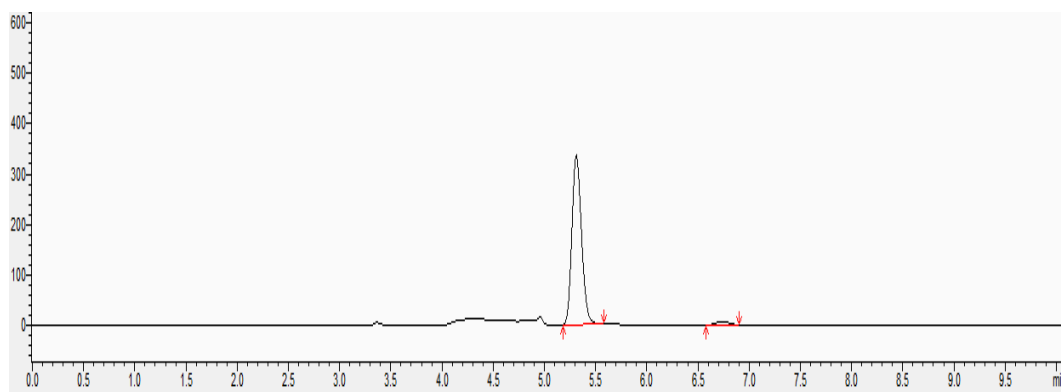
94% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak OD-H column, *n*-hexane/*i*-PrOH = 98/2, 1 mL/min, 254 nm.

Racemate:

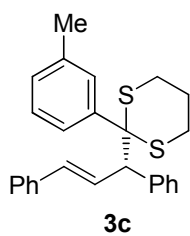


	Retention Time (min)	Area (%)
Peak 1	5.362	49.553
Peak 2	6.624	50.447

Chiral:

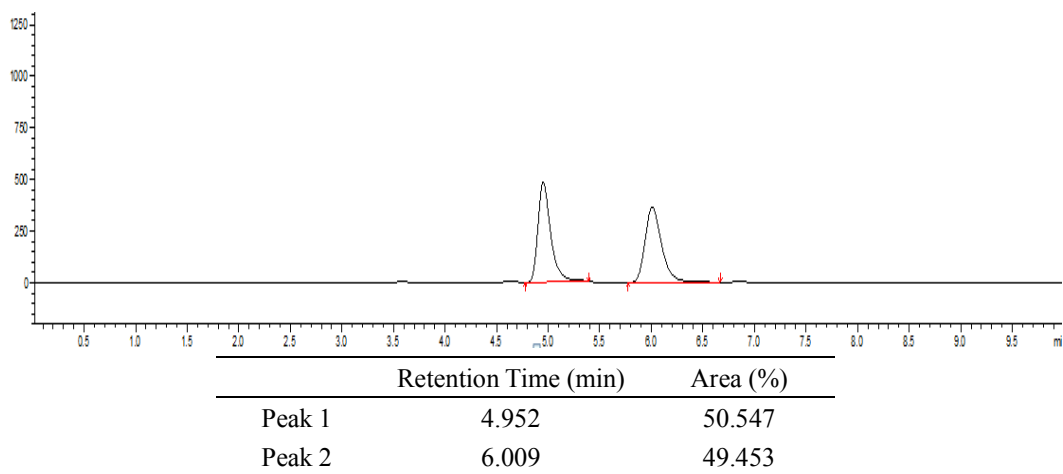


	Retention Time (min)	Area (%)
Peak 1	5.314	97.078
Peak 2	6.734	2.922

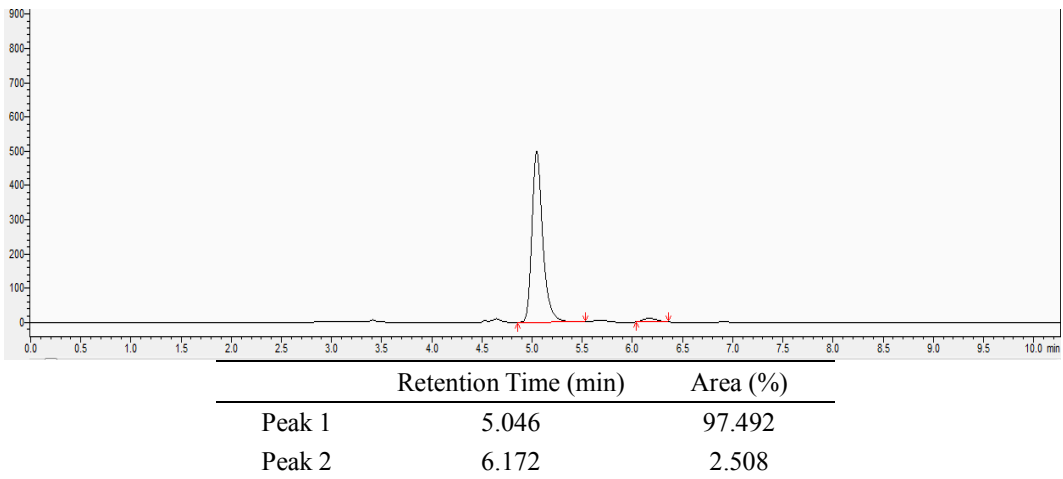


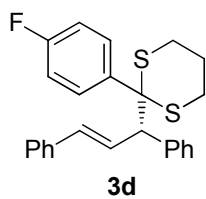
95% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak OD-H column, *n*-hexane/*i*-PrOH = 98/2, 1 mL/min, 254 nm.

Racemate:



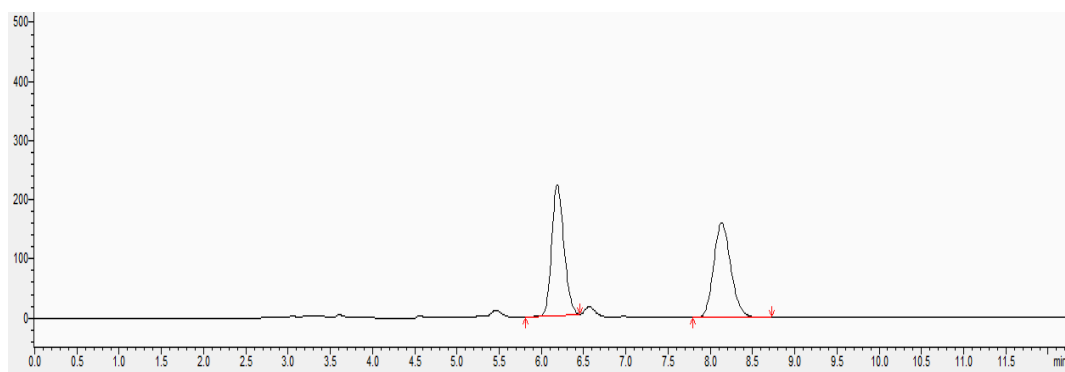
Chiral:





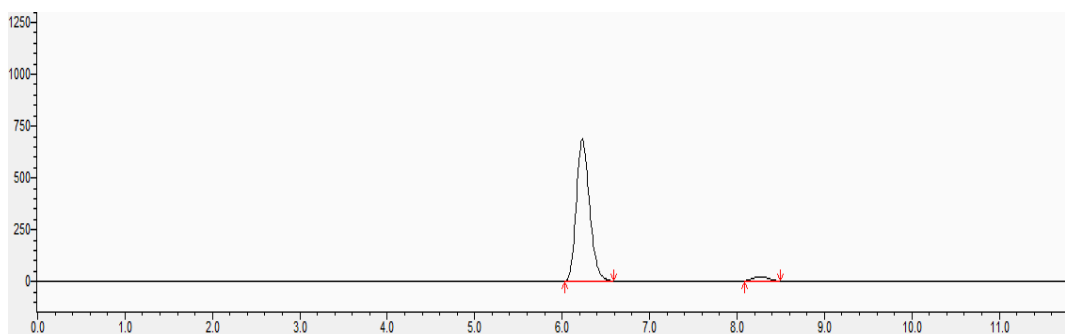
93% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 99/1, 0.7 mL/min, 254 nm.

Racemate:

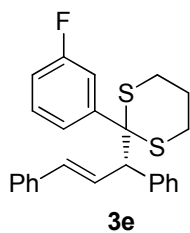


	Retention Time (min)	Area (%)
Peak 1	6.188	49.678
Peak 2	8.134	50.322

Chiral:

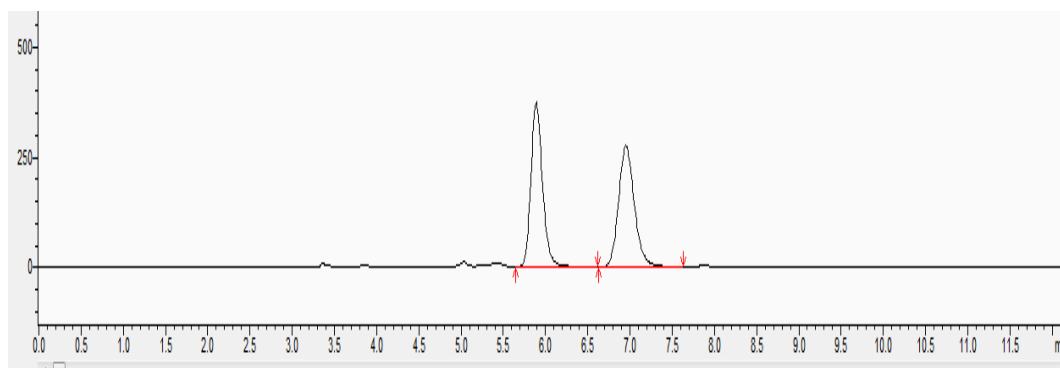


	Retention Time (min)	Area (%)
Peak 1	6.231	96.357
Peak 2	8.262	3.643



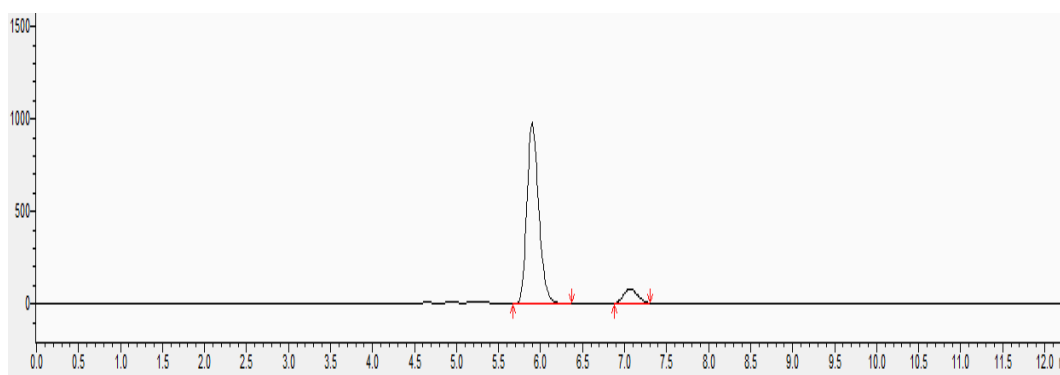
83% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak OD-H column, *n*-hexane/*i*-PrOH = 98/2, 1 mL/min, 254 nm.

