

Supplementary belongs to:

Biochemical characterization of the functional roles of residues in the active site of the β -galactosidase from *Bacillus circulans* ATCC 31382

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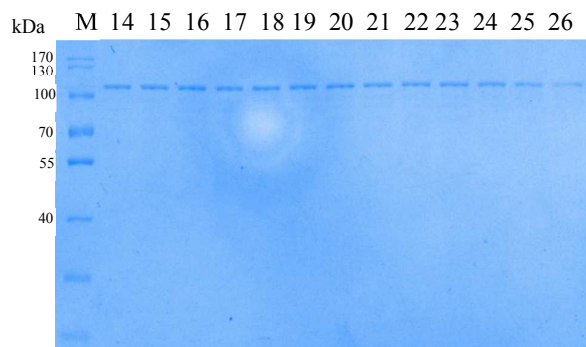
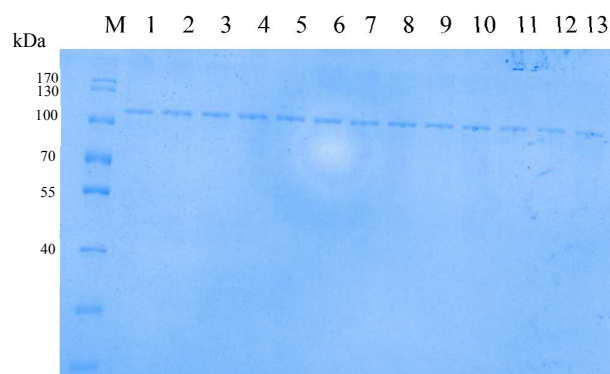
Table S1. The primer pairs used for site-directed mutagenesis of BgaD-D.

