

# Four resonance structures elucidate double-bond isomerisation of a biological chromophore

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Photoinduced double-bond isomerisation of the chromophore of photoactive yellow protein (PYP) is highly sensitive to chromophore-protein interactions. On the basis of high-level *ab initio* calculations, we scrutinise the effect of the chromophore-protein hydrogen bonds on the photophysical and photochemical properties of the chromophore. We identify four resonance structures – two closed-shell and two biradicaloid – that elucidate the electronic structure of the ground and first excited states involved in the isomerisation process. Changing the relative energies of the resonance structures by hydrogen-bonding interactions tunes *all* photochemical properties of the chromophore in an interdependent manner. Our study sheds new light on the role of the chromophore electronic structure in tuning in photosensors and fluorescent proteins.

## 1. INTRODUCTION

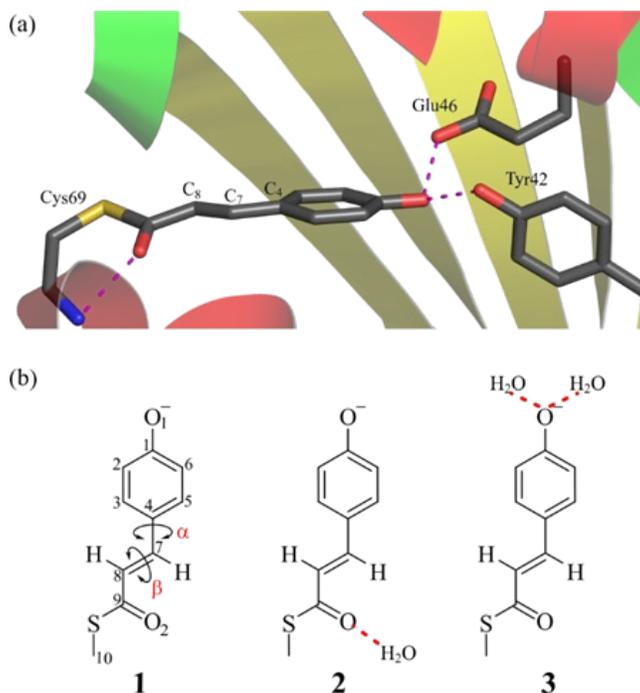
Photoactive yellow protein (PYP)<sup>1</sup> is a remarkable model system for studying double-bond isomerisation in photoreceptor proteins, as this photoreceptor has amply been characterised using a wide variety of advanced methods such as time-resolved X-ray crystallography,<sup>2-5</sup> neutron crystallography,<sup>6</sup> NMR,<sup>7</sup> and ultrafast spectroscopy.<sup>8-12</sup> These studies provided important insights into photoinduced double-bond isomerisation of the PYP chromophore that triggers the PYP photoresponse. The chromophore is derived from the anionic phenolate form of the *p*-coumaric thioester (pCTM<sup>-</sup>), which is in the *E(trans)*-configuration and stabilised by hydrogen bonds (H-bonds) with the protein (Fig. 1a).<sup>13</sup> The excited-state dynamics of the pCTM<sup>-</sup> anion in the protein is different from that in solution and the gas phase.<sup>14-15</sup> Despite numerous studies, the exact mechanism of the protein control of the chromophore's isomerisation remains elusive. In particular, it is not clear how to describe the impact of the protein on the chromophore at the electronic structure level.

The moderate molecular size and high sensitivity to intermolecular interactions render the pCTM<sup>-</sup> chromophore (model **1** in Fig. 1b) particularly suitable for accurate quantum-chemistry calculations addressing the chromophore's tuning by intermolecular interactions.<sup>16-22</sup> Similarly to other ionic chromophores of photoresponsive proteins, pCTM<sup>-</sup> undergoes a pronounced charge-transfer, from the phenolic moiety to the carbonyl fragment, upon photoexcitation.<sup>23</sup> Twisting the chromophore's central double bond (C<sub>7</sub>=C<sub>8</sub>) or single bond (C<sub>4</sub>-C<sub>7</sub>) results in localisation of the negative charge either on the phenolic or carbonyl moieties, respectively, in the excited (S<sub>1</sub>) state.<sup>17, 20, 24-25</sup> The localisation of the negative charge in the ground state (S<sub>0</sub>) is reversed to that in the S<sub>1</sub> state at the twisted geometries. Polar interactions of pCTM<sup>-</sup> with the protein or solvent profoundly influence the ratio of the S<sub>1</sub>-state trajectories returning to the S<sub>0</sub> state via the single-

and double-bond twists.<sup>16, 18, 24, 26</sup> Experimentally it has been established that PYP photoactivation is triggered by the double bond isomerisation yielding a *Z(cis)* isomer with a quantum yield of 0.35,<sup>27</sup> which proceeds via a one-bond flip (OBF) mechanism destabilizing the carbonyl H-bond.<sup>2-3, 9</sup> Activation of the single-bond rotation has been reported for a mutated variant with weakened phenolic H-bonds.<sup>3</sup> The mutation leads to a slight red shift of the absorption maximum,<sup>28</sup> reduced charge-transfer character of the  $S_0$ – $S_1$  transition,<sup>23</sup> and a so-called hula-twist (HT) isomerisation combining rotations around the single and double bonds.<sup>3</sup> In contrast to the OBF isomerisation, the HT isomerisation preserves the carbonyl H-bond in the resulting *cis* photoproduct, which impairs the signal transduction.<sup>3</sup>

Here, by performing quantum-chemistry calculations for models **1-3** (Fig. 1b) we study the impact of H-bonds on the  $S_0$  and  $S_1$  energy at the geometries of pCTM<sup>-</sup> constituting the single and double bond isomerisation pathways, including the geometries of the  $S_0/S_1$  conical intersections (CoIns). The accurate description of the models is attained by using a high-level *ab initio* method, the extended multi-state multi-configurational quasi-degenerate perturbation theory of second order (XMCQDPT2),<sup>29</sup> which accounts for both static and dynamical electron correlation. In this context, the present work critically extends the previous studies performed using the complete-active-space self-consistent field (CASSCF) method<sup>21, 24, 30</sup> and the single-reference second-order approximate coupled-cluster (CC2) method.<sup>17, 20, 22, 25</sup> To explain the trends revealed from contrasting the three models, we deduce four resonance structures, two closed-shell (CS) and two biradicaloid (BR), that describe charge delocalisation and charge transfer upon the  $S_0$ – $S_1$  excitation and the bond twists. Stabilisation and destabilisation of these structures by H-bonds account for interdependent changes of all photochemical properties of the pCTM<sup>-</sup> chromophore. We compare our findings to previously published results on PYP, fluorescent proteins and rhodopsin

photoreceptors noting multiple parallels among the properties of ionic chromophore in these proteins.



**Figure 1.** (a) Chromophore binding pocket (active site) of PYP featuring the anionic *E(trans)*-*p*-coumaroyl chromophore bound to PYP via a thioester linkage of the Cys69 residue and hydrogen-bonded to the backbone of Cys69 and side chains of Tyr42 and Glu46. (b) Three models considered here: the anionic *p*-coumaric thiomethyl (pCTM<sup>-</sup>) chromophore (1), pCTM<sup>-</sup> hydrogen-bonded with one water molecule at the carbonyl oxygen (2), mimicking the H-bond with the backbone of Cys69, and pCTM<sup>-</sup> hydrogen-bonded with two water molecules at the phenolic oxygen (3), mimicking the H-bonds with the Tyr42/Glu46 residues.

## 2. Computational details

The energies and geometries for models 1-3 in the  $S_0$  and  $S_1$  states were obtained from the XMCQDPT2 calculations. As a zero-order wave function employed in the XMCQDPT2 calculations, we used a state-averaged CASSCF wave function, SA2-CASSCF(12,11). The active space comprised 12 electrons over 11  $\pi$  molecular orbitals (MOs), as previously used by Boggio-Pasqua and Groenhof.<sup>21</sup> The state-averaging was done for the two lowest electronic states,  $S_0$  and

$S_1$ , with equal weights. The subsequent XMCQDPT2 calculations using the SA2-CASSCF(12,11) zero-order wave function further accounted for dynamical electron correlation, up to the second order of perturbation theory, and interaction of the  $S_0$  and  $S_1$  states, via their mixing in a  $2 \times 2$  Hamiltonian. The resolution of the identity (RI) approximation<sup>31-33</sup> implemented in Firefly version 8.2 afforded a substantial speed-up of the XMCQDPT2 computations and a decrease of memory requirements, significantly facilitating numerical evaluation of the energy gradients utilizing supercomputer resources. An intruder state avoidance (ISA) energy shift<sup>34</sup> of 0.02 Hartree was applied throughout. In all our calculations, the correlation consistent double-zeta basis set of Dunning (cc-pVDZ) was used,<sup>35-36</sup> resulting in 231 to 277 MOs depending on the model. All MOs were included in the MCSCF procedure, whereas the chemical-core MOs (1s of carbon and oxygen and 1s, 2s and 2p of sulphur) were not correlated in the XMCQDPT2 calculations.

The search of stationary points on the  $S_1$  and  $S_0$  PESs and points of  $S_1/S_0$  minimum energy conical intersections (CoIn) was performed using numerical XMCQDPT2 energy gradients computed with finite differencing of second order. In the search, internal coordinates were used without imposing constraints, i.e, the full geometry optimisation was performed. The root mean square (RMS) gradient for optimisation convergence was  $10^{-3}$  Hartree/Bohr. In the CoIn geometry optimisation, a penalty function method was used, with a penalty function as proposed by Martinez and co-workers.<sup>37</sup> The threshold of the energy gap in the CoIn optimisation was  $10^{-3}$  Hartree. Because of exceedingly high computational demands for numerical evaluation of the Hessians at the XMCQDPT2 level, vibrational analysis at the stationary points was not performed. The type of the stationary points was inferred based on previously published computations.<sup>17, 20</sup> For the saddle points, the existence of a negative curvature was confirmed by calculating relaxed energy scans along the coordinate for which the negative curvature was expected. The planar structures

of models **1** and **2** ( $E$ - $S_1$ -Sad) and the single-bond twisted structure of model **3** ( $\alpha$ - $S_1$ -Sad) were demonstrated to correspond to saddle points by computing relaxed-energy scans along the single-bond twisting coordinate ( $\alpha$ -torsion). The double-bond twisted barrier ( $\beta$ - $S_1$ ) in all the models was demonstrated to correspond to a saddle point by computing relaxed-energy scans along the double-bond twisting coordinate ( $\beta$ -torsion).

