Peptidic boronic acids are potent cell-permeable inhibitors of the malaria parasite egress serine protease SUB1

Elina Lidumniea, Chrislaine Withers-Martinezb, Fiona Hackettb, Christine R. Collinsb, Abigail J. Perrinb, Konstantinos Koussisb, Claudine Bissonc,d, Michael J. Blackmanb,e,1, and Aigars Jirgensonsa,1

Plasmodium falciparum, a single-celled parasite that invades and replicates within red blood cells (RBCs), eventually destroys the cells in a lytic population. The causative agent, a protozoan parasite, replicates in vitro at submicromolar concentrations. Our results validate SUB1 as a potential target for a new class of antimalarial drugs designed to prevent parasite replication and disease progression.

Malaria is a devastating infectious disease, which causes over 400,000 deaths per annum and impacts the lives of nearly half the world’s population. The causative agent, a protozoan parasite, replicates within red blood cells (RBCs), eventually destroying the cells in a lytic process called egress to release a new generation of parasites. These invade fresh RBCs to repeat the cycle. Egress is regulated by an essential parasite subtilisin-like serine protease called SUB1. Here, we describe the development and optimization of substrate-based peptidic boronic acids that inhibit Plasmodium falciparum SUB1 with low nanomolar potency. Structural optimization generated membrane-permeable, slow-off-rate inhibitors that prevent P. falciparum egress through direct inhibition of SUB1 activity and block parasite replication in vitro at submicromolar concentrations. Our results validate SUB1 as a potential target for a new class of antimalarial drugs designed to prevent parasite replication and disease progression.

Significance

Malaria remains a major global health threat. In the face of increasing resistance to available chemotherapeutics, new antimalarial drugs with new modes of action are urgently needed. The causative agent of malaria is a single-celled parasite that invades and replicates within red blood cells. Escape from the red cell, a process called egress, involves a proteolytic pathway triggered by an essential parasite subtilisin-like serine protease called SUB1. Here, we describe the development and rational optimization of a potent, membrane-permeable substrate-based boronic acid compounds that block egress and parasite proliferation by direct inhibition of SUB1 activity. The compounds could form the basis of a new type of antimalarial medicine that would both protect against infection and treat disease.

Author contributions: A.J. designed inhibitors; E.L., C.W.-M., F.H., C.R.C., A.J.P., K.K., and A.J. performed research; E.L., C.W.-M., F.H., C.R.C., A.J.P., and K.K. analyzed data; M.J.B. and A.J. wrote the paper; and C.B. generated the B11-EXP2-mNeonGreen parasite line. The authors declare no competing interest.

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proteases, here we have investigated peptidic boronic acids as PISUB1 inhibitors. These efforts have generated nanomolar PISUB1 inhibitors that can access PISUB1 in the intraerythrocytic parasite and prevent parasite replication through direct inhibition of egress.

Results

Discovery of Potent Substrate-Based Peptidyl Boronic Acid Inhibitors of PISUB1. We previously described the development of a fluorescence-based in vitro assay suitable for the evaluation of substrate-based PISUB1 inhibitors, using recombinant (rPISUB1) and fluorogenic peptide substrates based on cleavage sites within endogenous protein substrates of PISUB1 (13, 21). In our earlier work (13, 20), we used the assay to identify a substrate-based pentapeptidic α-ketoamide with a P4 Ile residue and P2 Gly residue as our most potent inhibitor (IC\textsubscript{50} ∼900 nM; Fig. 1). Unfortunately, this and related α-ketoamides showed no anti-parasite activity in vitro. This was perhaps unsurprising due to the high molecular mass and polar nature of these compounds, including the presence of a carboxylic acid moiety that was designed to mimic endogenous PISUB1 protein substrates by interacting with the basic S\textsuperscript{1} surface of the PISUB1 active-site cleft (16). Collectively, these features likely rendered the compounds poorly membrane penetrant.