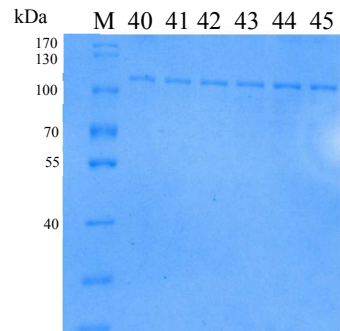
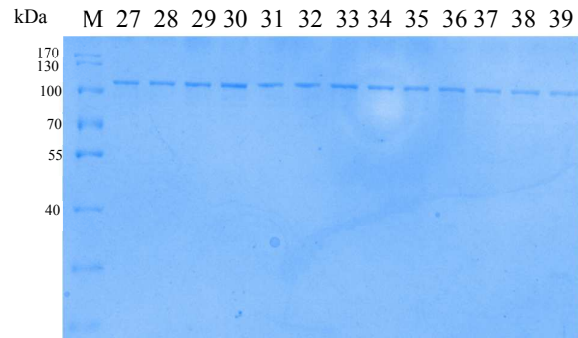
Primer name	DNA sequence (5' to 3')
Arg185-F	CACCCAGCCGAGCAGC <u>NN</u> KTGGTATTCGGGGAGCG
Arg185-R	CGCTCCCGAATACC <u>AMNN</u> GCTGCTCGGCTGGGTG
Asp481-F	GATCGGCGAG <u>NN</u> KAAAACCC
Asp481-R	GGGTTTT <u>MNN</u> CTCGCCGATC
Asp481Lys-F	GATCGGCGAGA <u>A</u> GAAAACCC
Asp481Lys-R	GGGTTTT <u>CTT</u> CTCGCCGATC
Asp481Arg-F	GATCGGCGAG <u>C</u> GGAAAACCC
Asp481Arg-R	GGGTTTT <u>CCG</u> CTCGCCGATC
Asp481Asn-F	GATCGGCGAGA <u>A</u> CAAAAACCC
Asp481Asn-R	GGGTTTT <u>GTT</u> CTCGCCGATC
Asp481Gln-F	GATCGGCGAG <u>C</u> AGAAAACCC
Asp481Gln-R	GGGTTTT <u>CTG</u> CTCGCCGATC
Asp481Leu-F	GATCGGCGAG <u>C</u> TGAAAACCC
Asp481Leu-R	GGGTTTT <u>CAG</u> CTCGCCGATC
Asp481Trp-F	GATCGGCGAG <u>T</u> GGAAAACCC
Asp481Trp-R	GGGTTTT <u>CCA</u> CTCGCCGATC
Asp481Gly-F	GATCGGCGAG <u>G</u> GGAAAACCC
Asp481Gly-R	GGGTTTT <u>CCC</u> CTCGCCGATC
Lys487-F	CGCGGAGAC <u>NN</u> KGTAAATGTTACAC
Lys487-R	GTGTAACATTTAC <u>MNN</u> GCTCTCCGCG
Tyr511-F	GGACTGAAC <u>NN</u> KAGCGAGAACAACACTATGATGGC
Tyr511-R	GCCATCATAGTTGTTCTCGCT <u>MNN</u> GTTTCAGTCC
Tyr511Phe-F	GGACTGAAC <u>T</u> ATAGCGAGAACAACACTATGATGGC
Tyr511Phe-R	GCCATCATAGTTGTTCTCGCT <u>T</u> ATAGTTTCAGTCC
Tyr511Trp-F	GGACTGAAC <u>T</u> GGAGCGAGAACAACACTATGATGGC
Tyr511Trp-R	GCCATCATAGTTGTTCTCGCT <u>CC</u> AGTTTCAGTCC
Trp570-F	GTCGGC <u>NN</u> KGGACGAACTGCAGAAG
Trp570-R	CTTCTGCAGTTCGTCC <u>MNN</u> GCCGAC
Trp570Tyr-F	GTCGGCT <u>AC</u> GGACGAACTGC
Trp570Tyr-R	GCAGTTC <u>CGT</u> CCGTAGCCGAC
Trp570Phe-F	GTCGGCT <u>TT</u> GGACGAACTGC
Trp570Phe-R	GCAGTTC <u>GT</u> CC <u>AA</u> AGCCGAC
Trp570Ala-F	GTCGGC <u>G</u> CTGGACGAACTGC
Trp570Ala-R	GCAGTTC <u>GT</u> CC <u>AG</u> CGCCGAC
Trp570Val-F	GTCGGC <u>G</u> TGGACGAACTGC
Trp570Val-R	GCAGTTC <u>GT</u> CC <u>AA</u> CGCCGAC
Trp570Leu-F	GTCGGC <u>CT</u> TGGACGAACTGC
Trp570Leu-R	GCAGTTC <u>GT</u> CC <u>AA</u> GCCGAC
Trp570Cys-F	GTCGGCT <u>G</u> CGGACGAACTGC
Trp570Cys-R	GCAGTTC <u>GT</u> CC <u>G</u> AGCCGAC
Trp593-F	ACCTGAAGCATATTGCAGGGCAATTTATC <u>NN</u> KACCGGCTTT
Trp593-R	GATTATATTGG CCAATATAATCAAAGCCGGT <u>MNN</u> GATAAATTGCCCTGCAA TATGCTTCAGGT
Glu601-F	CCGGCTTTGATTATATTGGC <u>NN</u> KCCGACGCCATATTATAAT TCC

