Supporting Information

Silver-Catalyzed C(sp³)–H Chlorination

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1. General method

AgOTf, 1,10-phenanthroline, and 'BuOCl were purchased from Wako Pure Chemical Industries, Ltd., Aldrich, and Tokyo Chemical Industry Co., Ltd. (TCI), respectively, which were used without any purification. NMR spectra were recorded on JEOL ECX-500 (500.16 MHz for 1 H NMR and 125.77 MHz for 13 C NMR), and JEOL ECS-400 (391.78 MHz for 1 H NMR, 98.52 MHz for 13 C NMR) spectrometers. For 1 H and 13 C NMR, chemical shifts were reported in the scale relative to the solvent used as an internal reference (δ = 7.26 and 77.00 ppm (CDCl₃), 2.04 and 29.80 ppm (acetone- d_6), 2.50 and 39.52 ppm (DMSO- d_6); 1 H and 13 C NMR respectively). Infrared (IR) spectra were recorded on a JASCO FT/IR 410 Fourier transform infrared spectrophotometer. ESI-MS spectra were measured on a Waters ZQ4000 spectrometer (for LRMS), and a JEOL JMS-T100LC AccuTOF spectrometer (for HRMS). Optical rotations were measured on a JASCO P-1010 polarimeter. HPLC was conducted by JASCO HPLC systems (pump: PU-2080; detector: UV-2075, measured at 254 nm). Column chromatography was performed with silica gel Merck 60 (230–400 mesh ASTM). NMR yields were calculated by 1 H NMR of crude products using CHCl₂CHCl₂ (δ = 5.9 ppm in CDCl₃) as an internal standard.

2. Preparation of Ag(phen)₂OTf¹

To a stirred mixture of AgOTf (256.9 mg, 1 mmol, 1 eq) in MeOH (5 mL) was added 1,10-phenanthroline anhydrate (360.4 mg, 2 mmol, 2 eq) in MeOH (10 mL). The colorless solution turned into yellow suspension on adding a 1,10-phenanthroline solution. After stirring for three hours the solid was collected on a Kiriyama filter and washed with MeOH, then dried under vacuum to give Ag(phen)₂OTf (542.0 mg, 88%) as yellow solid.

¹H NMR (400 MHz, DMSO- d_6) δ: 9.17 (4H, dd, J = 4.5 Hz, 1.8 Hz), 8.80 (4H, dd, J = 8.1 Hz, 1.8 Hz), 8.24 (4H, s), 8.02 (4H, dd, J = 8.1 Hz, 4.5 Hz); identical to the reported data.

3. Typical C(sp³)–H chlorination procedure

A) 2 mmol scale reaction

To a stirred mixture of substrate 1 (2 mmol, 1 eq) and $Ag(phen)_2OTf$ (2.5 mg, 0.004 mmol, 0.2 mol%) in MeCN (1 mL, 2 M to 1) was added 'BuOCl (678.6 μ L, 6 mmol, 3 eq)*, and then a cap was put on the flask. The color of the reaction mixture turned to brown by adding 'BuOCl. After stirring for 48 hours the reaction mixture was passed through a silica gel pad in a Pasteur pipet (about 4 cm, the eluent was Et_2O or EtOAc) to remove the silver catalyst and the volatiles were removed by rotavap. The residue was purified by silica gel column chromatography to obtain the chlorinated compound 2.

*In chlorination of a benzylic $C(sp^3)$ –H bond, we used 1) 2 eq of 'BuOCl to minimize dichlorination and 2) dry MeCN, and ran the reaction under argon atmosphere to avoid $S_N 2$ substitution of Cl by $H_2 O$; otherwise

¹ C. Santini, C. Pettinari, G. Gioia Lobbia, D. Leonesi, G. Valle, S. Calogero *Polyhedron* **1998**, *17*, 3201–3210.

significant amounts of acetophenones were produced.

B) 5 mmol scale reaction

The workup procedure was different from that of 2 mmol scale reaction. To a stirred reaction mixture cooled by ice bath was added $NaHSO_3$ aq and Et_2O . The organic layer was separated, dried over Na_2SO_4 , filtrated, and evaporated to give a crude mixture. The following procedure was the same as above.

4. Optimization of the reaction conditions

4.1. Chlorination of tert-C(sp3)-H bonds

The chlorination conditions were screened to maximize the yield using 0.2 mmol of 1a. First, we screened catalysts (Table S1). We found that AgOTf with a bidentate ligand catalyzed the chlorination in good yield (entry 5–7). Free Ag salts didn't work well (entry 1–4), and Cu(I) and Fe(II) salts, which are catalysts for Fenton-type reactions, failed efficient chlorination, either (entry 8–11). The ligand itself was unable to be the catalyst (entry 13–15), and almost no reaction proceeded without any catalyst (entry 12). The results suggested that Ag(phen)₂OTf was the best among the catalysts tested.

Table S1 The condition screening — catalysts.

	catalyst (1 mol%) Me tBuOCl (2 eq)	_	Me L C l
BzO 1a	Me dist. MeCN (1 M), Ar, r.t.,	24 h BzOʻ	2a Me
entry	catalyst	NMR yield	NMR rsm
1	AgOTf	12%	87%
2	AgNO ₃	7%	92%
3	AgOAc	8%	91%
4	AgTFA	16%	83%
5	Ag(1,10-phen) ₂ OTf	76 %	23%
6	Ag(2,2'-bpy)OTf	60%	39%
7	Ag(2,2'-bpy) ₂ OTf	69%	30%
8	CuCl (5 mol%)	24%	75%
9	CuCl + 1,10-phen (5+10 mol%)	51%	44%
10	CuCl + 2,2'-bpy (5+10 mol%)	24%	74%
11	Fe(OTf) ₂ (5 mol%)	< 10% (TLC)	_
12	(none)	6%	93%
13	1,10-phen (10 mol%)	5%	94%
14	2,2'-bpy (10 mol%)	13%	86%
15	4,4'-bpy (10 mol%)	1%	99%

Next, we screened the amount of the solvent and the catalyst (Table S2). The yield did not change a lot as the concentration was changed (entry 1–4), and 2 M should be the most proper concentration. The amount of the catalyst was able to decrease as low as 0.2 mol% (entry 6). Even though higher NMR yield was obtained in entry 7, the difference in the yields is not large considering 2.5 times as much as the catalyst was used.

Table S2 The condition screening — the amount of MeCN and Ag(phen)₂OTf.

		M - CN	NIME CALL	NMD
entry	Ag cat	MeCN	NMR yield	NMR rsm
1	1 mol%	0.5 M	66%	33%
2	1 mol%	1 M	76%	23%
3	1 mol%	2 M	77 %	22%
4	1 mol%	4 M	71%	28%
5	0.1 mol%	2 M	56%	43%
6	0.2 mol%	2 M	71%	28%
7	0.5 mol%*	2 M	74%	25%
8	1 mol%*	2 M	58%	41%
9	2 mol%*	2 M	59%	40%

^{*} Ag(phen)₂OTf did not dissolve completely in MeCN.

Then we screened solvents and oxidants (Table S3). Ag(phen) $_2$ OTf did not dissolve well in the solvents except for MeCN and DMF, which may be one reason for the yield to be decreased by replacing MeCN with other solvents (entry 1–8). Water contamination did not affect the yield (entry 1 vs 9), so MeCN without any purification (as purchased) was chosen as the best solvent. DMSO furiously reacted with t BuOCl so that no chlorination occurred on **1a**. Other oxidants like *N*-chlorosuccinimide or aqueous NaOCl didn't promote efficient chlorination.

Table S3 The condition screening — solvents and oxidants.

Me	Ag(phen) ₂ OTf (0.5 mol%) ^t BuOCl (x eq)	Me
BzO Me	solvent (2 M), Ar, r.t., 24 h	BzO Me

entry	^t BuOCl	solvent	NMR yield	NMR rsm
1	2 eq	dist. MeCN	78 %	21%
2	2 eq	dry EtOAc	50%	44%
3	2 eq	dry CH ₂ Cl ₂	37%	43%
4	2 eq	dry CHCl ₃	27%	58%
5	2 eq	dry ^t BuOH	49%	49%
6	2 eq	dry DMF	0% (TLC)	_
7	2 eq	dry DMSO	N.R. (TLC)	_
8	2 eq	PhCl	65%	25%
9	2 eq	MeCN/water (1/1)	79 %	12%
10	2 eq	MeCN ^a	76%	23%
11	NCS 2 eq	MeCN ^a	N.R. (TLC)	-
12	NaOCI 2 eq	MeCN ^a	3%	96%
13	3 eq	MeCN ^a	84%	11%
14	4 eq	MeCN ^a	87%	11%
15^{b}	2 eq	MeCN ^a	80%	19%
16 ^b	3 eq	MeCN ^a	92%	7 %

^a neither distilled nor anhydrous ^b 48 h

Finally, we tried the chlorination under air or O₂ atmosphere (Table S4). Oxygen had

adverse effects on the chlorination process, but aerobic conditions were acceptable if a small reaction vessel was used.

Table S4 Reaction atmosphere: Ar, O₂ or air.

entry	atmosphere	NMR yield ^a	NMR rsm ^a	NMR yield ^b	NMR rsm ^b
1	Ar	79%	18%	79%	20%
2	02	64%	35%	53%	46%
3	air	76%	23%	59%	40%

^a capped tube, ϕ 16 mm × 100 mm (12.5 mL) ^b test tube with balloon

4.2. Chlorination of benzylic C(sp³)–H bonds

When the optimized conditions above were applied to benzylic C(sp³)–H chlorination, two problems arose: ketone formation and dichlorination (Scheme S1). To improve the situation, we further optimized the reaction conditions using **1h**.

Scheme S1 Undesirable side reactions.

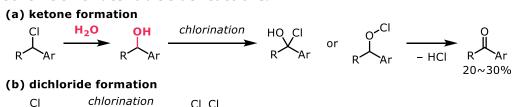
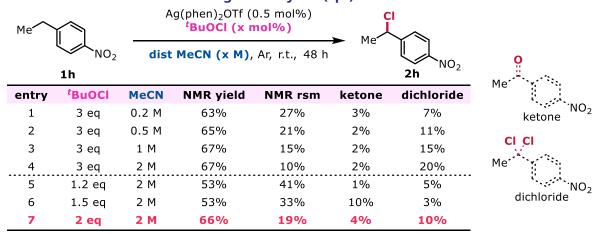


Table S5 The condition screening — benzylic C(sp³)-H chloriation.



After all we determined the optimal reaction conditions as below (Scheme S2). As there

was no problem for secondary C(sp³)–H chlorination under *tert*-C(sp³)–H chlorination conditions, no optimization was conducted further.

Scheme S2 The optimal reaction conditions.

Me
$$\frac{\text{Ag(phen)}_2\text{OTf (0.2 mol\%)}}{\text{dist. MeCN (2 M), Ar, r.t., 48 h}}$$
 Me $\frac{\text{Cl}}{\text{Me}}$ NO₂

5. Limitation in the substrate scope

Listed below are compounds that didn't give satisfactory yields in the C(sp³)–H chlorination.

1. A lot of C(sp³)-H bonds

The optimized chlorination conditions were so reactive that regionelective chlorination of $C(sp^3)$ –H bonds was not successful.

2. Nucleophilic attack to 'BuOCl

Anisole or naphthalene derivatives reacted directly with 'BuOCl irrespective of the Ag catalyst.

undesired aryl chlorides:

3. Direct oxidation of amide nitrogen atom by high-valent Ag

Amide nitrogen atoms seemed to be oxidized by a high-valent Ag that had formed from Ag catalyst and 'BuOCl. The resulting iminium cations should be attacked by water, giving C–N-bond-cleaved products.

4. Labile functional groups

Compounds listed below gave rather complicated reaction mixtures under the optimized chlorination conditions.

Me
$$AcO$$
 $(R = I, Bpin)$
 CO_2Me
 Me
 Me

6. Chlorination of a chiral compound

6.1. Synthesis of 1w from (-)- β -citronellol, and conversion of 2w to 2x

$$\begin{array}{c} \text{Me} & \text{Me} & \text{Me} \\ \text{HO} & \text{O}_2\text{N} & \text{Me} \\ \text{(-)-}\beta\text{-citronellol} & \text{O}_2\text{N} & \text{S1w} \\ \end{array}$$

(-)-β-citronellol to **S1w**

To a stirred mixture of (–)- β -citronellol (781 mg, 5 mmol, 1 eq) in pyridine (5 mL) was added 4-nitrobenzoyl chloride (1.39 g. 1.5 eq) at room temperature. HCl in water was added after 8 h, and the mixture was extracted with Et₂O. The combined organic layer was washed with HCl aq, dried over Na₂SO₄, and filtrated, and volatiles were removed by rotavap. The residue was purified by silica gel column chromatography (hexane/EtOAc = 20/1) so that **S1w** was obtained as yellow liquid (1.37 g, 90%).

S1w to S2w

A mixture of **S1w** (663 mg, 2.17 mmol, 1 eq) in CH_2Cl_2 (5 mL, 0.4 M) cooled by acetone/dry ice bath was treated with a stream of O_3/O_2 . After 5 min, the solution color turned blue, and then Ar was flushed through the mixture until the color disappeared. Me₂S (635 μ L, 8.68 mmol, 4 eq) was added, and the cold bath was removed. After 15 h, the reaction mixture was washed with HCl aq, and the organic layer was dried over Na₂SO₄. After filtration, volatiles were removed by rotavap. The residue was purified by silica gel column chromatography (hexane/EtOAc = 4/1 to 2/1 to 1/2) so that **S2w** was obtained as yellow liquid (561 mg, 92%), along with **1w** (27.7 mg).

S2w to 1w

To a stirred mixture of **S2w** (539 mg, 1.93 mmol, 1 eq), 2-methyl-2-butene (2.05 mL, 19.3 mmol, 10 eq), and Na₂HPO₄ (1.37 g, 9.65 mmol, 5 eq) in ^tBuOH/water (4 mL/4 mL, 0.25 M) was added NaClO₂ (655 mg, 5.79 mmol, 3 eq) at room temperature. After 1h, NaCl aq was added and the mixture was extracted with EtOAc. The organic layer was washed with HCl aq and dried over Na₂SO₄. After filtration, the volatiles were removed by rotavap. The

residue was purified by silica gel column chromatography (hexane/EtOAc = 3/1 to 1/2) so that **1w** was obtained as yellow liquid, which gradually solidified into yellow solid (495 mg, 87%).

[note] Minerals on silica gel sometimes eluted with the carboxylic acid 1w. An Et_2O solution of 1w was washed with HCl aq to remove the minerals in such a case.

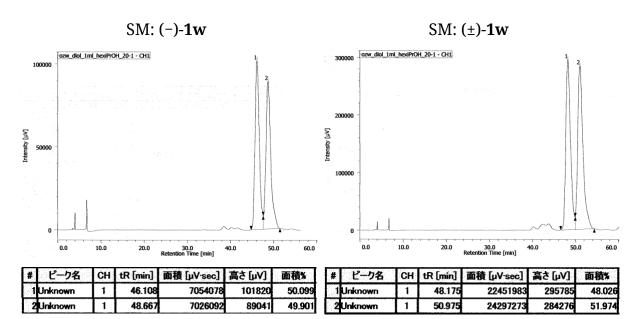
2w to 2x

To a solution of 2w (38.4 mg, 0.11 mmol, 1 eq) in toluene/MeOH (330 μ L/110 μ L, 0.25 M) was added 0.6 M hexane solution of trimethylsilyldiazomethane (200 μ L, 1.05 eq) at room temperature under air. After 10 min, AcOH (a few drops) was added to quench trimethylsilyldiazomethane. The volatiles were removed by rotavap, and the residue was purified by PTLC (hexane/EtOAc = 4/1) so that 2x was obtained as light yellow liquid (28.1 mg, 74%).

6.2. Chiral HPLC analysis of 2x

We analyzed **2x** using a chiral HPLC and the results are shown below. The charts indicated that the chirality disappeared during the chlorination.

HPLC conditions	
column:	CHIRALPAK® IC ϕ 4.6 mm × 250 mm
eluent:	hexane/2-propanol = 20/1
flow rate:	1.0 mL/min



7. Procedure for free tert-BuO radical reactions

A) N-hydroxyphthalimide (NHPI) conditions

To a stirred mixture of the substrate 1 (0.1 mmol, 1 eq) and NHPI (4.1 mg, 0.025 mmol, 25 mol%) in distilled MeCN (200 μ L, 0.5 M to 1) was added 'BuOCl (22.6 μ L, 0.2 mmol, 2 eq) under Ar atmosphere. The color of the reaction mixture turned into yellow first, and into dark yellow on adding 'BuOCl. After stirring for 24 h, Na₂SO₃ and water were added to the reaction mixture. The mixture was extracted with Et₂O and the volatiles were removed by rotavap. The NMR yields were calculated using 1,1,2,2-tetrachloroethane as an internal standard.

B) photoirradiation conditions

To a stirred solution of the substrate 1 (0.2 mmol, 1 eq) in distilled MeCN (400 μ L, 0.5 M to 1) was added 'BuOCl (45.2 μ L, 0.4 mmol, 2 eq) under Ar atmosphere. The color of the reaction mixture didn't change on adding 'BuOCl. Then the reaction was irrradiated with a white LED lump*. After stirring for 24 h, Na₂SO₃ and water were added to the reaction mixture. The mixture was extracted with Et₂O and the volatiles were removed by rotavap. The NMR yields were calculated using 1,1,2,2-tetrachloroethane as an internal standard.

* Irradiations were performed with VBL-SL150-WW as the photon source, white LED lamp for plant science research purchased from Valore Ltd., Kyoto, Japan.

8. Synthesis of authentic samples of 2e and 2j

Authentic samples for **2e** and **2j** were synthesized to identify the reaction products of **1e** and **1j**, because **2e** and **2j** were not detected in HRMS (ESI, DART, nor EI).

8.1. Synthesis of authentic 2e

A solution of ethyl benzoylacetate (192.2 mg, 1 mmol, 1 eq) in MeOH (2 mL, 0.5 M) was treated with NaBH₄ (113.5 mg, 3 mmol, 3 eq) at room temperature. After 1 h, water was added to the reaction mixture. The mixture was extracted with EtOAc, and the volatiles were removed by rotavap. The residue was purified by PTLC (hexane/EtOAc = 1/2) to give **S1e** (79.9 mg, 53%).

S1e (188.2 mg, 1.2 mmol, 1 eq) in toluene (2.4 mL, 0.5 M) was treated with HCl aq (12 N, 250 μ L, 3 mmol, 2.5 eq) at room temperature. After 15 h, water was added, the reaction mixture was extracted with Et₂O, and the volatiles were removed by rotavap. The residue was purified by silica gel column chromatography (hexane/EtOAc = 3/1) so that **S2e** was obtained as light dark orange liquid (132.3 mg, 64%).

A flask was charged with S2e (132.3 mg, 0.77 mmol, 1 eq) and dry CH_2Cl_2 (1.6 mL, 0.5 M)

under argon atmosphere. PBr₃ (36.7 μ L, 0.39 mmol, 0.5 eq) in dry CH₂Cl₂ (400 μ L) was added to the flask at room temperature. After 3 h, water and Et₂O were added. The organic layer was washed with water and NaCl aq, dried over Na₂SO₄, filtrated, and the volatiles were removed by rotavap. The residue was purified by silica gel column chromatography (hexane only) so that **2e** was obtained as light yellow liquid (58.4 mg, 32%).

8.2. Synthesis of authentic 2j

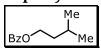
A solution of 4'-bromoacetophenone (99.5 mg, 0.5 mmol, 1 eq) in MeOH (500 μ L, 1 M) was treated with NaBH₄ (28.4 mg, 0.75 mmol, 1.5 eq) at room temperature. After 4 h, water was added to the reaction mixture. The mixture was extracted with EtOAc, and the volatiles were removed by rotavap to give **S1j** (108 mg, containing EtOAc). **S1j** was used in the next step without further purification.

S1j was treated with HCl aq (6 N, 1 mL, around 0.5 M to **S1j**) at room temperature. After 40 h, the reaction mixture was extracted with Et_2O , and the volatiles were removed by rotavap. The residue was purified by PTLC (hexane only) so that **2j** was obtained as light yellow liquid (64.5 mg, 59%).

9. Analytical data

¹H and ¹³C NMR data are described for **1**, which are known materials except for **1w**; full spectroscopic data for **S1w**, **S2w**, **1w** and **2** no matter whether they are new or known. HRMS for **2e** and **2j** are not described because no appropriate signals were detected. The amount of the chlorinated products is also described.

isopentyl benzoate (1a)

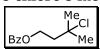


colorless liquid, $R_f = 0.50$ (hexane/EtOAc = 10/1)

¹H NMR (500 MHz, CDCl₃) δ: 8.04 (2H, d, J = 7.6 Hz), 7.55 (1H, t, J = 7.6 Hz), 7.43 (2H, t, J = 7.6 Hz), 4.36 (2H, t, J = 6.9 Hz), 1.75–1.85 (1H, m), 1.64–1.71 (2H, m), 0.98 (6H, d, J = 6.9 Hz)

¹³C NMR (126 MHz, CDCl₃) δ: 166.6, 132.8, 130.5, 129.5, 128.3, 63.6, 37.4, 25.2, 22.5

3-chloro-3-methylbutyl benzoate (2a)



pale yellow liquid, $R_f = 0.40$ (hexane/EtOAc = 10/1)

Yield: 370 mg, 82%; 791 mg (1 g scale), 70%; 853 mg (1 g scale, AgOTf and 1,10-phen as the catalyst), 75%

¹H NMR (500 MHz, CDCl₃) δ: 8.02–8.05 (2H, m), 7.53–7.58 (1H, m), 7.41–7.46 (2H, m), 4.56 (2H, t, J = 6.8 Hz), 2.25 (2H, t, J = 6.8 Hz), 1.68 (6H, s)

¹³C NMR (126 MHz, CDCl₃) δ: 166.4, 133.0, 130.1, 129.5, 128.3, 68.4, 62.0, 44.0, 32.9

IR (neat, cm⁻¹) ν: 2923, 1720, 1452, 1275, 1114, 712

HRMS (ESI): *m/z* calcd for C₁₂H₁₅ClO₂Na [M+Na]⁺ 249.0653, Found 249.0644

butyl benzoate (1b)

colorless liquid, $R_f = 0.46$ (hexane/EtOAc = 10/1)

¹H NMR (500 MHz, CDCl₃) δ: 8.05 (2H, dd, J = 8.0 H, 1.1 Hz), 7.52–7.57 (1H, m), 7.41–7.46 (2H, m), 4.33 (2H, t, J = 6.6 Hz), 1.70–1.79 (2H, m), 1.43–1.53 (2H, m), 0.98 (3H, t, J = 7.4 Hz)

¹³C NMR (126 MHz, CDCl₃) δ: 166.6, 132.7, 130.5, 129.5, 128.2, 64.8, 30.7, 19.2, 13.7

3-chlorobutyl benzoate (2b)



pale yellow liquid, $R_f = 0.34$ (hexane/EtOAc = 10/1)

Yield: 152 mg, 35%

¹H NMR (500 MHz, CDCl₃) δ: 8.04 (2H, dd, J = 8.0 Hz, 1.1 Hz), 7.54–7.58 (1H, m), 7.44 (2H, t, J = 7.7 Hz), 4.43–4.55 (2H, m), 4.20–4.28 (1H, m), 2.18–2.26 (1H, m), 2.07–2.15 (1H, m), 1.60 (3H, d, J = 6.9 Hz)

¹³C NMR (126 MHz, CDCl₃) δ: 166.3, 133.0, 130.0, 129.5, 128.3, 62.0, 54.8, 39.1, 25.4

IR (neat, cm⁻¹) v: 2974, 1720, 1452, 1274, 1114, 712

HRMS (ESI): *m*/z calcd for C₁₁H₁₃ClO₂Na [M+Na]⁺ 235.0496, Found 235.0491

pentyl benzoate (1c)

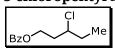


colorless liquid, $R_f = 0.58$ (hexane/EtOAc = 10/1)