To build on that work, we first explored a range of P4 substituents of the N-acetyl peptidyl α-ketoamide scaffold, maintaining the P1 Ala, P2 Gly, and P3 Thr sidechains unaltered. Replacement of the P4 Ile side chain with a cyclopentane improved potency, resulting in a peptidic α-ketoamide (2a) with an IC\textsubscript{50} ∼370 nM (SI Appendix, Table S1). Reasoning that the exploration of alternative warheads with known activity against serine proteases might prove fruitful, we replaced the α-ketoamide functionality of this compound with a boronic acid warhead, in the process removing the parasite activity in vitro. This was perhaps unsurprising due to the high molecular mass and polar nature of these compounds, including the presence of a carboxylic acid moiety that was designed to mimic endogenous PISUB1 protein substrates by interacting with the basic S\textsuperscript{1} surface of the PISUB1 active-site cleft (16). Collectively, these features likely rendered the compounds poorly membrane penetrant.

By designing and synthesizing a range of analogs of peptidic boronic acids (SI Appendix, Table S1) and re-evaluating them in our fluorescence-based in vitro assay, we found that compound 3a, which contains a cyclopentane side chain, possessed a significantly improved IC\textsubscript{50} of ∼130 nM (Fig. 1). Replacement of the P1 Thr side chain with an Arg residue preserved the IC\textsubscript{50} value at ∼130 nM (3b). Combining the features of the α-ketoamide and 3a by adding back the P4 cyclopentane improved potency by a further ∼13-fold, leading to the low nanomolar IC\textsubscript{50} compound 3b (Table 1). This compound was selected for further structural optimization.

To examine the importance of the stereochemistry of the aminoalcohol moiety of the P1 side chain, we used the same assay to evaluate the relative potency of compounds with different stereochemistry at the P1 position. While compound 3c, which contains a D-amino acid, was more than sixfold less growth inhibitory than compound 3b, the key feature was that while all the compounds inhibited parasite proliferation, with EC\textsubscript{50} values as low as 1.8 μM, there was a poor correlation between growth inhibition and the PISUB1 enzyme-inhibitory potency of the compounds (Table 1). In particular, the most potent inhibitor of PISUB1 enzymatic activity, compound 3e, was more than sixfold less growth inhibitory than compound 3b. We reasoned that the relative steric bulk was also important, and the L-amino acid in native substrates of PISUB1. Therefore, we investigated the stereochemistry of the α-ketoamide in the substrate-like manner, the P3 Thr side chain

Further work focused on enhancing the potency of the compound 3b structural template. Removal of the methyl side chain at the P1 position (compound 3d) reduced potency by eightfold. On the other hand, attempts to improve potency by exploring extended alkyl or phenyl substituents at the P1 side chain (compounds 3e, 3f, 3g, and 3h) met with only limited success, although compound 3e bearing a hydroxethyl substituent displayed a twofold increased potency over compound 3b. This appears to contradict earlier substrate specificity studies, which indicated a preference for the S1 subpocket of PISUB1 to accommodate polar sidechains (13). The observation may be explained by a preference of nucleophilic P1 side-chain residues to form cyclic boronic acids, preventing the polar hydroxyl group from engaging in interactions with the enzyme.

Fig. 1. Development of rationally designed peptidic PISUB1 inhibitors.

Consistent with the X-ray crystal structure of PISUB1, which includes its propeptide bound into the active-site groove of the catalytic domain in a substrate-like manner, the P3 Thr side chain
of the docked compounds 3b and 3e was observed to extend into solvent, with no significant contacts with the molecular surface of the PfSUB1 catalytic domain. Interestingly, however, in both docking poses we noticed potential for modifying and/or extending the P3 side chain (openness depicted in gray in the two-dimensional diagram) in order to promote hydrophobic interactions with the side-chain carbon atoms of Lys465 and Leu466 that line the side of the S3 pocket. In silico replacement of the P3 Thr with Val supported this, revealing potential hydrophobic interactions between the Val P3 side chain and the side chains of Leu466 and Lys465 (Fig. 2B).