Racemate:

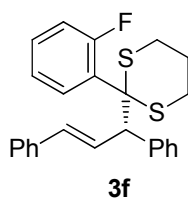


	Retention Time (min)	Area (%)
Peak 1	5.889	49.880
Peak 2	6.952	50.120

Chiral:

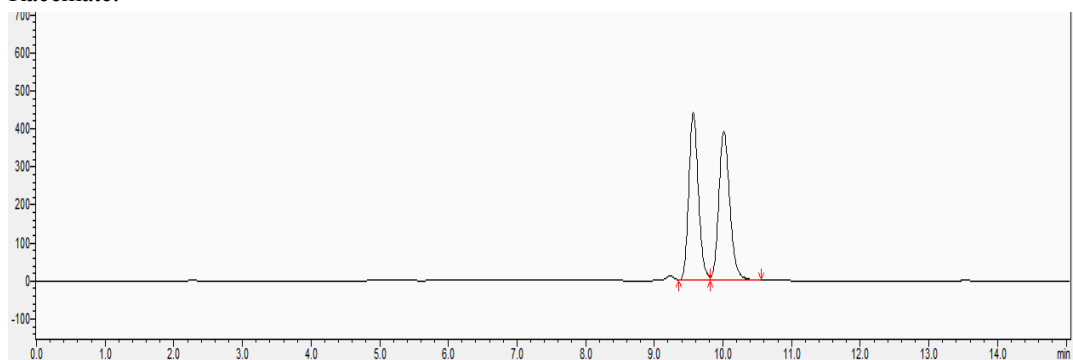


	Retention Time (min)	Area (%)
Peak 1	5.900	91.404
Peak 2	7.065	8.596



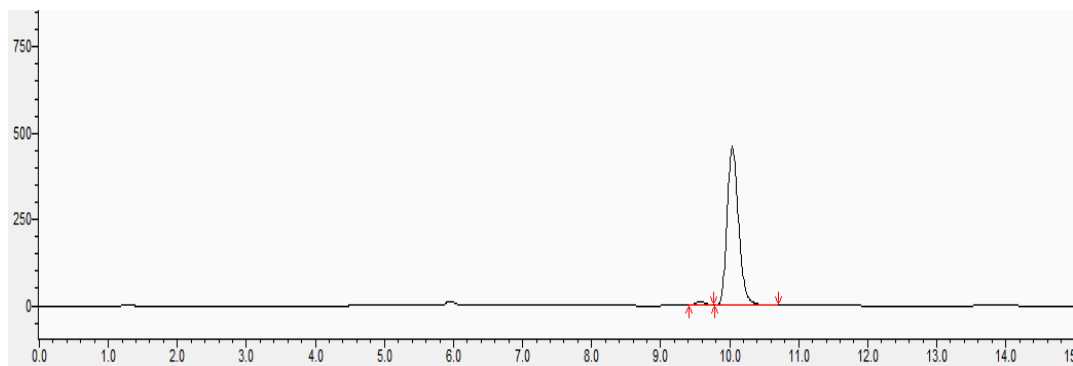
95% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 99/1, 0.7 mL/min, 254 nm.

Racemate:

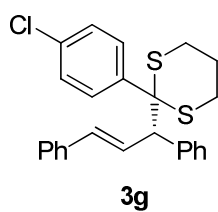


	Retention Time (min)	Area (%)
Peak 1	9.568	49.477
Peak 2	10.013	50.523

Chiral:

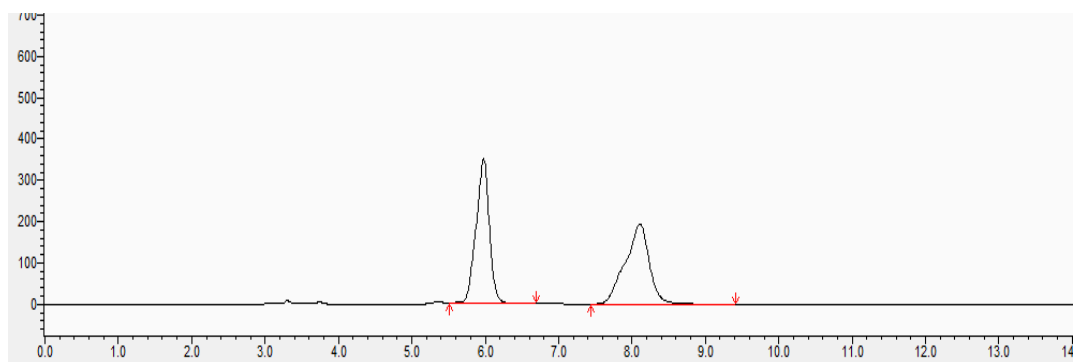


	Retention Time (min)	Area (%)
Peak 1	9.573	2.107
Peak 2	10.038	97.893



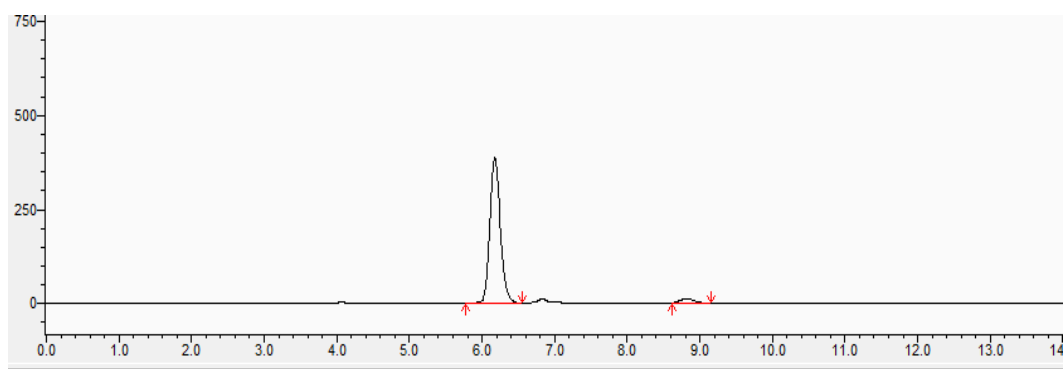
92% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak OD-H column, *n*-hexane/*i*-PrOH = 98/2, 1 mL/min, 254 nm.

Racemate:

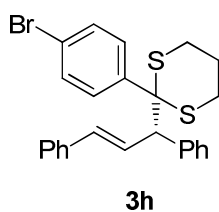


	Retention Time (min)	Area (%)
Peak 1	5.977	50.148
Peak 2	8.109	49.852

Chiral:

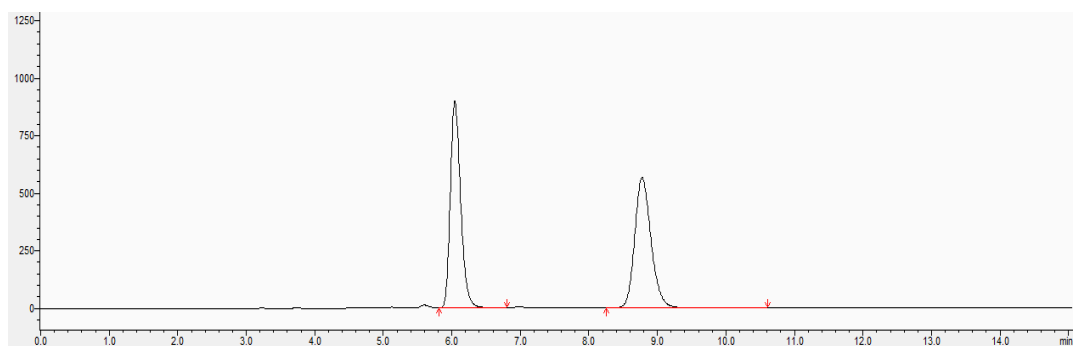


	Retention Time (min)	Area (%)
Peak 1	6.175	95.797
Peak 2	8.816	4.203



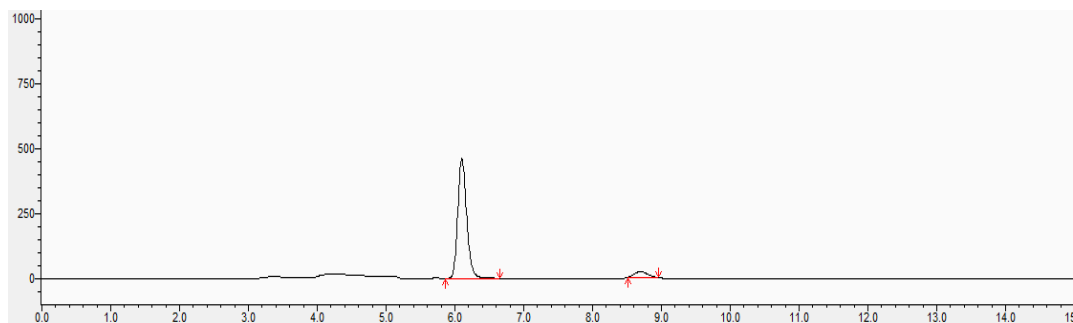
86% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak OD-H column, *n*-hexane/*i*-PrOH = 98/2, 1 mL/min, 254 nm.

Racemate:

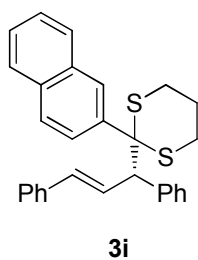


	Retention Time (min)	Area (%)
Peak 1	6.045	49.555
Peak 2	8.777	50.445

Chiral:

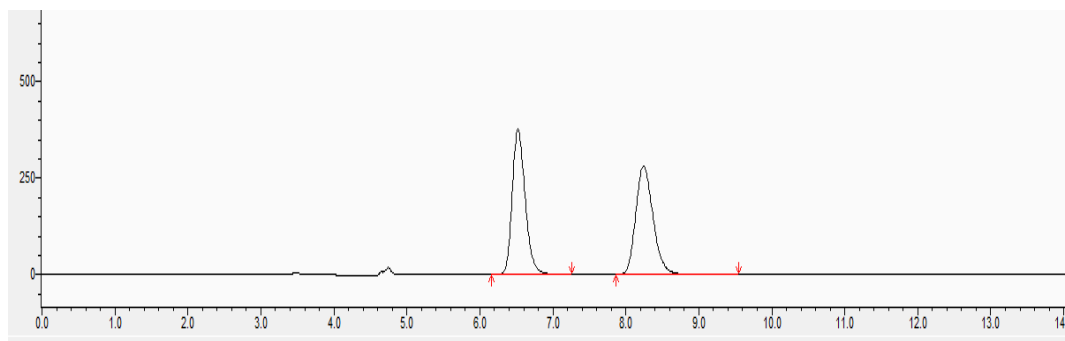


	Retention Time (min)	Area (%)
Peak 1	6.101	92.931
Peak 2	8.697	7.069



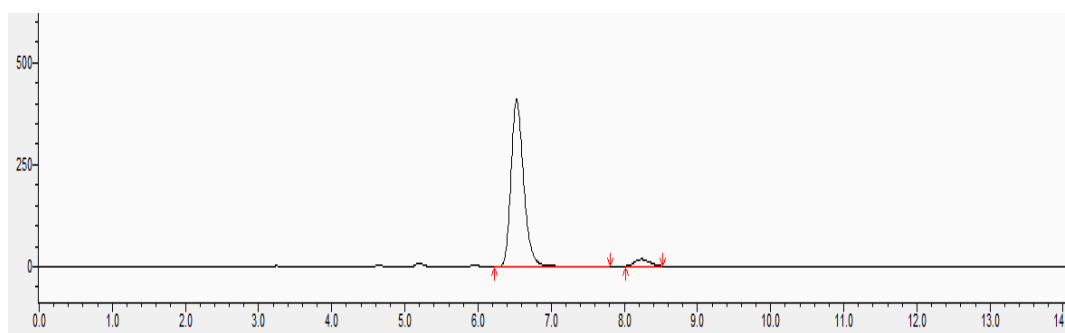
91% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak OD-H column, *n*-hexane /*i*-PrOH = 98/2, 1 mL/min, 254 nm.