For all the stationary points, the basis set superposition error (BSSE) corrections were computed for the  $S_0$  and  $S_1$  energies of models **2** and **3** using the protocol described in Ref <sup>38</sup>. Overall, the BSSE corrections for the relative/excitation energies are small, not exceeding 0.1 eV, with the largest correction (-0.085 eV) found for the  $\alpha$ - $S_1$ -Sad saddle point of model **3**. Hereby, the corrections for the twisted structures are larger than that for the planar structures, which obviously relates to the different charge distribution for the planar and twisted configurations. Importantly, accounting for the BSSE corrections does not lead to changes in the relative order of the stationary points, either within a particular model or the different models. In the following, we therefore neglect the BSSE corrections, presenting and discussing the plain XMCQDPT2 results.

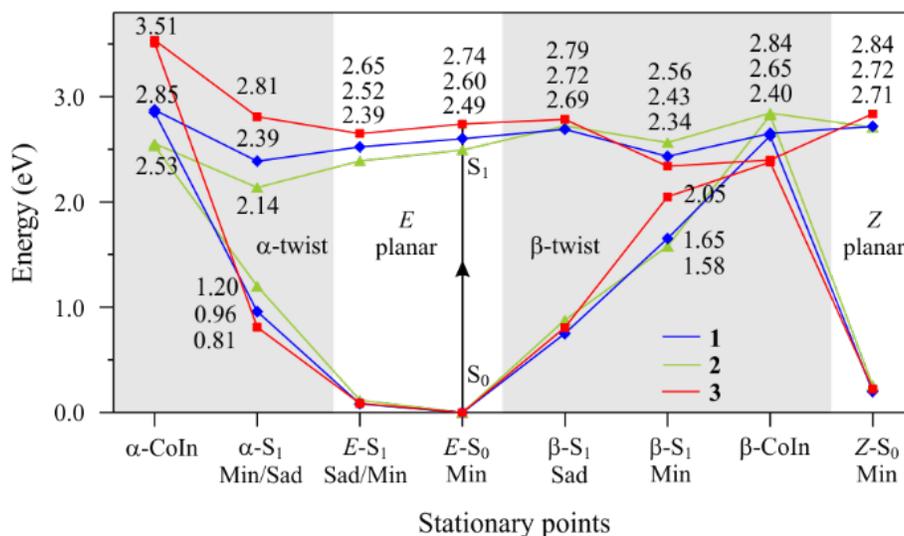
Oscillator strengths, Mulliken atomic charges and free valences on atoms (number of unpaired electrons)<sup>39</sup> were computed using the  $S_0$  and  $S_1$  zero-order QDPT electron densities. For analysis of the  $S_0$  and  $S_1$  PESs in the vicinity of CoIn, an approach suggested by Olivucci and co-workers was used.<sup>40-41</sup> A series of points circumscribing a CoIn point (loops) lying in the branching plane were computed. The branching plane is defined by the gradient difference ( $\mathbf{x}_1$ ) and derivative coupling ( $\mathbf{x}_2$ ) vectors.<sup>42</sup> We computed the  $\mathbf{x}_1$  and  $\mathbf{x}_2$  vectors with the SA2-CASSCF(12,11) method using Gaussian09,<sup>43</sup> with solving the coupled-perturbed equations in the MCSCF cycle being skipped (“nocpmcscf” approximation).<sup>44</sup> The error introduced by the latter decreases with the decrease of the  $S_0$ – $S_1$  energy gap; therefore it might be important at the Frank-Condon region and

negligible at CoIn. Except computations of the branching vectors, all calculations were performed using version 8.2 of the Firefly quantum chemistry package,<sup>45</sup> which is partially based on the Gamess-US source code.<sup>46</sup> The computations utilised resources of the high performance computing cluster JURECA.<sup>47</sup>

### 3. RESULTS AND DISCUSSION

#### 3.1. Characterisation of single- and double-bond twists

Figure 2 depicts cross-sections through the  $S_0$  and  $S_1$  potential energy surfaces (PESs) computed for models 1-3 along the single- and double-bond twisting pathways starting from the  $E(trans)$   $S_0$  minimum ( $E-S_0$ -Min). For each model, the cross-section is composed of several stationary points on the  $S_0$  and  $S_1$  PESs and points of  $S_1/S_0$  minimum-energy conical intersections (CoIns). Specifically, there are the  $E(trans)$  and  $Z(cis)$  planar minima in the ground  $S_0$  state, two minima and two saddle points of planar and twisted geometries in the  $S_1$  state, and two twisted CoIn points. All the points were obtained from a full geometry optimisation using the XMCQDPT2 method. In the following, we refer to the rotation around the  $C_4-C_7$  bond as  $\alpha$ -twist and around the  $C_7-C_8$  bond as  $\beta$ -twist (Fig. 1b) according to the previously adopted notation.<sup>17, 20, 25</sup> The geometries of the models at the stationary points are depicted in Fig. S1 in the Supporting Information (SI). The energies given in Fig. 2 are not corrected for the basis set superposition error (BSSE), which according to our estimation introduces only marginal changes in the cross-section energies (Table S3 in SI).



**Figure 2.** Cross-sections of  $S_0$  and  $S_1$  PESs for models 1 (blue), 2 (green) and 3 (red) for the photoinduced  $\alpha$  and  $\beta$  twisting. The following notations are used to indicate the geometry and type of the stationary points. Planar ground-state minima featuring  $E(trans)$  and  $Z(cis)$  configurations with respect to the central  $-C_7=C_8-$  double bond,  $E-S_0$ -Min and  $Z-S_0$ -Min, respectively; planar excited-state saddle point and minimum,  $E-S_1$ -Sad and  $E-S_1$ -Min, respectively;  $\alpha$ -twisted excited-state minimum and saddle point,  $\alpha-S_1$ -Min and  $\alpha-S_1$ -Sad, respectively;  $\alpha$ -twisted CoIn,  $\alpha$ -CoIn;  $\beta$ -twisted saddle point,  $\beta-S_1$ -Sad;  $\beta$ -twisted minimum,  $\beta-S_1$ -Min;  $\beta$ -twisted CoIn,  $\beta$ -CoIn. All geometries are shown in Fig. S1 in SI. The total and relative energies of the points are listed in Tables S1-S2 in SI.

At first, we consider the results for model 1, which relate to the intrinsic properties of the pCTM<sup>-</sup> chromophore (blue cross-section in Fig. 2). At the *trans* configuration  $E-S_0$ -Min minimum, the  $S_0$ - $S_1$  vertical excitation energy (VEE) is 2.60 eV and the corresponding oscillator strength is close to unity. From the Franck-Condon (FC) point, the chromophore first undergoes a bond-length alteration (BLA) geometry relaxation toward the planar  $E-S_1$ -Sad saddle point followed by the  $\alpha$ -twist relaxation toward the 90°-twisted  $\alpha-S_1$ -Min minimum. Structurally close to the  $\alpha-S_1$  minimum but higher in energy, there is a  $S_1/S_0$   $\alpha$ -CoIn, mediating decay of the excited-state population back to the  $S_0$  state. The excited-state  $\alpha$ -twist was previously implicated in internal conversion.<sup>20, 22, 24</sup> Activation of the  $\beta$ -twist involves passing a small barrier,  $\beta-S_1$ -Sad, which is 0.12 eV above VEE, and is associated with the change of the BLA coordinate and  $\beta$  torsion.

Beyond the barrier, the chromophore relaxes by undergoing further  $\beta$ -twist toward the  $90^\circ$ -twisted  $\beta$ -S<sub>1</sub>-Min minimum. Structurally close to the  $\beta$ -S<sub>1</sub> minimum but higher in energy, there is a S<sub>1</sub>/S<sub>0</sub>  $\beta$ -CoIn, through which the chromophore decays back to the S<sub>0</sub> state either forward to the Z-S<sub>0</sub>-Min minimum or backward to the E-S<sub>0</sub> minimum. The ratio of the Z and E products depends on many factors, one of which is the topology of the S<sub>0</sub> PES in the vicinity of  $\beta$ -CoIn. Both the  $\alpha$ - and  $\beta$ -CoIns demonstrate a sloped topology, as each of them was found in the vicinity of a  $90^\circ$ -twisted S<sub>1</sub> energy minimum.

The results of models **2** and **3** (green and red cross-sections, respectively, in Fig. 2) demonstrate that the H-bonds lead to considerable energy changes and, moreover, to a change of the S<sub>1</sub> PES topology in the case of model **3**. Specifically, the carbonyl H-bond in **2** reduces VEE, stabilises the  $\alpha$ -pathway and destabilises the  $\beta$ -pathway. In contrast, the phenolic H-bonds in **3** increase VEE, destabilise the  $\alpha$ -twist and stabilises the  $\beta$ -twist. Notably, the  $\alpha$ -twist energies are more affected by the H-bonds than that of the  $\beta$ -twist. In **3**, destabilisation of the  $\alpha$ -twist results in appearance of the planar E-S<sub>1</sub>-Min minimum, with the  $\alpha$ -S<sub>1</sub>  $\alpha$ -twisted geometry becoming a saddle point (red cross-section in Fig. 2).