In accord with this, we prepared compounds 3i and 3j in which the P3 Thr of compound 3b was replaced, respectively, with an Ala and Val side chain (Table 1). The new compounds showed slightly improved (3i; IC_{50} ~7.8 nM) or nearly twofold improved (3j; IC_{50} ~5.7 nM) potency relative to compound 3b in the in vitro PfSUB1 enzyme assay. Significantly, compounds 3i and 3j displayed ~10-fold improved growth inhibitory potency in the SYBR Green I parasite growth assay, likely due to their increased lipophilicity, which was expected to confer better membrane permeability.

Table 1. PfSUB1 enzyme inhibitory and parasite growth inhibitory potency of peptidic boronic acids

<table>
<thead>
<tr>
<th>Entry</th>
<th>Compound</th>
<th>Structure</th>
<th>IC_{50} (nM) (rPfSUB1)*</th>
<th>EC_{50} (µM) (parasite growth)†</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>3a</td>
<td><img src="image1" alt="Structure" /></td>
<td>69.4 ± 1.2</td>
<td>1.8 ± 0.6</td>
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<tr>
<td>2</td>
<td>3b</td>
<td><img src="image2" alt="Structure" /></td>
<td>9.3 ± 0.5</td>
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<tr>
<td>3</td>
<td>3c</td>
<td><img src="image3" alt="Structure" /></td>
<td>60.1 ± 2.1</td>
<td>18.4 ± 1.8</td>
</tr>
<tr>
<td>4</td>
<td>3d</td>
<td><img src="image4" alt="Structure" /></td>
<td>54.3 ± 1.1</td>
<td>1.9 ± 0.4</td>
</tr>
<tr>
<td>5</td>
<td>3e</td>
<td><img src="image5" alt="Structure" /></td>
<td>4.6 ± 0.1</td>
<td>15.0 ± 3.6</td>
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<tr>
<td>6</td>
<td>3f</td>
<td><img src="image6" alt="Structure" /></td>
<td>204.2 ± 7.5</td>
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</tr>
<tr>
<td>7</td>
<td>3g</td>
<td><img src="image7" alt="Structure" /></td>
<td>18.7 ± 1.3</td>
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</tr>
<tr>
<td>8</td>
<td>3h</td>
<td><img src="image8" alt="Structure" /></td>
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<td>N.D.</td>
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<tr>
<td>9</td>
<td>3i</td>
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<td>7.8 ± 0.3</td>
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<tr>
<td>10</td>
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<td><img src="image10" alt="Structure" /></td>
<td>5.7 ± 0.1</td>
<td>0.26 ± 0.06</td>
</tr>
</tbody>
</table>