¹H NMR (400 MHz, CDCl₃) δ: 8.05 (2H, d, J = 7.4 Hz), 7.55 (1H, t, J = 7.4 Hz), 7.44 (2H, t, J = 7.4 Hz), 4.32 (2H, t, J = 6.7 Hz), 1.71–1.84 (2H, m), 1.32–1.49 (4H, m), 0.93 (3H, t, J = 7.0 Hz)

¹³C NMR (126 MHz, CDCl₃) δ: 166.7, 132.8, 130.5, 129.5, 128.3, 65.1, 28.4, 28.1, 22.4, 14.0

3-chloropentyl benzoate (2c)



pale yellow liquid, $R_f = 0.50$ (hexane/EtOAc = 10/1)

Yield: 288 mg (2c + 2c'), 64%

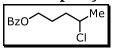
¹H NMR (500 MHz, CDCl₃) δ: 8.01–8.05 (2H, m), 7.54–7.59 (1H, m), 7.42–7.47 (2H, m), 4.44–4.58 (2H, m), 4.01–4.09 (1H, m), 2.20–2.28 (1H, m), 2.05–2.15 (1H, m), 1.73–1.92 (2H, m), 1.07 (3H, t, J = 7.4 Hz)

 13 C NMR (126 MHz, CDCl₃) δ: 166.4, 133.0, 130.1, 129.5, 128.4, 62.1, 61.6, 37.0, 31.6, 10.8

IR (neat, cm⁻¹) ν: 2970, 1720, 1452, 1274, 1114, 712

HRMS (ESI): m/z calcd for $C_{12}H_{15}ClO_2Na$ [M+Na]⁺ 249.0653, Found 249.0661

4-chloropentyl benzoate (2c')



pale yellow liquid, $R_f = 0.46$ (hexane/EtOAc = 10/1)

¹H NMR (500 MHz, CDCl₃) δ: 8.02–8.06 (2H, m), 7.54–7.58 (1H, m), 7.43–7.48 (2H, m), 4.32–4.37 (m, 2H), 4.06–4.13 (1H, m), 1.99–2.05 (1H, m), 1.82–1.94 (3H, m), 1.55 (3H, d, *I* = 6.3 Hz)

¹³C NMR (126 MHz, CDCl₃) δ: 166.5, 132.9, 130.2, 129.5, 128.3, 64.3, 58.1, 36.8, 26.0, 25.4

IR (neat, cm⁻¹) ν: 2964, 1719, 1451, 1274, 1112, 711

HRMS (ESI): m/z calcd for $C_{12}H_{15}ClO_2Na$ [M+Na]⁺ 249.0653, Found 249.0643

3-phenylpropyl acetate (1d)

Ph OAc

pale yellow liquid, $R_f = 0.52$ (hexane/EtOAc = 5/1)

¹H NMR (500 MHz, CDCl₃) δ: 7.27-7.33 (2H, m), 7.17-7.23 (3H, m), 4.10 (2H, t, J = 6.6 Hz), 2.70 (2H, t, J = 7.7 Hz), 2.06 (3H, s), 1.94-2.01 (2H, m)

¹³C NMR (126 MHz, CDCl₃) δ: 171.1, 141.1, 128.4, 128.3, 125.9, 63.8, 32.1, 30.1, 20.9

3-chloro-3-phenylpropyl acetate (2d)



pale yellow liquid, $R_f = 0.46$ (hexane/EtOAc = 5/1)

Yield: 319 mg, 75%

¹H NMR (500 MHz, CDCl₃) δ: 7.29–7.42 (5H, m), 4.98–5.03 (1H, m), 4.21–4.28 (1H, m), 4.12–4.18 (1H, m), 2.31–2.48 (2H, m), 2.05 (3H, s)

¹³C NMR (126 MHz, CDCl₃) δ: 170.8, 140.9, 128.7, 128.5, 126.9, 61.5, 59.8, 38.6, 20.8

IR (neat, cm⁻¹) v: 2961, 1741, 1365, 1237, 1038, 761, 698

HRMS (ESI): m/z calcd for $C_{11}H_{13}ClO_2Na$ [M+Na]⁺ 235.0496, Found 235.0504

(3-bromopropyl)benzene (1e)



colorless liquid, $R_f = 0.54$ (hexane/EtOAc = 1/0)

¹H NMR (500 MHz, CDCl₃) δ: 7.29-7.35 (2H, m), 7.20-7.25 (3H, m), 3.41 (2H, t, J = 6.6 Hz), 2.80 (2H, t, J = 7.4 Hz), 2.15-2.22 (2H, m)

¹³C NMR (126 MHz, CDCl₃) δ: 140.5, 128.52, 128.47, 126.1, 34.1, 33.9, 33.1

(3-bromo-1-chloropropyl)benzene (2e)



light yellow liquid, $R_f = 0.44$ (hexane/EtOAc = 1/0)

Yield: 234 mg, 50%

¹H NMR (500 MHz, CDCl₃) δ: 7.31-7.44 (5H, m), 5.11-5.15 (1H, m), 3.56-3.62 (1H, m), 3.39-3.45 (1H, m), 2.59-2.68 (1H, m), 2.44-2.52 (1H, m)

¹³C NMR (126 MHz, CDCl₃) δ: 140.4, 128.8, 128.6, 127.0, 61.1, 42.3, 30.1

IR (neat, cm⁻¹) ν: 3031, 2967, 1492, 1454, 1258, 757, 697

HRMS (ESI, DART, EI): not detected

1-phenylpropane-1,3-diol (S1e)



light yellow liquid, $R_f = 0.42$ (hexane/EtOAc = 1/2)

¹H NMR (400 MHz, CDCl₃) δ: 7.21–7.36 (5H, m), 4.86 (1H, dd, J = 8.5 Hz, 4.0 Hz), 3.12–4.32 (2H, m), 3.68–3.80 (2H, m), 1.79–1.98 (2H, m)

¹³C NMR (100 MHz, CDCl₃) δ: 144.2, 128.3, 127.3, 125.6, 73.5, 60.7, 40.3

3-chloro-3-phenylpropan-1-ol (S2e)



dark orange liquid, $R_f = 0.77$ (hexane/EtOAc = 1/2)

¹H NMR (400 MHz, CDCl₃) δ: 7.28-7.43 (5H, m), 5.13 (1H, dd, J = 9.4 Hz, 5.4 Hz), 3.82-3.90 (1H, m), 3.68-3.76 (1H, m), 2.20-2.39 (2H, m), 2.14 (1H, bs)

¹³C NMR (100 MHz, CDCl₃) δ : 141.4, 128.7, 128.3, 126.9, 60.3, 59.6, 42.1

(3-bromo-1-chloropropyl)benzene (2e, authentic sample)



light yellow liquid, $R_f = 0.44$ (hexane/EtOAc = 1/0)

¹H NMR (400 MHz, CDCl₃) δ: 7.31-7.44 (5H, m), 5.13 (1H, dd, J = 9.0 Hz, 5.4 Hz), 3.54-3.64 (1H, m), 3.38-3.46 (1H, m), 2.58-2.70 (1H, m), 2.42-2.54 (1H, m)

¹³C NMR (100 MHz, CDCl₃) δ: 140.4, 128.8, 128.6, 127.0, 61.1, 42.3, 30.1

IR (neat, cm⁻¹) ν: 3032, 2968, 1493, 1454, 1258, 758, 697

4-phenylbutyronitrile (1f)



colorless liquid, $R_f = 0.48$ (hexane/EtOAc = 5/1)

¹H NMR (400 MHz, CDCl₃) δ: 7.29–7.36 (2H, m), 7.17–7.28 (3H, m), 2.79 (2H, t, J = 7.4 Hz), 2.32 (2H, t, J = 7.2 Hz), 1.94–2.03 (2H, m)

¹³C NMR (100 MHz, CDCl₃) δ: 139.6, 128.5, 128.3, 126.4, 119.4, 34.2, 26.8, 16.2

4-chloro-4-phenylbutyronitrile (2f)



yellow liquid; , $R_f = 0.40$ (hexane/EtOAc = 5/1)

Yield: 255 mg, 71%

¹H NMR (400 MHz, CDCl₃) δ: 7.32–7.43 (5H, m), 4.96–5.01 (1H, m), 2.28–2.64 (4H, m)

¹³C NMR (100 MHz, CDCl₃) δ: 139.6, 128.9, 126.7, 118.4, 61.1, 35.3, 15.2

IR (neat, cm⁻¹) ν: 2934, 2247, 1492, 1455, 1247, 761

HRMS (ESI): *m/z* calcd for C₁₀H₁₀ClNNa [M+Na]⁺ 202.0394, Found 202.0397

3-phenylpropyl pivalate (1g)

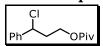


colorless liquid, $R_f = 0.54$ (hexane/EtOAc = 10/1)

¹H NMR (500 MHz, CDCl₃) δ: 7.30 (2H, t, J = 7.4 Hz), 7.20 (3H, t, J = 8.3 Hz), 4.09 (2H, t, J = 6.6 Hz), 2.71 (2H, t, J = 7.7 Hz), 1.94–2.01 (2H, m), 1.23 (9H, s)

¹³C NMR (126 MHz, CDCl₃) δ: 178.5, 141.2, 128.39, 128.36, 125.9, 63.5, 38.7, 32.1, 30.3, 27.1

3-chloro-3-phenylpropyl pivalate (2g)



light yellow liquid, $R_f = 0.48$ (hexane/EtOAc = 10/1)

Yield: 406 mg, 80%

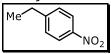
¹H NMR (500 MHz, CDCl₃) δ: 7.32-7.41 (5H, m), 4.99 (1H, dd, J = 8.6 Hz, 5.7 Hz), 4.20-4.26 (1H, m), 4.08-4.14 (1H, m), 2.32-2.48 (2H, m), 1.21 (9H, s)

¹³C NMR (100 MHz, CDCl₃) δ: 178.2, 140.9, 128.8, 128.5, 126.8, 61.4, 59.9, 38.8, 38.7, 27.1

IR (neat, cm⁻¹) v: 2972, 1730, 1479, 1283, 1154, 1035, 759, 698

HRMS (ESI): m/z calcd for $C_{14}H_{19}ClO_2Na$ [M+Na]⁺ 277.0966, Found 277.0956

1-ethyl-4-nitrobenzene (1h)

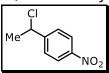


yellow liquid, $R_f = 0.65$ (hexane/EtOAc = 5/1)

¹H NMR (500 MHz, CDCl₃) δ: 8.08–8.13 (2H, m), 7.32 (2H, d, J = 8.6 Hz), 2.74 (2H, q, J = 7.7 Hz), 1.26 (3H, t, J = 7.7 Hz)

¹³C NMR (126 MHz, CDCl₃) δ: 152.0, 146.1, 128.5, 123.5, 28.7, 14.9

1-(1-chloroethyl)-4-nitrobenzene (2h)



yellow liquid, $R_f = 0.52$ (hexane/EtOAc = 5/1)

Yield: 231 mg, 62%

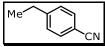
¹H NMR (500 MHz, CDCl₃) δ: 8.21 (2H, d, J = 8.9 Hz), 7.59 (2H, d, J = 8.9 Hz), 5.13 (1H, q, J = 6.9 Hz), 1.86 (3H, d, J = 6.9 Hz);

¹³C NMR (126 MHz, CDCl₃) δ: 149.6, 147.5, 127.5, 123.9, 56.8, 26.3

IR (neat, cm⁻¹) v: 2982, 1606, 1523, 1349, 856, 781, 698

HRMS (ESI): *m*/*z* calcd for C₈H₈ClNO₂Na [M+Na]⁺ 208.0136, Found 208.0146

4-ethylbenzonitrile (1i)

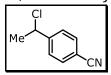


yellow liquid, $R_f = 0.60$ (hexane/EtOAc = 5/1)

¹H NMR (500 MHz, CDCl₃) δ: 7.55 (2H, d, J = 8.3 Hz), 7.28 (2H, d, J = 8.3 Hz), 2.69 (2H, q, J = 7.4 Hz), 1.24 (3H, t, J = 7.4 Hz)

¹³C NMR (126 MHz, CDCl₃) δ: 149.7, 132.0, 128.6, 119.1, 109.3, 28.9, 14.9

4-(1-chloroethyl)benzonitrile (2i)



light yellow liquid, $R_f = 0.50$ (hexane/EtOAc = 5/1)

Yield: 200 mg, 60%

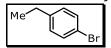
¹H NMR (500 MHz, CDCl₃) δ: 7.64 (2H, d, J = 8.3 Hz), 7.52 (2H, d, J = 8.3 Hz), 5.07 (1H, q, J = 6.9 H), 1.83 (3H, d, J = 6.9 Hz)

¹³C NMR (126 MHz, CDCl₃) δ: 147.7, 132.4, 127.3, 118.4, 111.9, 57.2, 26.2

IR (neat, cm⁻¹) v: 2981, 2360, 2229, 1922, 1608, 1505, 1415, 1232, 1047, 838, 668

HRMS (ESI): *m/z* calcd for C₉H₈ClNNa [M+Na]⁺ 188.0238, Found 188.0230

1-bromo-4-ethylbenzene (1j)



colorless liquid, $R_f = 0.66$ (hexane/EtOAc = 1/0)

¹H NMR (500 MHz, CDCl₃) δ: 7.41 (2H, d, J = 8.0 Hz), 7.08 (2H, d, J = 8.0 Hz), 2.61 (2H, q, J = 7.5 Hz), 1.23 (3H, t, J = 7.5 Hz)

¹³C NMR (126 MHz, CDCl₃) δ: 143.1, 131.3, 129.6, 119.2, 28.3, 15.4

1-bromo-4-(1-chloroethyl)benzene (2j)

light yellow liquid, $R_f = 0.52$ (hexane/EtOAc = 1/0)

Yield: 200 mg, 46%

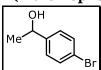
¹H NMR (500 MHz, CDCl₃) δ: 7.47-7.50 (2H, m), 7.29-7.31 (2H, m), 5.04 (1H, q, J = 6.9 Hz), 1.82 (3H, d, J = 6.9 Hz)

¹³C NMR (126 MHz, CDCl₃) δ: 141.8, 131.7, 128.2, 122.1, 57.8, 26.4

IR (neat, cm⁻¹) ν: 2978, 2926, 1592, 1489, 1406, 1226, 1074, 1011, 825, 786, 717

HRMS (ESI, DART, EI): not detected

1-(4-bromophenyl)ethan-1-ol (S1j)

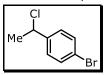


Light yellow liquid, $R_f = 0.42$ (hexane/EtOAc = 3/1)

¹H NMR (500 MHz, CDCl₃) δ: 7.46 (2H, d, J = 7.2 Hz), 7.23 (2H, d, J = 7.2 Hz), 4.84 (1H, q, J = 6.7 Hz), 2.09 (bs, 1H), 1.45 (3H, d, J = 6.7 Hz)

¹³C NMR (126 MHz, CDCl₃) δ: 144.7, 131.5, 127.1, 121.1, 69.7, 25.2

1-bromo-4-(1-chloroethyl)benzene (2j, authentic sample)



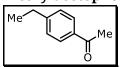
light yellow liquid, $R_f = 0.52$ (hexane/EtOAc = 1/0)

¹H NMR (500 MHz, CDCl₃) δ: 7.49 (2H, d, J = 8.6 Hz), 7.30 (2H, d, J = 8.6 Hz), 5.04 (1H, q, J = 6.9 Hz), 1.83 (3H, d, J = 6.9 Hz)

¹³C NMR (126 MHz, CDCl₃) δ: 141.8, 131.7, 128.2, 122.1, 57.8, 26.4

IR (neat, cm⁻¹) ν: 2976, 2926, 1592, 1489, 1406, 1226, 1074, 1011, 825, 785, 717

4'-ethylacetophenone (1k)

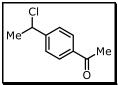


colorless liquid, $R_f = 0.54$ (hexane/EtOAc = 5/1)

¹H NMR (500 MHz, CDCl₃) δ: 7.88 (2H, d, J = 8.3 Hz), 7.27 (2H, d, J = 8.3 Hz), 2.70 (2H, q, J = 7.5 Hz), 2.57 (3H, s), 1.25 (3H, t, J = 7.5 Hz)

¹³C NMR (126 MHz, CDCl₃) δ: 197.8, 150.0, 134.8, 128.5, 128.0, 28.9, 26.5, 15.1

1-(4-(1-chloroethyl)phenyl)ethan-1-one (2k)



light yellow liquid, $R_f = 0.44$ (hexane/EtOAc = 5/1)

Yield: 274 mg, 75%

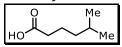
¹H NMR (500 MHz, CDCl₃) δ: 7.94 (2H, d, J = 8.3 Hz), 7.50 (2H, d, J = 8.3 Hz), 5.10 (1H, q, J = 6.9 Hz), 2.59 (3H, s), 1.84 (3H, d, J = 6.9 Hz)

¹³C NMR (126 MHz, CDCl₃) δ : 197.4, 147.7, 136.8, 128.7, 126.7, 57.7, 26.6, 26.3

IR (neat, cm⁻¹) v: 2979, 1685, 1608, 1415, 1359, 1267, 1047, 838, 683

HRMS (ESI): *m/z* calcd for C₁₀H₁₁ClONa [M+Na]⁺ 205.0391, Found 205.0397

5-methylhexanoic acid (11)

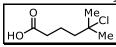


colorless liquid, $R_f = 0.52$ (hexane/EtOAc = 2/1)

¹H NMR (500 MHz, CDCl₃) δ: 10.82 (1H, bs), 2.33 (2H, t, J = 7.4 Hz), 1.60–1.67 (2H, m), 1.51–1.60 (1H, m), 1.18–1.24 (2H, m), 0.88 (6H, d, J = 6.3 Hz)

¹³C NMR (126 MHz, CDCl₃) δ: 180.6, 38.2, 34.3, 27.7, 22.5, 22.4

5-chloro-5-methylhexanoic acid (21)



yellow liquid, $R_f = 0.36$ (hexane/EtOAc = 2/1)

Yield: 281 mg, 85%

¹H NMR (500 MHz, CDCl₃) δ: 9.45 (1H, bs), 2.38 (2H, t, J = 7.2 Hz), 1.73–1.87 (4H, m), 1.56 (6H, s)

¹³C NMR (126 MHz, CDCl₃) δ : 179.5, 70.3, 45.0, 33.8, 32.3, 20.3

IR (neat, cm⁻¹) v: 2972, 2667, 1714, 1281, 1115, 938, 826

HRMS (ESI): m/z calcd for $C_7H_{13}ClO_2Na$ [M+Na]⁺ 187.0496, Found 187.0503

2-butylisoindoline-1,3-dione (1m)

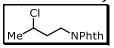


white solid, $R_f = 0.60$ (hexane/EtOAc = 3/1)

¹H NMR (500 MHz, CDCl₃) δ: 7.80 (2H, dd, J = 5.1 Hz, 3.1 Hz), 7.67 (2H, dd, J = 5.1 Hz, 3.1 Hz), 3.65 (2H, t, J = 7.4 Hz), 1.58–1.66 (2H, m), 1.29–1.38 (2H, m), 0.91 (3H, t, J = 7.2 Hz)

¹³C NMR (126 MHz, CDCl₃) δ: 168.3, 133.7, 132.1, 123.0, 37.7, 30.5, 20.0, 13.6

2-(3-chlorobutyl)isoindoline-1,3-dione (2m)



white solid, $R_f = 0.48$ (hexane/EtOAc = 3/1)

Yield: 295 mg, 62%

¹H NMR (500 MHz, CDCl₃) δ: 7.79–7.85 (2H, m), 7.67–7.73 (2H, m), 3.98–4.07 (1H, m), 3.76–3.90 (2H, m), 1.99–2.13 (2H, m), 1.54 (3H, d, J = 6.3 Hz)

¹³C NMR (126 MHz, CDCl₃) δ: 168.2, 133.9, 132.0, 123.2, 55.5, 38.6, 35.6, 25.2

IR (neat, cm⁻¹) v: 2975, 1773, 1712, 1397, 720

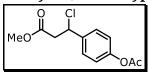
HRMS (ESI): m/z calcd for C₁₂H₁₂ClNO₂Na [M+Na]⁺ 260.0449, Found 260.0445

methyl 3-(4-acetoxyphenyl)propanoate (1n)

colorless liquid, R_f = 0.52 for hexane/EtOAc = 2/1; 0.48 for hexane/EtOAc = 10/1 × 4 times **1H NMR (400 MHz, CDCl₃) δ**: 7.20 (2H, d, J = 8.5 Hz), 6.99 (2H, d, J = 8.5 Hz), 3.66 (3H, s), 2.93 (2H, t, J = 7.8 Hz), 2.62 (2H, t, J = 7.8 Hz), 2.28 (3H, s)

¹³C NMR (100 MHz, CDCl₃) δ: 173.1, 169.6, 149.0, 138.0, 129.2, 121.5, 51.6, 35.5, 30.2, 21.0

methyl 3-(4-acetoxyphenyl)-3-chloropropanoate (2n)



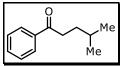
white solid, R_f = 0.52 for hexane/EtOAc = 2/1; 0.40 for hexane/EtOAc = 10/1 × 4 times **Yield:** 30 mg, 58% (0.2 mmol scale)

¹H NMR (400 MHz, CDCl₃) δ: 7.42 (2H, dt, J = 9.2 Hz, 2.5 Hz), 7.08 (2H, dt, J = 9.2 Hz, 2.5 Hz), 5.34 (1H, dd, J = 9.3 Hz, 5.7 Hz), 3.70 (3H, s), 3.16 (1H, dd, J = 16.2 Hz, 9.3 Hz), 3.01 (1H, dd, J = 16.2 Hz, 5.7 Hz), 2.29 (3H, s)

¹³C NMR (100 MHz, CDCl₃) δ: 169.9, 169.2, 150.6, 137.7, 128.1, 121.9, 57.3, 52.1, 44.6, 21.1 IR (neat, cm⁻¹) ν: 2954, 1741, 1508, 1437, 1370, 1201, 1016, 910

HRMS (ESI): m/z calcd for $C_{12}H_{13}ClO_4Na$ [M+Na]⁺ 279.0395, Found 279.0394

4-methyl-1-phenylpentan-1-one (10)²

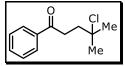


colorless liquid, $R_f = 0.62$ (hexane/EtOAc = 10/1)

¹H NMR (400 MHz, CDCl₃) δ: 7.94–7.98 (2H, m), 7.52–7.57 (1H, m), 7.42–7.48 (2H, m), 2.96 (2H, t, J = 7.4 Hz), 1.59–1.70 (3H, m), 0.95 (6H, d, J = 6.3 Hz)

¹³C NMR (100 MHz, CDCl₃) δ: 200.7, 137.0, 132.8, 128.5, 128.0, 36.6, 33.2, 27.8, 22.4

4-chloro-4-methyl-1-phenylpentan-1-one (20)



white solid, $R_f = 0.44$ (hexane/EtOAc = 10/1)

Yield: 214 mg, 51%

¹H NMR (500 MHz, CDCl₃) δ: 8.00 (2H, d, J = 7.4 Hz), 7.57 (1H, t, J = 7.4 Hz), 7.48 (2H, t, J = 7.4 Hz), 3.24 (2H, t, J = 7.7 Hz), 2.20 (2H, t, J = 7.7 Hz), 1.64 (6H, s)

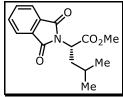
¹³C NMR (126 MHz, CDCl₃) δ: 199.4, 136.8, 133.1, 128.6, 128.1, 70.4, 39.6, 34.7, 32.6

IR (neat, cm⁻¹) v: 2972, 1686, 1448, 1371, 1215, 1110, 742, 689

HRMS (ESI): m/z calcd for $C_{12}H_{15}ClONa$ [M+Na]⁺ 233.0704, Found 233.0706

² Liang, Y.-F.; Zhou, X.-F.; Tang, S.-Y.; Huang, Y.-B.; Feng, Y.-S.; Xu, H.-J. RSC Adv. 2013, 3, 7739–7742.