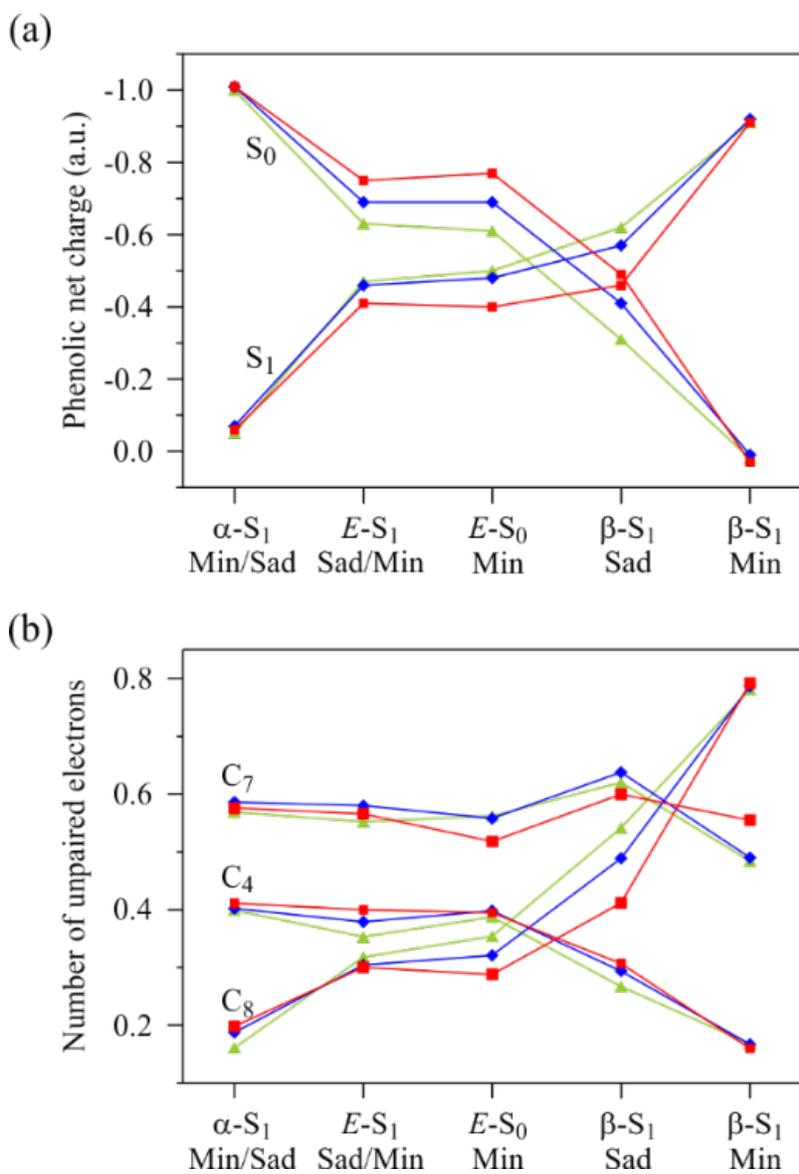
### 3.2. Four resonance structures rationalizing effect of H-bonds

In the previous studies, the effect of the H-bonds on the S<sub>0</sub> and S<sub>1</sub> energies has been explained by stabilisation and destabilisation of the negative charge either on the phenolic or carbonyl groups of pCTM<sup>-</sup>.<sup>17, 20, 24-25</sup> At the planar E-S<sub>0</sub> minimum, the charge de-localisation is explained by the resonance of two closed-shell (CS) anionic forms.<sup>17, 25</sup> At the same time, the origin of the reversed negative charge localisation at the  $\alpha$ - and  $\beta$ -twisted S<sub>1</sub> minima has not been linked to any resonance forms so far.<sup>17, 20, 25</sup> To elucidate this issue, we analyzed distribution of the negative charge and

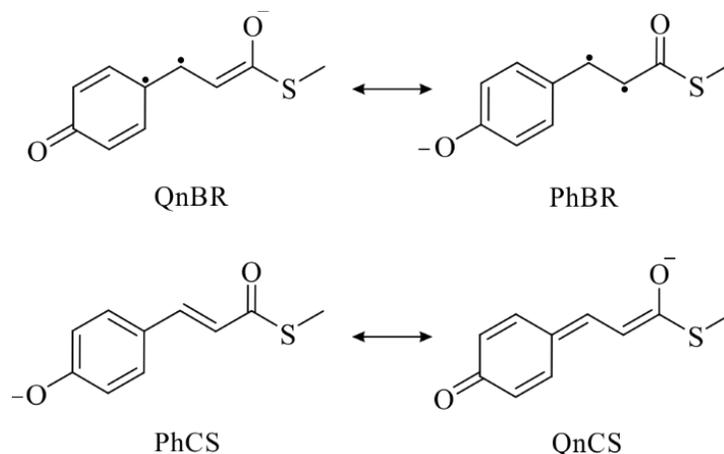
the number of unpaired electrons (NUE)<sup>39</sup> at the different geometries (stationary points) in the S<sub>0</sub> and S<sub>1</sub> state (Fig. 3, Tables S5-S7 in SI). The analysis indicates that the S<sub>0</sub>–S<sub>1</sub> electronic excitation results in a significant charge transfer and increase of NUE, which is consistent with a biradicaloid (BR) structure of the S<sub>1</sub> state (Table S8 in SI). The key result of our analysis is realizing that the changes of the phenolic charge and NUE at all the stationary points (Fig. 3) can be accounted for by changing contributions of *four* resonance structures – two CS structures, characterizing the S<sub>0</sub> state as mentioned above, and two BR structures, characterizing the S<sub>1</sub> state (Scheme 1). The resonance structures are denoted according to the presence of the phenolic (Ph) or quinonic (Qn)  $\pi$ -system. Noteworthy, QnBR relates to PhCS, and PhBR relates to QnCS via an electron excitation. Precisely, QnBR describes a single-electron charge-transfer excitation of PhCS, whereas PhBR describes a single-electron charge-transfer excitation of QnCS. To the best of our knowledge, the QnBR and PhBR structures have never been explicitly considered for pCTM<sup>-</sup> before.

It is worth noting that the four resonance structures are consistent with those introduced in the biradicaloid theory of Bonačić-Koutecký et al.<sup>48-49</sup> and theory of twisted intermolecular charge-transfer (TICT) of Rettig and co-workers.<sup>50-51</sup> The TICT theory treats the “hole-pair” (*hp*) configuration of a polar chromophore as electron-acceptor and electron-donor moieties connected via a central single bond. The electron transfer from the donor to the acceptor upon the HOMO-LUMO excitation (typically corresponding to the S<sub>0</sub>–S<sub>1</sub> transition) yields a “dot-dot” (*dd*) structure with a translocated molecular charge. By twisting the linking single bond, the HOMO and LUMO involved in the charge-transfer transition become localised on the mutually orthogonal molecular fragments, and thus do not interact. Therefore, the twisting decreases the energy of the *dd*

configuration and increases the energy of the  $hp$  configuration, which altogether significantly reduces the  $S_0$ – $S_1$  energy difference and even leads to  $S_0/S_1$  state crossing.<sup>48, 51</sup>



**Figure 3.** Distribution of (a) the net negative charge on the phenolic moiety in the  $S_0$  and  $S_1$  states and (b) the number of unpaired electrons (NUE) at the  $C_4$ ,  $C_7$  and  $C_8$  atoms in the  $S_1$  state in models 1-3 at the different stationary points along the  $\alpha$ - and  $\beta$ -twisting pathways. The colour code is the same as in Fig. 2. The atomic and net charges on the phenolic and carbonyl moieties, as well as NUE are given in Tables S5-S7 in SI.

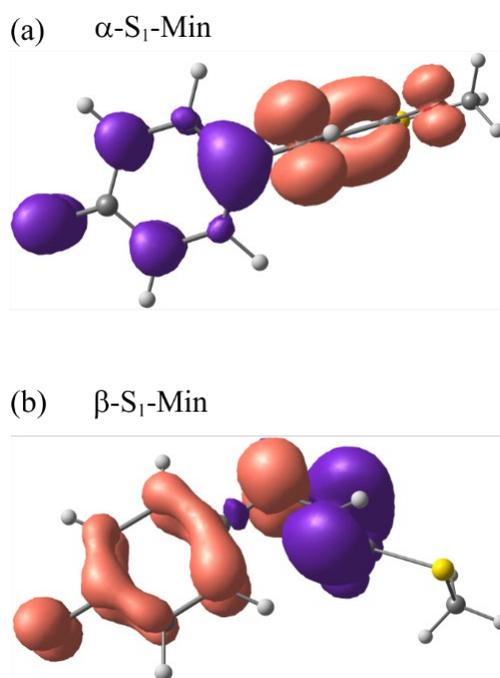


**Scheme 1.** Resonance of two closed-shell (CS), phenolic (PhCS) and quinonic (QhCS), and two biradicaloid (BR), qiononic (QnBR) and phenolic (PnBR) structures elucidating photophysical and photochemical properties of the anionic PYP chromophore.

It is worth noting that the four resonance structures are consistent with those introduced in the biradicaloid theory of Bonačić-Koutecký et al.<sup>48-49</sup> and theory of twisted intermolecular charge-transfer (TICT) of Rettig and co-workers.<sup>50-51</sup> The TICT theory treats the “hole-pair” (*hp*) configuration of a polar chromophore as electron-acceptor and electron-donor moieties connected via a central single bond. The electron transfer from the donor to the acceptor upon the HOMO-LUMO excitation (typically corresponding to the  $S_0$ – $S_1$  transition) yields a “dot-dot” (*dd*) structure with a translocated molecular charge. By twisting the linking single bond, the HOMO and LUMO involved in the charge-transfer transition become localised on the mutually orthogonal molecular fragments, and thus do not interact. Therefore, the twisting decreases the energy of the *dd* configuration and increases the energy of the *hp* configuration, which altogether significantly reduces the  $S_0$ – $S_1$  energy difference and even leads to  $S_0/S_1$  state crossing.<sup>48, 51</sup>

In Scheme 1, PhCS and QnCS correspond to conjugated *hp* configurations featuring the phenolic and carbonyl negatively charged electron-donor moieties, respectively. Electron excitation of the PhCS and QnCS *hp* configurations populates the QnBR and PhBR *dd* configurations, respectively.