Racemate:

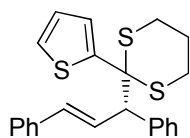


	Retention Time (min)	Area (%)
Peak 1	6.522	49.910
Peak 2	8.243	50.090

Chiral:



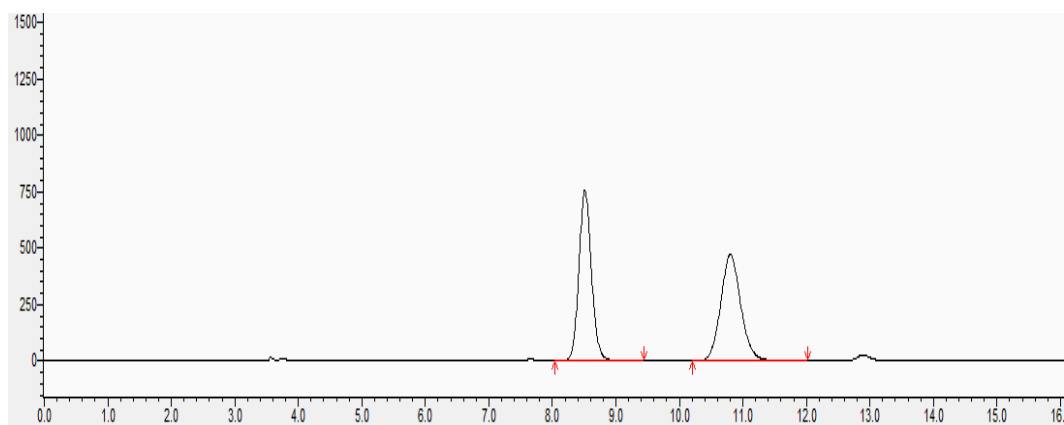
	Retention Time (min)	Area (%)
Peak 1	6.528	95.354
Peak 2	8.237	4.646



3j

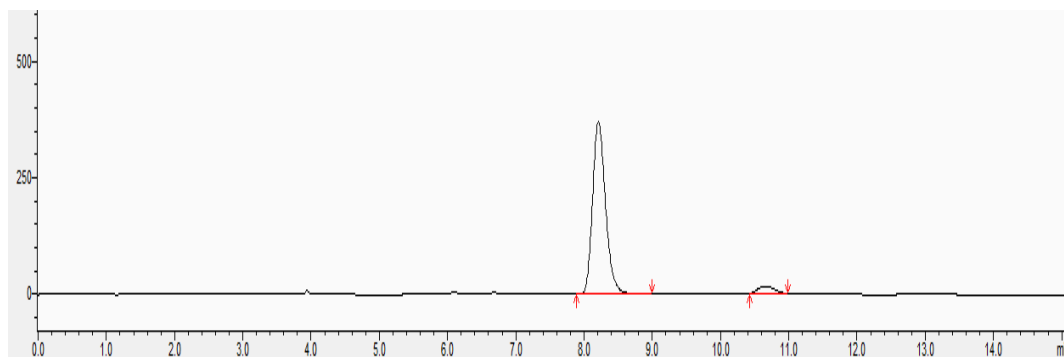
90% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak OD-H column, *n*-hexane /*i*-PrOH = 98/2, 1 mL/min, 254 nm.

Racemate:

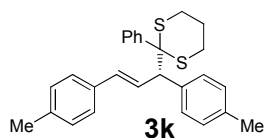


	Retention Time (min)	Area (%)
Peak 1	8.511	49.907
Peak 2	10.803	50.093

Chiral:

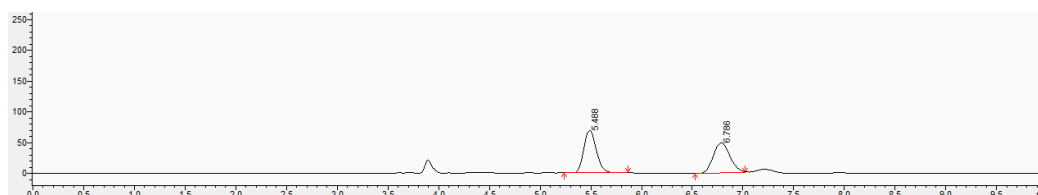


	Retention Time (min)	Area (%)
Peak 1	8.212	94.794
Peak 2	10.660	5.206



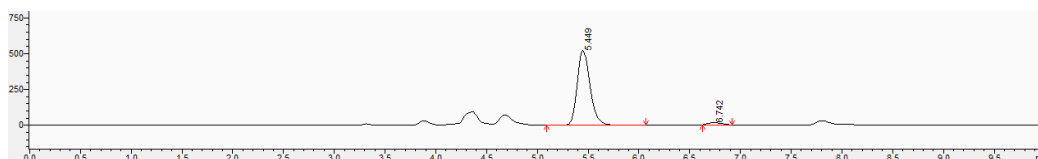
94% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak OD-H column, *n*-hexane /*i*-PrOH = 98/2, 1 mL/min, 254 nm.

Racemate:

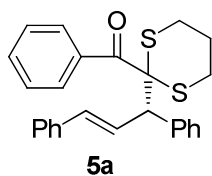


	Retention Time (min)	Area (%)
Peak 1	5.488	50.111
Peak 2	6.786	49.889

Chiral:

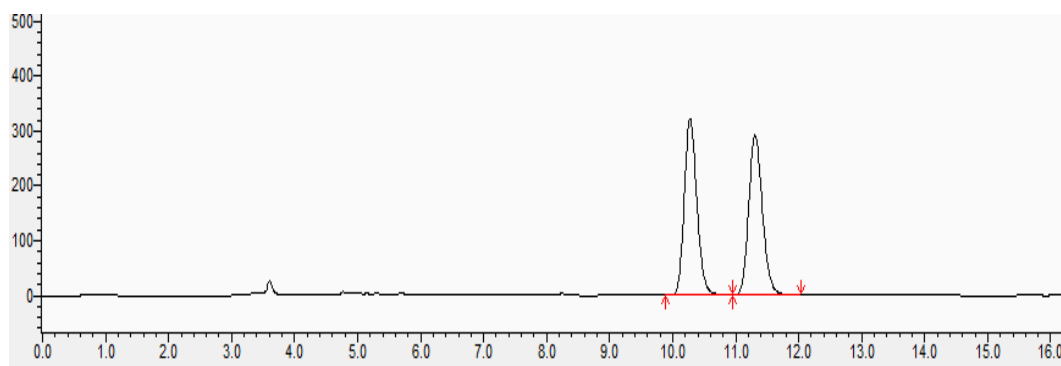


	Retention Time (min)	Area (%)
Peak 1	5.449	97.082
Peak 2	6.742	2.918



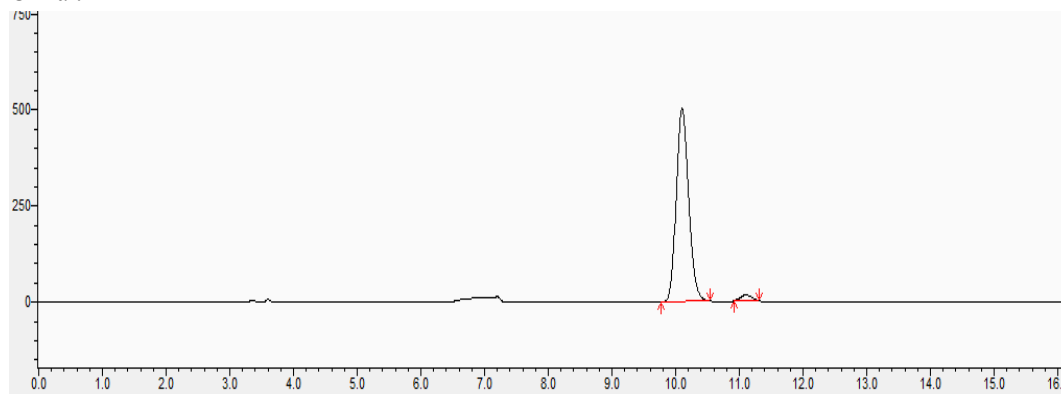
95% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column,
n-hexane /*i*-PrOH = 98/2, 1 mL/min, 254 nm.

Racemate:

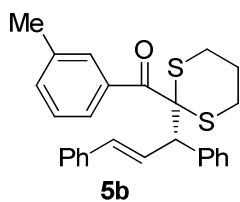


	Retention Time (min)	Area (%)
Peak 1	10.277	50.184
Peak 2	11.308	49.816

Chiral:

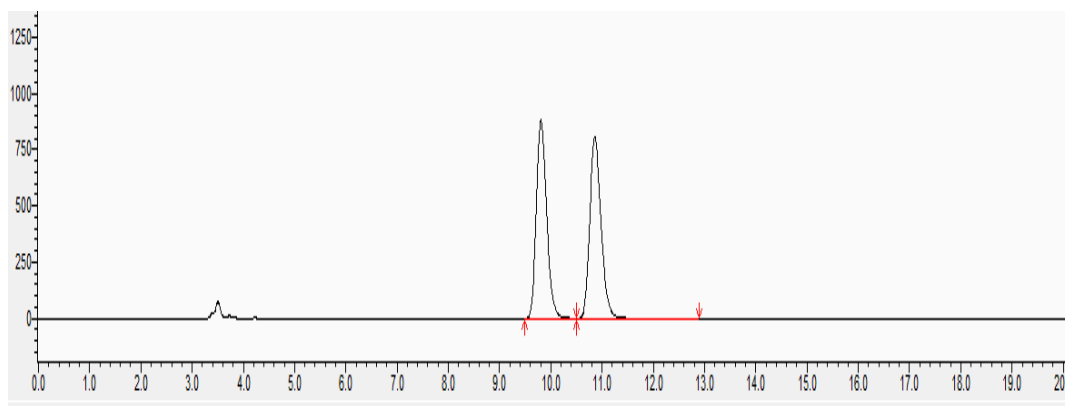


	Retention Time (min)	Area (%)
Peak 1	10.099	97.333
Peak 2	11.103	2.667



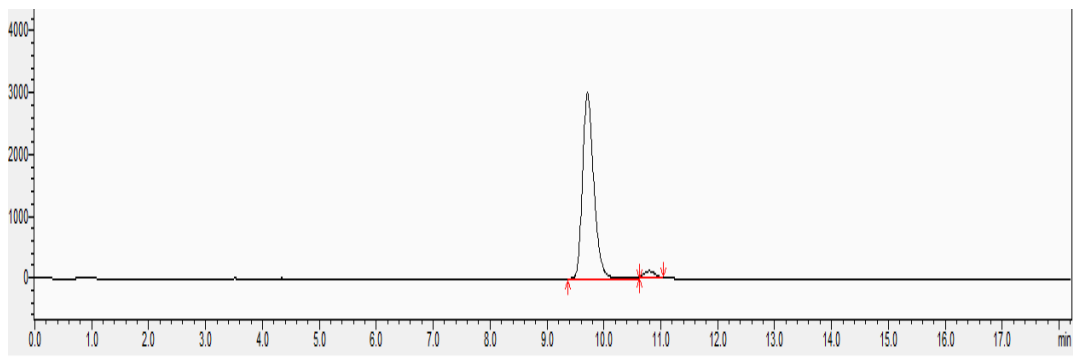
94% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column,
n-hexane /*i*-PrOH = 98/2, 1 mL/min, 254 nm.