methyl (S)-2-(1,3-dioxoisoindolin-2-yl)-4-methylpentanoate (1p)

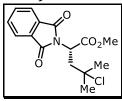


light yellow liquid, $R_f = 0.37$ (hexane/EtOAc = 5/1), $[\alpha]_D^{19} = -27.2$ (c = 1.08, MeOH)

¹H NMR (400 MHz, CDCl₃) δ: 7.85 (2H, dd, J = 5.5 Hz, 3.0 Hz), 7.73 (2H, dd, J = 5.5 Hz, 3.0 Hz), 4.93 (1H, dd, J = 11.4 Hz, 4.3 Hz), 3.71 (3H, s), 2.27–2.37 (1H, m), 1.89–1.99 (1H, m), 1.40–1.53 (1H, m), 0.92 (6H, dd, J = 10.5 Hz, 6.5 Hz)

¹³C NMR (100 MHz, CDCl₃) δ: 170.2, 167.7, 134.1, 131.7, 123.5, 52.7, 50.5, 37.2, 25.0, 23.1, 20.9

methyl (S)-4-chloro-2-(1,3-dioxoisoindolin-2-yl)-4-methylpentanoate (2p)



white solid, $R_f = 0.34$ (hexane/EtOAc = 3/1), $[\alpha]_D^{19} = -24.7$ (c = 0.930, MeOH)

Yield: 585 mg, 94%

¹H NMR (500 MHz, CDCl₃) δ: 7.83-7.90 (2H, m), 7.71-7.76 (2H, m), 5.20 (1H, dd, J = 9.6 Hz, 2.9 Hz), 3.70-3.74 (3H, m), 2.69-2.85 (2H, m), 1.64 (3H, s), 1.55 (3H, s)

¹³C NMR (126 MHz, CDCl₃) δ: 169.6, 167.6, 134.2, 131.6, 123.6, 68.1, 53.1, 49.2, 42.8, 33.2, 31.9 IR (neat, cm⁻¹) ν: 2977, 1746, 1715, 1389, 1271, 1138, 720

HRMS (ESI): m/z calcd for $C_{15}H_{16}ClNO_4Na$ [M+Na]⁺ 332.0660, Found 332.0664

2-methylbenzonitrile (1q)



colorless liquid, $R_f = 0.65$ (hexane/EtOAc = 5/1)

¹H NMR (400 MHz, CDCl₃) δ: 7.58 (1H, dd, J = 7.6 Hz, 0.9 Hz), 7.47 (1H, td, J = 7.7 Hz, 1.2 Hz), 7.31 (1H, dd, J = 7.9 Hz, 0.6 Hz), 7.24–7.28 (1H, m), 2.54 (3H, s)

¹³C NMR (100 MHz, CDCl₃) δ: 141.9, 132.6, 132.4, 130.2, 126.2, 118.1, 112.7, 20.4

2-(chloromethyl)benzonitrile (2q)



light yellow solid, $R_f = 0.50$ (hexane/EtOAc = 5/1)

Yield: 128 mg, 42%

¹H NMR (500 MHz, CDCl₃) δ: 7.67 (1H, d, J = 7.6 Hz), 7.56–7.65 (2H, m), 7.44 (1H, td, J = 7.4 Hz, 1.6 Hz), 4.75 (2H, s)

¹³C NMR (126 MHz, CDCl₃) δ: 140.6, 133.2, 133.0, 130.0, 128.9, 116.7, 112.2, 43.1

IR (neat, cm⁻¹) ν: 3021, 2225, 1597, 1489, 1452, 1271, 1217, 832, 754, 673

HRMS (ESI): *m/z* calcd for C₈H₆ClNNa [M+Na]⁺ 174.0081, Found 174.0074

o-tolyl methanesulfonate (1r)



colorless liquid, $R_f = 0.50$ (hexane/EtOAc = 3/1)

¹H NMR (500 MHz, CDCl₃) δ: 7.19–7.30 (4H, m), 3.18 (3H, s), 2.37 (3H, s)

¹³C NMR (126 MHz, CDCl₃) δ: 147.8, 131.8, 131.2, 127.2, 122.0, 38.0, 16.5

2-(chloromethyl)phenyl methanesulfonate (2r)



colorless liquid, $R_f = 0.48$ (hexane/EtOAc = 3/1)

Yield: 14 mg, 32% (0.2 mmol scale)

¹H NMR (500 MHz, CDCl₃) δ: 7.52 (1H, dd, J = 7.6 Hz, 1.7 Hz), 7.37–7.44 (2H, m), 7.33 (1H, td, J = 7.3 Hz, 16 Hz), 4.69 (2H, s), 3.27 (3H, s)

¹³C NMR (126 MHz, CDCl₃) δ: 146.9, 131.5, 130.4, 130.2, 127.6, 122.3, 40.5, 38.3 IR (neat, cm⁻¹) ν: 3030, 2939, 1585, 1490, 1363, 1153, 1090, 970, 876, 805, 670 HRMS (ESI): m/z calcd for C₈H₉ClO₃SNa [M+Na]⁺ 242.9853, Found 242.9861

ethyl 2-methylbenzoate (1s)



colorless liquid, $R_f = 0.56$ (hexane/EtOAc = 10/1)

¹H NMR (500 MHz, CDCl₃) δ: 7.91 (1H, dd, J = 8.0 Hz, 1.3 Hz), 7.39 (1H, td, J = 7.5 Hz, 1.3 Hz), 7.22–7.27 (2H, m), 4.36 (2H, q, J = 7.2 Hz), 2.60 (3H, s), 1.39 (3H, t, J = 7.2 Hz)

¹³C NMR (126 MHz, CDCl₃) δ : 167.7, 139.9, 131.8, 131.6, 130.4, 129.9, 125.6, 60.6, 21.7, 14.3

ethyl 2-(chloromethyl)benzoate (2s)



light yellow liquid, $R_f = 0.42$ (hexane/EtOAc = 10/1)

Yield: 177 mg, 45%

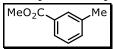
¹H NMR (500 MHz, CDCl₃) δ: 7.98 (1H, d, J = 7.4 Hz), 7.49–7.55 (2H, m), 7.37–7.41 (1H, m), 5.04 (2H, s), 4.40 (2H, q, J = 7.2 Hz), 1.41 (3H, t, J = 7.2 Hz)

¹³C NMR (126 MHz, CDCl₃) δ: 166.7, 138.6, 132.3, 131.0, 130.8, 129.4, 128.3, 61.2, 44.4, 14.2

IR (neat, cm⁻¹) ν: 2981, 1716, 1602, 1448, 1366, 1269, 1131, 1080, 714, 679

HRMS (ESI): *m*/*z* calcd for C₁₀H₁₁ClO₂Na [M+Na]⁺ 221.0340, Found 221.0331

methyl 3-methylbenzoate (1t)



colorless liquid, $R_f = 0.52$ (hexane/EtOAc = 10/1)

¹H NMR (500 MHz, CDCl₃) δ: 7.82–7.87 (2H, m), 7.30–7.38 (2H, m), 3.91 (3H, s), 2.40 (3H, s)

¹³C NMR (126 MHz, CDCl₃) δ: 167.3, 138.1, 133.6, 130.1, 130.0, 128.2, 126.7, 52.0, 21.2

methyl 3-(chloromethyl)benzoate (2t)

light yellow liquid, $R_f = 0.40$ (hexane/EtOAc = 10/1)

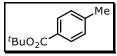
Yield: 14 mg, 38% (0.2 mmol scale)

¹H NMR (500 MHz, CDCl₃) δ: 8.07 (1H, bs), 8.00 (1H, d, J = 7.4 Hz), 7.59 (1H, d, J = 7.4 Hz), 7.45 (1H, t, J = 7.7 Hz), 4.62 (2H, s), 3.93 (3H, s)

¹³C NMR (126 MHz, CDCl₃) δ: 166.6, 137.8, 133.0, 130.7, 129.6, 129.5, 128.9, 52.2, 45.5

IR (neat, cm⁻¹) v: 2952, 1724, 1591, 1434, 1291, 1205, 1108, 990, 759, 708 HRMS (ESI): m/z calcd for $C_9H_9ClO_2Na$ [M+Na]⁺ 207.0183, Found 207.0174

tert-butyl 4-methylbenzoate (1u)

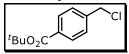


colorless liquid, $R_f = 0.48$ (hexane/EtOAc = 10/1)

¹H NMR (400 MHz, CDCl₃) δ: 7.88 (2H, d, J = 8.3 Hz), 7.21 (2H, d, J = 8.3 Hz), 2.39 (3H, s), 1.59 (9H, s)

¹³C NMR (100 MHz, CDCl₃) δ: 165.9, 142.9, 129.4, 129.2, 128.8, 80.6, 28.2, 21.5

tert-butyl 4-(chloromethyl)benzoate (2u)



white solid, $R_f = 0.42$ (hexane/EtOAc = 10/1)

Yield: 17 mg, 36% (0.2 mmol scale)

¹H NMR (500 MHz, CDCl₃) δ: 7.98 (2H, d, J = 8.0 Hz), 7.43 (2H, d, J = 8.0 Hz), 4.60 (2H, s), 1.59 (9H, s)

¹³C NMR (126 MHz, CDCl₃) δ: 165.2, 141.7, 132.0, 129.8, 128.3, 81.2, 45.4, 28.1

IR (neat, cm⁻¹) ν: 2979, 1714, 1612, 1414, 1368, 1294, 1167, 1119, 1019, 849, 713

HRMS (ESI): *m/z* calcd for C₁₂H₁₅ClO₂Na [M+Na]⁺ 249.0653, Found 249.0652

1-tetralone (1v)



yellow liquid, $R_f = 0.38$ (hexane/EtOAc = 5/1)

¹H NMR (500 MHz, CDCl₃) δ: 8.02 (1H, d, J = 8.0 Hz), 7.45 (1H, td, J = 7.4 Hz, 1.1 Hz), 7.29 (1H, t, J = 7.4 Hz), 7.24 (1H, d, J = 8.0 Hz), 2.95 (2H, t, J = 6.3 Hz), 2.64 (2H, t, J = 6.3 Hz), 2.09–2.16 (2H, m)

¹³C NMR (126 MHz, CDCl₃) δ: 198.3, 144.4, 133.3, 132.5, 128.7, 127.0, 126.5, 39.1, 29.6, 23.2

4-chloro-1-tetralone (2v)



yellow liquid, $R_f = 0.34$ (hexane/EtOAc = 5/1)

Yield: 219 mg, 61%

¹H NMR (500 MHz, CDCl₃) δ: 8.05 (1H, d, J = 8.6 Hz), 7.58 (1H, t, J = 8.3 Hz), 7.42–7.51 (2H, m), 5.38 (1H, t, J = 4.0 Hz), 3.11–3.20 (1H, m), 2.49–2.74 (3H, m)

¹³C NMR (126 MHz, CDCl₃) δ: 196.3, 142.2, 134.1, 131.0, 129.3, 129.0, 127.3, 56.6, 33.9, 31.8

IR (neat, cm⁻¹) ν: 2959, 1687, 1599, 1455, 1285, 1234, 921, 881, 777, 720

HRMS (ESI): m/z calcd for $C_{10}H_9ClONa$ [M+Na]⁺ 203.0234, Found 203.0234

(S)-3,7-dimethyloct-6-en-1-yl 4-nitrobenzoate (S1w)

yellow liquid, $R_f = 0.60$ (hexane/EtOAc = 5/1), $[\alpha]_D^{22} = -1.9$ (c = 1.45, MeOH)

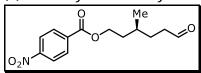
¹H NMR (500 MHz, CDCl₃) δ: 8.27 (2H, d, J = 8.9 Hz), 8.19 (2H, d, J = 8.9 Hz), 5.07 (1H, t, J = 7.2 Hz), 4.34–4.46 (2H, m), 1.92–2.07 (2H, m), 1.78–1.87 (1H, m), 1.55–1.71 (2H, m), 1.65 (3H, s), 1.59 (3H, s), 1.34–1.44 (1H, m), 1.19–1.30 (1H, m), 0.97 (3H, d, J = 6.3 Hz)

¹³C NMR (126 MHz, CDCl₃) δ: 164.6, 150.4, 135.8, 131.4, 130.6, 124.4, 123.4, 64.4, 36.8, 35.3, 29.5, 25.6, 25.3, 19.4, 17.6

IR (neat, cm⁻¹) ν: 2925, 1726, 1608, 1529, 1349, 1276, 1102, 719

HRMS (ESI): m/z calcd for $C_{17}H_{23}NO_4Na$ [M+Na]⁺ 328.1519, Found 328.1523

(S)-3-methyl-6-oxohexyl 4-nitrobenzoate (S2w)

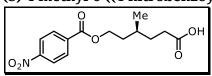


yellow liquid, $R_f = 0.48$ (hexane/EtOAc = 2/1), $[\alpha]_D^{21} = -3.9$ (c = 2.17, MeOH)

¹H NMR (500 MHz, CDCl₃) δ: 9.77 (1H, s), 8.26 (2H, d, J = 8.0 Hz), 8.17 (2H, d, J = 8.0 Hz), 4.36–4.46 (2H, m), 2.42–2.55 (2H, m), 1.59–1.87 (4H, m), 1.47–1.56 (1H, m), 0.97 (3H, d, J = 6.3 Hz) (13C NMR (126 MHz, CDCl₃) δ: 202.2, 164.6, 150.4, 135.6, 130.6, 123.5, 64.0, 41.4, 35.2, 29.5, 28.6, 19.1

IR (neat, cm⁻¹) ν: 2960, 2723, 1724, 1607, 1528, 1350, 1278, 1104, 875, 720 **HRMS (ESI)**: *m/z* calcd for C₁₄H₁₇NO₅Na [M+Na]⁺ 302.0999, Found 302.0995

(S)-4-methyl-6-((4-nitrobenzoyl)oxy)hexanoic acid (1w)



yellow solid, $R_f = 0.40$ (hexane/EtOAc = 1/1), $[\alpha]_D^{21} = 0$ (c = 1.69, MeOH)

¹H NMR (400 MHz, CDCl₃) δ: 8.28 (2H, d, J = 8.5 Hz), 8.19 (2H, d, J = 8.5 Hz), 4.35–4.48 (2H, m), 2.31–2.50 (2H, m), 1.47–1.90 (5H, m), 0.99 (3H, d, J = 6.3 Hz)

¹³C NMR (100 MHz, CDCl₃) δ: 180.0, 164.7, 150.4, 135.7, 130.6, 123.5, 64.0, 35.1, 31.5, 31.3, 29.3, 19.0

IR (neat, cm⁻¹) ν: 3112, 2961, 1723, 1608, 1528, 1350, 1277, 1103, 876, 719

HRMS (ESI): m/z calcd for $C_{14}H_{17}NO_6Na$ [M+Na]⁺ 318.0948, Found 318.0938

DL-4-chloro-4-methyl-6-((4-nitrobenzoyl)oxy)hexanoic acid (2w)

white solid, $R_f = 0.32$ (hexane/EtOAc = 1/1)

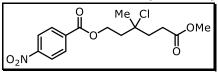
Yield: 38.4 mg (0.3 mmol scale), 37%

¹H NMR (500 MHz, CDCl₃) δ: 8.28 (2H, d, J = 8.6 Hz), 8.20 (2H, d, J = 8.6 Hz), 4.63 (2H, t, J = 6.3 Hz), 2.59–2.73 (2H, m), 2.10–2.36 (4H, m), 1.64 (3H, s)

¹³C NMR (126 MHz, CDCl₃) δ: 178.9, 164.5, 150.5, 135.3, 130.7, 123.5, 70.3, 62.5, 42.3, 38.7, 29.7, 29.6

IR (neat, cm⁻¹) v: 3426, 1724, 1608, 1528, 1413, 1350, 1276, 1104, 874, 719 HRMS (ESI): m/z calcd for $C_{14}H_{16}ClNO_6Na$ [M+Na]⁺ 352.0558, Found 352.0566

DL-3-chloro-6-methoxy-3-methyl-6-oxohexyl 4-nitrobenzoate (2x)



light yellow liquid, $R_f = 0.50$ (hexane/EtOAc = 2/1)

¹H NMR (500 MHz, CDCl₃) δ: 8.28 (2H, d, J = 8.3 Hz), 8.19 (2H, d, J = 8.3 Hz), 4.60 (2H, t, J = 6.6 Hz), 3.68 (3H, s), 2.52–2.63 (2H, m), 2.09–2.34 (4H, m), 1.61 (3H, s)

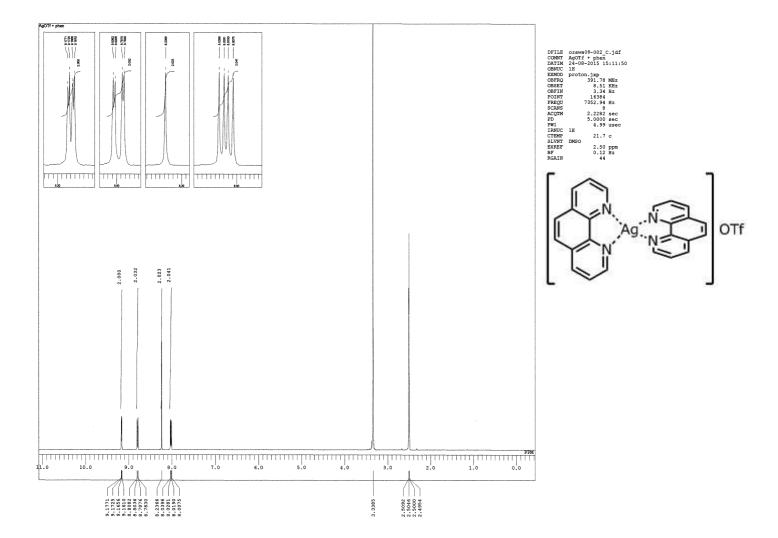
¹³C NMR (100 MHz, CDCl₃) δ: 173.2, 164.4, 150.5, 135.3, 130.6, 123.5, 70.4, 62.4, 51.8, 42.2, 39.0, 29.58, 29.55

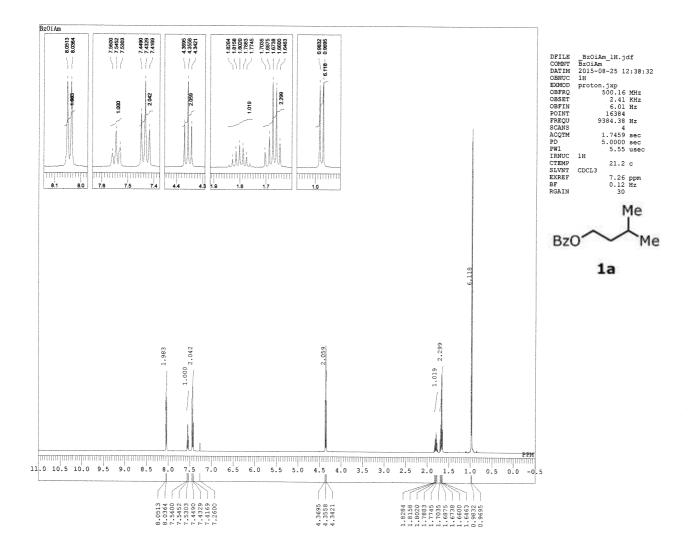
IR (neat, cm⁻¹) v: 2953, 1730, 1607, 1530, 1436, 1271, 1102, 874, 718

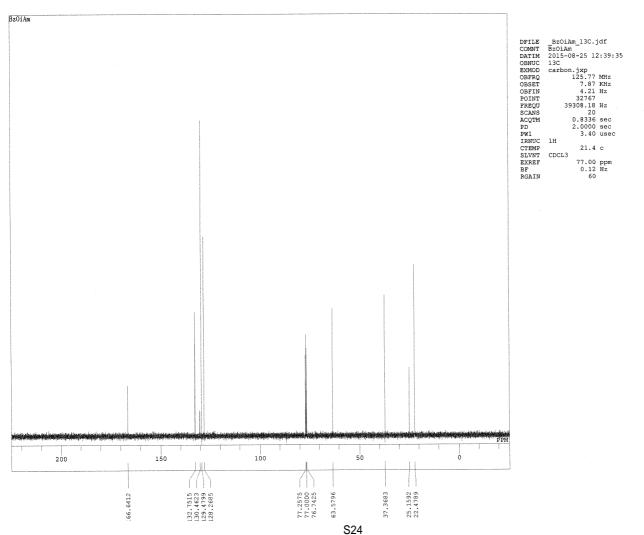
HRMS (ESI): m/z calcd for $C_{15}H_{18}ClNO_6Na$ [M+Na]⁺ 366.0715, Found 366.0706

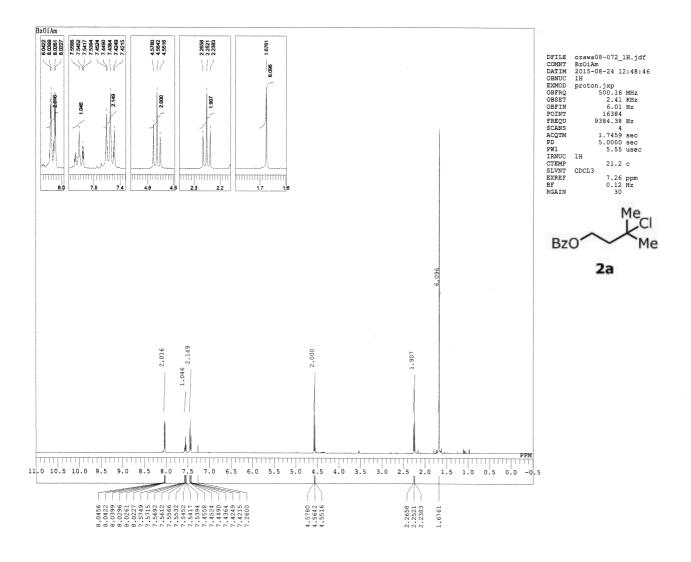
10. NMR spectra

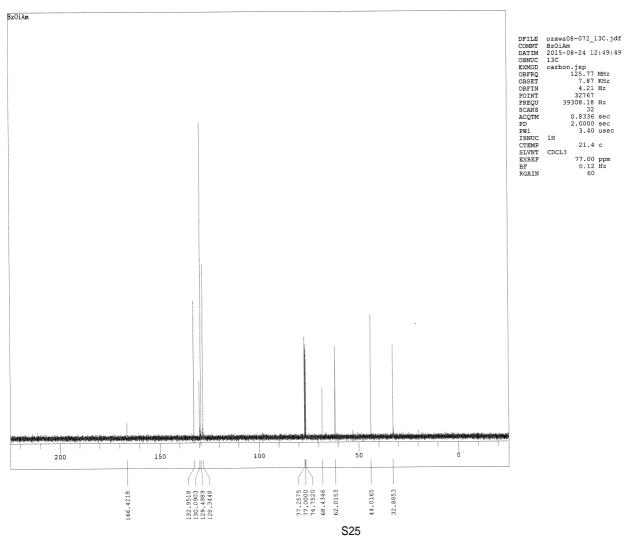
A ¹H NMR spectrum for Ag(phen)₂OTf, and ¹H and ¹³C NMR spectra for the compounds in **9. Analytical data** are appended below.