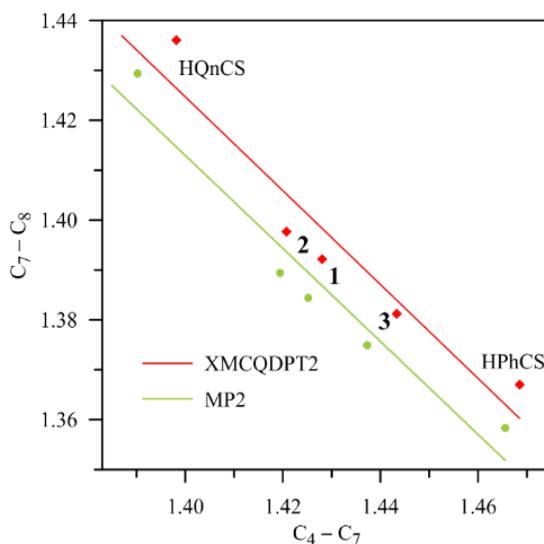
At the same time, the  $\alpha$ - ( $\beta$ -) twist leads to the QnBR (PhBR) structure in the  $S_1$  state, and the PhCS (QnCS) structure in the  $S_0$  state. This picture is fully confirmed by Fig 4, which displays the difference of the  $S_1$  and  $S_0$  total electron densities at the  $\alpha$ - $S_1$  and  $\beta$ - $S_1$  twisted minima. As can be clearly seen in the figure, the negative charge at the  $\alpha$ - $S_1$  minimum resides on the alkene-carbonyl fragment in the  $S_1$  state and on the phenolic fragment in the  $S_0$  state (Fig. 4a), whereas it is *vice versa* at the  $\beta$ - $S_1$  minimum (Fig. 4b). The same holds for models 2 and 3.



**Figure 4.** Difference of the  $S_1$  and  $S_0$  electron densities at the  $\alpha$ -twisted (a) and  $\beta$ -twisted (b) minima of pCTM<sup>-</sup> (model 1). Orange and purple correspond to  $S_1$  and  $S_0$  electron densities, respectively.

Resonance of the PhCS and QnCS structures determines the single- and double-bond lengths at the planar  $E$ - $S_0$ -Min minimum and the charge distribution in the  $S_0$  state. Figure 5 shows the BLA of the  $C_4$ - $C_7$ - $C_8$  methine bridge in the anionic models in comparison to that in the protonated phenol and enol isomers of pCTM. The shorter  $C_7$ - $C_8$  distance in comparison to  $C_4$ - $C_7$  (Fig. 5)

and the amount of the negative charge on the phenolate (Fig. 3a) indicate the dominance of PhCS over QnCS in the three models. The prevailing phenolic character of the anionic models suggests that PhCS has a lower energy than QnCS in all the models under consideration. The decrease of the phenolic charge and increase of the C<sub>7</sub>–C<sub>8</sub> bond length in the order **3-1-2** indicate the increase of the QnCS weight, and hence the increase of the PhCS and QnCS mixing in the same order.



**Figure 5.** Bond length alteration (BLA) of the C<sub>4</sub>–C<sub>7</sub>–C<sub>8</sub> methine bridge in models **1-3** in comparison to the protonated isomers of pCTM, HPhCS and HQnCS. In addition to the XMCQDPT2 results (red filled diamonds, red line), the MP2 results (green points, green line) are also shown.

At the FC point in the S<sub>1</sub> state, QnBR dominates over PhBR as it follows from the low negative charge on the phenolate (Fig. 3a) and higher NUE at C<sub>4</sub> and C<sub>7</sub> than at C<sub>8</sub> (Fig. 3b). The weight of QnBR increases in the order **2-1-3**, as indicated by a simultaneous decrease of the C<sub>8</sub> NUE and phenolic negative charge in this order. In the same order, the S<sub>0</sub>–S<sub>1</sub> charge-transfer character increases, as demonstrated by the increase of the S<sub>1</sub>–S<sub>0</sub> phenolate charge difference (Fig. 3a) and the permanent dipole moment difference (Table S1 in SI). The increase of VEE in the same order of models (Fig. 2) is explained by the higher VEE of PhCS in comparison to that of QnCS.

Thus, mixing of PhCS and QnCS determines de-localisation of the molecular charge in the  $S_0$  state, the extent of BLA at the  $S_0$  equilibrium geometry, the  $S_0$ – $S_1$  VEE and the extent of the charge transfer upon  $S_0$ – $S_1$  excitation. Obviously, the more PhCS prevails in the  $S_0$  state (small PhCS/QnCS mixing), the larger the BLA and phenolic charge in the  $S_0$  state and the larger the  $S_0$ – $S_1$  VEE and the extent of the charge-transfer. In contrast, the less the PhCS prevails in the  $S_0$  wave function (large PhCS/QnCS mixing), the smaller the BLA and phenolic charge in the  $S_0$  state, and the smaller the  $S_0$ – $S_1$  VEE and the charge-transfer upon excitation.

### **3.3 Energies of the resonance structures – impact on the $S_1$ state and $S_1/S_0$ conical intersections**

The energies of the resonance structures can be estimated for the  $90^\circ$ -twisted geometries.<sup>52</sup> Indeed, the four resonance structures are uncoupled at the  $\alpha$ - $S_1$  and  $\beta$ - $S_1$  twisted minima, with the negative charge being completely localised either on the phenolic or carbonyl fragment, as evident in Fig 3a. At the  $\alpha$ - $S_1$  minimum, the  $S_0$  state corresponds to the PhBR structure, whereas the  $S_1$  state is represented by the QnBR structure, with the negative charge being localised on the phenolic moiety and carbonyl fragment in the  $S_0$  and  $S_1$  states, respectively (Fig. 3a and 4a), and the  $C_8$  NUE has the smallest value (Fig. 3b). At the  $\beta$ - $S_1$  minimum, the  $S_0$  state is represented by the QnCS structure, while the  $S_1$  state is associated with the QnBR structure, with the negative charge being localised on the carbonyl fragment and phenolic moiety in the  $S_0$  and  $S_1$  states, respectively (Fig. 3a and 4b), and the  $C_8$  NUE has the largest value (Fig. 3b).

By comparing the energies at the  $90^\circ$ -twisted geometries (Fig. 2), we infer that PhCS has the lowest energy among the four resonance structures in all the models, which is in agreement with the analysis of the resonance mixing at the planar geometries described above. The carbonyl H-

bond stabilises QnCS and QnBR and destabilises PhCS and PhBR, whereas the phenolic H-bonds stabilise PhCS and PhBR and destabilise QnCS and QnBR. Among the four resonance forms, the energy of QnBR demonstrates the largest variation due to the H-bonding interactions.

As the geometries of the stationary points demonstrate, the BLA,  $\alpha$ -twist and  $\beta$ -twist are the main coordinates driving the relaxation of the pCTM<sup>-</sup> chromophore in the S<sub>1</sub> excited state. If the QnBR energy is lower than VEE, the initial relaxation from the FC point involves the BLA coordinate followed by the  $\alpha$ -twist. If that condition is not fulfilled, which is the case for model **3** (see Fig.2), the planar *E*-S<sub>1</sub> minimum appears on the S<sub>1</sub> PES.

At the planar *E*-S<sub>1</sub>-Min minimum (model **3**), the S<sub>0</sub>-S<sub>1</sub> oscillator strength is 0.9, and the computed radiative rate is  $\sim 0.26 \text{ ns}^{-1}$ , suggesting a sizable emission with a Stokes shift defined by the extent of the BLA relaxation. At the twisted  $\alpha$ -S<sub>1</sub> minimum in models **1** and **2**, the computed radiative rate is several orders of magnitude smaller,  $\sim 0.51 \text{ s}^{-1}$ , because of the substantial decrease of the S<sub>0</sub>-S<sub>1</sub> energy and oscillator strength. Based on these results, we predict that the dynamics of the fluorescence signal in PYP correlates with the dynamics of the chromophore H-bonds stabilizing/destabilizing the planar *E*-S<sub>1</sub> minimum. The smaller BLA variation at the planar S<sub>1</sub> minimum as compared to that at the S<sub>0</sub> minimum is a consequence of the chromophore instability with respect to the  $\alpha$ -twist. As the  $\alpha$ -twist virtually quenches the fluorescence, the variation of the emission maximum in colour-tuned PYP mutants should be significantly smaller than that of the absorption maximum, which indeed was experimentally observed by Hoff and co-workers.<sup>28</sup>

Finally, the energies of the resonance structures at the  $\alpha$ -S<sub>1</sub> and  $\beta$ -S<sub>1</sub> minima determine the energies of  $\alpha$ -CoIn and  $\beta$ -CoIn, which in turn define whether the excited chromophore returns to the S<sub>0</sub> state via a single- or double-bond twist. As Fig 2 demonstrates, the CoIn energies correlate with the S<sub>0</sub>-S<sub>1</sub> energy gap at the 90°-twisted geometries, i.e., the difference in energy of the

corresponding CS and BR resonance structures. The smaller the energy gap at the 90°-twisted  $S_1$  minimum (or saddle point), the lower the CoIn energy and the smaller the distortion of the CoIn geometry in comparison to the twisted minimum (or saddle point). As an example, we consider model **3**, for which the PhCS and QnBR energy difference is rather high, and consistently the high-energy  $\alpha$ -CoIn in this model demonstrates a substantial  $\beta$ -twist in addition to the  $\alpha$ -twist (Fig. S1 in SI). In contrast, stabilisation of PhBR and destabilisation of QnCS by the phenolic H-bonds lowers the energy of the  $\beta$ -CoIn and reduces its distortion with respect to the  $\beta$ - $S_1$  minimum. In models **1** and **2**, the PhCS and QnBR energy difference is smaller than in model **3**, the  $\alpha$ -CoIn energy therefore decreases, whereas the PhBR and QnCS energy difference is larger, and hence the  $\beta$ -CoIn energy increases. The variations of the  $\alpha$ -CoIn energy among the models suggests that accessibility of  $\alpha$ -CoIn and therefore activation of the single bond rotation and internal conversion via the single-bond twist can be efficiently controlled by chromophore-protein interactions. Stabilisation of PhCS and PhBR by these interactions reduces the resonance mixing and favours the double-bond OBF isomerisation.