Racemate:

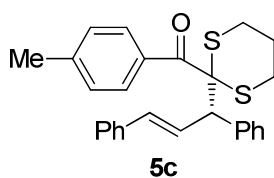


	Retention Time (min)	Area (%)
Peak 1	9.807	49.866
Peak 2	10.859	50.134

Chiral:

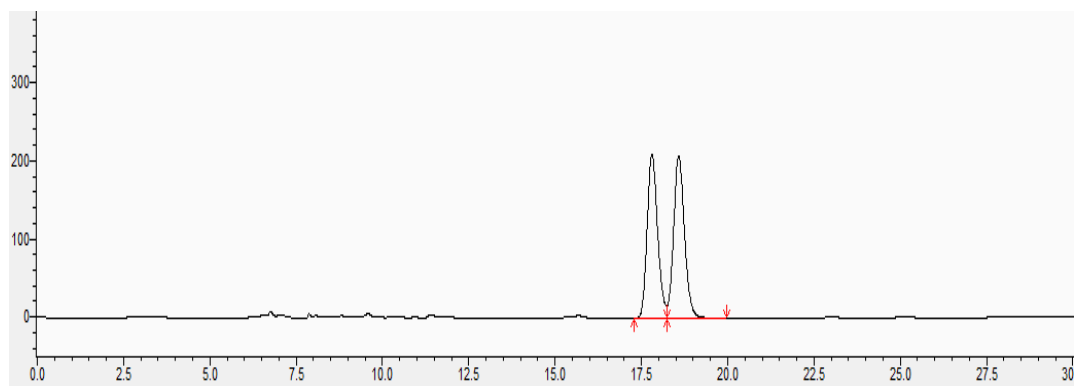


	Retention Time (min)	Area (%)
Peak 1	9.711	96.911
Peak 2	10.795	3.089



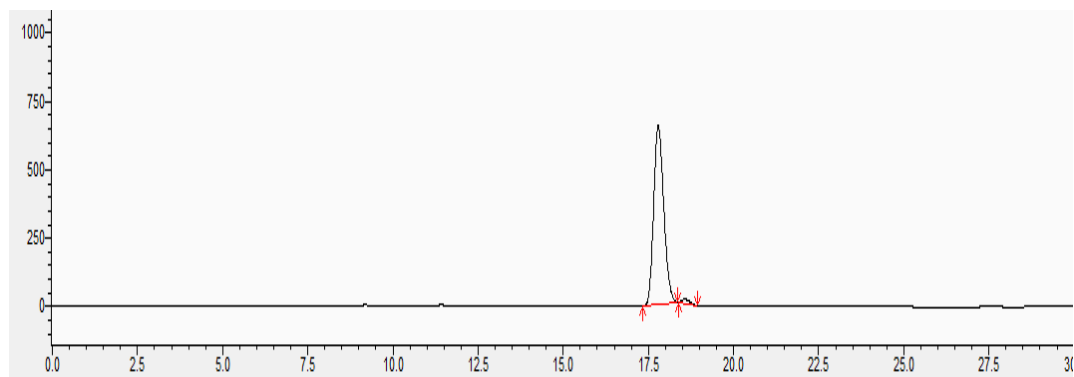
96% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column, *n*-hexane /*i*-PrOH = 97/3, 0.5 mL/min, 254 nm.

Racemate:

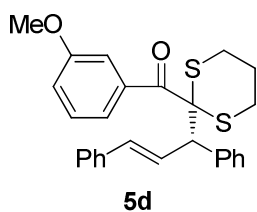


	Retention Time (min)	Area (%)
Peak 1	17.806	49.197
Peak 2	18.581	50.803

Chiral:

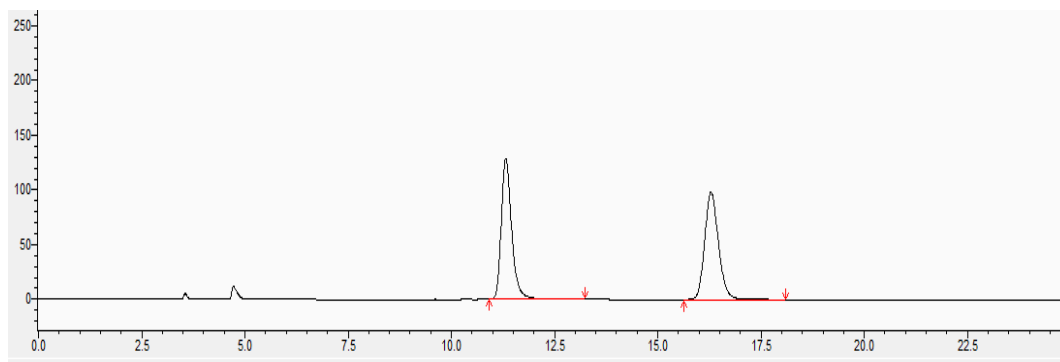


	Retention Time (min)	Area (%)
Peak 1	17.792	98.204
Peak 2	18.570	1.796



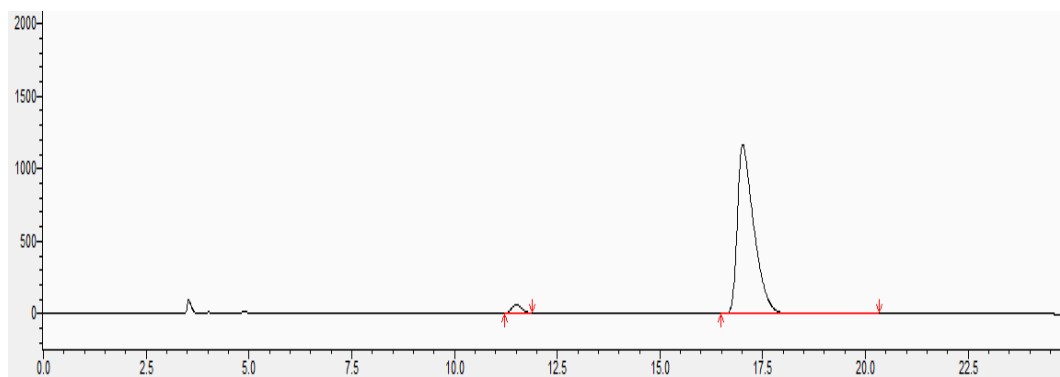
94% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IE column,
n-hexane /*i*-PrOH = 95/5, 1 mL/min, 254 nm.

Racemate:

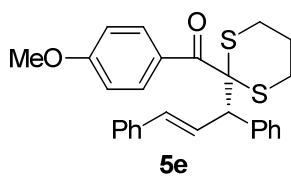


	Retention Time (min)	Area (%)
Peak 1	11.319	49.840
Peak 2	16.288	50.160

Chiral:

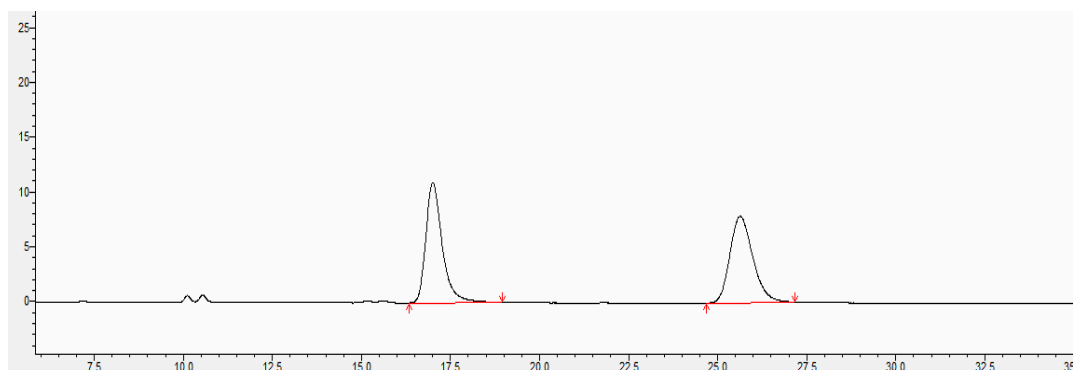


	Retention Time (min)	Area (%)
Peak 1	11.506	3.090
Peak 2	17.013	96.910



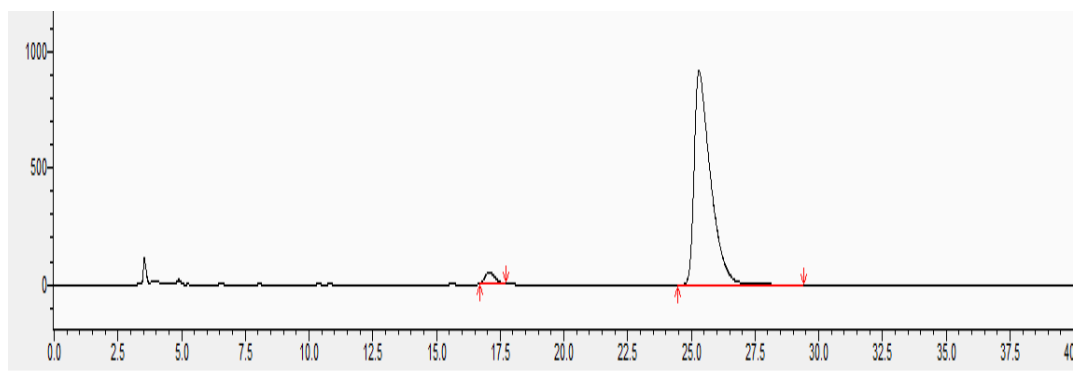
94% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IE column, *n*-hexane /*i*-PrOH = 95/5, 1 mL/min, 254 nm.

Racemate:

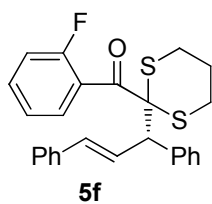


	Retention Time (min)	Area (%)
Peak 1	17.005	49.967
Peak 2	25.629	50.033

Chiral:

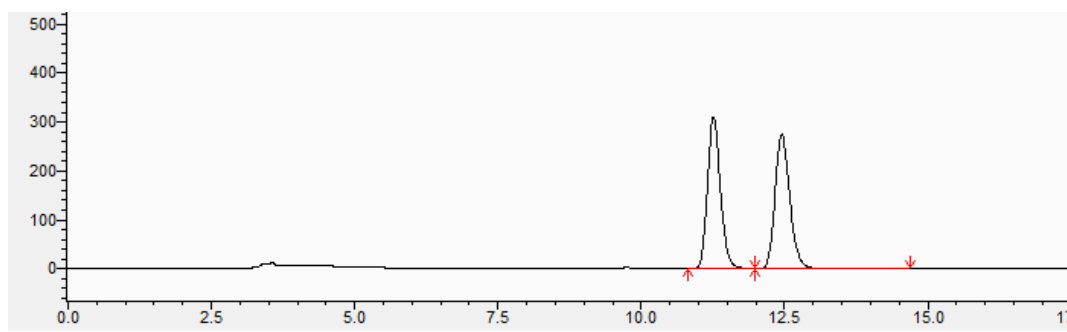


	Retention Time (min)	Area (%)
Peak 1	17.077	3.170
Peak 2	25.302	96.830



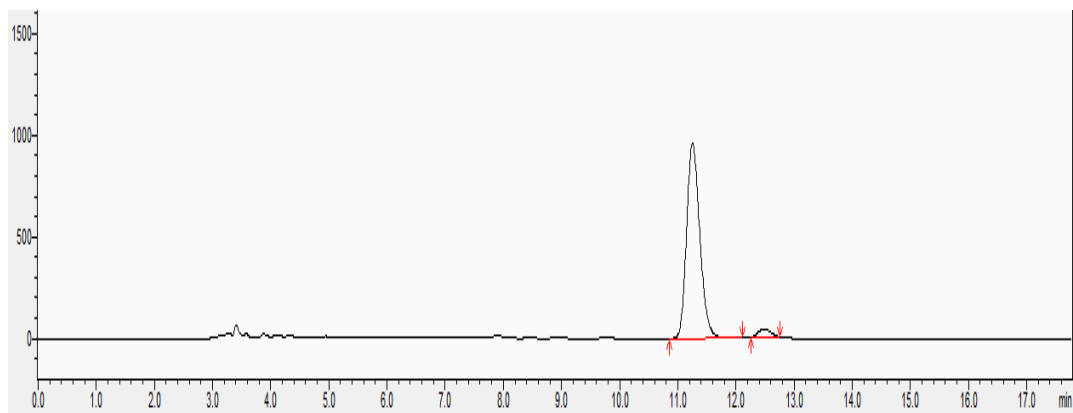
92% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column, *n*-hexane /*i*-PrOH = 97/3, 1 mL/min, 254 nm.