*IC_{50} values were determined by quantifying inhibition of rPfSUB1-mediated proteolytic cleavage of a fluorogenic peptide substrate. Values are mean averages from at least three independent measurements ± SD.
†EC_{50} values were obtained by quantifying inhibition of P. falciparum growth in vitro over a period of 96 h (two erythrocytic growth cycles) using the DNA-binding fluorescent dye SYBR Green I to measure parasite replication (22). Values are mean averages from at least three independent measurements ± SD. N.D., not determined.
Fig. 2. Substrate-based development of peptidic boronic acid inhibitors of PFSUB1. (A) ICM-Pro best docking poses for the PFSUB1-inhibitory compounds 3b (ICM-Pro score −27), 3e (ICM-Pro score −32), and 3j (ICM-Pro score −34) in the active site of PFSUB1 (4LVN). The inhibitors are represented as colored balls and sticks. Hydrogen atoms are shown, while hydrogen bond interactions are indicated (dotted lines). The enzyme molecular surface is shown in transparent mode (green, hydrophobic; blue, hydrogen bond donor; and red, hydrogen bond acceptor). ICM docking score values below −32 (unitless) are considered good docking scores. (B) Corresponding two-dimensional interaction diagrams, with green shading for hydrophobic regions, blue shading for hydrogen bond acceptors, gray dashed arrows for hydrogen bonds (including length in Å), broken thick lines around ligand shape for accessible surfaces, and gray parabolas for large accessible surface areas. Interacting PFSUB1 residues are labeled and enclosed in oval shapes, the size of which varies depending on the degree of residue contribution. In all cases, the ligand boron atom is shown covalently bound to the active-site Ser606 Oγ through a boron ester bond, and positions of the S1 and S4 PFSUB1 active-site pockets are indicated. The P3 position is annotated in red in B.
Inhibition of *P. falciparum* Egress by Selective Peptidic Boronic Acids that Access PSUB1 in Intracellular Parasites. To determine their mode of action, the four most potent growth inhibitory compounds were next evaluated using very short-term cell-based assays focused on the narrow window within the asexual blood-stage lifecycle during which the parasite undergoes egress from host RBCs and invades into fresh cells. For this, *P. falciparum* cultures containing synchronous, highly mature schizonts were supplemented with compounds 3b, 3e, 3i, and 3j at a range of dilutions, then allowed to undergo egress and invasion for just 4 h in the continued presence of the compounds, before assessing formation of newly invaded “ring” stage parasites by flow cytometry. This confirmed a dose-dependent inhibitory effect on the transition from schizont to ring stage, with the relatively lipophilic compounds 3i and 3j displaying similar EC50 values that were significantly lower than those of 3b and 3e (Fig. 3A). Microscopic examination of the cultures revealed schizonts arrested by compounds 3i and 3j, confirming inhibition of schizont rupture. Importantly, the arrested schizonts were morphologically indistinguishable from those arrested by the reversible PKG inhibitor 4-[7-[(dimethylamino)methyl]-2-(4-fluorophenyl)imidazol[1,2-\(a\)]pyridine-3-yl]pyrimidin-2-amine (C2), appearing as segmented forms trapped within an apparently intact PVM and RBC membrane (Fig. 3B). This egress-arrest phenotype is similar to that obtained by genetic disruption of PSUB1 and was clearly different from that following arrest by the cysteine protease inhibitor E64 (Fig. 3B), which does not inhibit PSUB1 directly but which blocks a cysteine protease-dependent step in egress following SUB1 discharge and PVM rupture (12).

Examination of the inhibitory activity of compound 3j against the mammalian trypsin-family serine proteases trypsin, chymotrypsin, and elastase revealed a high degree of selectivity for PSUB1 (10X100-fold; SI Appendix, Figs. S1–S3), encouraging us to focus subsequent work on this compound. To directly visualize the inhibitory effects of compound 3j on parasite egress and to examine the reversibility of inhibition, we used live time-lapse video microscopy to observe the behavior of schizonts exposed to the compound for just 1 h immediately prior to egress. For this, we used a transgenic parasite line expressing a PVM protein (EXP2) fused with the green fluorescent protein mNeon Green, facilitating real-time visualization of PVM integrity as previously reported by Glushakova and colleagues (23). As shown in Fig. 3A and Movie S1, this clearly demonstrated significant inhibition of PVM rupture and egress in parasites treated with 3j, with no signs of egress even 30 min following washout of the compound. Importantly, 3j-treated parasites remained viable, as shown by their continued capacity to incorporate the vital mitochondrial dye MitoTracker Red CMXRos (24) (SI Appendix, Fig. S4), but showed no signs of the PVM rounding and other morphological changes that typically precede egress (23, 25), indicating a complete and selective block in the egress pathway. These egress-associated transitions were also absent from PSUB1-null parasites (12), indicating that the effects of 3j closely mimic genetic disruption of PSUB1. Further extended incubation of the treated, washed schizonts with fresh RBCs resulted in only very limited appearance of new ring stage parasites (Fig. 4B). This confirmed that even short-term treatment with compound 3j could dramatically impede parasite escape from the host RBC and that the egress inhibition over these timescales was effectively irreversible.