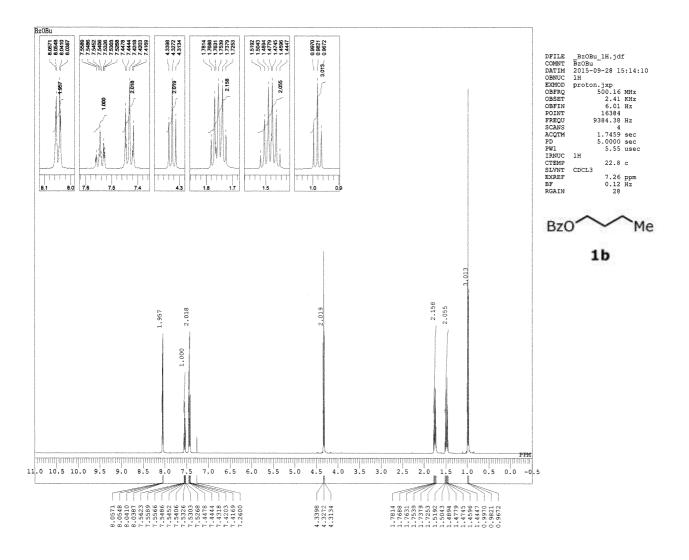


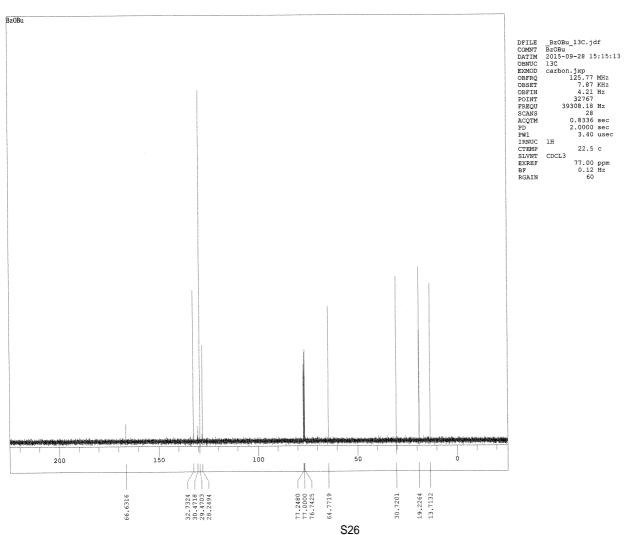


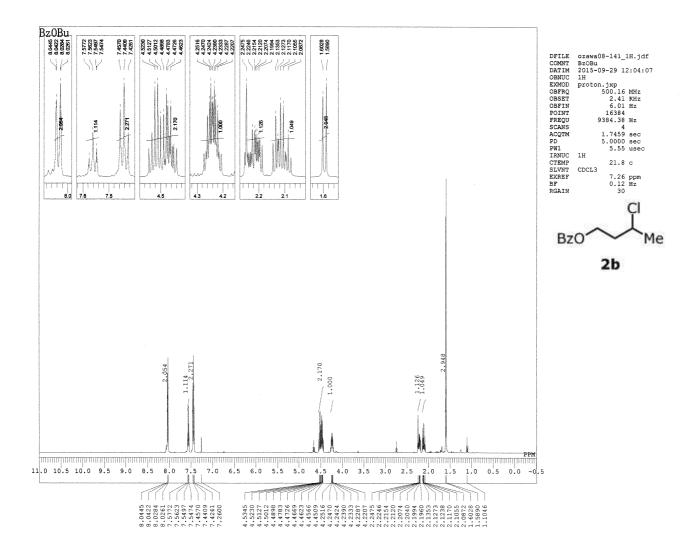


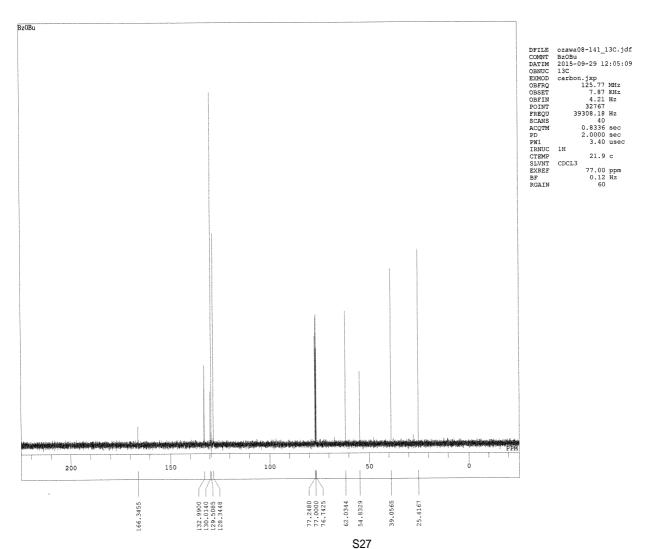


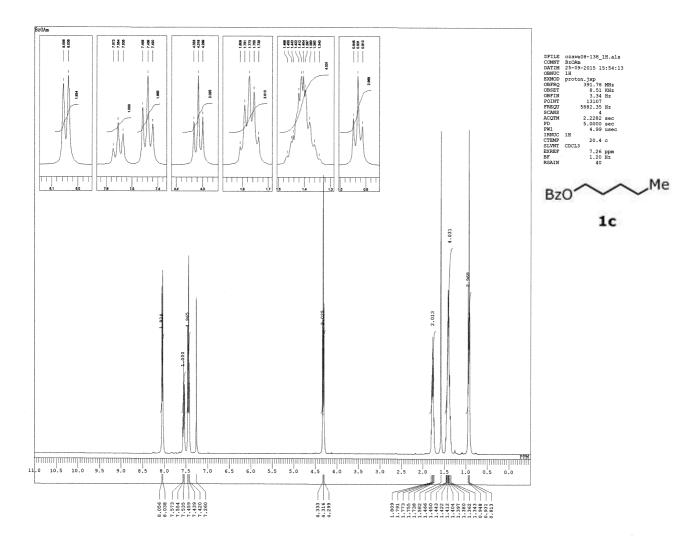


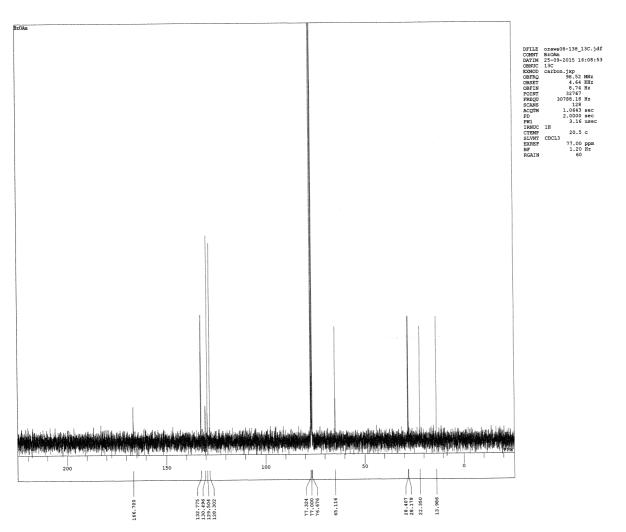


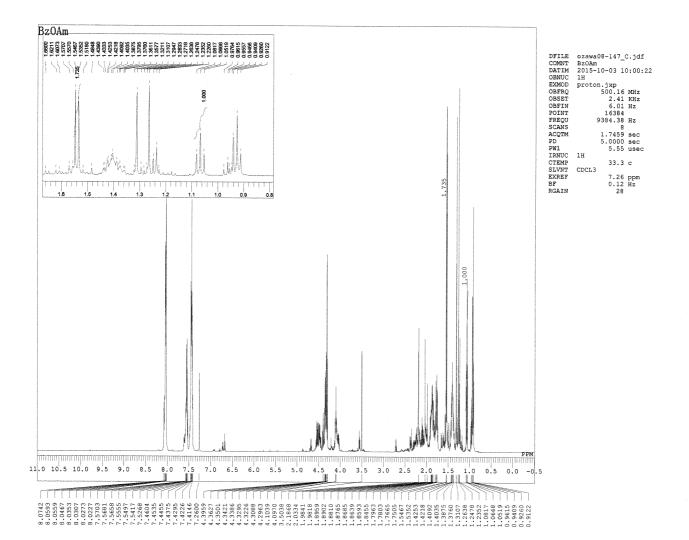






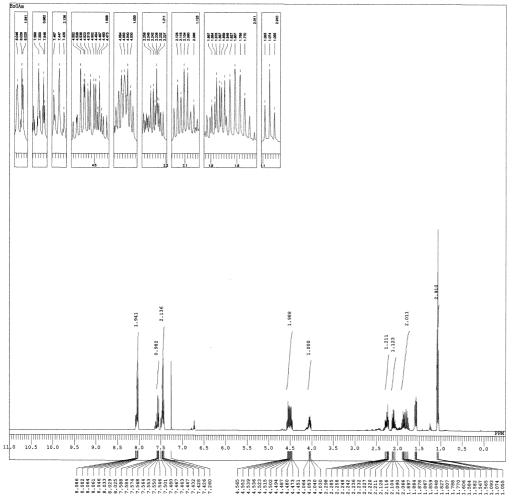


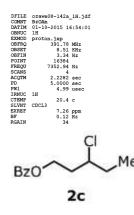


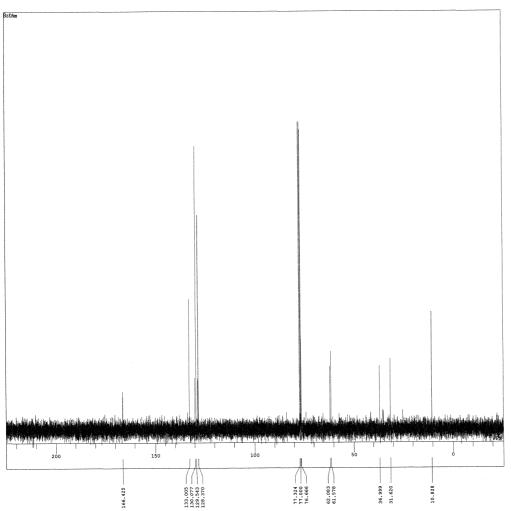


¹H NMR of crude mixture.

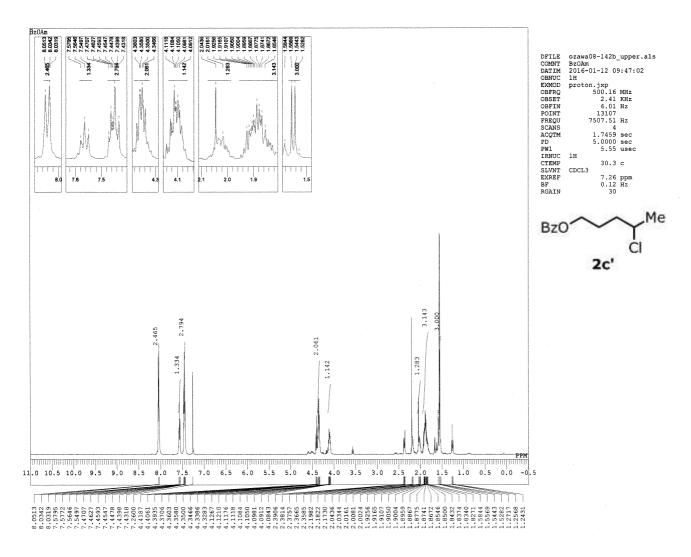
$$BzO \longrightarrow Me \qquad \underbrace{\begin{array}{c} Standard\ conditions \\ \textbf{2c}: \textbf{2c'} = 1:1.7 \end{array}}_{\textbf{BzO} \longrightarrow \textbf{Me}} \qquad + \qquad \underbrace{\begin{array}{c} CI \\ BzO \longrightarrow \textbf{CI} \\ \textbf{2c'} \end{array}}_{\textbf{CI}} Me$$

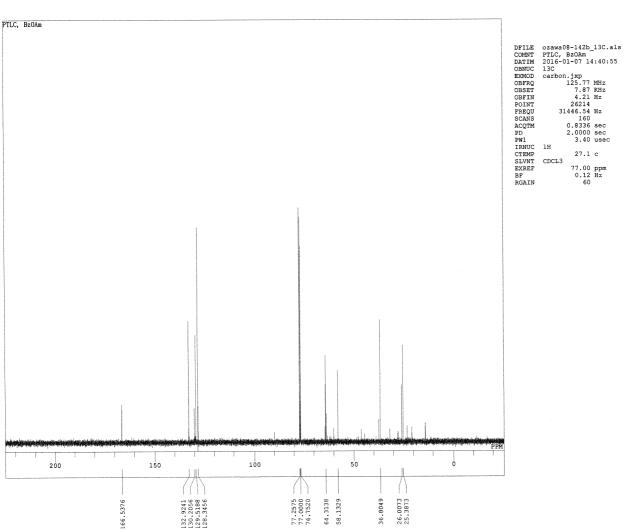




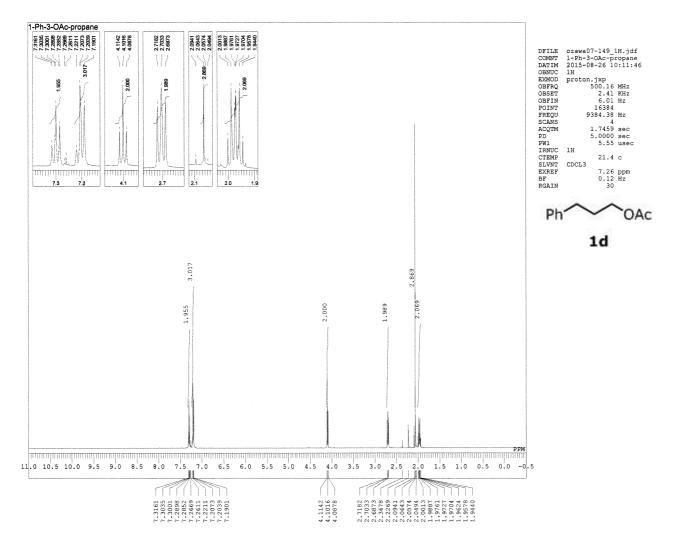


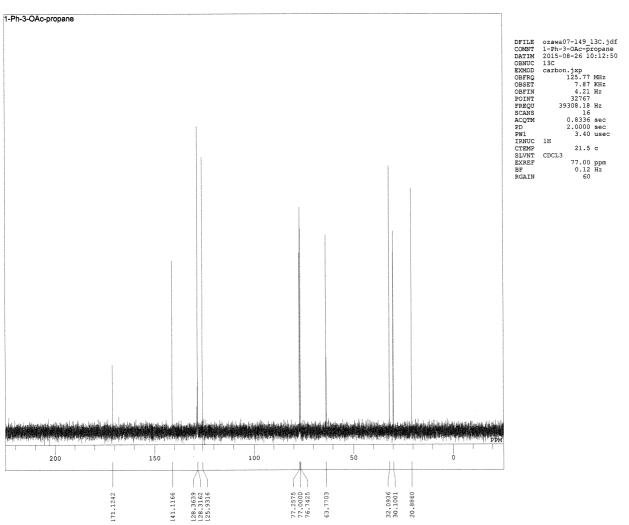
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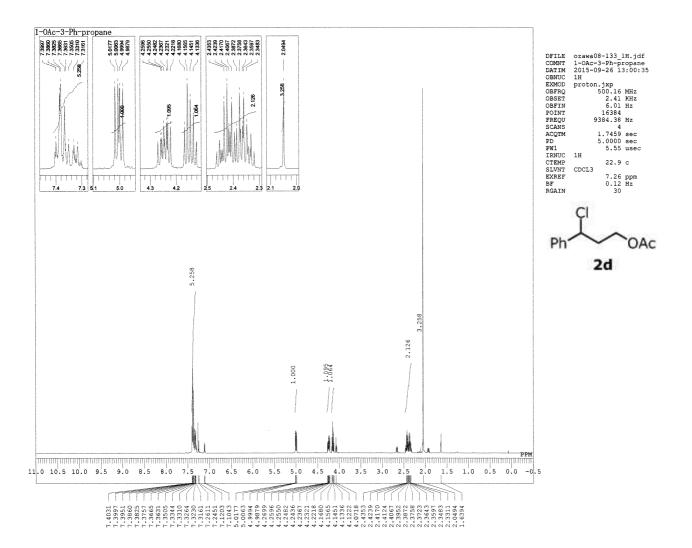