Racemate:

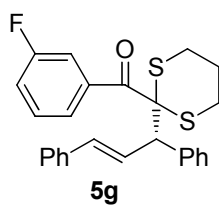


	Retention Time (min)	Area (%)
Peak 1	11.260	49.862
Peak 2	12.452	50.138

Chiral:

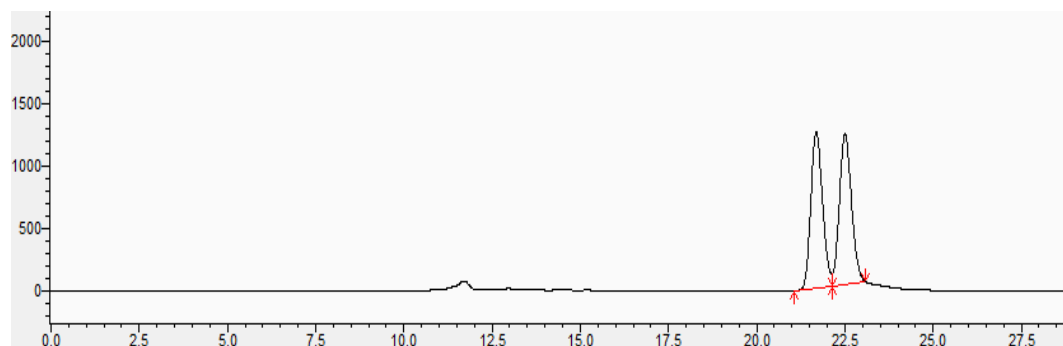


	Retention Time (min)	Area (%)
Peak 1	11.254	96.060
Peak 2	12.486	3.940



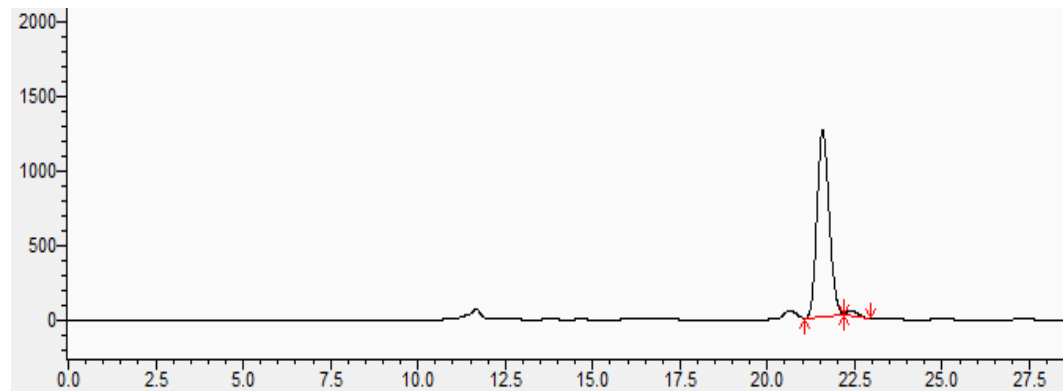
95% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column, *n*-hexane /*i*-PrOH = 98/2, 0.3 mL/min, 254 nm.

Racemate:

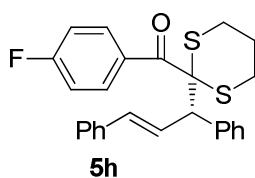


	Retention Time (min)	Area (%)
Peak 1	21.688	49.720
Peak 2	22.503	50.280

Chiral:

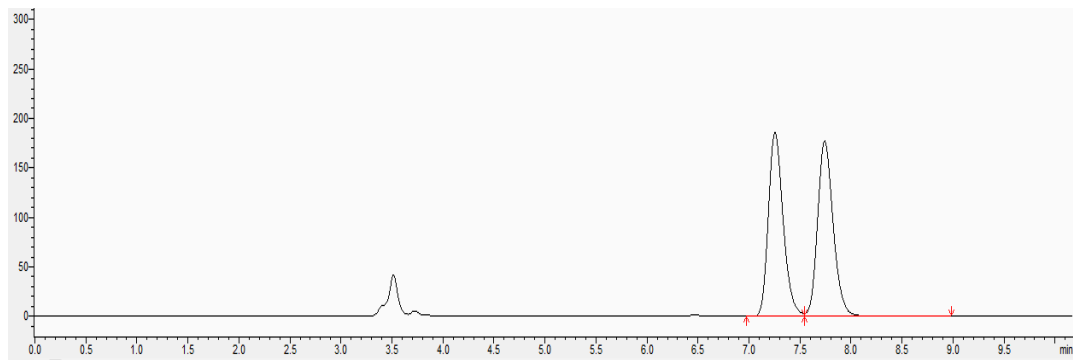


	Retention Time (min)	Area (%)
Peak 1	21.579	97.348
Peak 2	22.404	2.652



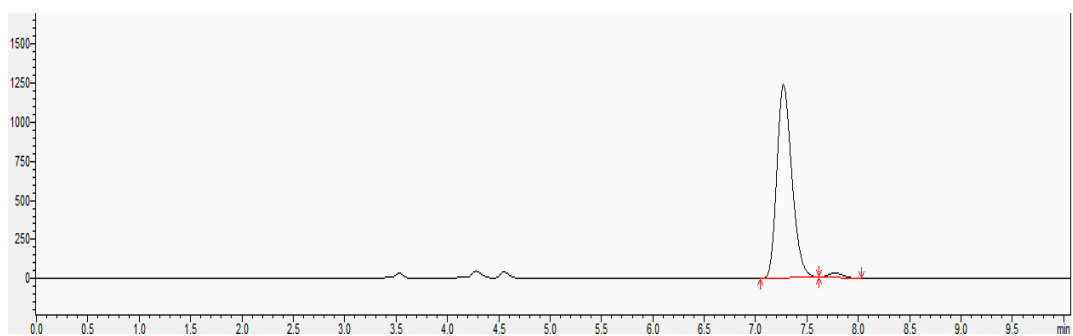
95% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column, *n*-hexane /*i*-PrOH = 98/2, 1 mL/min, 254 nm.

Racemate:

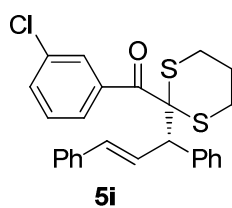


	Retention Time (min)	Area (%)
Peak 1	7.254	49.700
Peak 2	7.739	50.300

Chiral:

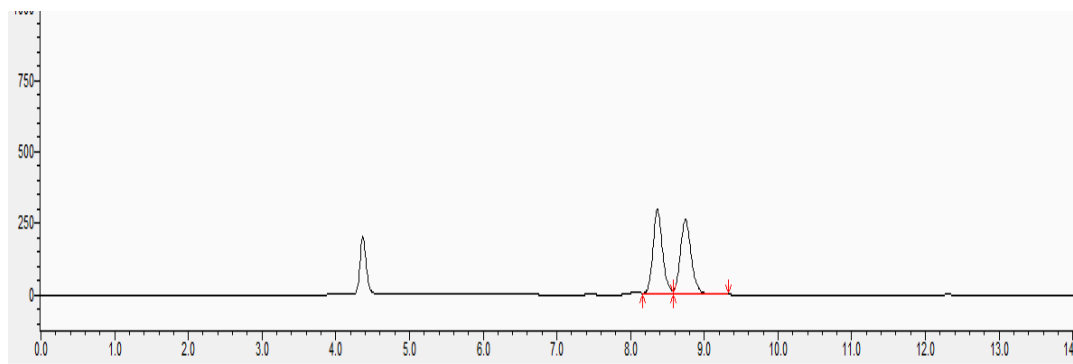


	Retention Time (min)	Area (%)
Peak 1	7.270	97.654
Peak 2	7.771	2.346



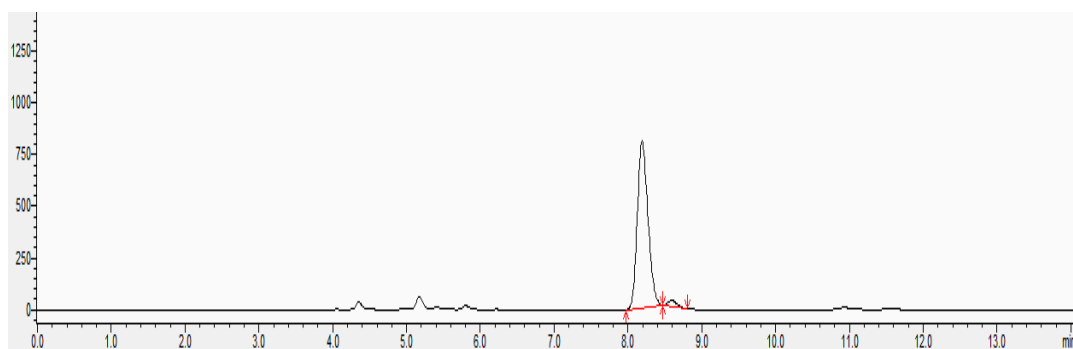
94% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column, *n*-hexane /*i*-PrOH = 98/2, 0.8 mL/min, 254 nm.

Racemate:

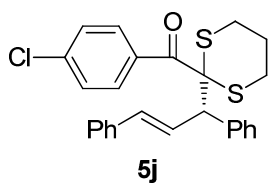


	Retention Time (min)	Area (%)
Peak 1	8.364	50.038
Peak 2	8.743	49.962

Chiral:

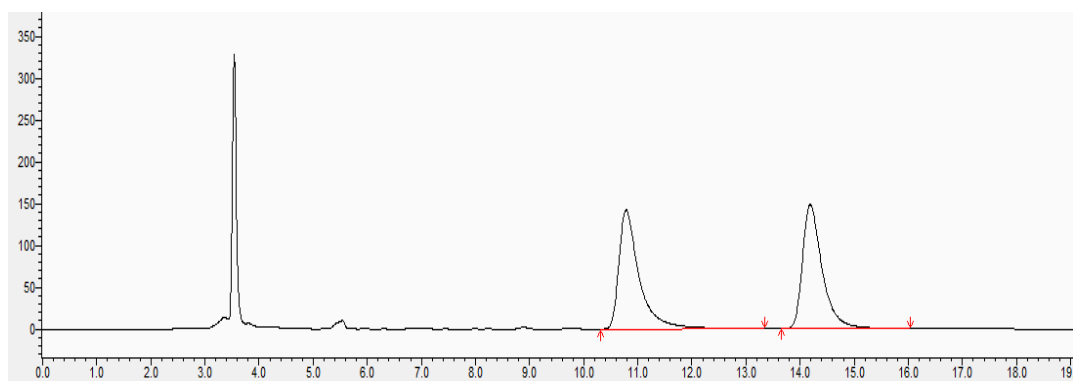


	Retention Time (min)	Area (%)
Peak 1	8.193	96.828
Peak 2	8.592	3.172



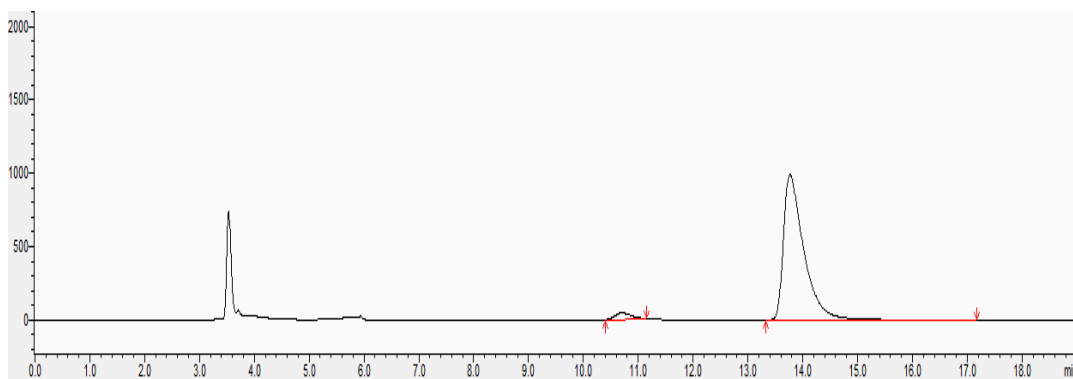
93% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IE column,
n-hexane /*i*-PrOH = 98/2, 1 mL/min, 254 nm.