### **3.4 Double-bond isomerisation of the pCTM<sup>-</sup> chromophore**

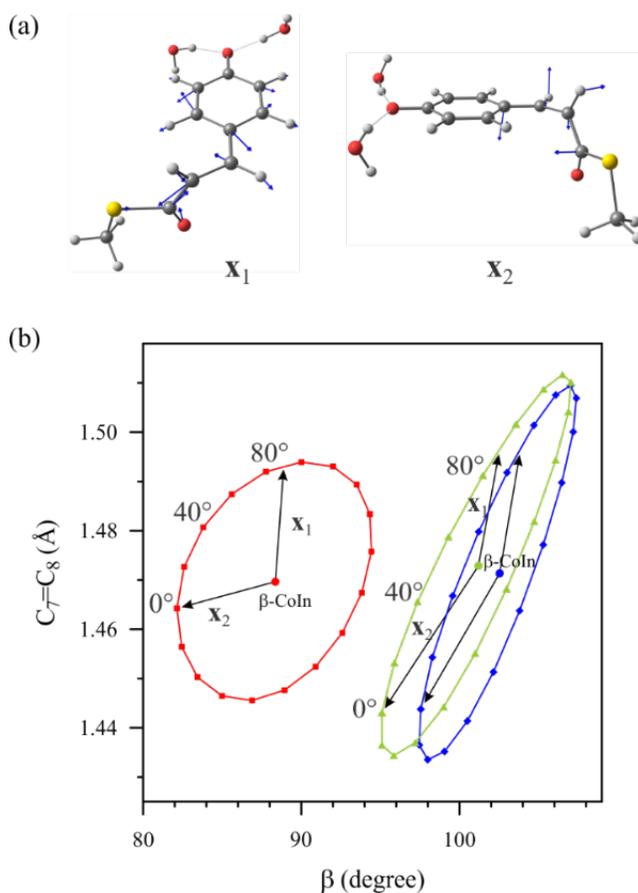
The  $\beta$ -CoIn provides the excited-state decay channel for the OBF double-bond isomerisation of pCTM<sup>-</sup>. We remind that a minimum energy CoIn geometry corresponds to a local energy minimum on the  $3N-8$  dimensional hypersurface ( $N$  is the number of atoms) where the  $S_1$  and  $S_0$  states are degenerate.<sup>42</sup> The degeneracy is lifted in the first order along the gradient difference vector ( $\mathbf{x}_1$ ) and the derivative coupling vector ( $\mathbf{x}_2$ ), forming a so-called branching plane.<sup>53-55</sup> Inspection of the energies in the branching plane unveils relaxation on the  $S_0$  PES after a non-adiabatic  $S_1$ - $S_0$  transition at the  $\beta$ -CoIn. We computed  $\mathbf{x}_1$  and  $\mathbf{x}_2$  at the  $\beta$ -CoIn geometries and

constructed branching-plane loops following the procedure suggested by Olivucci and co-workers.<sup>56</sup> In all our models,  $\mathbf{x}_1$  is dominated by BLA, whereas  $\mathbf{x}_2$  features the  $\beta$ -torsion and, in addition, hydrogen out-of-plane (HOOP)<sup>20, 22, 57</sup> excursions, which mix with the BLA changes in models **1** and **2** (see Fig. S2 in SI). At the  $\beta$ -CoIn geometries, the HOOP value tends to be “ahead” of  $\beta$ -torsion, i.e., smaller than  $\beta$ -torsion, when  $\beta > 90^\circ$  (in models **1** and **2**), whereas it is “concerted” with  $\beta$ -torsion (close to  $\beta$ -torsion) when  $\beta < 90^\circ$  (model **3**) (Fig. S3 in SI). Fig. 6a presents the  $\mathbf{x}_1$  and  $\mathbf{x}_2$  vectors for model **3** (for the other models see Fig. S2 in SI). Fig. 6b demonstrates that  $\beta$ -CoIn has the  $\beta$ -torsion value slightly smaller than  $90^\circ$  in model **3**, whereas it shifts toward a larger  $\beta$ -torsion value (i.e., closer to the initial  $E(trans)$  geometry) in models **1** and **2**. The smaller  $\beta$ -torsion value at the  $\beta$ -CoIn, explained by stabilisation of PhBR and destabilisation of QnCS in model **3**, is favourable for the  $E(trans)$ - $Z(cis)$  isomerisation. The  $\beta$ -CoIn branching plane of pCTM<sup>-</sup> is similar to that previously reported for the double-bond isomerisation of the retinal protonated Schiff base (RPSB) chromophore and analogues in rhodopsins.<sup>58-59</sup>

Figure 7 shows the energy and property changes along the branching plane loops. In the  $\mathbf{x}_1$  direction ( $\theta=90^\circ, 270^\circ$ ), the  $S_0$  and  $S_1$  wave functions change from one resonance structure to another. Specifically, the charge distribution and NUE (Fig. 7b) indicate that the  $S_0$  wave function changes from QnCS to PhBR whereas the  $S_1$  wave function changes from PhBR to QnCS. The  $\mathbf{x}_2$  direction ( $\theta=0^\circ, 180^\circ$ ) corresponds to mixing of PhBR and QnCS and, to a lesser extent, of PhCS, as indicated by the changes of the phenolic charge and NUE (Fig. 7b). The presence of PhCS follows from the steeper phenolic charge curve in comparison to the NUE curve. The PhCS weight significantly increases as the chromophore becomes more planar.

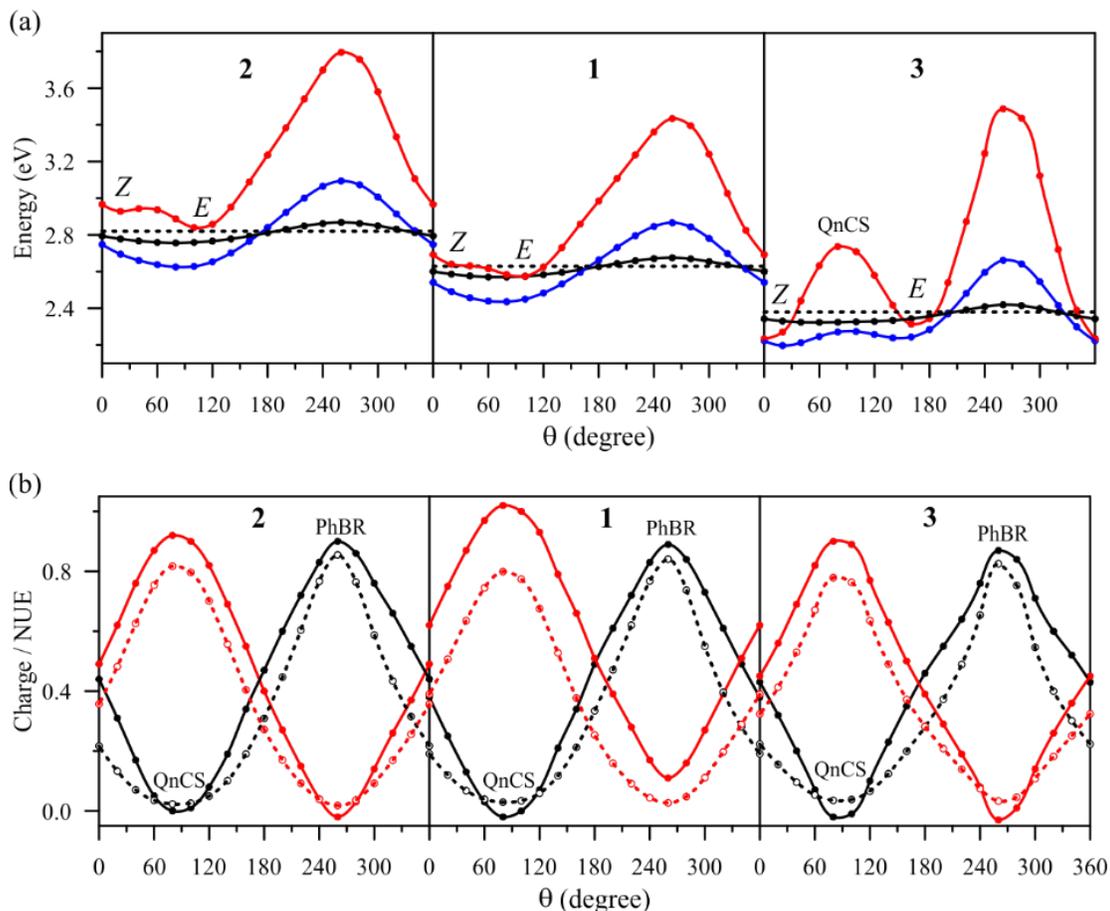
Two  $S_0$  energy minima emerge along  $\mathbf{x}_2$  (Fig. 7a) as decay channels towards the  $Z(cis)$  and  $E(trans)$  minima. Notably, the  $Z(cis)$  decay channel lies below the  $\beta$ -CoIn energy only in **3** (at

$\theta=0^\circ$ , Fig. 7a), correlating with the smaller  $\beta$ -torsion value at  $\beta$ -CoIn in this model. In addition, there is a high-energy barrier, associated with the QnCS structure, separating the two channels in model **3**, which is explained by the higher QnCS energy in this model. Overall, the topology of the  $S_0$  PES along the loops suggests that the double-bond isomerisation quantum yield is higher in **3** as compared to **1** and **2**.



**Figure 6.** Characterisation of the  $\beta$ -twisted CoIn: (a) The branching plane vectors  $x_1$  and  $x_2$  for model **3** (see also Fig. S2 presenting  $x_1$  and  $x_2$  for all three models) (b) Changes in the  $\beta$ -torsion and  $C_7=C_8$  bond length along the  $0.005\text{-\AA}$  loops for models **1-3**. The values of the angular coordinate  $\theta$  at some points of the loops, and the orientation of the  $x_1$  and  $x_2$  vectors are indicated. The colour code is the same as in Fig. 2.

Finally, we discuss on some aspects related to the sloped character<sup>60</sup> of the  $\beta$ -CoIn. There are two stationary points associated with the  $\beta$ -CoIn, the  $\beta$ -S<sub>1</sub> minimum on the S<sub>1</sub> PES, and a saddle point on the S<sub>0</sub> PES, which are of the PhBR and QnCS electronic structure, respectively. The S<sub>0</sub> saddle point is associated with the aforementioned QnCS energy barrier separating *Z(cis)* and *E(trans)* channel (Fig. 7a). Our preliminary calculations for model **3** have identified such a point.