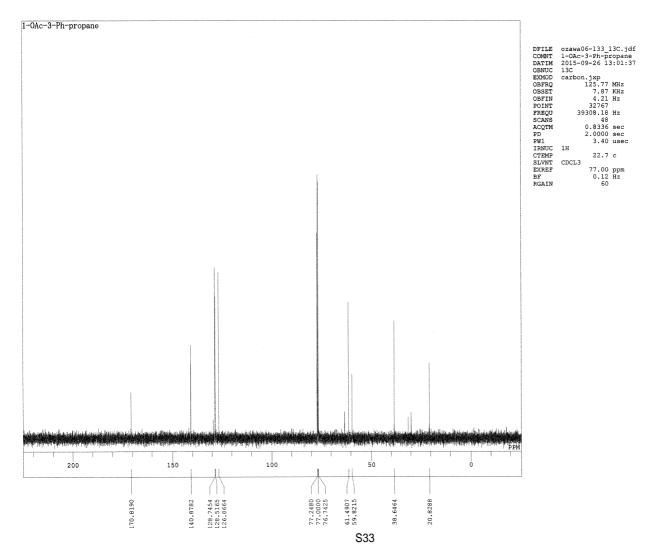
S31

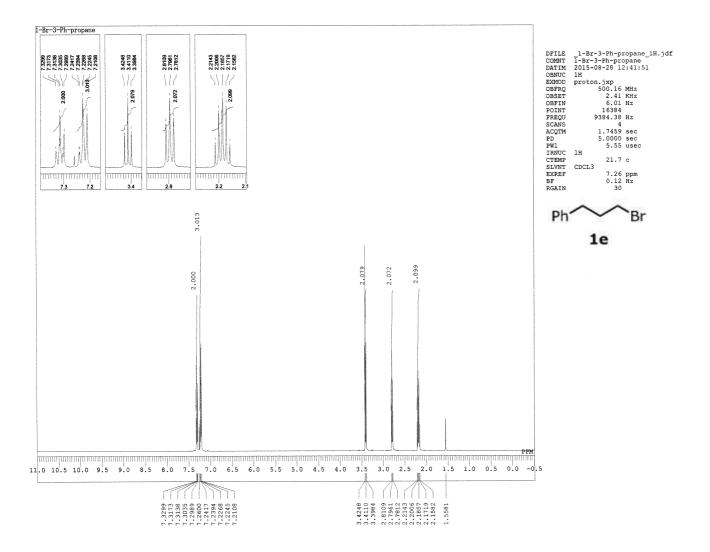


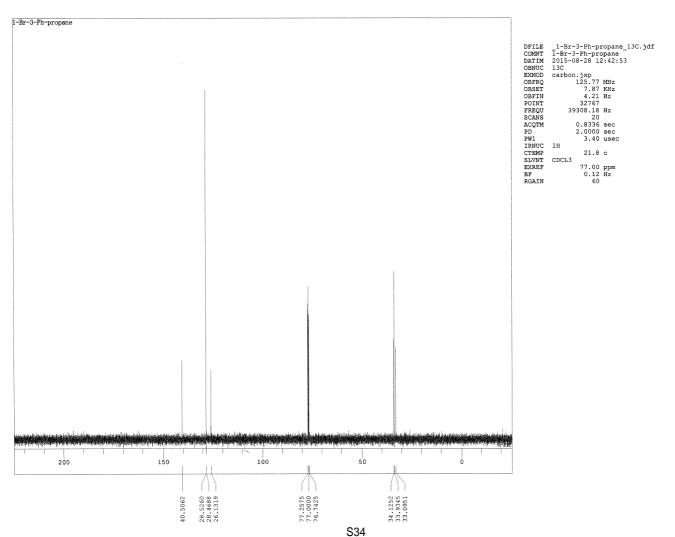


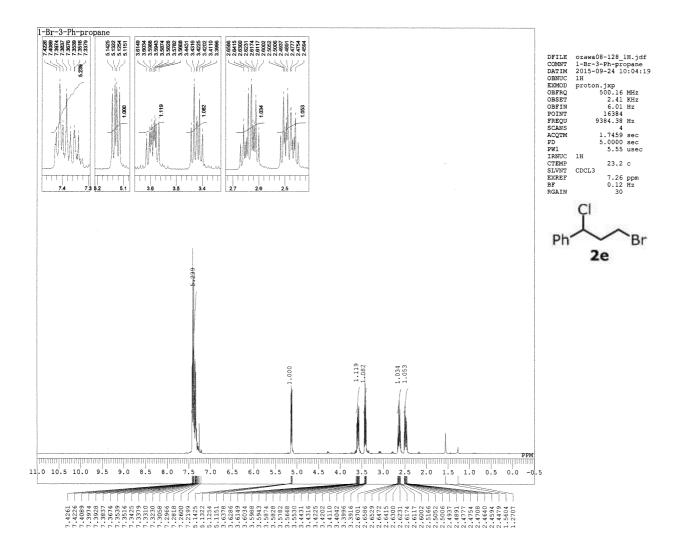
S32

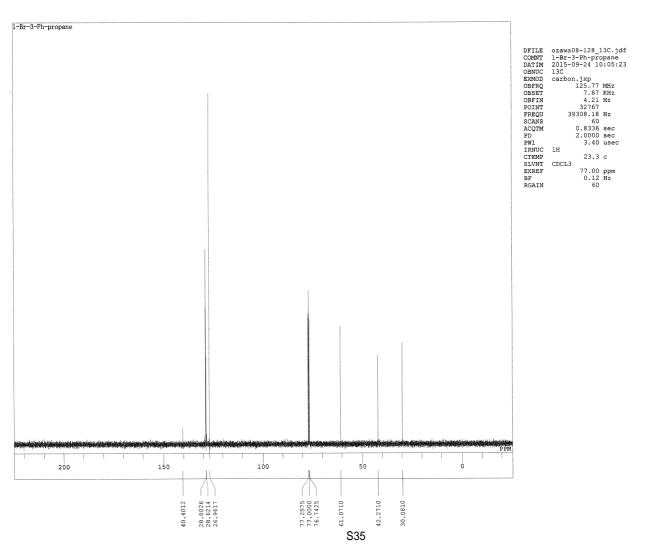


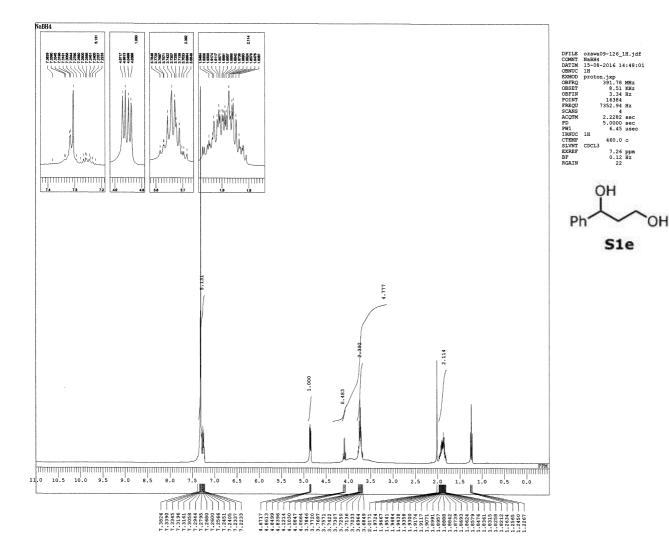


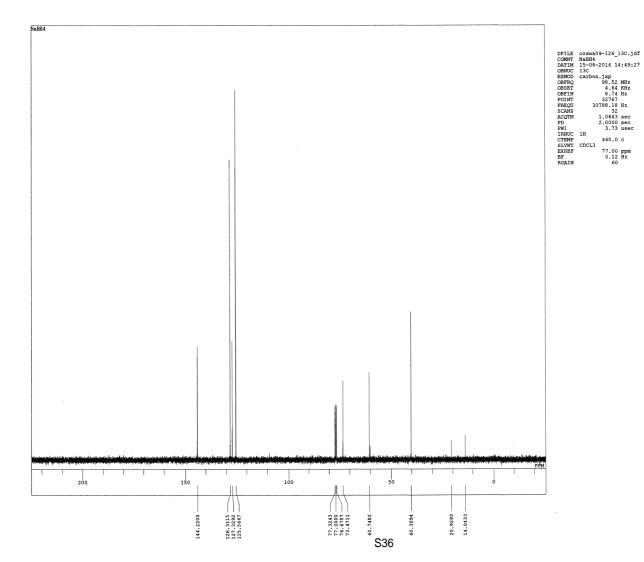


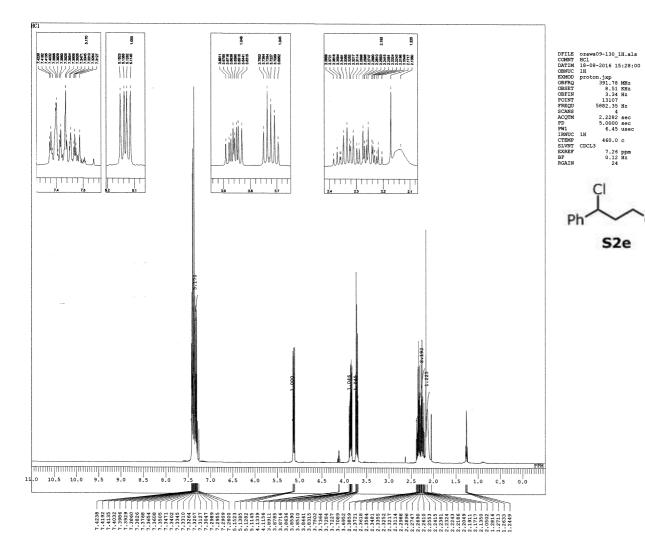




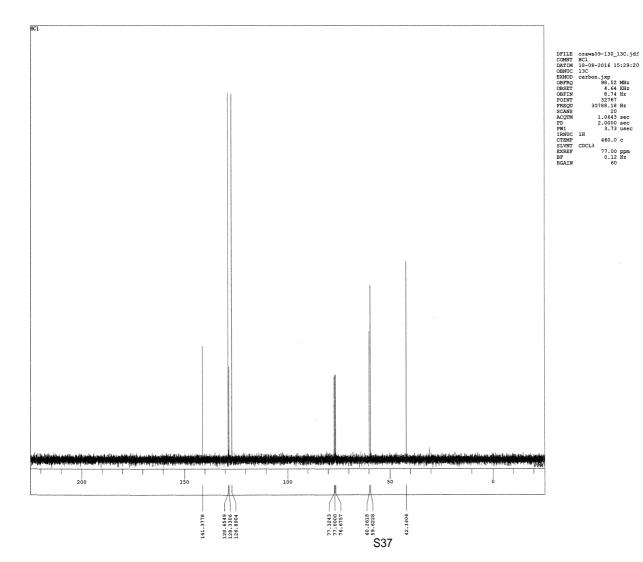


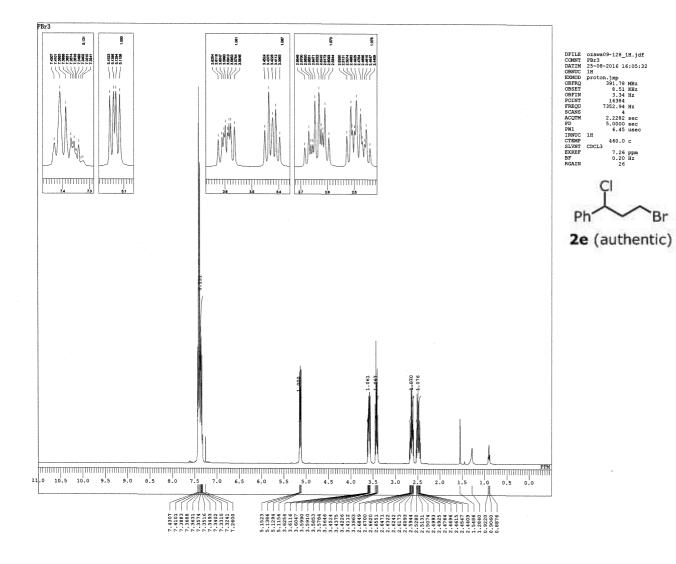


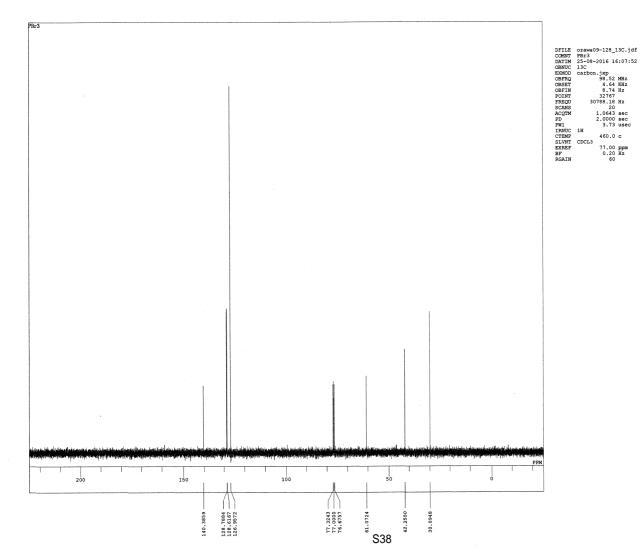


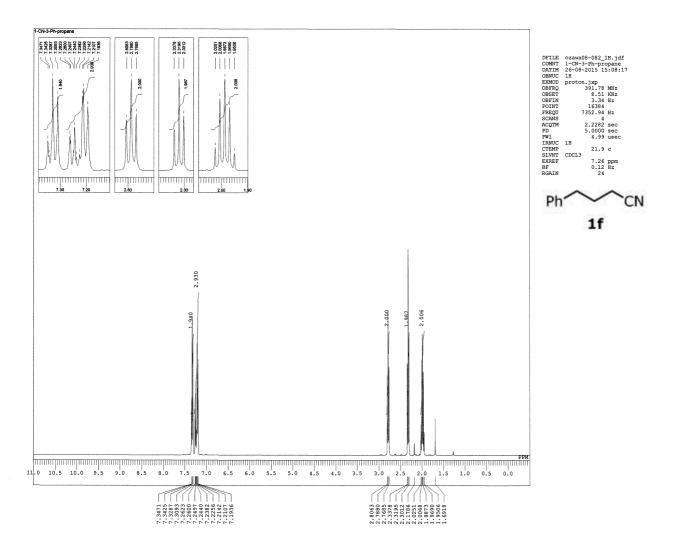


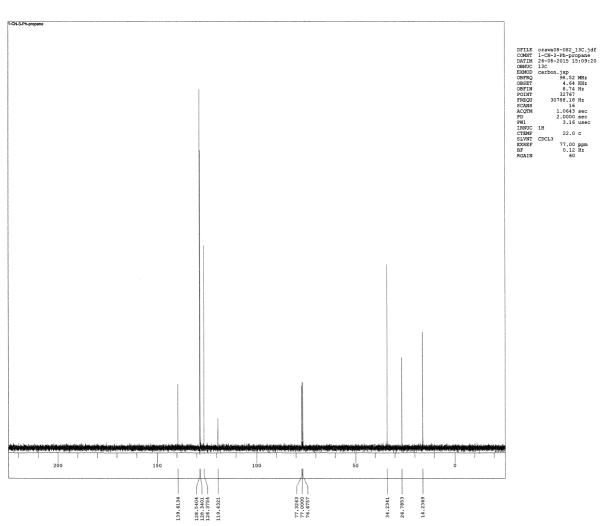
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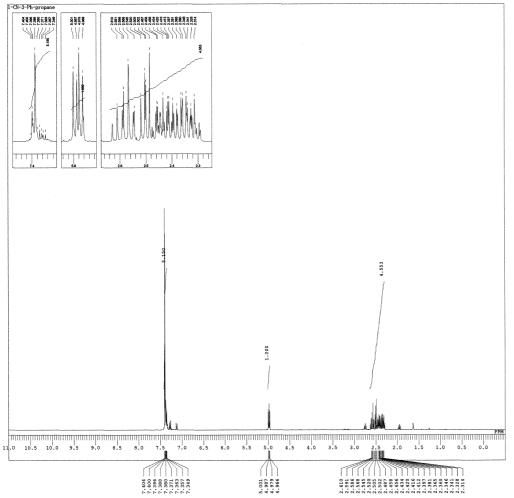


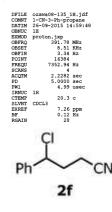


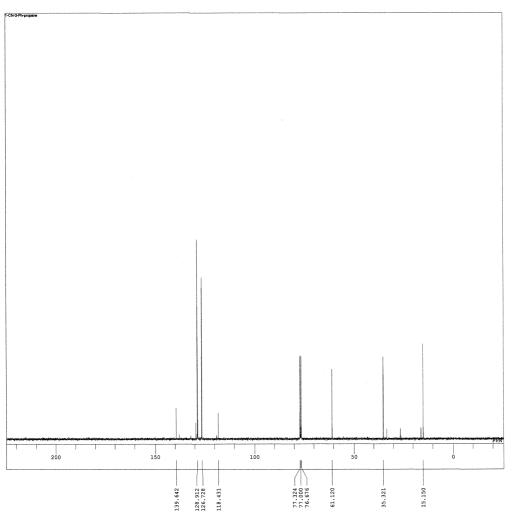


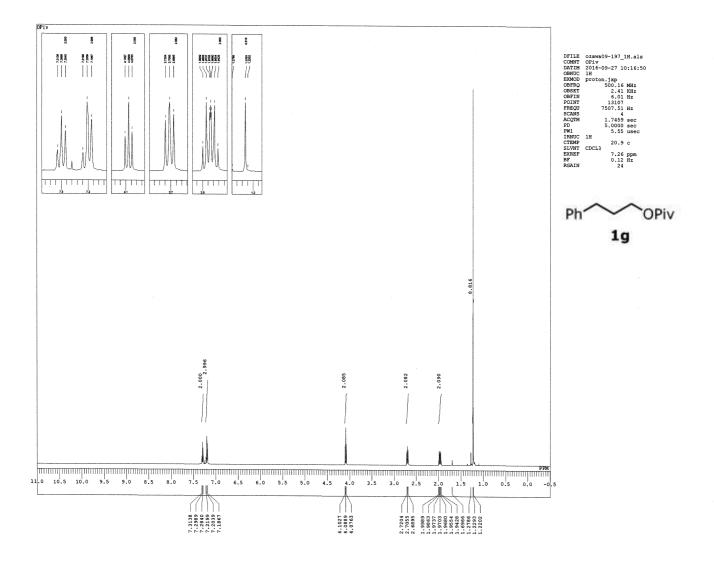


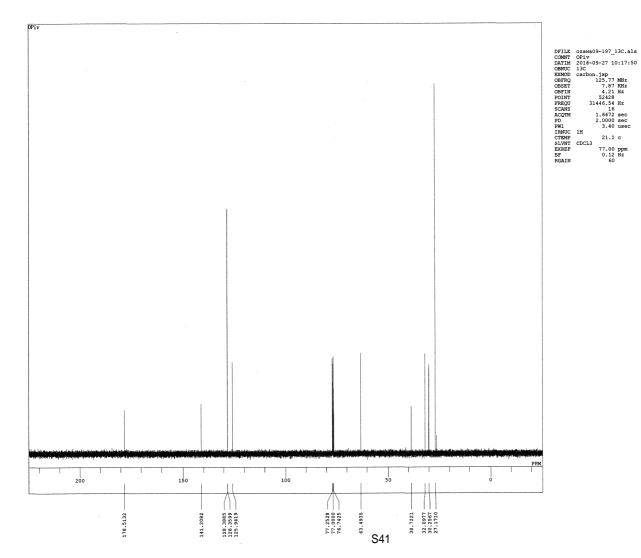


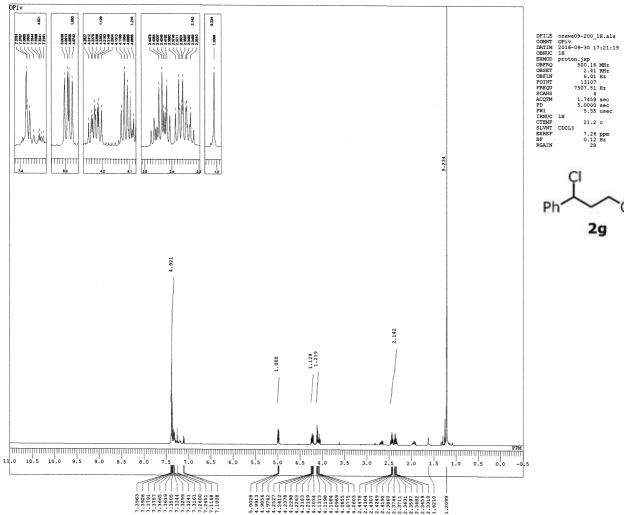




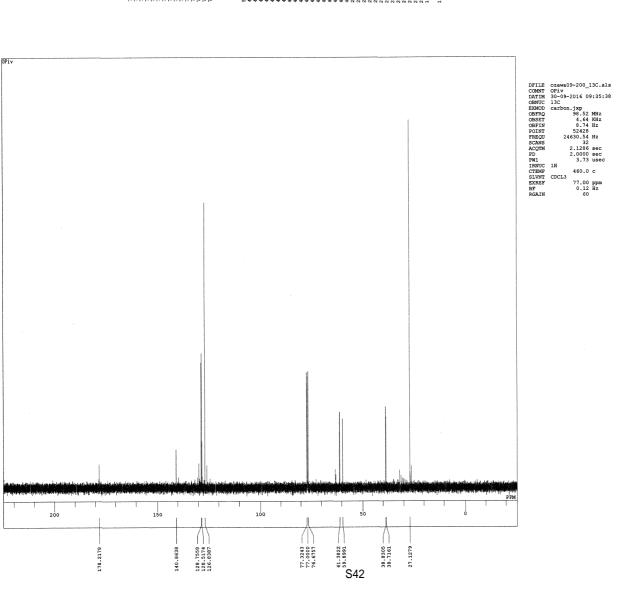


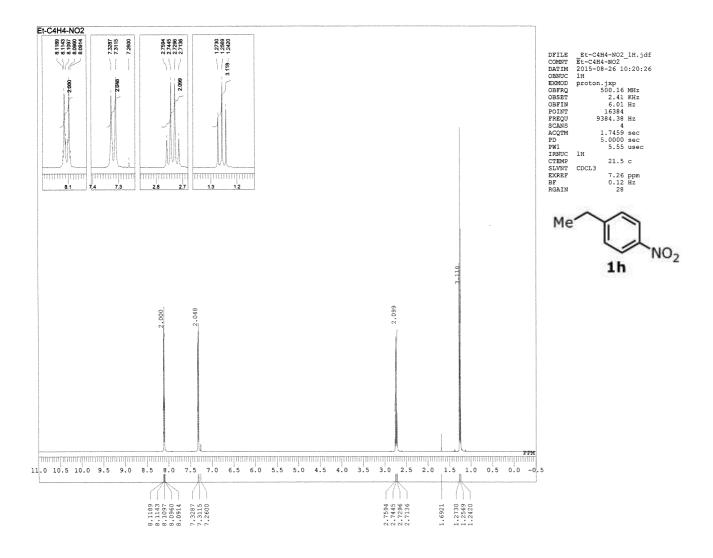


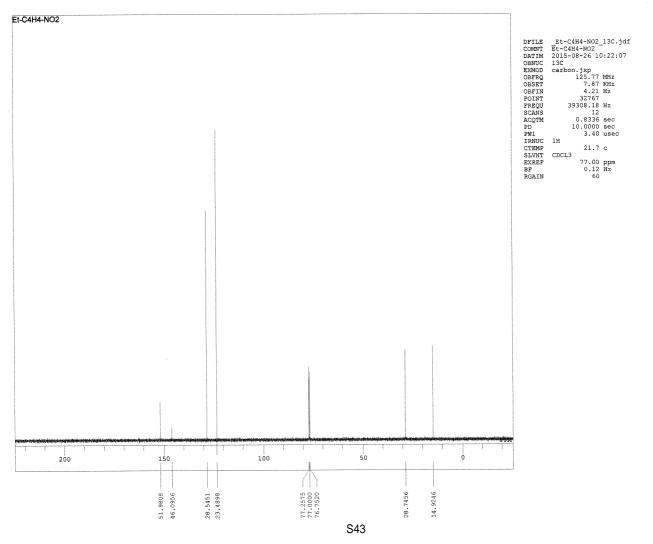


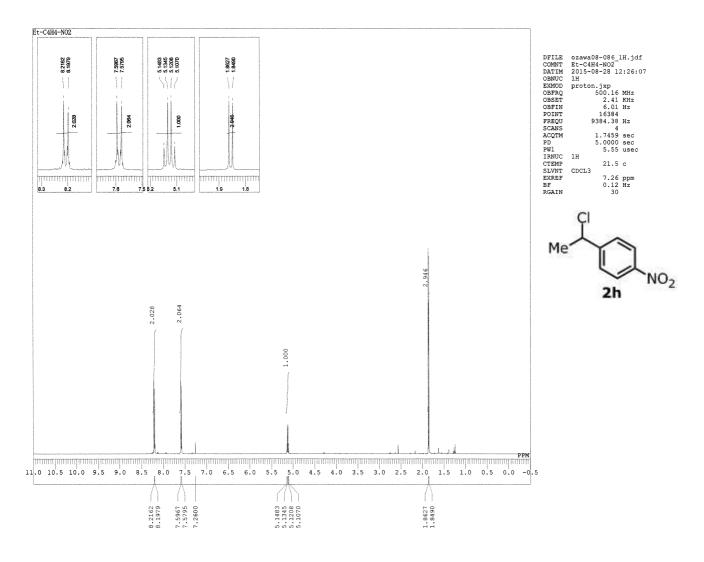


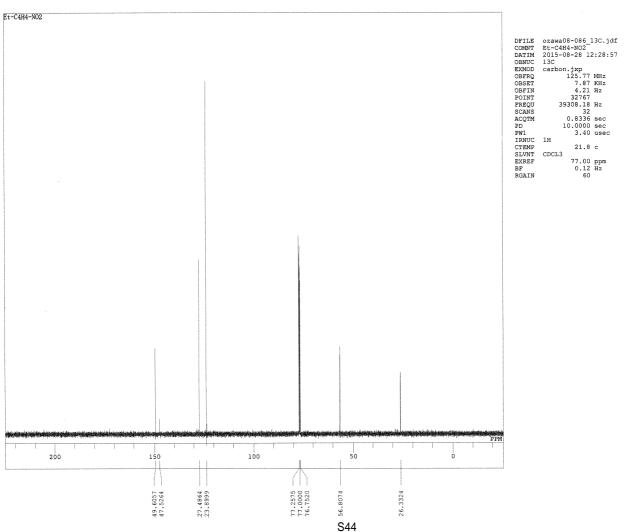
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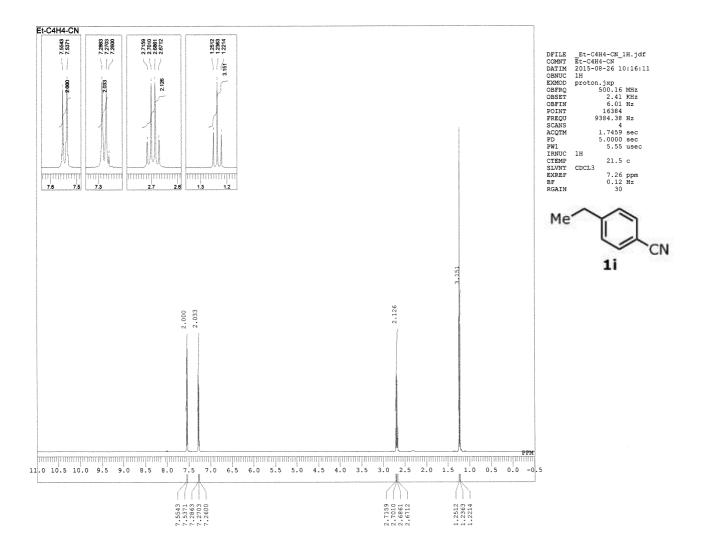