Racemate:

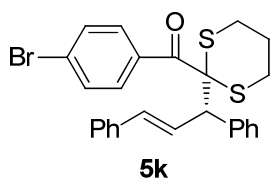


	Retention Time (min)	Area (%)
Peak 1	10.784	49.801
Peak 2	14.186	50.199

Chiral:

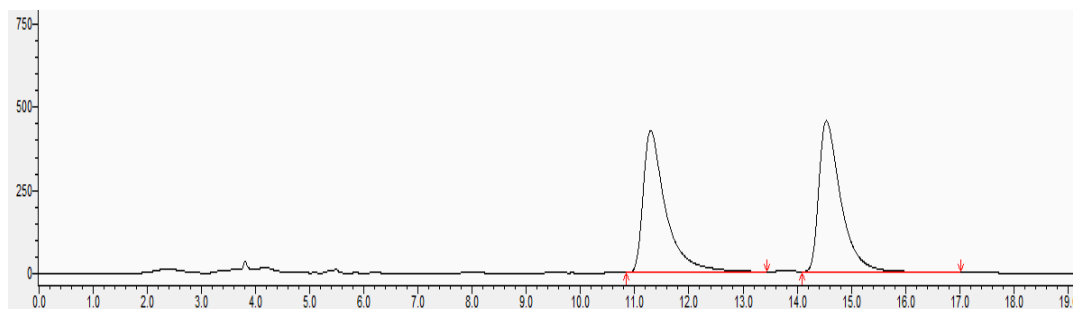


	Retention Time (min)	Area (%)
Peak 1	10.706	3.411
Peak 2	13.768	96.589



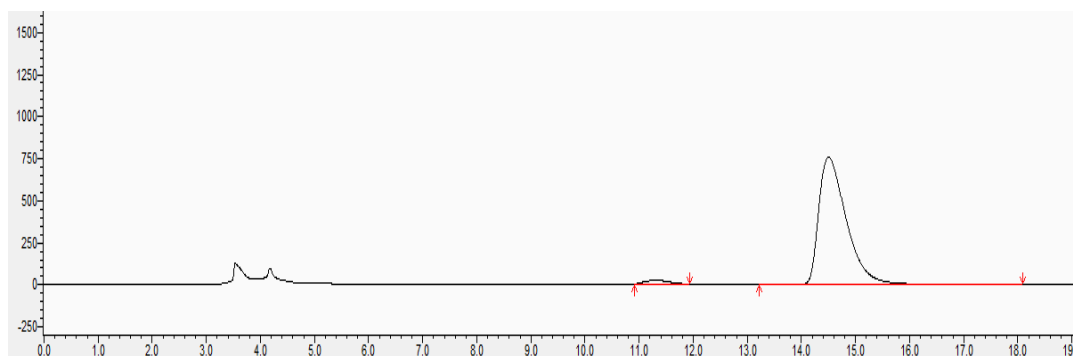
95% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IE column,
n-hexane /*i*-PrOH = 98/2, 1 mL/min, 254 nm.

Racemate:

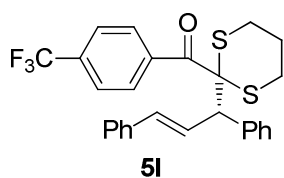


	Retention Time (min)	Area (%)
Peak 1	11.300	49.241
Peak 2	14.538	50.759

Chiral:

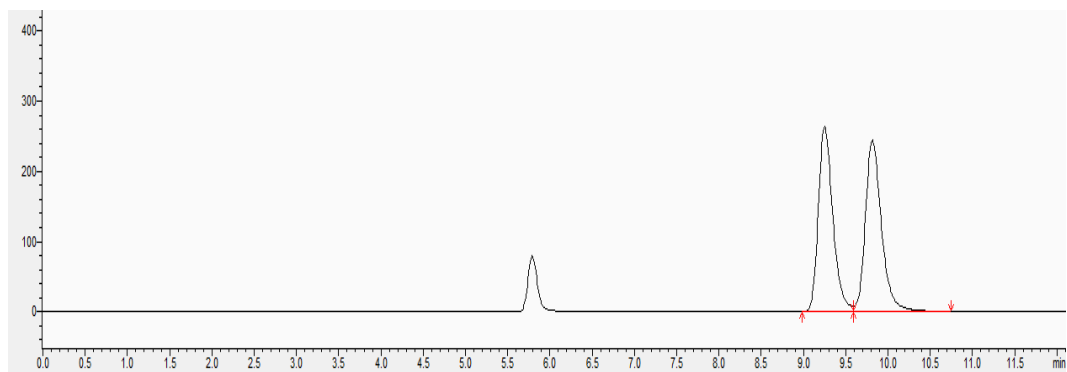


	Retention Time (min)	Area (%)
Peak 1	11.309	2.530
Peak 2	14.503	97.470



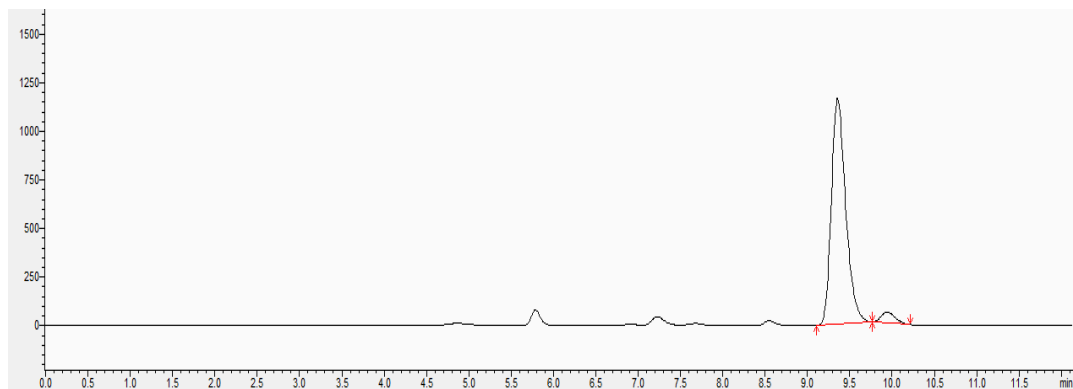
91% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column, *n*-hexane /*i*-PrOH = 99/1, 0.6 mL/min, 254 nm.

Racemate:

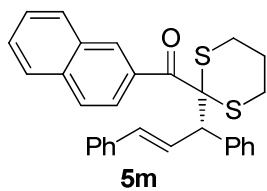


	Retention Time (min)	Area (%)
Peak 1	9.251	49.240
Peak 2	9.816	50.760

Chiral:

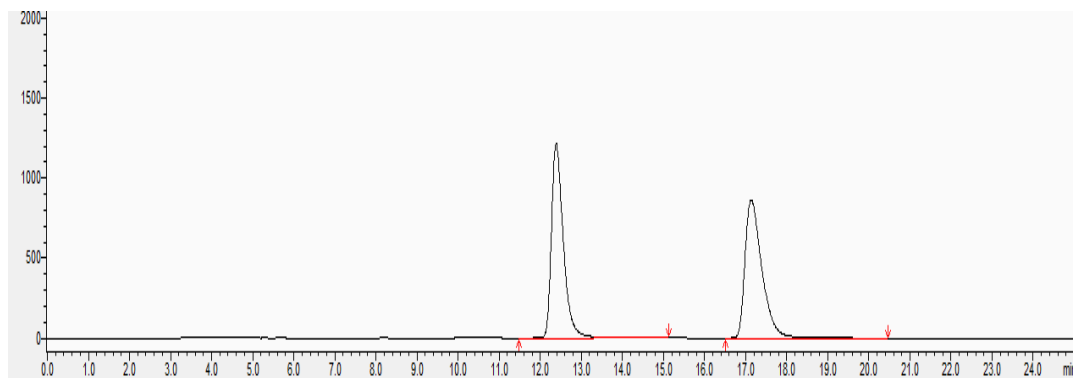


	Retention Time (min)	Area (%)
Peak 1	9.356	95.482
Peak 2	9.942	4.518



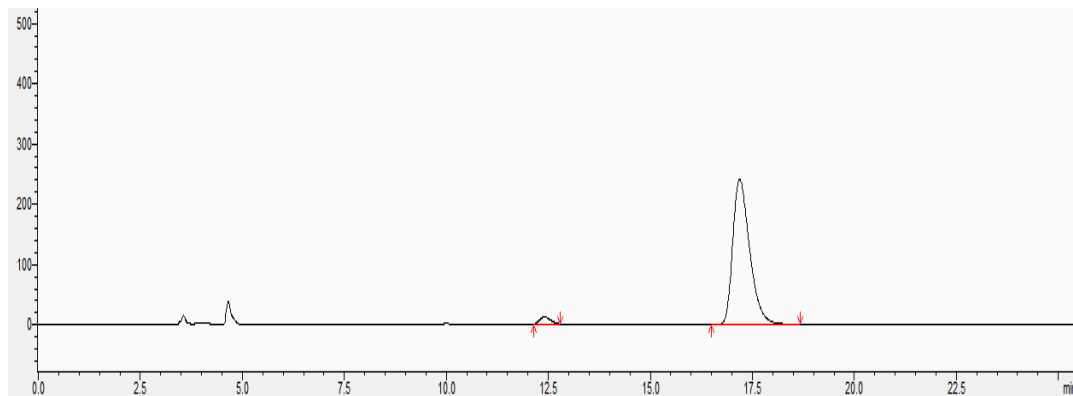
94% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IE column, *n*-hexane /*i*-PrOH = 95/5, 1 mL/min, 254 nm.

Racemate:

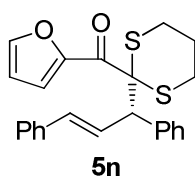


	Retention Time (min)	Area (%)
Peak 1	12.388	49.886
Peak 2	17.136	50.114

Chiral:

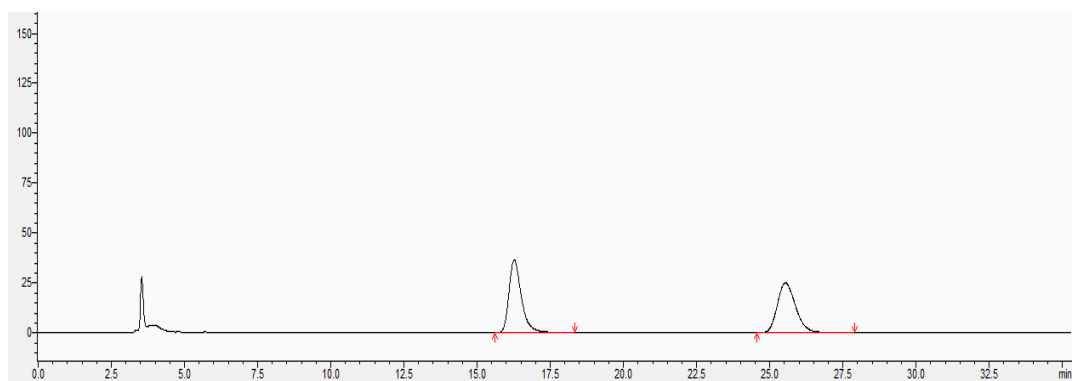


	Retention Time (min)	Area (%)
Peak 1	12.410	3.033
Peak 2	17.190	96.967



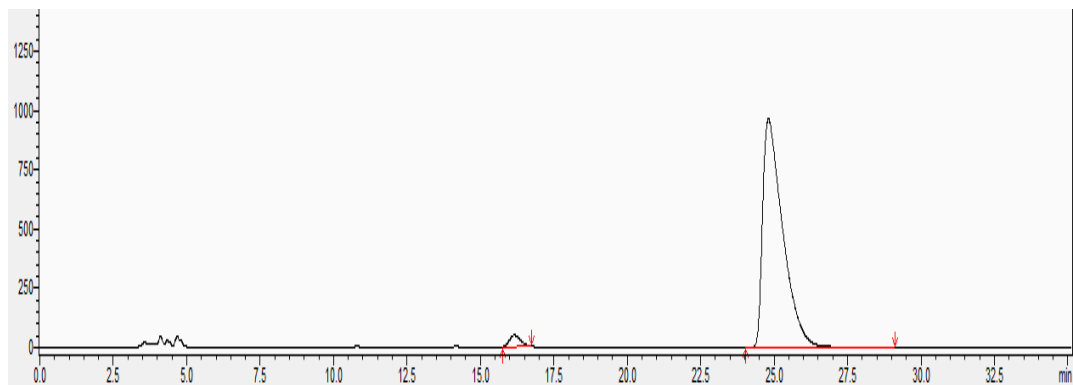
95% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IE column,
n-hexane /*i*-PrOH = 95/5, 1 mL/min, 254 nm.