**Figure 7.** Energies and properties computed at the geometries constituting the loops circumventing the  $\beta$ -CoIn in the branching plane for models **1-3**.  $\theta$  represents the angular coordinate of the loops.  $\theta=0^\circ$  and  $\theta=180^\circ$  define the derivative coupling vector  $\mathbf{x}_2$ .  $\theta=90^\circ$  and  $\theta=270^\circ$  define the gradient difference vector  $\mathbf{x}_1$ . (a) S<sub>0</sub> energies for the loops of radii 0.001 Å (black), 0.005 Å (blue) 0.0011 Å (red). The dashed line indicates the  $\beta$ -CoIn energy. (b) Net charge on the phenolic fragment (solid lines with filled symbols) and NUE at the C<sub>8</sub> atom (dashed lines with empty symbols) in the S<sub>0</sub> (black) and S<sub>1</sub> (red) states.

This is the only saddle point connecting the  $Z(cis)$  and  $E(trans)$  minima on the  $S_0$  PES, and hence it mediates the *thermal* isomerisation. The sloped type of  $\beta$ -CoIn implies that the aforementioned two stationary points lie on the *same* side from the  $\beta$ -CoIn along  $x_1$ . This is different from the peaked CoIn found for the RPSB chromophore and its analogues, where two saddle points of different electronic structure lying on the  $S_0$  PES flank the CoIn along the BLA coordinate.<sup>61-64</sup> Noteworthy, for the rhodopsin photoreceptor, the energy of the saddle point controlling the thermal isomerisation was found to correlate with VEE, such that the blue-shifted absorption maximum corresponds to a reduced thermal activation (higher energy of the saddle points).<sup>61</sup> Despite different  $\beta$ -CoIn topology, a similar correlation as for the RPSB chromophore should be expected for the pCTM<sup>-</sup> chromophore, as the increase of VEE correlates with the increase of the QnCS energy.

### 3.5 Comparison to other computational studies

Although our models **1-3** do not account for the real environment of the pCTM<sup>-</sup> chromophore in PYP, they correctly grasp the effects of the Cys69 and Tyr42/Glu46 H-bonds (Fig. 1a). The energy changes presented in Fig. 2 are qualitatively consistent with those predicted by the hybrid quantum-mechanics/molecular mechanics (QM/MM) models of PYP<sup>16, 18-19, 26</sup> and solvated chromophore.<sup>24</sup> Specifically, the carbonyl H-bond lowers the  $\alpha$ -CoIn energy, whereas the phenolic H-bonds lower the  $\beta$ -CoIn energy in the QM/MM models.<sup>24</sup> In the surface-hopping molecular dynamics simulations of PYP,<sup>26</sup> the presence of the protonated (positively charged) Arg52 stabilizing the phenolic negative charge, that is the PhCS and PhBR resonance structures with respect to the QnCS and QnBR ones, leads to stabilisation of the  $\beta$ -CoIn, and hence facilitates the OBF isomerisation.

We compare the energies of model **1** with the results of Boggio-Pasqua and Groenhof<sup>21</sup> obtained using the CASSCF(12,11) method, and the energies of model **3** with the results of Gromov et al.<sup>17, 20</sup> obtained using the CC2 method. It is instructive to consider the energies of the resonance structures at the 90°-twisted geometries. Both CC2 and CASSCF favour QnBR over PhBR more than predicted by the XMCQDPT2 method. Consistent with this energy trend, the  $\beta$ -CoIn is higher in energy than  $\alpha$ -CoIn at the CASSCF level of theory, whereas it is vice versa at the XMCQDPT2 level. Thus, employing the CASSCF method to describe the pCTM<sup>-</sup> chromophore in surface-hopping dynamics of PYP<sup>16, 26</sup> underestimates the efficiency of the  $\beta$ -twist in the S<sub>1</sub> state. For a similar anionic chromophore of the green fluorescent proteins (GFPs), it has indeed been reported that the CASSCF method underestimates the decay of the S<sub>1</sub> state via the  $\beta$ -twist as compared to the more accurate XMSCASPT2 method.<sup>65</sup> In the QM/MM simulations of PYP,<sup>26</sup> this underestimation seems to be compensated by the effect of the positively charged residues, such as protonated Arg52 stabilizing, the PhCS and PhBR structures.

### 3.6 Similarity with other biological chromophores

Similar to pCTM<sup>-</sup>, the anionic p-hydroxybenzylideneimidazolinole (pHBDI<sup>-</sup>) chromophore of GFPs is described by resonance interactions of the PhCS and QnCS forms coupled along the BLA reaction coordinate.<sup>66-67</sup> Recently, Boxer and co-workers<sup>67</sup> suggested that the energy difference of the PhCS and QnCS forms can serve as a linear scale for quantitative prediction of the GFP photophysical properties using a small number of parameters intrinsic to the pHBDI<sup>-</sup> chromophore. This energy difference was implicated to explain all strong correlations of properties among a large number of systematically tuned GFP variants.<sup>67</sup> We note that the charge-transfer GFP model of Boxer and co-workers<sup>67</sup> is consistent with our computational results for the anionic

pCTM<sup>-</sup> except that it does not invoke the QnBR and PhBR structures of pHBDI<sup>-</sup>. By analogy to pCTM<sup>-</sup>, we suggest that the energies and properties of the QnBR and PhBR resonance forms should be considered when addressing the S<sub>1</sub> PES of pHBDI<sup>-</sup> in GFPs, in particular its photoactivation. Applying this prediction, stabilisation of PhBR in pHBDI<sup>-</sup> favours the OBF double-bond isomerisation, whereas stabilisation of QnBR gives a chance to the single-bond activation and HT isomerisation.<sup>68</sup>

Mixing of the resonance forms describes charge delocalisation in protonated cationic chromophores such as the RPSB chromophore of rhodopsins and tetrapyrrole chromophores of phytochromes. The extent of mixing explains different sensitivities of these chromophores to electrostatic interactions with their proteins. The dominant resonance structure of RPSB features a positive charge localised on the Schiff-base fragment, giving rise to considerable charge-transfer character of the S<sub>0</sub>-S<sub>1</sub> transition and high sensitivity to electrostatic interactions with the protein. These interactions in turn, efficiently control the contributions of the resonance forms, as suggested by the correlations of the increased/decreased BLA and increased/decreased VEE.<sup>69-71</sup> In contrast, the tetrapyrrole positive charge is redistributed among the four pyrrole rings described by at least four resonance forms.<sup>72</sup> This charge delocalisation reduces the charge-transfer character of the S<sub>0</sub>-S<sub>1</sub> transition and subsequently reduces the electrostatic effect of the protein on the chromophore.

#### **4. CONCLUSIONS AND OUTLOOK**

Given the prominent role of ionic chromophores in light activation of naturally evolved photoreceptor proteins, it is important to rationalise the effect of chromophore-protein interactions that are crucial for the light sensory function. Here, by analysing how the anionic pCTM<sup>-</sup>

chromophore of the PYP bacterial photoreceptor is influenced by phenolic or carbonyl H-bonds we derived a qualitative model explaining interdependent changes of all photochemical properties related to the PYP photoactivation. To account for electronic correlation effects, the state-of-the-art XMCQDPT2 method was employed to compute the  $S_0$  and  $S_1$  PES cross-sections. To rationalise the obtained trends, two pairs of resonance structures, each pair comprised of a CS structure and corresponding to it charge-transfer BR structure were invoked (Scheme 1). Delocalisation of the negative charge and other properties of the  $S_0$  and  $S_1$  states are described by chemical resonance of the structures with the opposite charge localisation. Contributions of the resonance structures depend on their energies that we compare at the  $90^\circ$  single- and double-bond twisted geometries where chemical resonance vanishes.

From the computational results, the following picture of tuning pCTM<sup>-</sup> by intermolecular interactions emerges. Stabilisation of the phenolic CS and BR structures by the phenolic H-bonds leads to the larger difference in the lengths of the central single and double bonds (i.e. larger BLA) at the  $S_0$  minimum-energy geometry; there is also an increase of the excitation energy and stabilisation of the planar  $S_1$  minimum. Moreover, isomerisation around the central double bond becomes more favourable than the phenolic ring rotation around the single bond, and the energy barrier controlling the thermal double-bond isomerisation increases. In contrast, stabilisation of the quinonic structures by the carbonyl H-bond leads to delocalisation of the negative charge and a smaller BLA in the  $S_0$  state, smaller  $S_0$ – $S_1$  excitation energy and smaller charge transfer character, as well as activation of the single-bond rotation in the  $S_1$ , increasing the probability of the HT photoisomerisation. Overall, our computational results predict that stabilisation of the pCTM<sup>-</sup> phenolic resonance structures, i.e. reduction of chemical resonance, is crucial for PYP photoactivation via the OBF double-bond isomerisation.

Comparison of our findings and conclusions to the previously published computational and theoretical results characterizing other biological chromophores identifies many common features. In particular, our model derived from the high-level quantum-chemistry calculations in its essence is similar to a model proposed by Boxer and co-workers<sup>67, 73</sup> explaining spectral properties of systematically mutated GFP proteins. Yet, our model extends Boxer's model to the twisted geometries and invokes BR structures to describe the  $S_1$  state properties. The broad similarities among the ionic chromophores imply that the concept of chemical resonance is instructive in experimental and especially computational studies addressing photochemical mechanisms of photosensory proteins operating via the double-bond isomerisation.