These results suggested that compound 3j can access and inhibit PSUB1 in an intracellular location (i.e., within the intracytoplasmic parasite, or in the PV, or both). To seek unambiguous confirmation that PSUB1 is the intracellular target of compound 3j, we examined the effects of the compound on the PSUB1-mediated proteolytic processing of the established endogenous PSUB1 substrate SERA5, an abundant parasite PV protein that only becomes accessible to cleavage upon discharge of PSUB1 into the PV in the minutes leading up to egress and is then released in a processed form into culture supernatants (6, 11, 12). As shown in Fig. 4C, treatment with compound 3j reproducibly prevented proteolytic processing and release of SERA5 into culture supernatants in a dose-dependent manner, even following compound washout. Crucially, at higher concentrations of the drug where egress and release of processed SERA5 was completely blocked, no intracellular...
processing of SERA5 was evident in the intact egress-arrested schizonts. Quantitation of egress in these same washout assays by measuring release of residual hemoglobin from the rupturing schizonts showed an EC50 of ~2 μM (Fig. 4D), similar to the EC50 value for new ring generation previously determined by flow cytometry (Fig. 3A). It was concluded that compound 3j prevents egress and parasite proliferation through direct inhibition of intracellular PfSUB1.

An Optimized Membrane-Penetrant Peptidic Boronic Acid Displays Time-Dependent, Slowly Reversible Binding Kinetics to PfSUB1. Boronic acids form reversible covalent bonds with serine and threonine proteases (26). Inhibition is generally time dependent, and the covalent nature of the binding can result in relatively long target occupancy times despite the reversibility of the bond. That this might be the case with compound 3j binding to PfSUB1 was initially suggested by our washout experiments (Fig. 4), which showed that egress inhibition by compound 3j following washout was much longer lasting than the rapidly reversible egress inhibition mediated by C2. To analyze the kinetic characteristics of the interaction between 3j and rPfSUB1, we used progress curve analysis to continuously monitor rPfSUB1-mediated cleavage of a fluorogenic substrate in the presence of a range of concentrations of 3j. As shown in SI Appendix, Fig. S5, under conditions where substrate cleavage in the absence of inhibitor (control reaction) displayed a linear relationship with time, indicating negligible substrate depletion, progress curves in the presence of compound 3j became progressively nonlinear, characteristic of slow-binding (time-dependent) inhibition. Under such conditions, fit of the progress curves by nonlinear regression to Eq. 1 (see Materials and Methods) allows determination of koff, the pseudo first-order rate constant for onset of inhibition. The koff value was 3.6 × 10^{-4} s^{-1}, allowing calculation of an apparent equilibrium inhibition constant K_{eq} (k_{on}/k_{off}) of 1.0 nM. It was concluded that compound 3j is a potent, slowly reversible inhibitor of PfSUB1, completely consistent with the washout data.

Discussion

Prior to parasite egress from the confines of its host RBC, SUB1 is stored in membrane-bound merozoite secretory organelles called exonemes before its discharge into the PV lumen minutes before egress to encounter its endogenous substrates. As a result, in order to gain access to the intracellular enzyme prior to egress...
substrate cleavage, exogenously applied inhibitory compounds likely need to cross at least two and as many as four distinct biological membranes: the RBC membrane, the PVM, the parasite plasma membrane, and the exoneme membrane (Fig. 5). This poses particular challenges for the design of substrate-based inhibitors. In the case of covalent modifying compounds, such as those described here, access to the exoneme-resident enzyme could potentially allow inactivation of the stored SUB1 long before its PKG-regulated discharge into the PV. In this work, we did not determine the intracellular site of PISUB1 inhibition, so we cannot state whether inhibition took place within the PV, or the exonemes, or both. Regardless, by gradual optimization of the structure and lipophilicity of our compounds we have now successfully developed potent PISUB1-inhibitory compounds that can functionally inactivate PISUB1 within intact, parasite-infected RBCs and block egress.