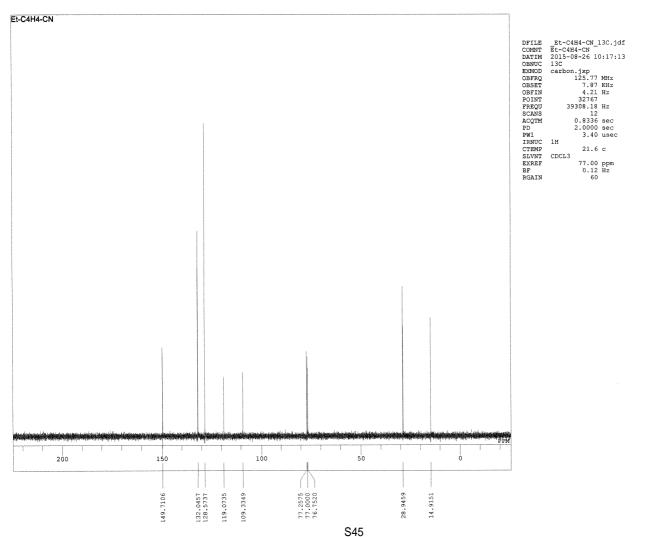


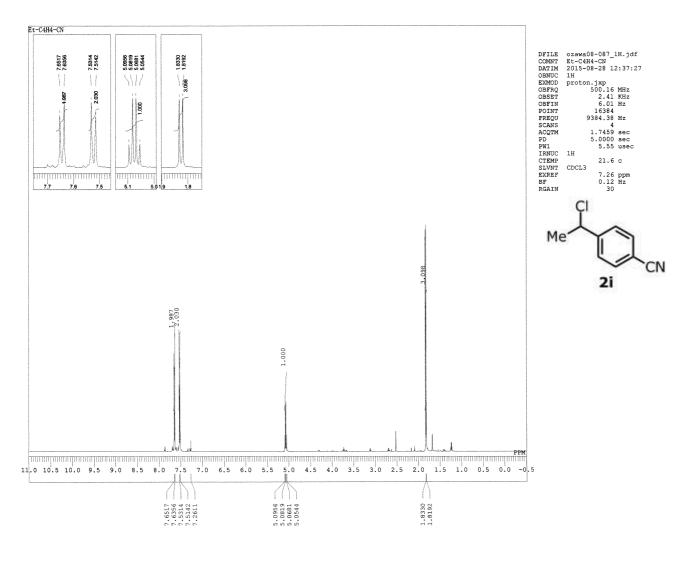


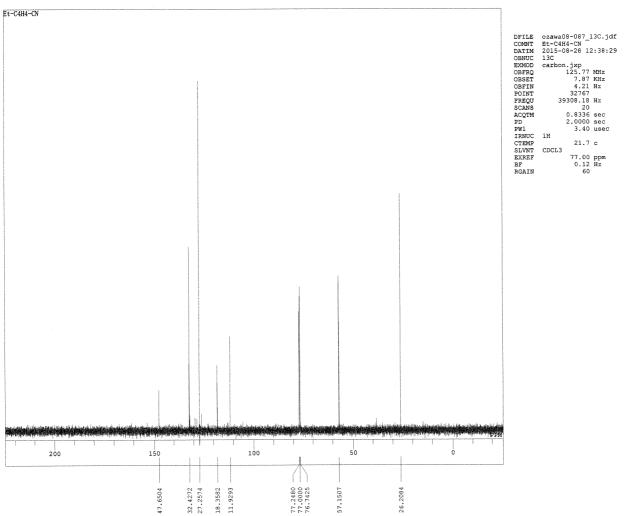


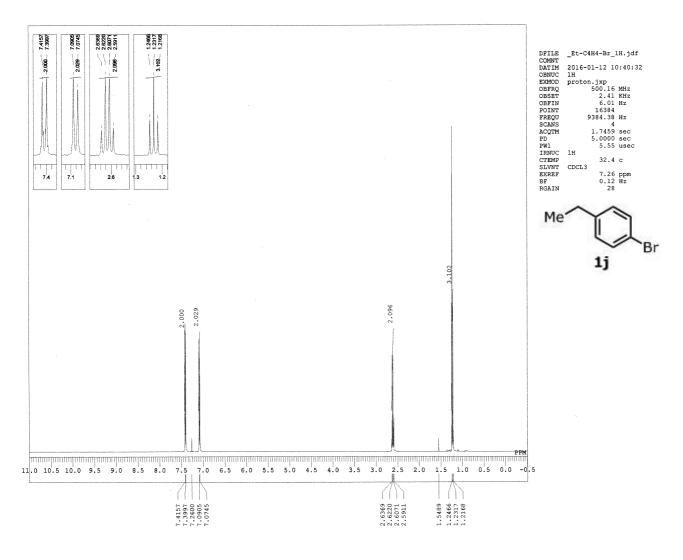


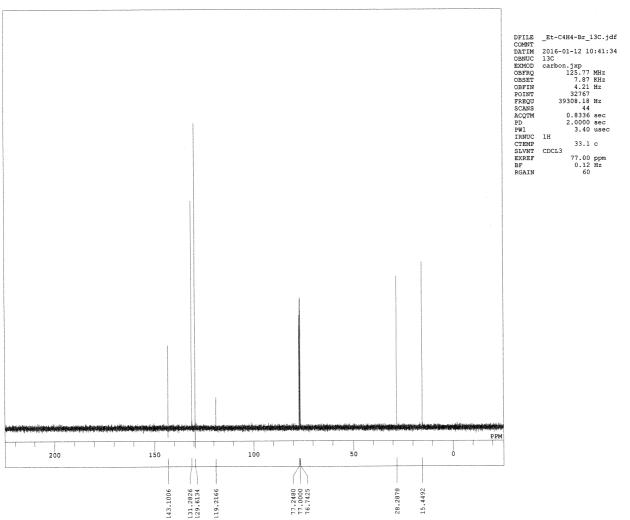


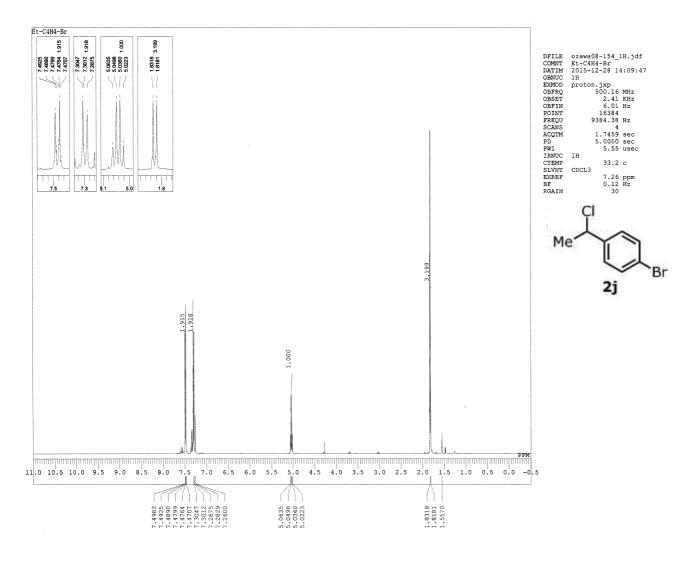


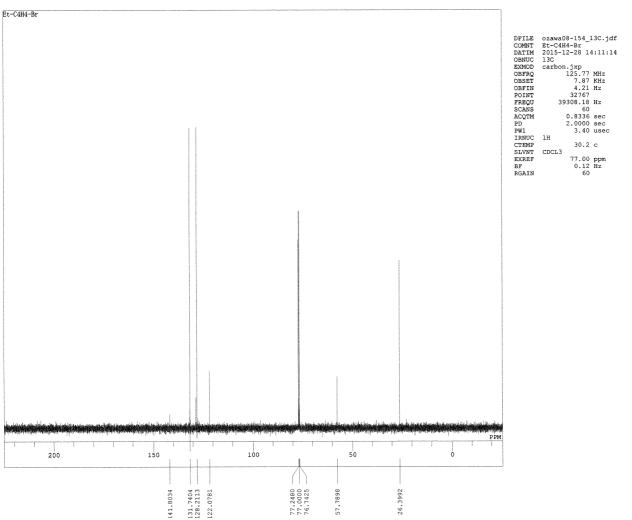


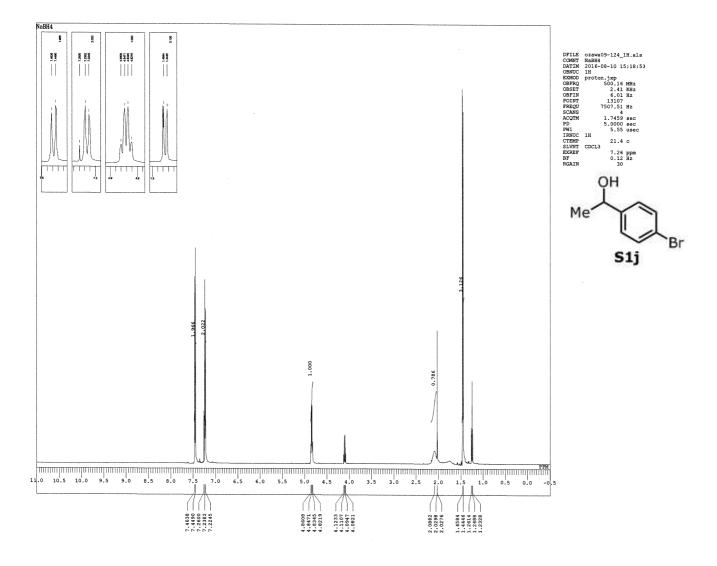


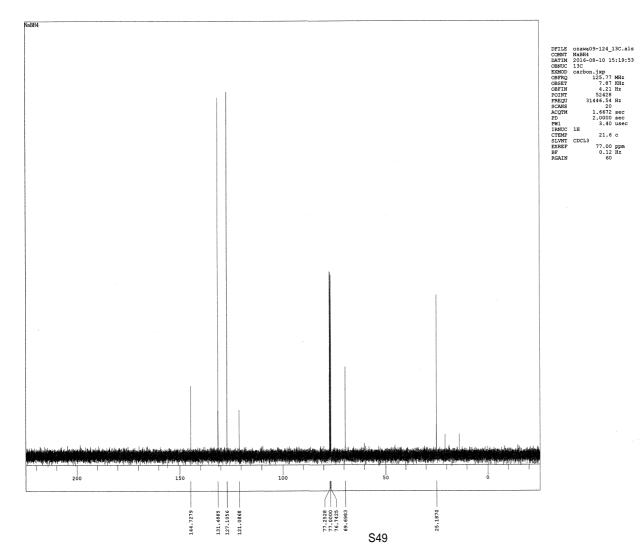


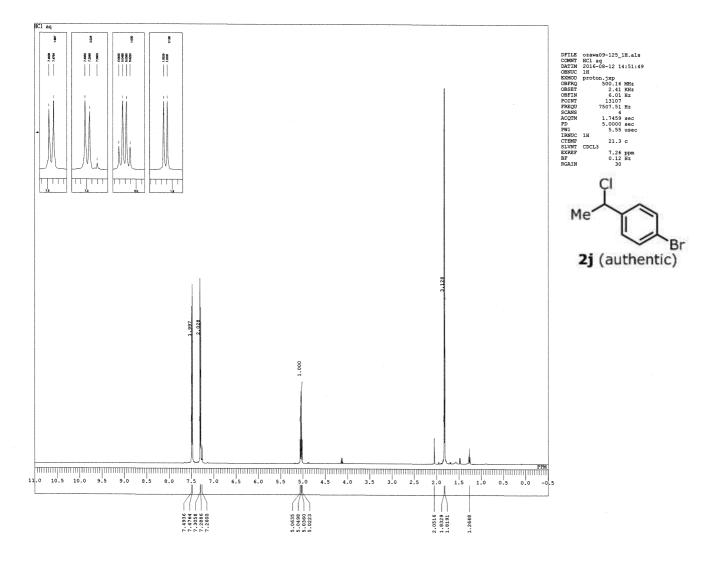


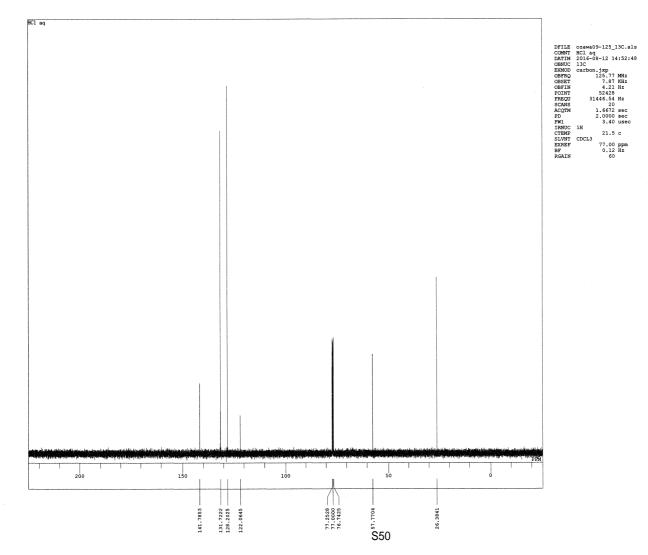


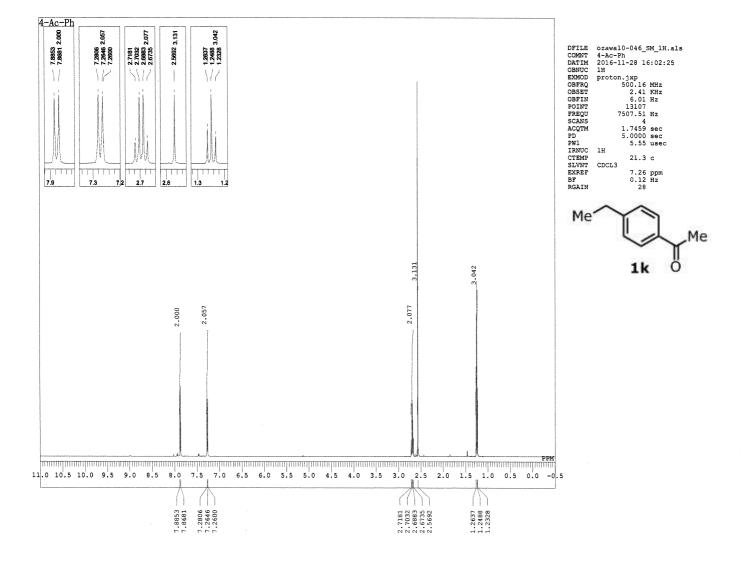


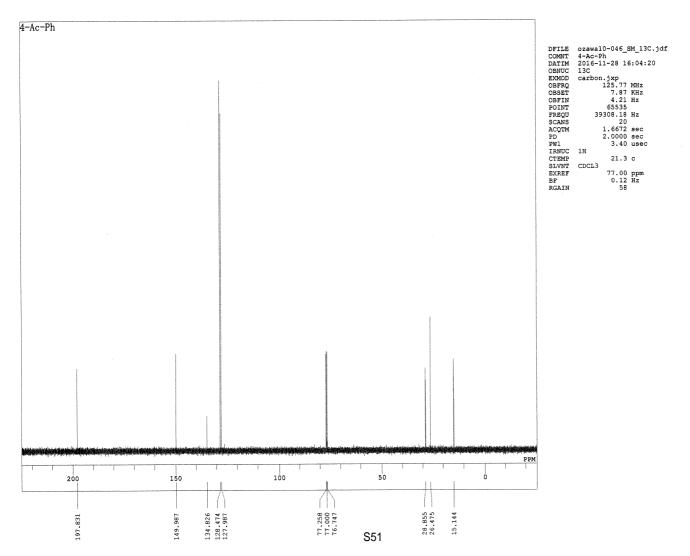


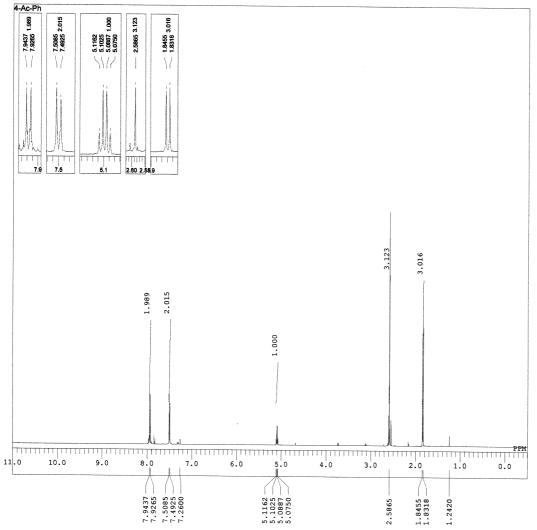


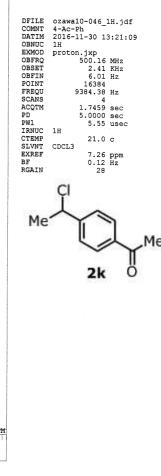


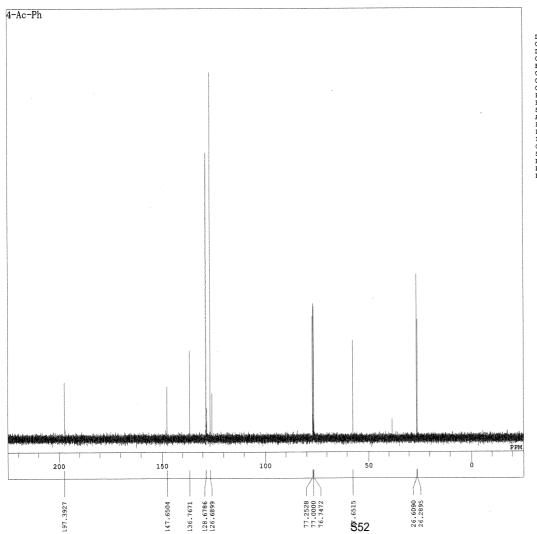




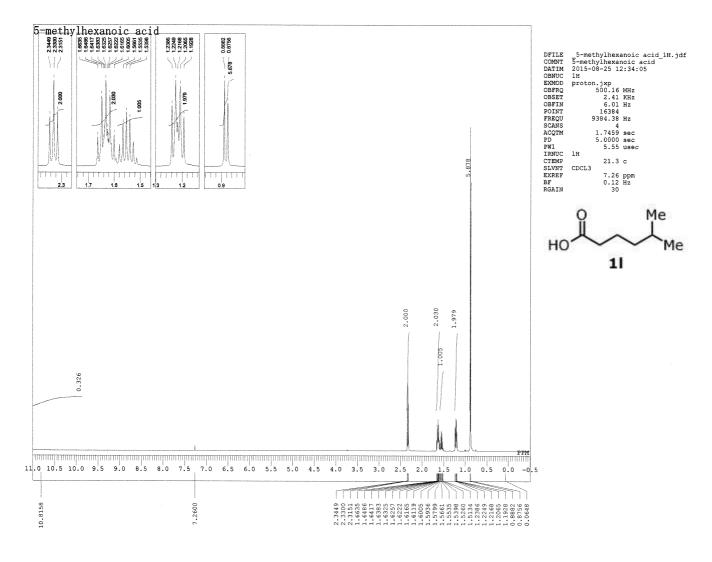


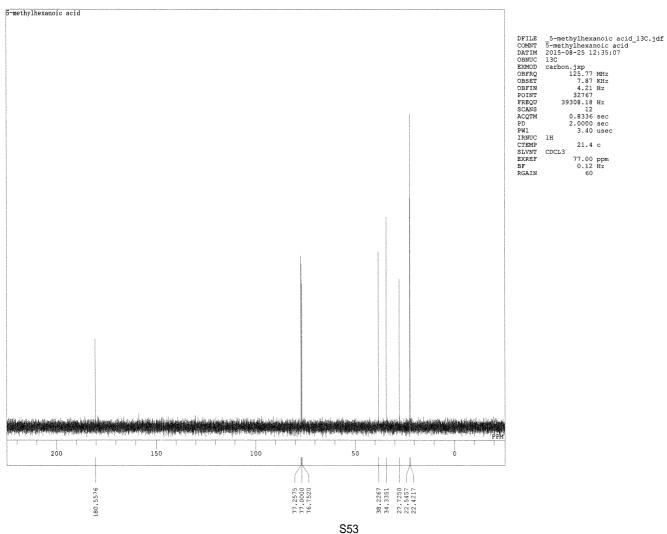


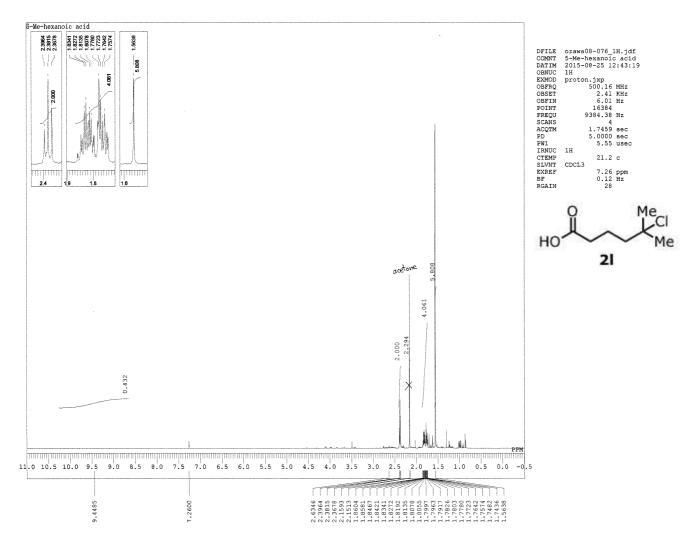


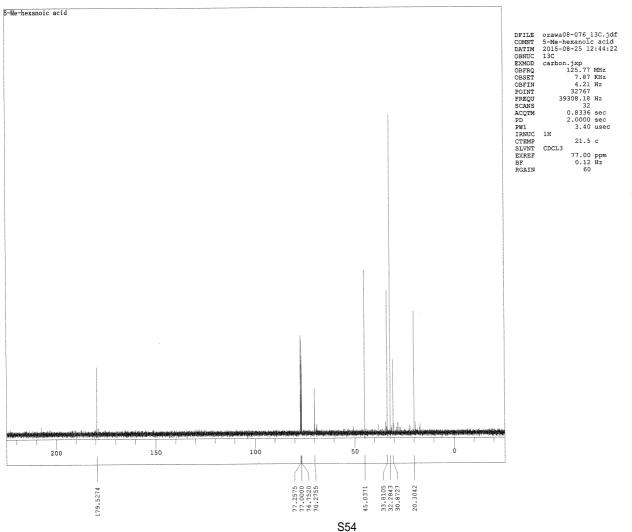


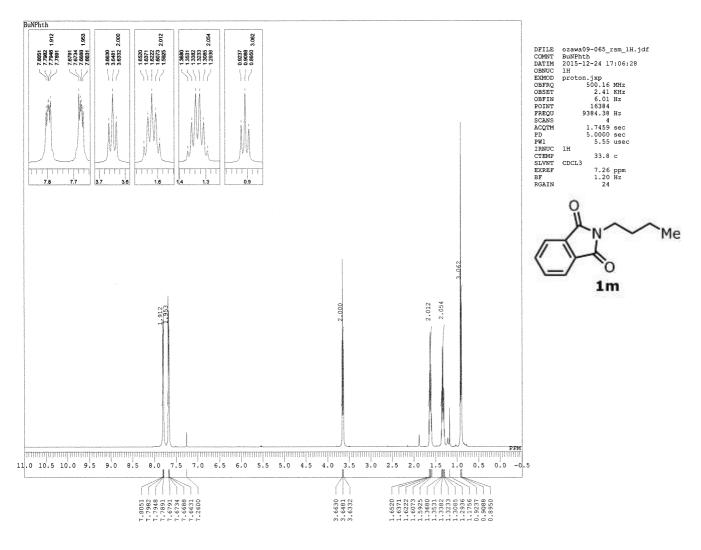
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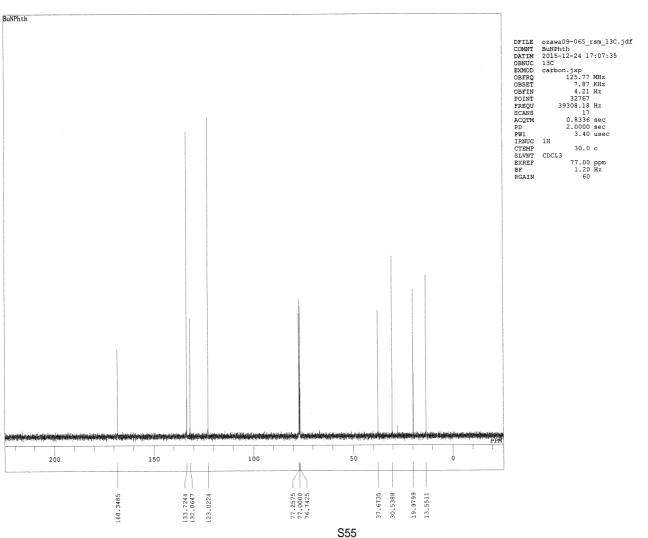