Racemate:

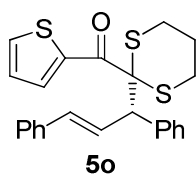


	Retention Time (min)	Area (%)
Peak 1	16.267	49.826
Peak 2	25.539	50.174

Chiral:

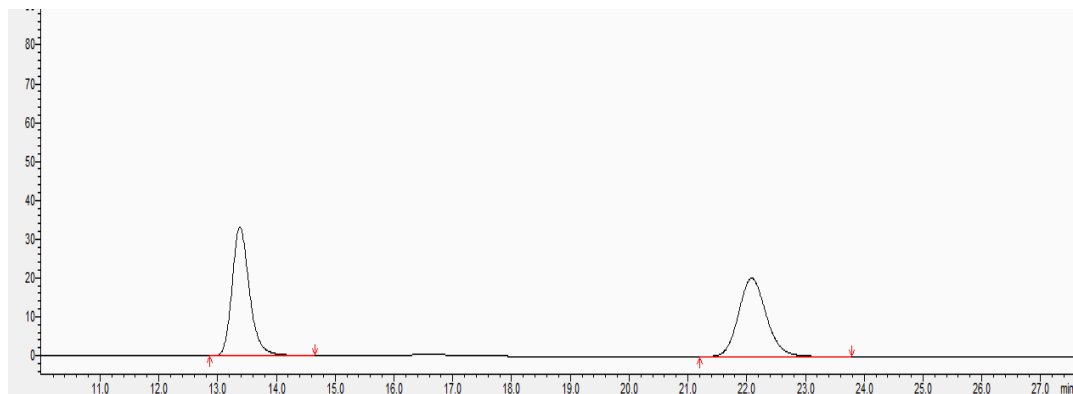


	Retention Time (min)	Area (%)
Peak 1	16.160	2.714
Peak 2	24.806	97.286



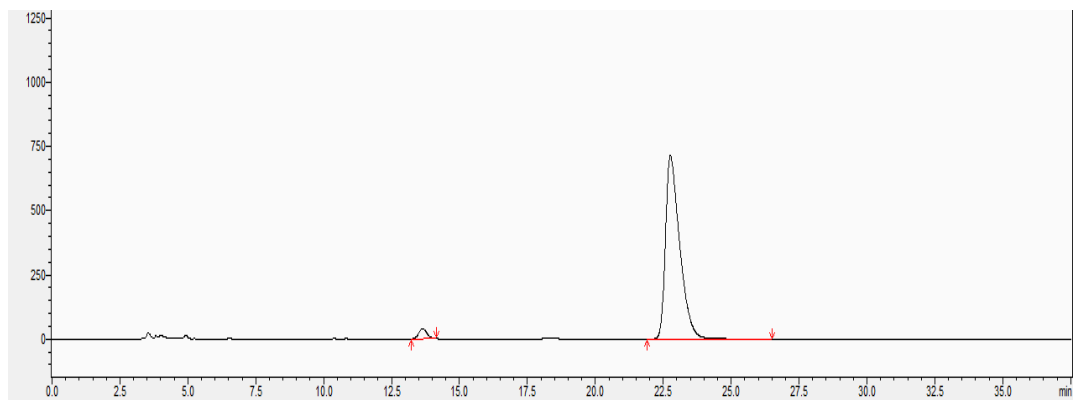
94% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IE column,
n-hexane /*i*-PrOH = 95/5, 1 mL/min, 254 nm.

Racemate:

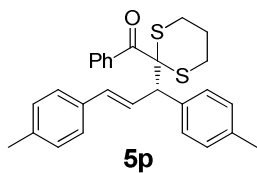


	Retention Time (min)	Area (%)
Peak 1	13.380	50.511
Peak 2	22.085	49.489

Chiral:

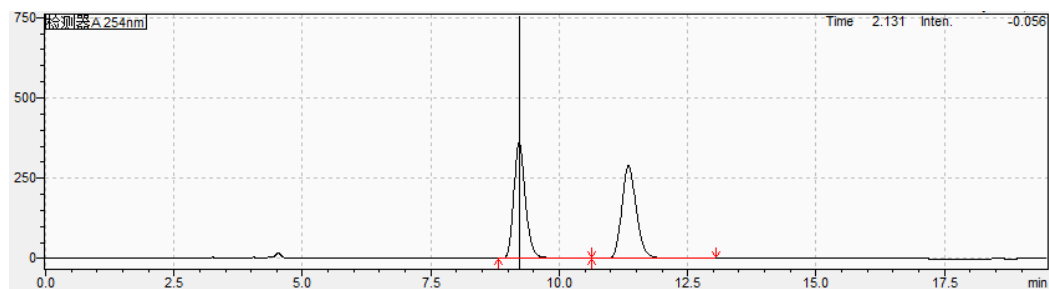


	Retention Time (min)	Area (%)
Peak 1	13.640	3.128
Peak 2	22.767	96.872



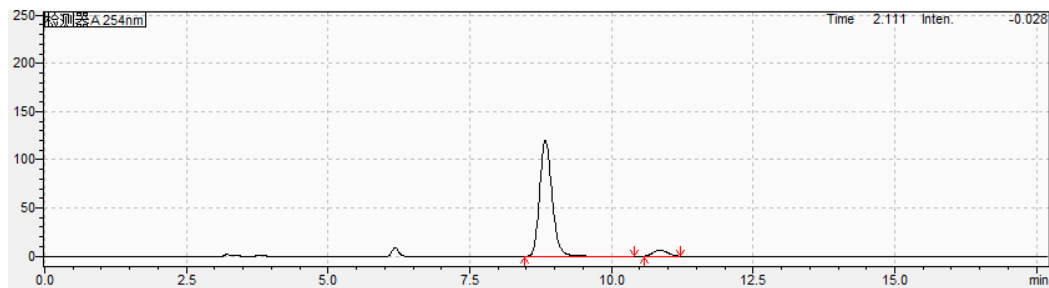
89% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column, *n*-hexane /*i*-PrOH = 98/2, 1 mL/min, 254 nm.

Racemate:

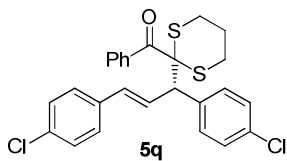


	Retention Time (min)	Area (%)
Peak 1	9.218	50.022
Peak 2	11.348	49.978

Chiral:

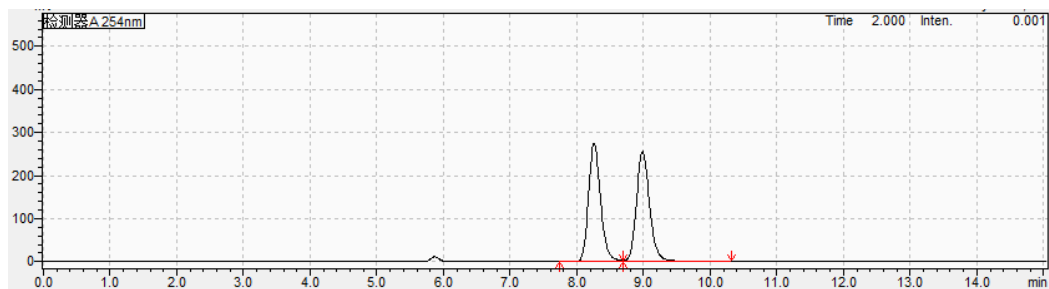


	Retention Time (min)	Area (%)
Peak 1	8.835	94.453
Peak 2	10.853	5.547



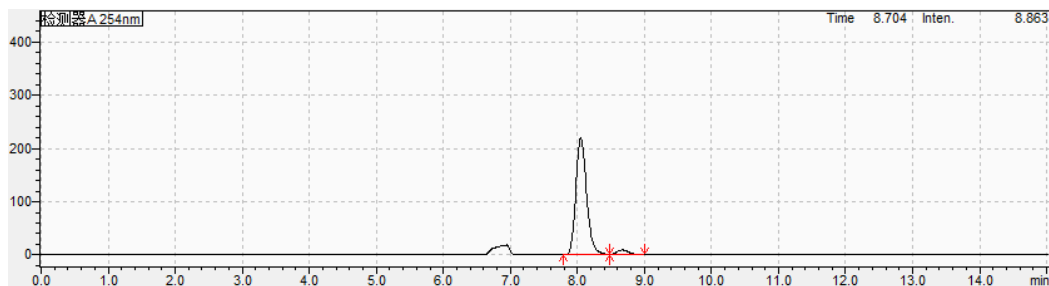
92% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column, *n*-hexane /*i*-PrOH = 98/2, 1 mL/min, 254 nm.