Glu601-R	GGAATTATAATATGGCGTCGGM <u>NN</u> NGCCAATATAATCAAAG CCGG
Glu601Asp-F	CCGGCTTTGATTATATTGGCG <u>AC</u> CCGACGCCATATTATAAT TCC
Glu601Asp-R	GGAATTATAATATGGCGTCGG <u>GTC</u> GCCAATATAATCAAAG CCGG
Phe616-F	GCAAAAAGCTCCTAT <u>NN</u> KGGTGCTGTGGATACGG
Phe616-R	CCGTATCCACAGCAC <u>CM</u> NNATAGGAGCTTTTTGC
Phe616Trp-F	GCAAAAAGCTCCTAT <u>TGG</u> GGTGCTGTGGATACGG
Phe616Trp-R	CCGTATCCACAGCAC <u>CCA</u> ATAGGAGCTTTTTGC
Phe616His-F	GCTCCTAT <u>CAT</u> GGTGCTGTGGATAC
Phe616His-R	GTATCCACAGCAC <u>CA</u> TGATAGGAGC
Phe616Ser-F	GCTCCTAT <u>TC</u> AGGTGCTGTGGATAC
Phe616Ser-R	GTATCCACAGCAC <u>CTGA</u> ATAGGAGC
Phe616Thr-F	GCTCCTAT <u>AC</u> AGGTGCTGTGGATAC
Phe616Thr -R	GTATCCACAGCAC <u>CTGT</u> ATAGGAGC
Phe616Asn-F	GCTCCTAT <u>AA</u> TGGTGCTGTGGATAC
Phe616Asn-R	GTATCCACAGCAC <u>CA</u> TATAGGAGC
Phe616Cys-F	GCTCCTAT <u>TGC</u> GGTGCTGTGGATAC
Phe616Cys-R	GTATCCACAGCAC <u>CGCA</u> ATAGGAGC
Phe616Pro-F	GCTCCTAT <u>CC</u> AGGTGCTGTGGATAC
Phe616Pro-R	GTATCCACAGCAC <u>CTGG</u> ATAGGAGC
Phe616Ala-F	GCTCCTAT <u>GCT</u> GGTGCTGTGGATAC
Phe616Ala-R	GTATCCACAGCAC <u>AGCA</u> TAGGAGC
Phe616Ile-F	GCTCCTAT <u>ATC</u> GGTGCTGTGGATAC
Phe616Ile-R	GTATCCACAGCAC <u>CGAT</u> ATAGGAGC
Phe616Met-F	GCTCCTAT <u>ATG</u> GGTGCTGTGGATAC
Phe616Met-R	GTATCCACAGCAC <u>CCAT</u> ATAGGAGC
Phe616Tyr-F	GCTCCTAT <u>TAC</u> GGTGCTGTGGATAC
Phe616Tyr-R	GTATCCACAGCAC <u>CGTA</u> ATAGGAGC

The “N” base in the primers stands for a “A or T or C or G” base, and “K” stands for a “G or T” base. “NNK” codons are commonly used in screens for codon substitutions to reduce the presence of some codon-rich amino acids, thereby reducing their over representation in a particular library. Additionally, using NNK codons removes 2 out of 3 possible stop codons, which also limits the number of sequences in a library that produce unwanted truncated gene products¹.

Figure S1. SDS-PAGE gel analysis of the wild-type BgaD-D and the mutant enzymes (loaded with 5 μ L of 0.1 mg/mL protein solutions). M, Marker proteins; 1, WT; 2, Trp570Gly; 3, Trp570Thr; 4, Trp570Arg; 5, Trp570Glu; 6, Trp570Tyr; 7, Trp570Phe; 8, Trp570Ala; 9, Trp570Val; 10, Trp570Cys; 11, Trp570Leu; 12, Trp593Tyr; 13, Trp593Phe; 14, Phe616Val; 15, Phe616Glu; 16, Phe616Gly; 17, Phe616Lys; 18, Phe616Gln; 19, Phe616Arg; 20, Phe616Asp; 21, Phe616Leu; 22, Phe616Trp; 23, Phe616His; 24, Phe616Ser; 25, Phe616Thr; 26, Phe616Asn; 27, Phe616Cys; 28, Phe616Pro; 29, Phe616Ala; 30, Phe616Ile; 31, Phe616Met; 32, Phe616Tyr; 33, Asp481Glu; 34, Asp481His; 35, Asp481Ser; 36, Asp481Asn; 37, Asp481Gln; 38, Lys487Met; 39, Lys487Phe; 40, Lys487Leu; 41, Lys487Gln; 42, Lys487Ser; 43, Lys487Gly; 44, Lys487Asn; 45, Lys487Cys.





Reference:

- 1 Tang, L., Gao, H., Zhu, X., Wang, X., Zhou, M., and Jiang, R. (2012) Construction of “small-intelligent” focused mutagenesis libraries using well-designed combinatorial degenerate primers. *Biotechniques*, 52 (3), 149–158.