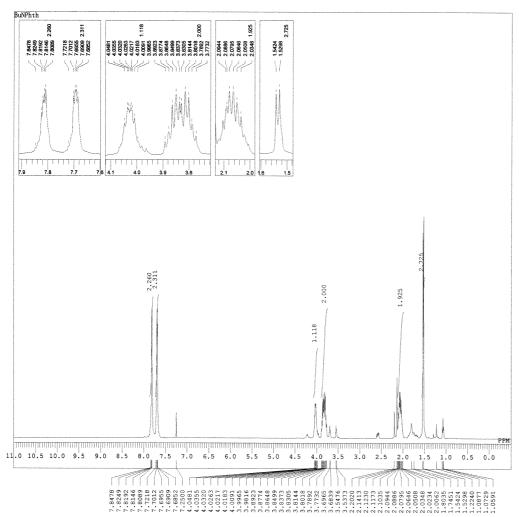


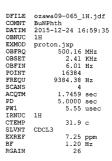


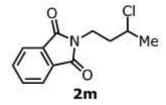


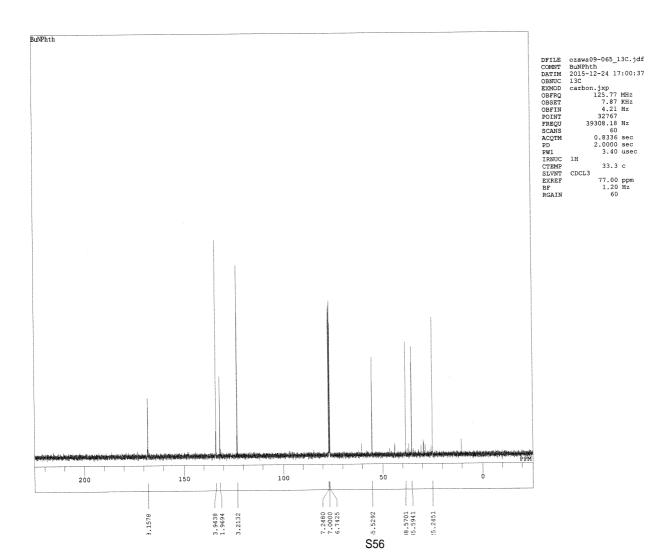


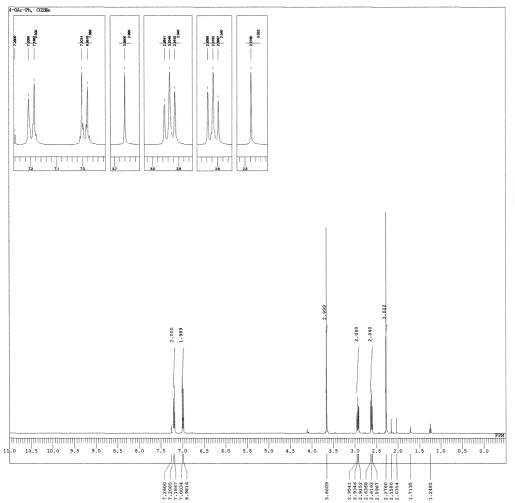




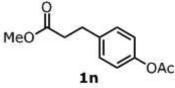


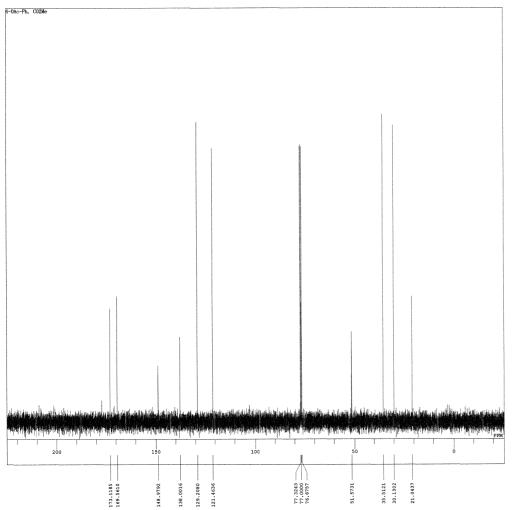




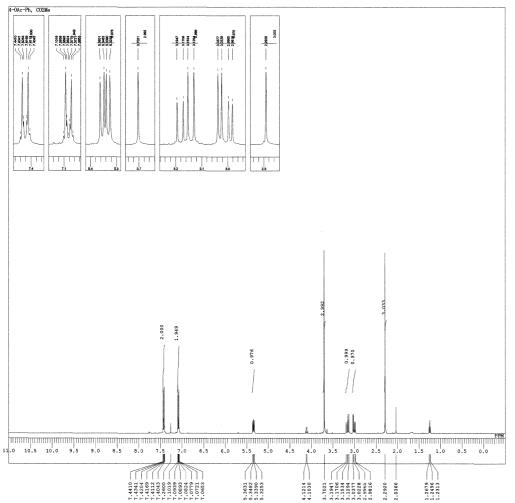


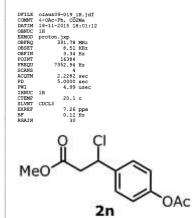


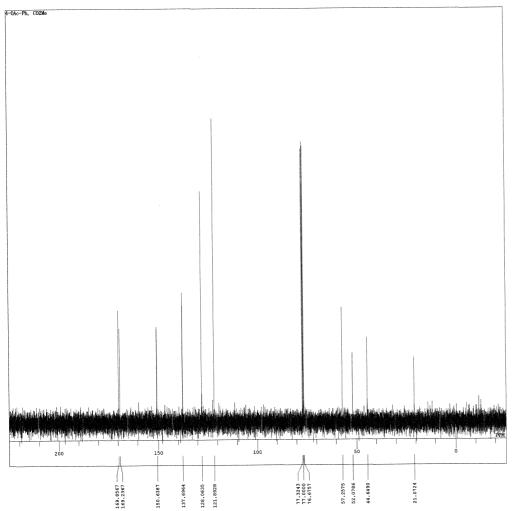




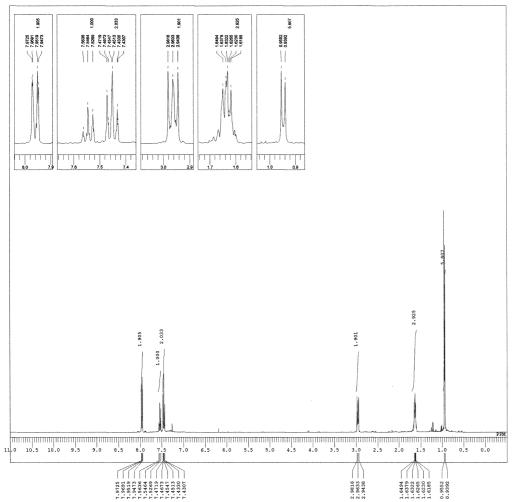
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COMN: 4-OAC-Ph, COZMe
DATM 18-11-2015 17:36:31
DATM 18-11-2015 17:36:31
EXMOD Carbon.]xp
OBERO
OBSE? 4.64 KHz
OBFIN 2.744 Hz
SCANS
SCANS
SCANS
SCANS
SCANS
SCANS
LOGATION 1.0643 sc
PD 2.0000 sec
PD 2.0000 sec
PD 2.0000 sec
LTBUUC 1H 20.3 C
SCHOTT COCLS
EXERT 77.00 PPM
EFF 77.00 PPM
EFF 0.12 Hz
RGAIN 66

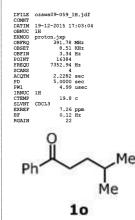


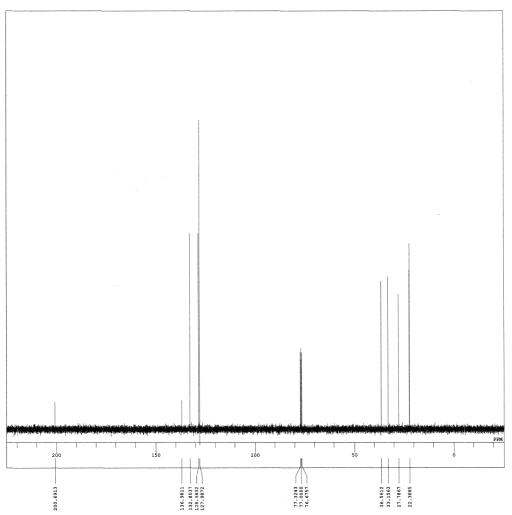




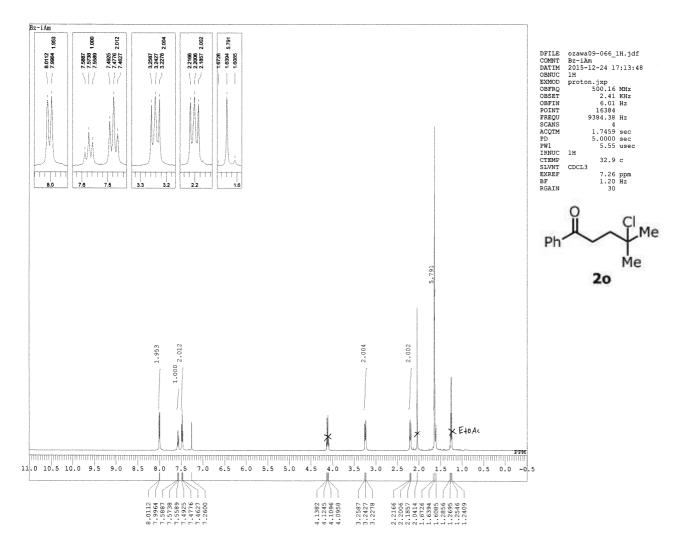
DFILE 0:08409-019 13C.jdf
COMPT 4-0AC-Ph. COZMe
4-0AC-Ph. COZMe
13C.1-2015 16:02:14
OBNUC 13C.
OBNU

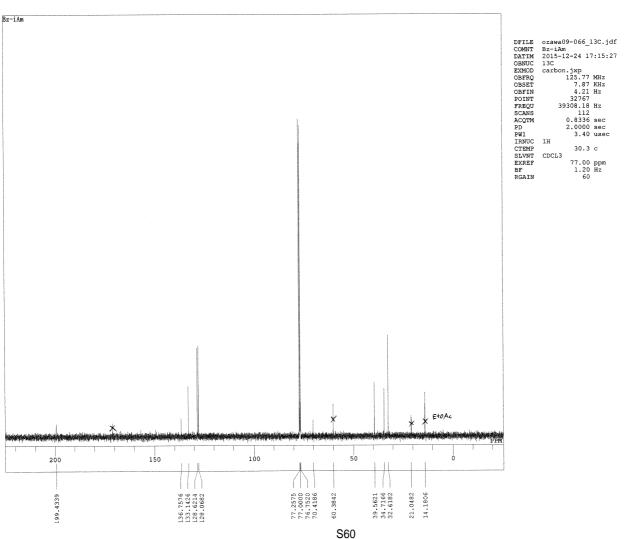


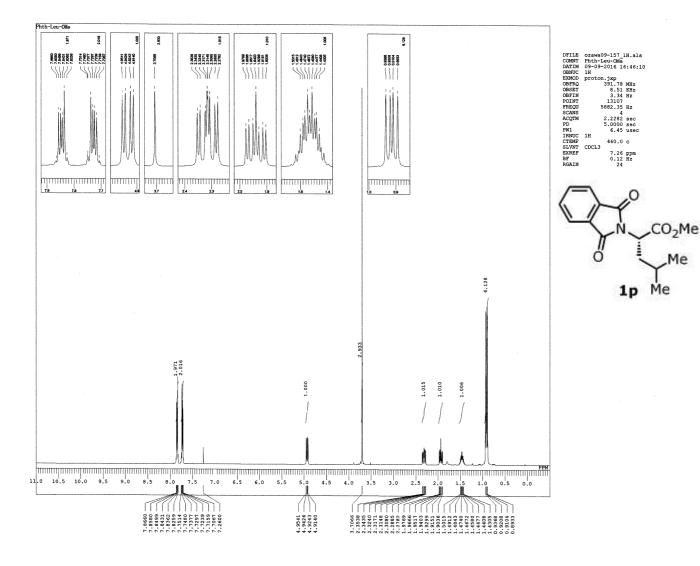




DFILE OZAWA09-059_13C.jdf
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DBMUC 13C
EXMOD Carbon.jxp
OBFRO 98.52 MHz
OBFIN 98.52 MHz
OBFIN 14.64 KHz
OBFIN 2.744 Hz
SCANS 30785.18 Hz
SCANS 10.6643 sec
PD 2.0000 sec
PD 2.0000 sec
PD 1.0643 sec
PD 1.0643 sec
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PD 2.0000 sec
SCANS 10.0643 sec
PD 2.0000 sec
PD 1000 Sec
PD 1

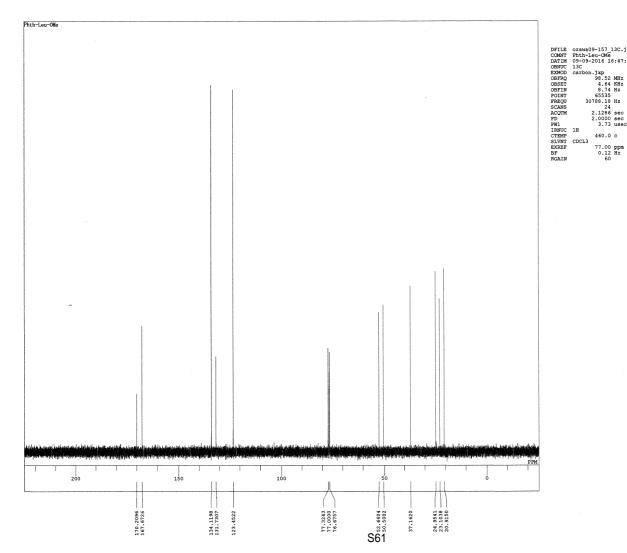


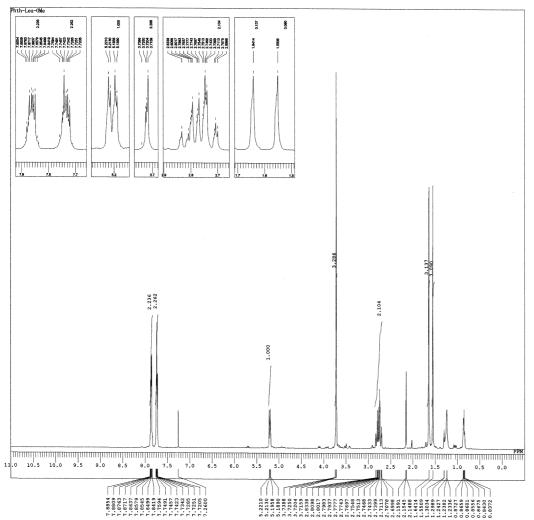


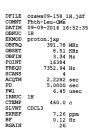


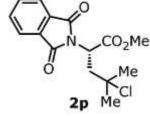
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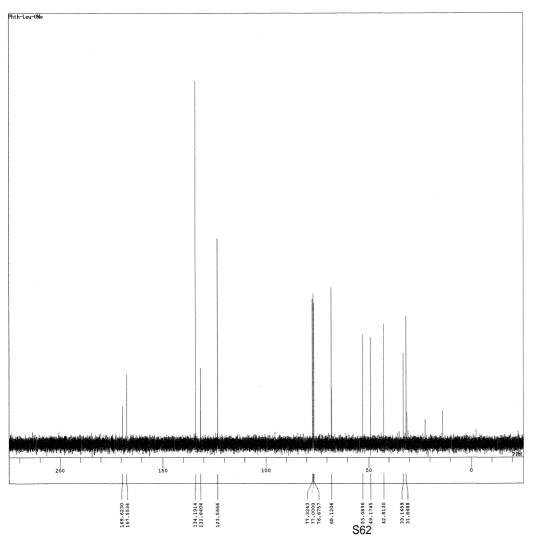
Ozawa09-157 13C.jdf
PRth-Leu-CNG
(99-09-2016 16:47.38
13C
carbon.jap
98.52 MHz
4.64 KHz
8.74 Hz
65535
30788.18 Hz
2.1286 sec
2.0000 sec
3.73 usec
1H



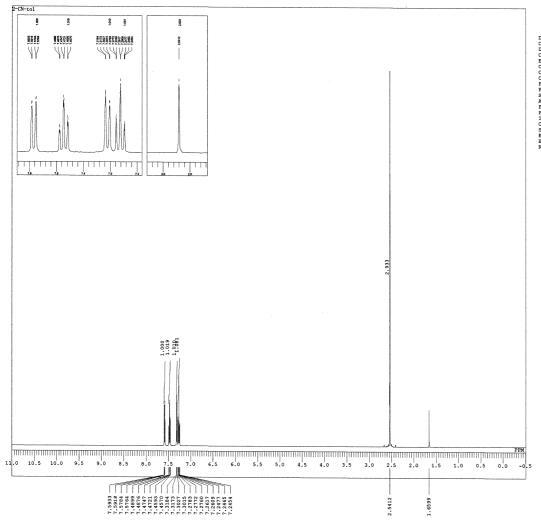


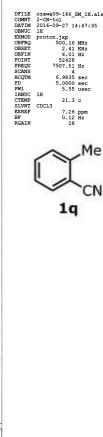






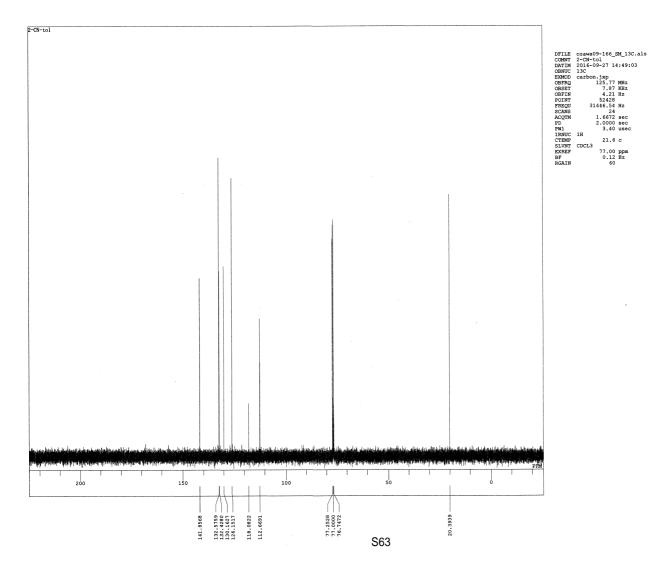


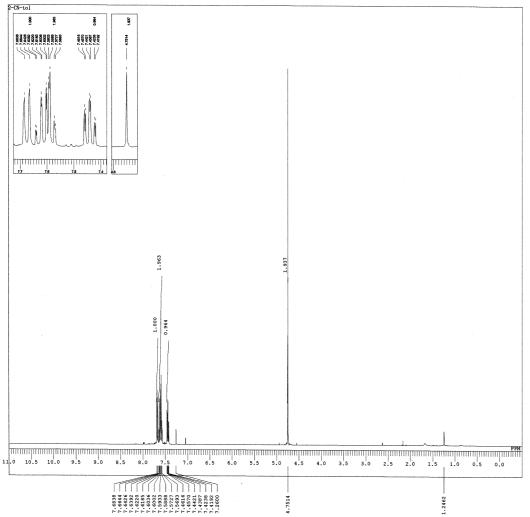


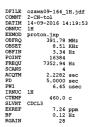


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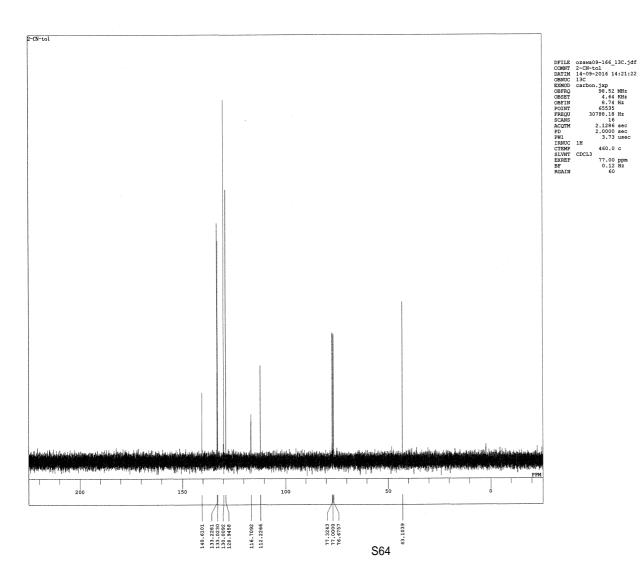
ozawa09-166_SM_1H.als 2-CN-tol 2016-09-27 14:47:35 1H