Racemate:

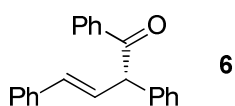


	Retention Time (min)	Area (%)
Peak 1	8.257	49.612
Peak 2	8.985	50.388

Chiral:

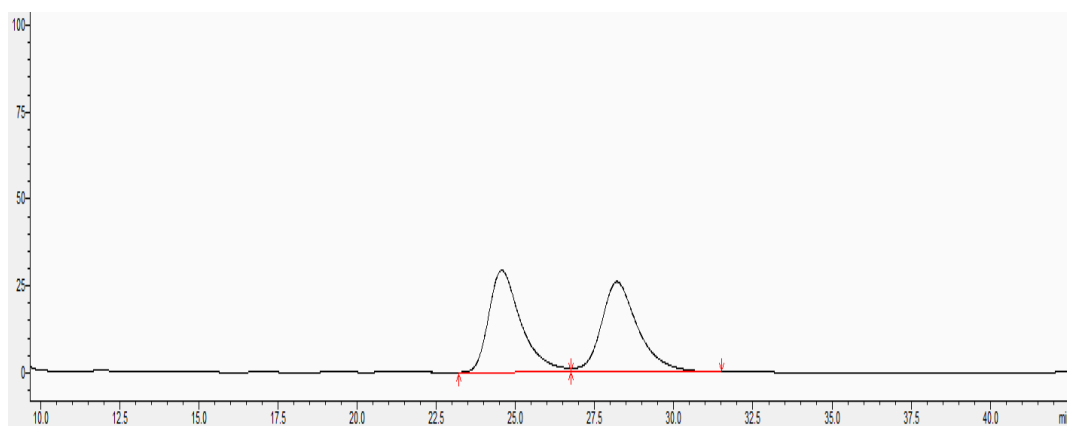


	Retention Time (min)	Area (%)
Peak 1	8.050	96.185
Peak 2	8.673	3.815



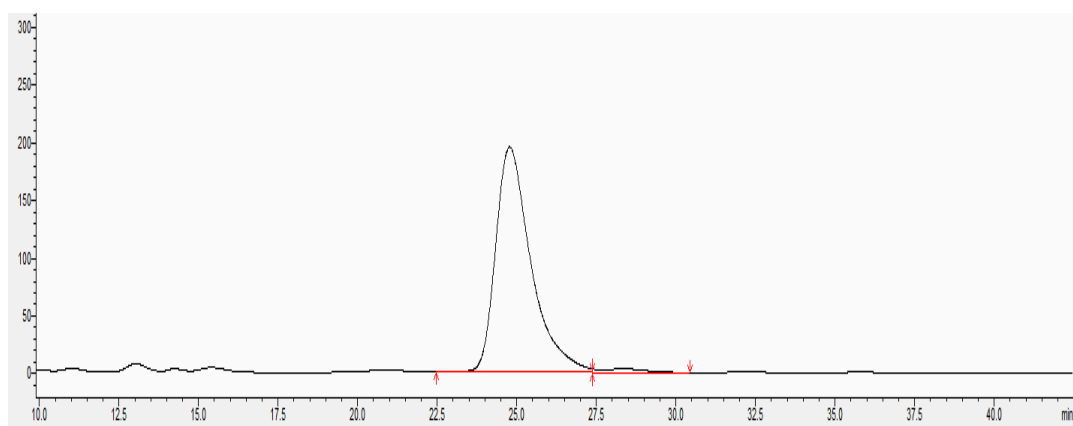
96% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak AD-H column, *n*-hexane /*i*-PrOH = 98/2, 1 mL/min, 254 nm.

Racemate:

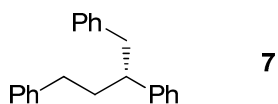


	Retention Time (min)	Area (%)
Peak 1	24.564	49.886
Peak 2	28.205	50.114

Chiral:

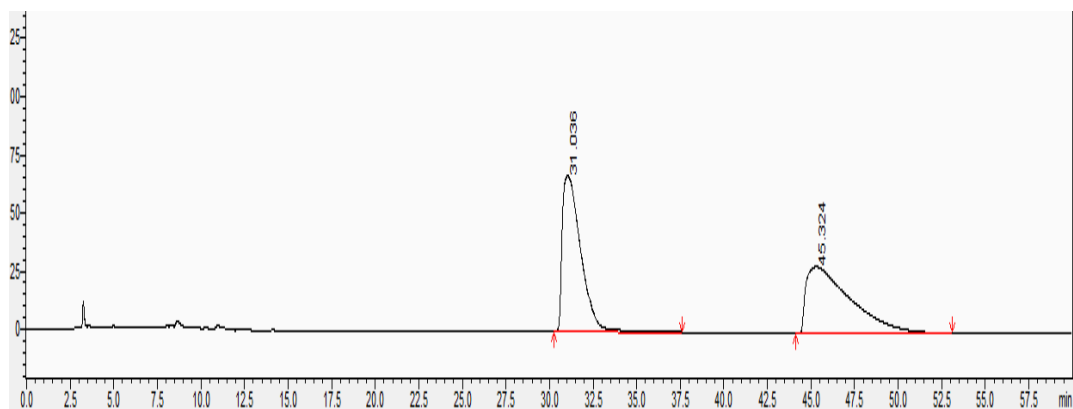


	Retention Time (min)	Area (%)
Peak 1	24.777	97.954
Peak 2	28.359	2.046



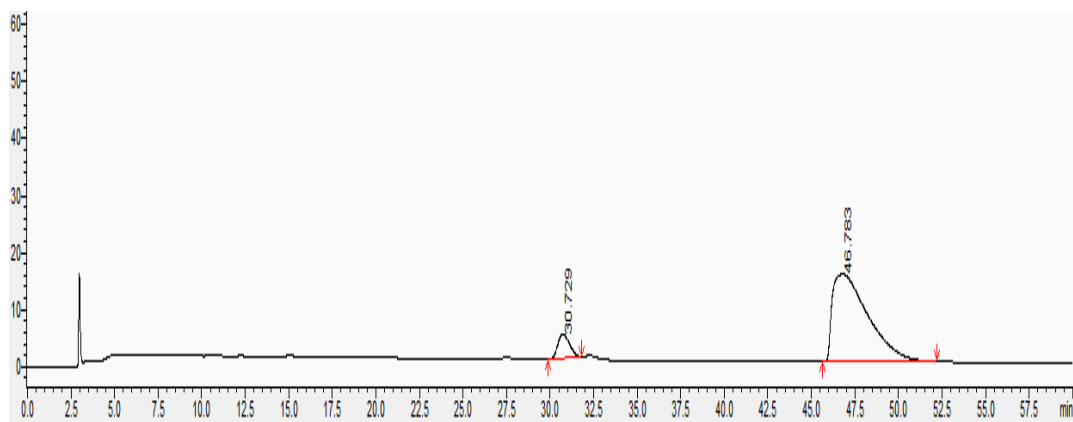
83% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak OD-H column, *n*-hexane /*i*-PrOH = 100/0, 1 mL/min, 220 nm.

Racemate:

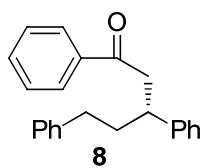


	Retention Time (min)	Area (%)
Peak 1	31.036	49.640
Peak 2	45.324	50.360

Chiral:

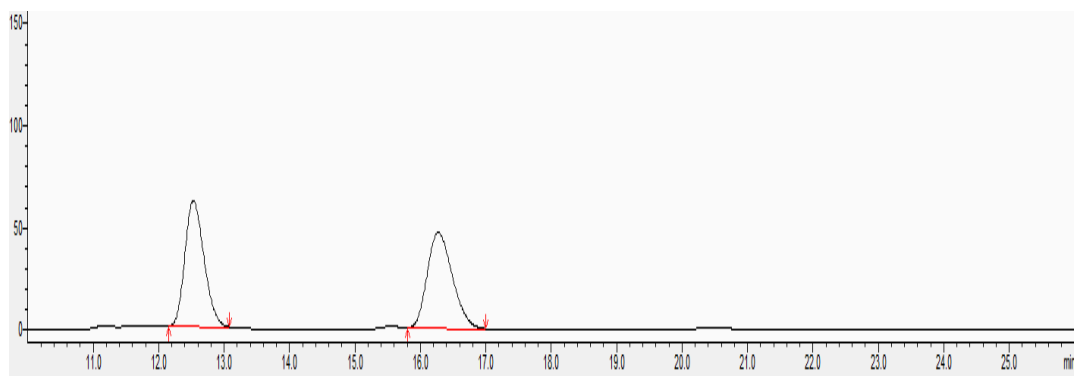


	Retention Time (min)	Area (%)
Peak 1	30.729	8.546
Peak 2	46.783	91.454



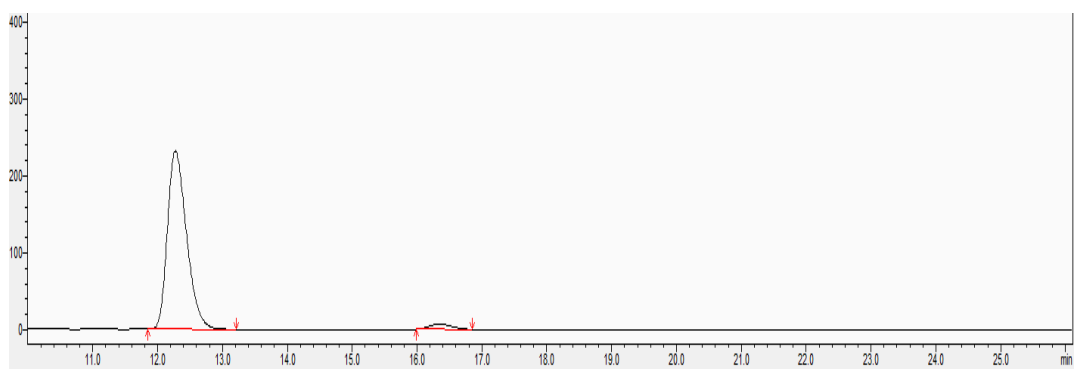
93% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak OD-H column, *n*-hexane /*i*-PrOH = 98/2, 1 mL/min, 254 nm.

Racemate:

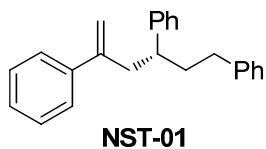


	Retention Time (min)	Area (%)
Peak 1	12.529	50.644
Peak 2	16.273	49.356

Chiral:

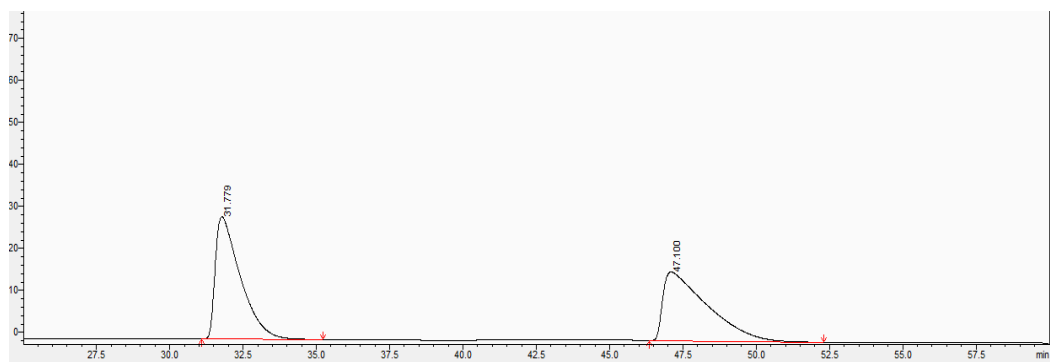


	Retention Time (min)	Area (%)
Peak 1	12.275	96.651
Peak 2	16.345	3.349



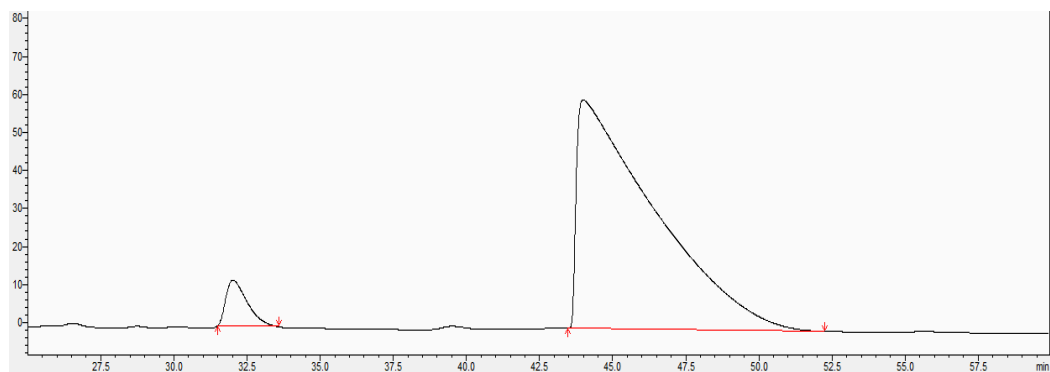
90% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak OD-H column, Hexane/*i*-PrOH = 100/0, 1 mL/min, 254 nm.

Racemate:



	Retention Time (min)	Area (%)
Peak 1	31.779	50.239
Peak 2	47.100	49.761

Chiral:



	Retention Time (min)	Area (%)
Peak 1	32.021	5.138
Peak 2	43.995	94.862