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## REFERENCES

1. Hellingwerf, K. J.; Hendriks, J.; Gensch, T., Photoactive Yellow Protein, a new type of photoreceptor protein: Will this "yellow lab" bring us where we want to go? *J Phys Chem A* **2003**, *107* (8), 1082-1094.
2. Schotte, F.; Cho, H. S.; Kaila, V. R.; Kamikubo, H.; Dashdorj, N.; Henry, E. R.; Graber, T. J.; Henning, R.; Wulff, M.; Hummer, G.; Kataoka, M.; Anfinrud, P. A., Watching a signaling protein function in real time via 100-ps time-resolved Laue crystallography. *Proc Natl Acad Sci U S A* **2012**, *109* (47), 19256-61.
3. Jung, Y. O.; Lee, J. H.; Kim, J.; Schmidt, M.; Moffat, K.; Srajer, V.; Ihee, H., Volume-conserving trans-cis isomerization pathways in photoactive yellow protein visualized by picosecond X-ray crystallography. *Nat Chem* **2013**, *5* (3), 212-20.
4. Tenboer, J.; Basu, S.; Zatsepin, N.; Pande, K.; Milathianaki, D.; Frank, M.; Hunter, M.; Boutet, S.; Williams, G. J.; Koglin, J. E.; Oberthuer, D.; Heymann, M.; Kupitz, C.; Conrad, C.; Coe, J.; Roy-Chowdhury, S.; Weierstall, U.; James, D.; Wang, D.; Grant, T.; Barty, A.; Yefanov, O.; Scales, J.; Gati, C.; Seuring, C.; Srajer, V.; Henning, R.; Schwander, P.; Fromme, R.; Ourmazd, A.; Moffat, K.; Van Thor, J. J.; Spence, J. C.; Fromme, P.; Chapman, H. N.; Schmidt, M., Time-resolved serial crystallography captures high-resolution intermediates of photoactive yellow protein. *Science* **2014**, *346* (6214), 1242-6.
5. Pande, K.; Hutchison, C. D.; Groenhof, G.; Aquila, A.; Robinson, J. S.; Tenboer, J.; Basu, S.; Boutet, S.; DePonte, D. P.; Liang, M.; White, T. A.; Zatsepin, N. A.; Yefanov, O.; Morozov, D.; Oberthuer, D.; Gati, C.; Subramanian, G.; James, D.; Zhao, Y.; Koralek, J.; Brayshaw, J.; Kupitz, C.; Conrad, C.; Roy-Chowdhury, S.; Coe, J. D.; Metz, M.; Xavier, P. L.; Grant, T. D.; Koglin, J. E.; Ketawala, G.; Fromme, R.; Srajer, V.; Henning, R.; Spence, J. C.; Ourmazd, A.; Schwander, P.; Weierstall, U.; Frank, M.; Fromme, P.; Barty, A.; Chapman, H. N.; Moffat, K.; van Thor, J. J.; Schmidt, M., Femtosecond structural dynamics drives the trans/cis isomerization in photoactive yellow protein. *Science* **2016**, *352* (6286), 725-9.
6. Yonezawa, K.; Shimizu, N.; Kurihara, K.; Yamazaki, Y.; Kamikubo, H.; Kataoka, M., Neutron crystallography of photoactive yellow protein reveals unusual protonation state of Arg52 in the crystal. *Sci Rep* **2017**, *7* (1), 9361.
7. Sigala, P. A.; Tsuchida, M. A.; Herschlag, D., Hydrogen bond dynamics in the active site of photoactive yellow protein. *Proc Natl Acad Sci USA* **2009**, *106* (23), 9232-9237.
8. Larsen, D. S.; van Grondelle, R., Initial photoinduced dynamics of the photoactive yellow protein. *Chemphyschem* **2005**, *6* (5), 828-837.
9. van Wilderen, L. J.; van der Horst, M. A.; van Stokkum, I. H.; Hellingwerf, K. J.; van Grondelle, R.; Groot, M. L., Ultrafast infrared spectroscopy reveals a key step for successful entry into the photocycle for photoactive yellow protein. *Proc Natl Acad Sci U S A* **2006**, *103* (41), 15050-5.
10. Mendonca, L.; Hache, F.; Chagnenet-Barret, P.; Plaza, P.; Chosrowjan, H.; Taniguchi, S.; Imamoto, Y., Ultrafast Carbonyl Motion of the Photoactive Yellow Protein Chromophore Probed by Femtosecond Circular Dichroism. *J Am Chem Soc* **2013**, *135* (39), 14637-14643.
11. Creelman, M.; Kumauchi, M.; Hoff, W. D.; Mathies, R. A., Chromophore Dynamics in the PYP Photocycle from Femtosecond Stimulated Raman Spectroscopy. *Journal of Physical Chemistry B* **2014**, *118* (3), 659-667.

12. Kuramochi, H.; Takeuchi, S.; Yonezawa, K.; Kamikubo, H.; Kataoka, M.; Tahara, T., Probing the early stages of photoreception in photoactive yellow protein with ultrafast time-domain Raman spectroscopy. *Nature Chemistry* **2017**, *9* (7), 660-666.
13. Genick, U. K.; Soltis, S. M.; Kuhn, P.; Canestrelli, I. L.; Getzoff, E. D., Structure at 0.85 Å resolution of an early protein photocycle intermediate. *Nature* **1998**, *392* (6672), 206-9.
14. Vengris, M.; van der Horst, M. A.; Zgrablic, G.; van Stokkum, I. H.; Haacke, S.; Chergui, M.; Hellingwerf, K. J.; van Grondelle, R.; Larsen, D. S., Contrasting the excited-state dynamics of the photoactive yellow protein chromophore: protein versus solvent environments. *Biophys J* **2004**, *87* (3), 1848-57.
15. Lee, I. R.; Lee, W.; Zewail, A. H., Primary steps of the photoactive yellow protein: isolated chromophore dynamics and protein directed function. *Proc Natl Acad Sci U S A* **2006**, *103* (2), 258-62.
16. Groenhof, G.; Bouxin-Cademartory, M.; Hess, B.; De Visser, S. P.; Berendsen, H. J. C.; Olivucci, M.; Mark, A. E.; Robb, M. A., Photoactivation of the photoactive yellow protein: Why photon absorption triggers a trans-to-cis isomerization of the chromophore in the protein. *J Am Chem Soc* **2004**, *126* (13), 4228-4233.
17. Gromov, E. V.; Burghardt, I.; Hynes, J. T.; Koppel, H.; Cederbaum, L. S., Electronic structure of the photoactive yellow protein chromophore: Ab initio study of the low-lying excited singlet states. *J Photoch Photobio A* **2007**, *190* (2-3), 241-257.
18. Ko, C.; Virshup, A. M.; Martinez, T. J., Electrostatic control of photoisomerization in the photoactive yellow protein chromophore: Ab initio multiple spawning dynamics. *Chem Phys Lett* **2008**, *460* (1-3), 272-277.
19. Coto, P. B.; Marti, S.; Oliva, M.; Olivucci, M.; Merchan, M.; Andres, J., Origin of the absorption maxima of the photoactive yellow protein resolved via ab initio multiconfigurational methods. *Journal of Physical Chemistry B* **2008**, *112* (24), 7153-7156.
20. Gromov, E. V.; Burghardt, I.; Koppel, H.; Cederbaum, L. S., Photoinduced Isomerization of the Photoactive Yellow Protein (PYP) Chromophore: Interplay of Two Torsions, a HOOP Mode and Hydrogen Bonding. *J Phys Chem A* **2011**, *115* (33), 9237-9248.
21. Boggio-Pasqua, M.; Groenhof, G., On the use of reduced active space in CASSCF calculations. *Comput Theor Chem* **2014**, *1040*, 6-13.
22. Gromov, E. V., Unveiling the mechanism of photoinduced isomerization of the photoactive yellow protein (PYP) chromophore. *J Chem Phys* **2014**, *141* (22), 224308-224320.
23. Premvardhan, L. L.; van der Horst, M. A.; Hellingwerf, K. J.; van Grondelle, R., Stark spectroscopy on photoactive yellow protein, E46Q, and a nonisomerizing derivative, probes photo-induced charge motion. *Biophysical Journal* **2003**, *84* (5), 3226-3239.
24. Boggio-Pasqua, M.; Robb, M. A.; Groenhof, G., Hydrogen Bonding Controls Excited-State Decay of the Photoactive Yellow Protein Chromophore. *J Am Chem Soc* **2009**, *131* (38), 13580-13581.
25. Gromov, E. V.; Burghardt, I.; Koppel, H.; Cederbaum, L. S., Native hydrogen bonding network of the photoactive yellow protein (PYP) chromophore: Impact on the electronic structure and photoinduced isomerization. *J Photoch Photobio A* **2012**, *234*, 123-134.
26. Groenhof, G.; Schafer, L. V.; Boggio-Pasqua, M.; Grubmüller, H.; Robb, M. A., Arginine52 controls the photoisomerization process in photoactive yellow protein. *J Am Chem Soc* **2008**, *130* (11), 3250-1.
27. Groot, M. L.; van Wilderen, L. J. G. W.; Larsen, D. S.; van der Horst, M. A.; van Stokkum, I. H. M.; Hellingwerf, K. J.; van Grondelle, R., Initial steps of signal generation in