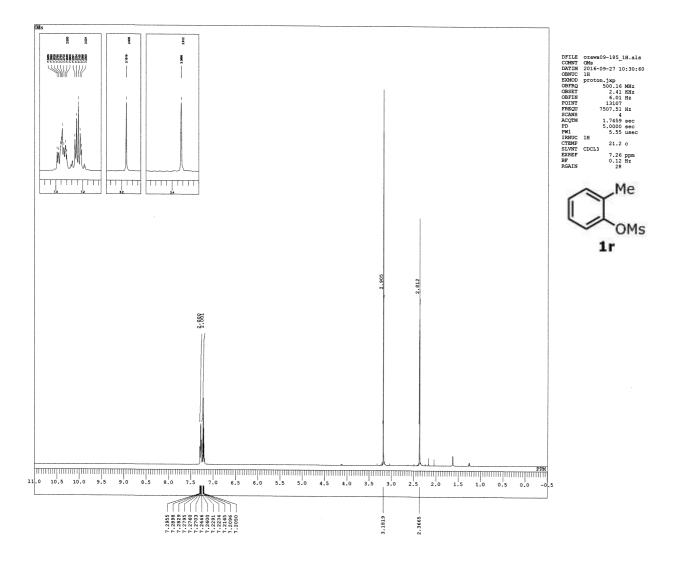


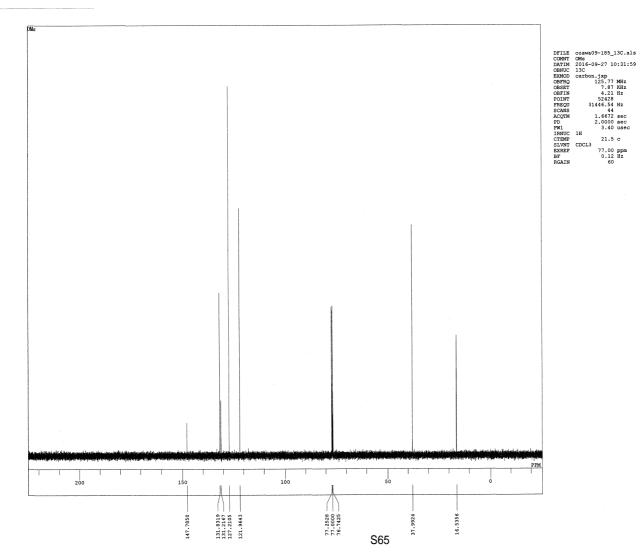


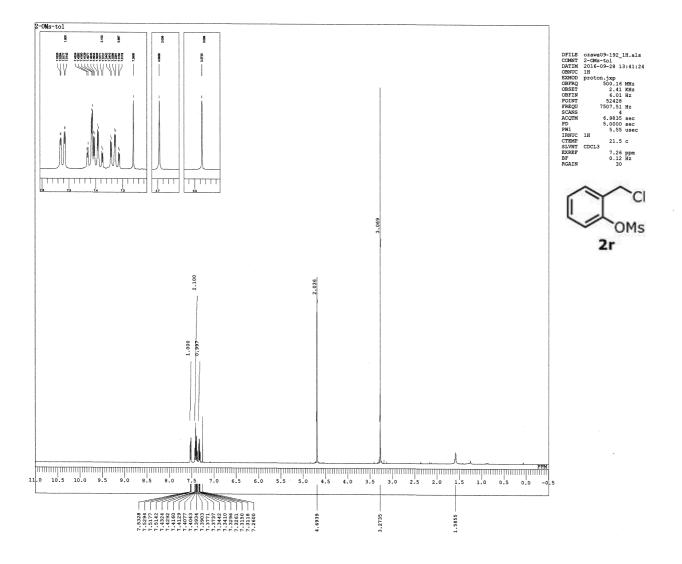


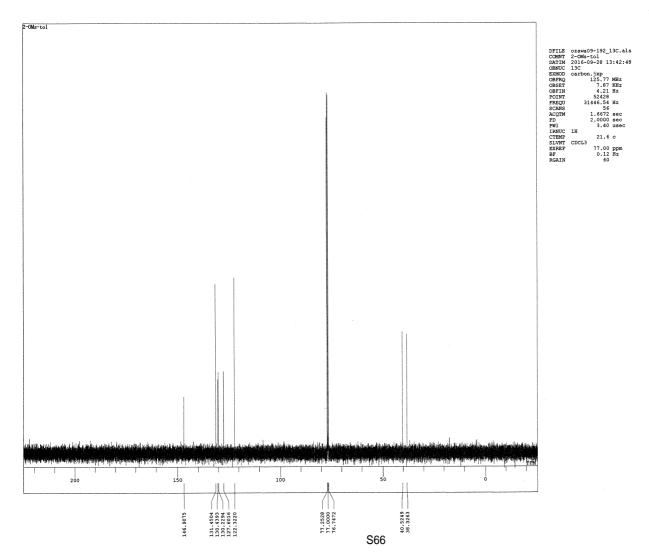


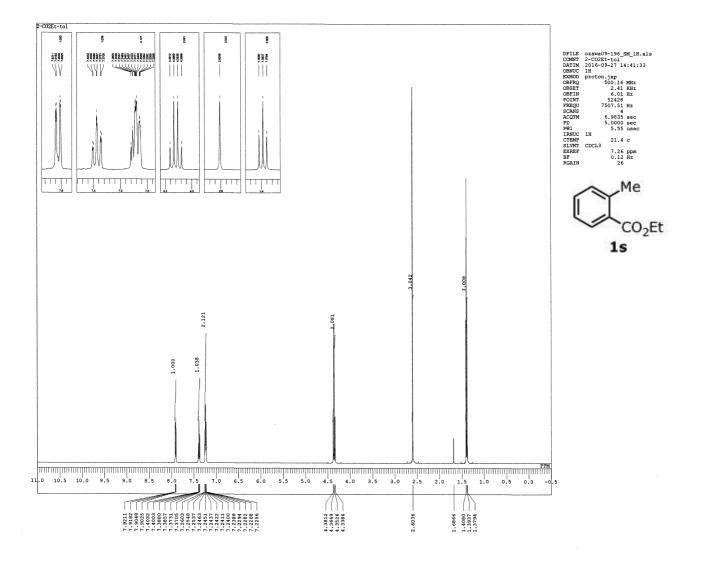


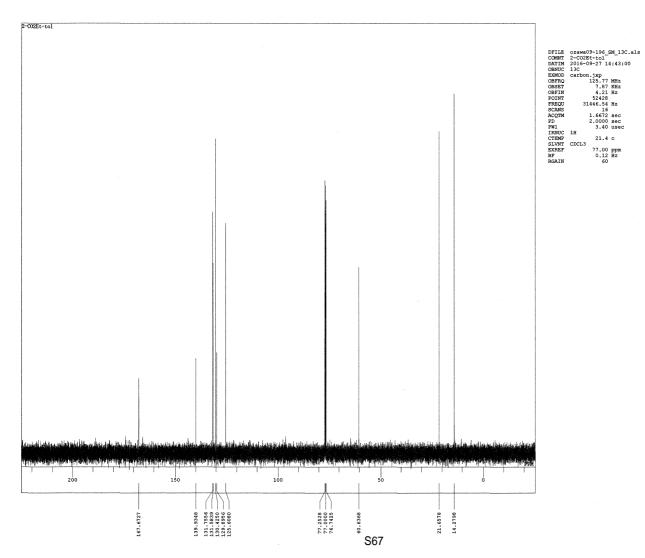


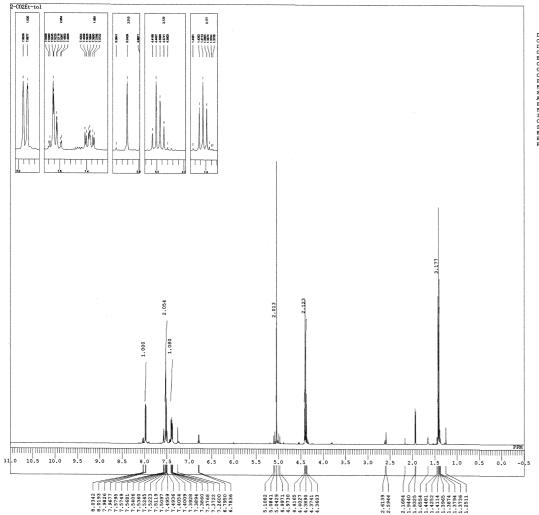




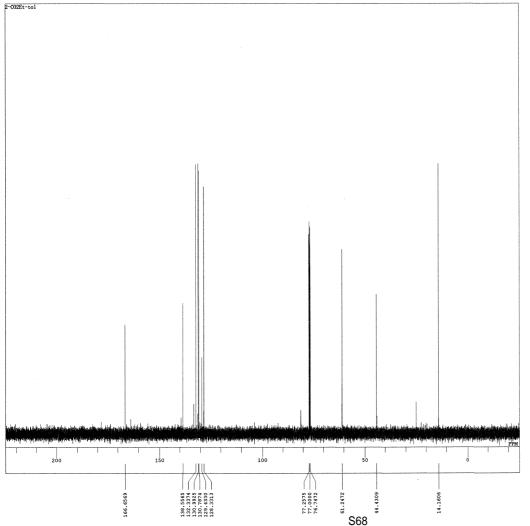


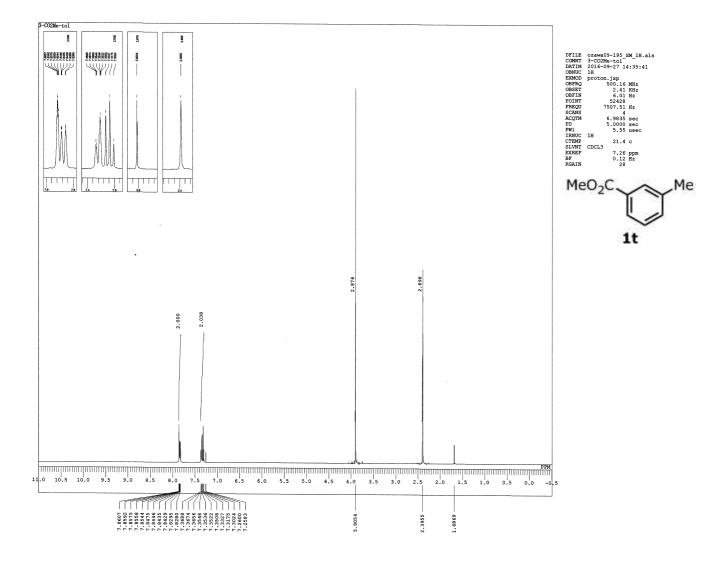


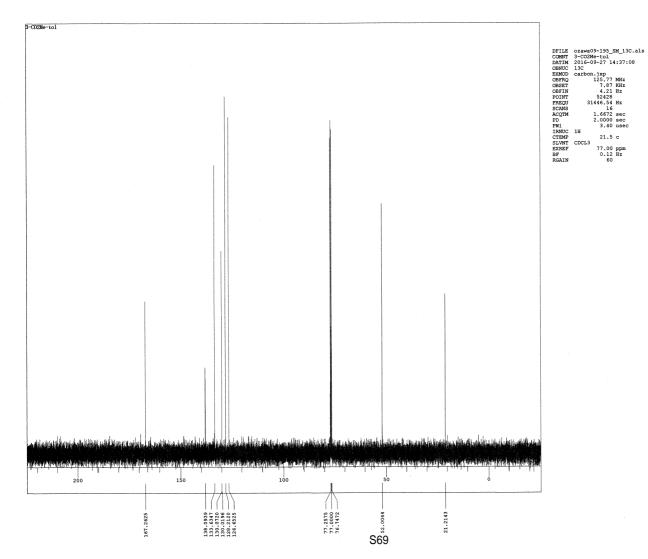


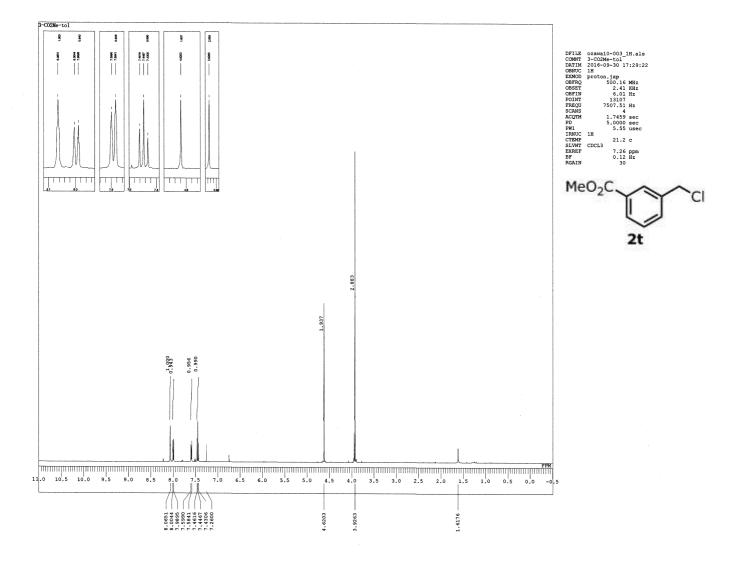


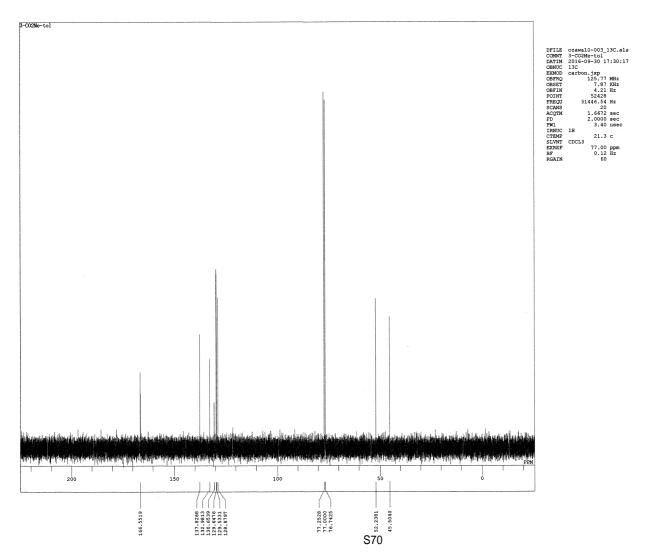


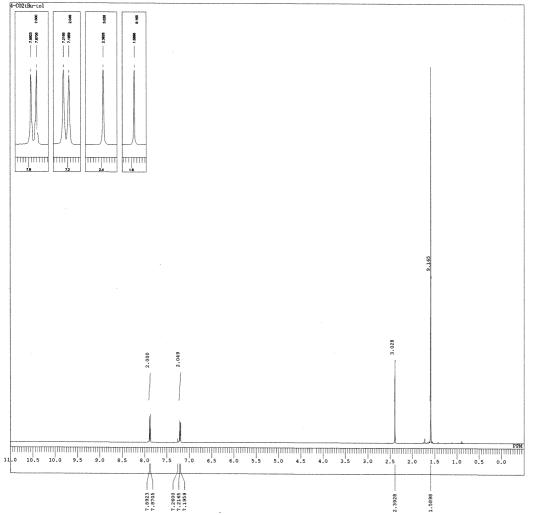




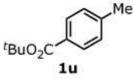


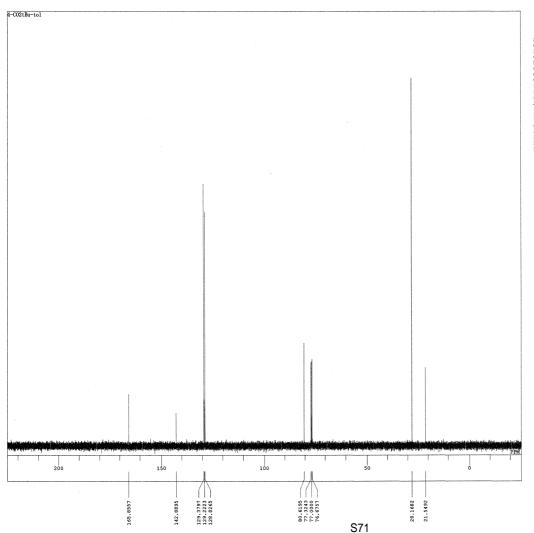




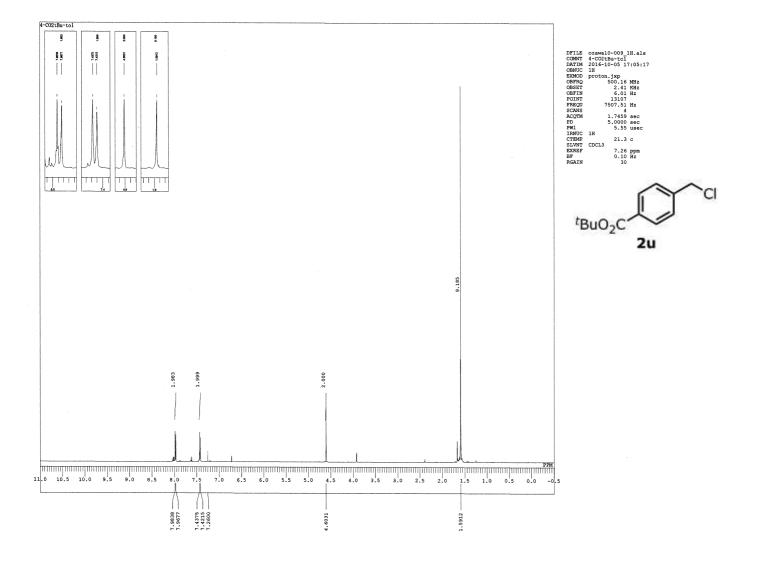


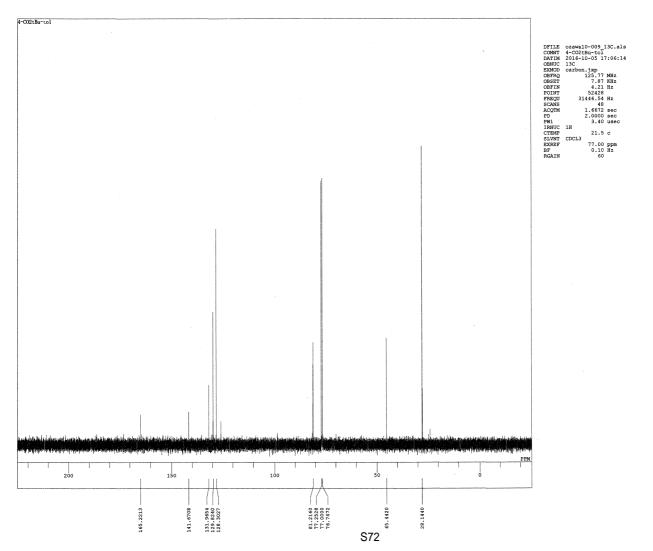


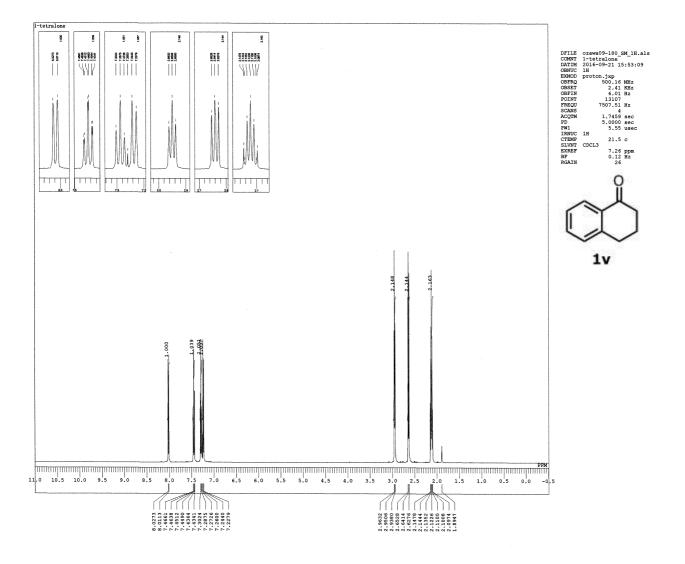


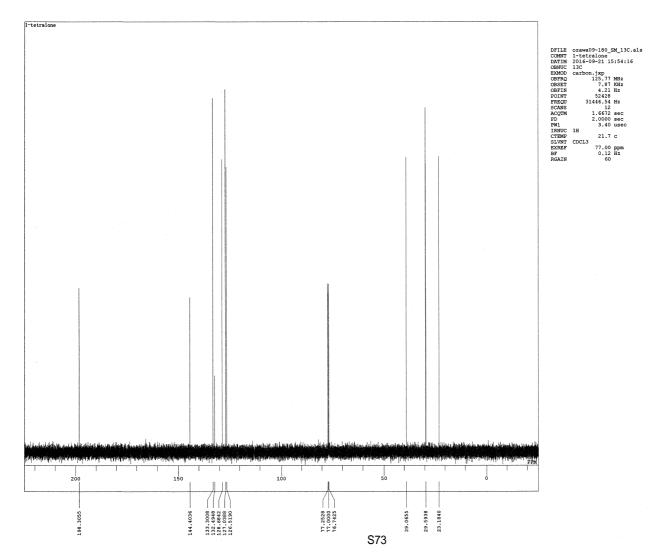


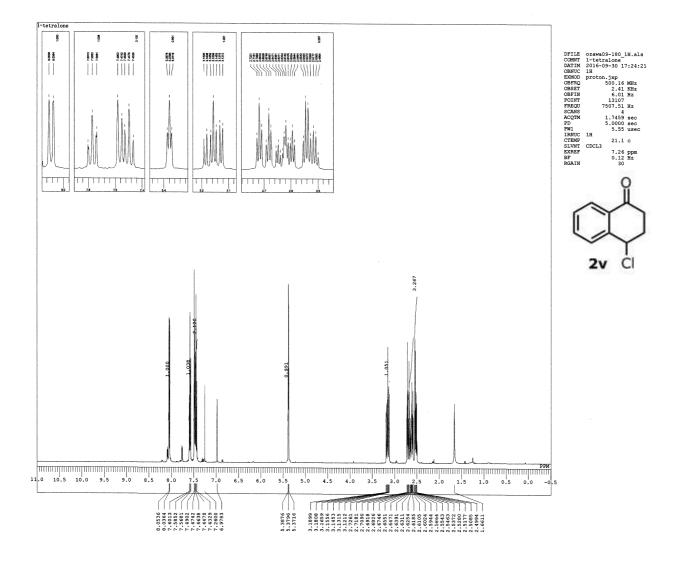
| DFILE | OZAWA10-002_13C-1-1.jdf | COMPT | 4-CQZ+Bu-t-01 | 4-CQZ+Bu-t-01 | 4-CQZ+Bu-t-01 | 4-CQZ+Bu-t-01 | 4-CQZ+Bu-t-01 | 4-CQZ+Bu-t-02 | 4-

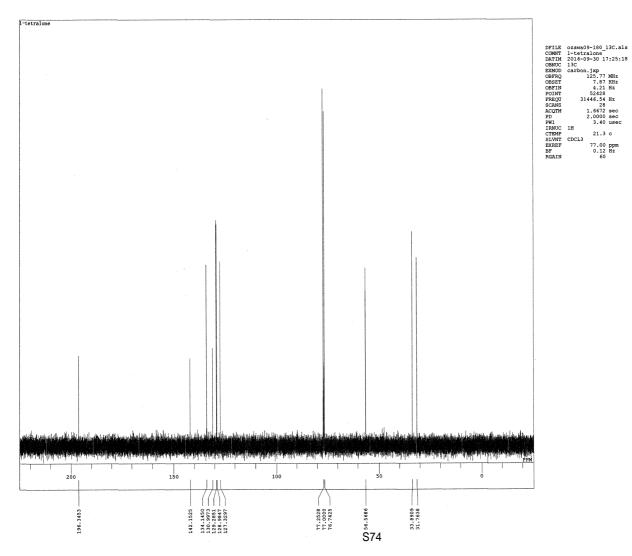


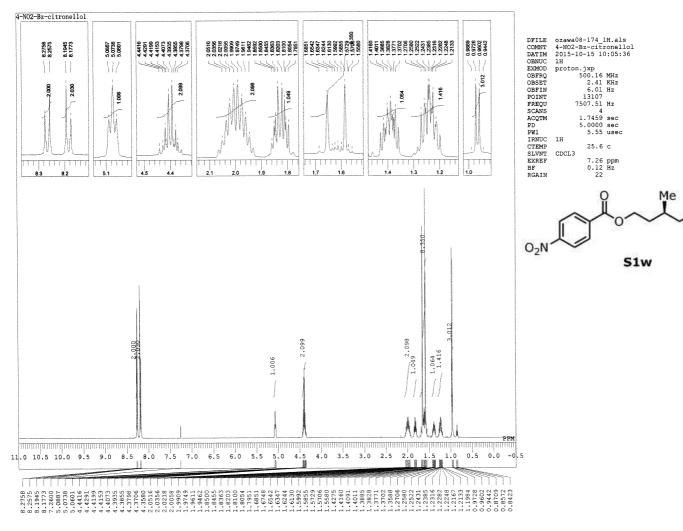




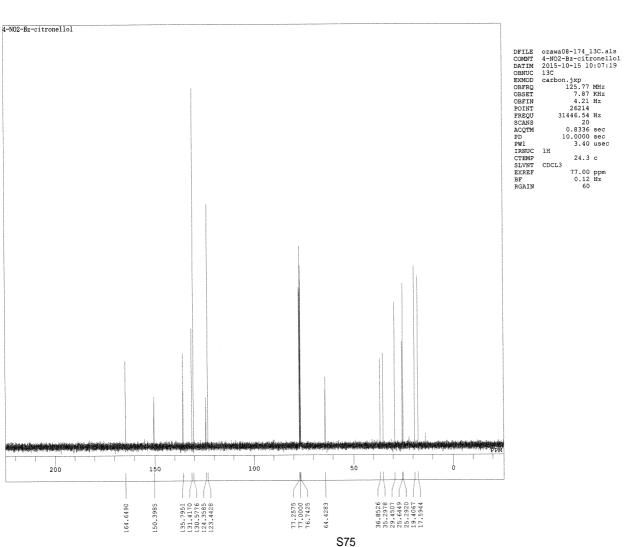


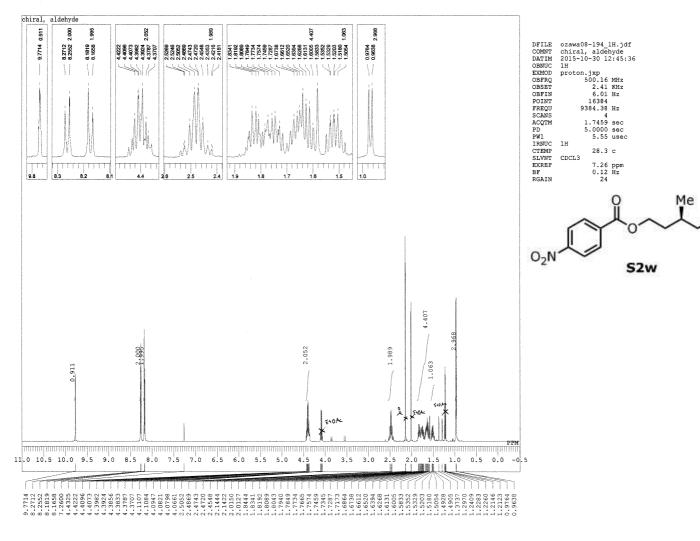


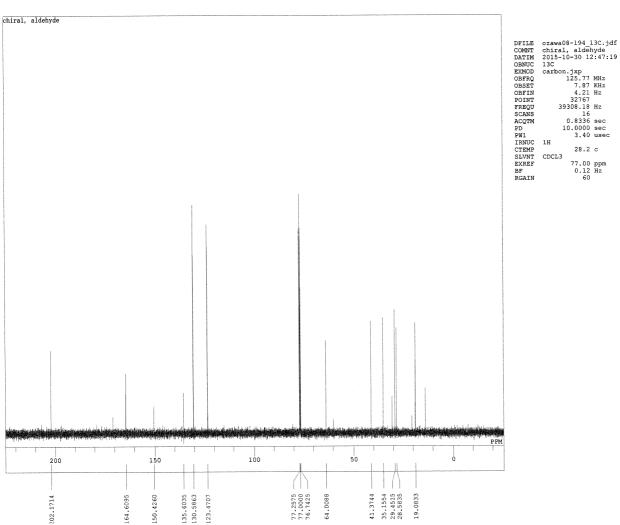


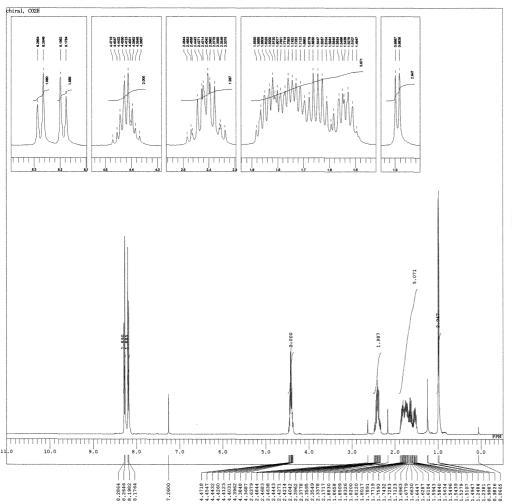


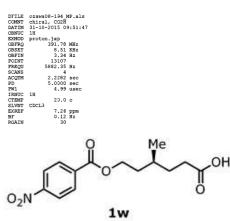
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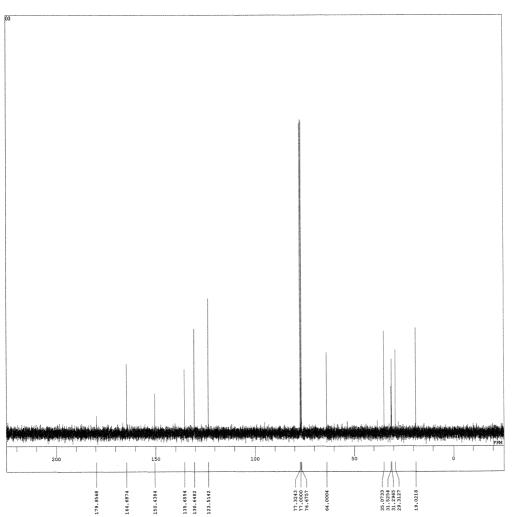












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DNTHM 33-10-2015 10:20:05
EMMOD 08FR0 98.52 MHz
OBSTH 4.64 KHz
OBSTH 6.74 Hz
FREGUI 30788.18 Hz
SCANS
ACCTM 1.0643 sec
PD 10.0000 sec
PD 10.0000 sec
FMUU 14 20.3 c
SUMNT CEMBE
EXERS 77.00 ppm
EXERF 77.00 ppm
RGAIN 60