- photoactive yellow protein revealed with femtosecond mid-infrared spectroscopy. *Biochemistry-Us* **2003**, *42* (34), 10054-10059.
28. Philip, A. F.; Nome, R. A.; Papadantonakis, G. A.; Scherer, N. F.; Hoff, W. D., Spectral tuning in photoactive yellow protein by modulation of the shape of the excited state energy surface. *P Natl Acad Sci USA* **2010**, *107* (13), 5821-5826.
  29. Granovsky, A. A., Extended multi-configuration quasi-degenerate perturbation theory: The new approach to multi-state multi-reference perturbation theory. *J Chem Phys* **2011**, *134* (21).
  30. Coto, P. B.; Roca-Sanjuan, D.; Serrano-Andres, L.; Martin-Pendas, A.; Marti, S.; Andres, J., Toward Understanding the Photochemistry of Photoactive Yellow Protein: A CASPT2/CASSCF and Quantum Theory of Atoms in Molecules Combined Study of a Model Chromophore in Vacuo. *J Chem Theory Comput* **2009**, *5* (11), 3032-3038.
  31. Vahtras, O.; Almlöf, J.; Feyereisen, M. W., Integral Approximations for Lcao-Scf Calculations. *Chem Phys Lett* **1993**, *213* (5-6), 514-518.
  32. Eichkorn, K.; Treutler, O.; Ohm, H.; Haser, M.; Ahlrichs, R., Auxiliary Basis-Sets to Approximate Coulomb Potentials. *Chem Phys Lett* **1995**, *240* (4), 283-289.
  33. Ahlrichs, R., Efficient evaluation of three-center two-electron integrals over Gaussian functions. *Phys Chem Chem Phys* **2004**, *6* (22), 5119-5121.
  34. Witek, H. A.; Choe, Y. K.; Finley, J. P.; Hirao, K., Intruder state avoidance multireference Moller-Plesset perturbation theory. *J Comput Chem* **2002**, *23* (10), 957-965.
  35. Dunning, T. H., Gaussian-Basis Sets for Use in Correlated Molecular Calculations .1. The Atoms Boron through Neon and Hydrogen. *J Chem Phys* **1989**, *90* (2), 1007-1023.
  36. Woon, D. E.; Dunning, T. H., Gaussian-Basis Sets for Use in Correlated Molecular Calculations .3. The Atoms Aluminum through Argon. *J Chem Phys* **1993**, *98* (2), 1358-1371.
  37. Levine, B. G.; Coe, J. D.; Martinez, T. J., Optimizing conical intersections without derivative coupling vectors: Application to multistate multireference second-order perturbation theory (MS-CASPT2). *Journal of Physical Chemistry B* **2008**, *112* (2), 405-413.
  38. Sherill, C. D., Counterpoise Correction and Basis Set Superposition Error <http://vergil.chemistry.gatech.edu/notes/cp.pdf>. **2010**.
  39. Mayer, I., Bond order and valence indices: A personal account. *J Comput Chem* **2007**, *28* (1), 204-221.
  40. Gozem, S.; Melaccio, F.; Valentini, A.; Filatov, M.; Huix-Rotllant, M.; Ferre, N.; Frutos, L. M.; Angeli, C.; Krylov, A. I.; Granovsky, A. A.; Lindh, R.; Olivucci, M., Shape of Multireference, Equation-of-Motion Coupled-Cluster, and Density Functional Theory Potential Energy Surfaces at a Conical Intersection. *J Chem Theory Comput* **2014**, *10* (8), 3074-3084.
  41. Tuna, D.; Lefrancois, D.; Wolanski, L.; Gozem, S.; Schapiro, I.; Andruniow, T.; Dreuw, A.; Olivucci, M., Assessment of Approximate Coupled-Cluster and Algebraic-Diagrammatic-Construction Methods for Ground- and Excited-State Reaction Paths and the Conical-Intersection Seam of a Retinal-Chromophore Model. *J Chem Theory Comput* **2015**, *11* (12), 5758-5781.
  42. Atchity, G. J.; Xantheas, S. S.; Ruedenberg, K., Potential-Energy Surfaces near Intersections. *J Chem Phys* **1991**, *95* (3), 1862-1876.

43. Frisch, M. J.; al., e., Gaussian 09, Revision E.01. *Gaussian, Inc., Wallingford, CT, 2016* **2009**.
44. Schmidt, M. W.; Gordon, M. S., The construction and interpretation of MCSCF wavefunctions. *Annu Rev Phys Chem* **1998**, *49*, 233-266.
45. Granovsky, A. A., Firefly version 8, www <http://classic.chem.msu.su/gran/firefly/index.html>.
46. Schmidt, M. W.; Baldrige, K. K.; Boatz, J. A.; Elbert, S. T.; Gordon, M. S.; Jensen, J. H.; Koseki, S.; Matsunaga, N.; Nguyen, K. A.; Su, S. J.; Windus, T. L.; Dupuis, M.; Montgomery, J. A., General Atomic and Molecular Electronic-Structure System. *J Comput Chem* **1993**, *14* (11), 1347-1363.
47. Krause, D.; Thörnig, P., JURECA: Modular supercomputer at Jülich Supercomputing Centre. *Journal of large-scale research facilities* **2018**, *4*, A132.
48. Bonacic-Koutecky, V.; Koutecky, J.; Michl, J., Neutral and Charged Biradicals, Zwitterions, Funnels in S1, and Proton Translocation - Their Role in Photochemistry, Photophysics, and Vision. *Angew Chem Int Edit* **1987**, *26* (3), 170-189.
49. Bonacic-Koutecky, V.; Schoffel, K.; Michl, J., Critically Heterosymmetric Biradicaloid Geometries of Protonated Schiff-Bases - Possible Consequences for Photochemistry and Photobiology. *Theor Chim Acta* **1987**, *72* (5-6), 459-474.
50. Grabowski, Z. R.; Rotkiewicz, K.; Rettig, W., Structural changes accompanying intramolecular electron transfer: Focus on twisted intramolecular charge-transfer states and structures. *Chem Rev* **2003**, *103* (10), 3899-4031.
51. Dekhtyar, M.; Rettig, W., Charge-transfer transitions in twisted stilbenoids: Interchangeable features and generic distinctions of single- and double-bond twists. *J Phys Chem A* **2007**, *111* (11), 2035-2039.
52. Dormans, G. J. M.; Groenenboom, G. C.; Vandorst, W. C. A.; Buck, H. M., A Quantum Chemical Study on the Mechanism of Cis Trans Isomerization in Retinal-Like Protonated Schiff-Bases. *J Am Chem Soc* **1988**, *110* (5), 1406-1415.
53. Robb, M. A.; Bernardi, F.; Olivucci, M., Conical Intersections as a Mechanistic Feature of Organic-Photochemistry. *Pure Appl Chem* **1995**, *67* (5), 783-789.
54. Bernardi, F.; Olivucci, M.; Robb, M. A., Potential energy surface crossings in organic photochemistry. *Chem Soc Rev* **1996**, *25* (5), 321-328.
55. Robb, M. A., Conical Intersections in Organic Photochemistry. In *Conical Intersections, Theory, Computation, and Experiment*, Domcke, W.; Yarkony, D. R.; Köppel, H., Eds. World Scientific Singapore: 2011; Vol. 17, pp 3-50.
56. Coto, P. B.; Sinicropi, A.; De Vico, L.; Ferre, N.; Olivucci, M., Characterization of the conical intersection of the visual pigment rhodopsin at the CASPT2//CASSCF/AMBER level of theory. *Mol Phys* **2006**, *104* (5-7), 983-991.
57. Boggio-Pasqua, M.; Burmeister, C. F.; Robb, M. A.; Groenhof, G., Photochemical reactions in biological systems: probing the effect of the environment by means of hybrid quantum chemistry/molecular mechanics simulations. *Phys Chem Chem Phys* **2012**, *14* (22), 7912-28.
58. Molnar, F.; Ben-Nun, M.; Martinez, T. J.; Schulten, K., Characterization of a conical intersection between the ground and first excited state for a retinal analog. *J Mol Struct-Theochem* **2000**, *506*, 169-178.
59. Sen, S.; Schapiro, I., A comprehensive benchmark of the XMS-CASPT2 method for the photochemistry of a retinal chromophore model. *Mol Phys* **2018**, *116* (19-20), 2571-2582.

60. Hall, K. F.; Boggio-Pasqua, M.; Bearpark, M. J.; Robb, M. A., Photostability via sloped conical intersections: a computational study of the excited states of the naphthalene radical cation. *J Phys Chem A* **2006**, *110* (50), 13591-9.
61. Gozem, S.; Schapiro, I.; Ferre, N.; Olivucci, M., The Molecular Mechanism of Thermal Noise in Rod Photoreceptors. *Science* **2012**, *337* (6099), 1225-1228.
62. Rinaldi, S.; Melaccio, F.; Gozem, S.; Fanelli, F.; Olivucci, M., Comparison of the isomerization mechanisms of human melanopsin and invertebrate and vertebrate rhodopsins. *Proc Natl Acad Sci U S A* **2014**, *111* (5), 1714-9.
63. Luk, H. L.; Bhattacharyya, N.; Montisci, F.; Morrow, J. M.; Melaccio, F.; Wada, A.; Sheves, M.; Fanelli, F.; Chang, B. S.; Olivucci, M., Modulation of thermal noise and spectral sensitivity in Lake Baikal cottoid fish rhodopsins. *Sci Rep* **2016**, *6*, 38425.
64. Gozem, S.; Luk, H. L.; Schapiro, I.; Olivucci, M., Theory and Simulation of the Ultrafast Double-Bond Isomerization of Biological Chromophores. *Chem Rev* **2017**, *117* (22), 13502-13565.
65. Park, J. W.; Shiozaki, T., Analytical Derivative Coupling for Multistate CASPT2 Theory. *J Chem Theory Comput* **2017**, *13* (6), 2561-2570.
66. Bravaya, K. B.; Grigorenko, B. L.; Nemukhin, A. V.; Krylov, A. I., Quantum chemistry behind bioimaging: insights from ab initio studies of fluorescent proteins and their chromophores. *Acc Chem Res* **2012**, *45* (2), 265-75.
67. Lin, C. Y.; Romei, M. G.; Oltrogge, L. M.; Mathews, I. I.; Boxer, S. G., Unified Model for Photophysical and Electro-Optical Properties of Green Fluorescent Proteins. *J Am Chem Soc* **2019**, *141* (38), 15250-15265.
68. Chang, J.; Romei, M. G.; Boxer, S. G., Structural Evidence of Photoisomerization Pathways in Fluorescent Proteins. *J Am Chem Soc* **2019**, *141* (39), 15504-15508.
69. Sekharan, S.; Katayama, K.; Kandori, H.; Morokuma, K., Color Vision: "OH-Site" Rule for Seeing Red and Green. *J Am Chem Soc* **2012**, *134* (25), 10706-10712.
70. Sekharan, S.; Wei, J. N.; Batista, V. S., The Active Site of Melanopsin: The Biological Clock Photoreceptor. *J Am Chem Soc* **2012**, *134* (48), 19536-19539.
71. Pal, R.; Sekharan, S.; Batista, V. S., Spectral Tuning in Halorhodopsin: The Chloride Pump Photoreceptor. *J Am Chem Soc* **2013**, *135* (26), 9624-9627.
72. Maximowitsch, E.; Domracheva, T., Submitted for publication. 2020.
73. Romei, M. G.; Lin, C. Y.; Mathews, I. I.; Boxer, S. G., Electrostatic control of photoisomerization pathways in proteins. *Science* **2020**, *367* (6473), 76-79.