Our conclusion that the intracellular inhibition of PISUB1 mediated by compound 3j is directly and causally responsible for the observed block in egress is most clearly supported by the phenotype of the arrested schizonts, which was indistinguishable from that resulting from conditional genetic disruption of the PISUB1 (12) or PKG gene (27), or following treatment with the PKG inhibitor C2, with no signs of the morphological changes that typically precede egress such as PVM rounding or PV rupture. We cannot rule out the possibility of effects on other parasite enzymes at the concentrations used to obtain complete egress inhibition, even in the short-term assays designed to focus on the short window of the parasite life cycle over which egress occurs. However, we consider off-target effects unlikely given that no other parasite serine protease has been implicated in egress; the only two other subtilisin-like enzymes expressed in the parasite, SUB2 and SUB3, are respectively dispensable for egress (28) or nonessential in blood stages (29). The ~10-fold higher potency of 3j in the long-term SYBR Green–based parasite growth assay (EC50 0.26 μM; Table 1) may be a result of the fact that this assay captures the combined effects of egress inhibition over the course of two erythrocytic cycles in the continuous presence of the drug, although again off-target effects cannot be formally ruled out. We anticipate that further optimization of the PISUB1-inhibitory potency and membrane permeability of 3j is highly feasible. Work is already underway to determine the atomic structure of the 3j-PISUB1 complex to facilitate structure-based inhibitor improvement.

Peptidic boronic acids have long-established therapeutic potential, as best exemplified by the widespread clinical use for multiple myeloma of the proteasome inhibitors bortezomib (Velcade) and ixazomib, the latter of which is orally bioavailable in its citric acid form, Ninlaro. The clinical success of these compounds is in part due to the long drug target residence times that can be obtained with slowly reversible covalent inhibitors. Target binding by boronic acid protease inhibitors is generally time dependent, perhaps further explaining the differences in potency we observed between the long-term and short-term cellular assays with compounds 3i and 3j. However, our wash-out experiments with compound 3j suggest that, once bound to PISUB1 in the exonemes and/or PV, it takes at least 30 min for the level of target engagement to fall below a threshold that allows successful egress. Examination of the capacity of schizonts treated with saturating levels of 3j to productively egress and form new rings following compound washout showed that the egress block under these conditions was effectively irreversible.

An alternative explanation for this apparently irreversible inhibition of egress by 3j is that the inhibitor is not easily washed out due to its accumulation in the parasite (or infected RBC) at high concentrations. We cannot formally rule out this possibility. However, our ex vivo kinetic analysis of the inhibition of rPISUB1 by compound 3j fully supports a slow off rate, with an estimated bound half-life (t1/2(off)) of ~30 min, very similar to that of the interaction between bortezomib and the β5 chymotrypsin-like subunit of the human proteasome (30–32). Even if cellular accumulation does contribute to the prolonged egress inhibition exerted by 3j, this is only likely to favor efficacy; indeed, intracellular accumulation is an important component of the mode of action of the important antimalarial 4-aminooquinoline chloroquine (33). While peptide-based drug development can present challenges for in vivo applications due to metabolic instability, covalent compounds can be effective even with relatively short plasma half-lives, since target residence time can be longer than plasma half-life. Peptidyl boronic acids can anyway have excellent pharmacodynamic properties; for example, the terminal half-life of azamib is ~5.5 h, a fact that allows weekly dosing of patients for treatment of multiple myeloma; ref. 34), which is nearly five times the duration of the P. falciparum asexual blood-stage lifecycle.

SUB1 has an unusual substrate specificity, which differs subtly between different Plasmodium species, suggesting that the enzyme and its multiple cognate parasite substrates have coevolved to ensure optimal cleavage efficiency (13). As a result, inhibitors of PISUB1 are unlikely to show similar potency against SUB1 orthologs from rodent malaria parasite species such as Plasmodium berghei, making these parasite species unsuitable as model systems for assessing the in vivo efficacy of our compounds. Importantly, SUB1 also lacks structural resemblance to any known human serine protease (16), reducing the likelihood of substrate-based SUB1 inhibitors displaying toxicity due to off-target activity against host enzymes. In support of this, we found here that 3j is only poorly potent against the mammalian serine proteases examined. Toxicity can be especially problematic where long-term or life-long treatment regimens are required due to chronic infection (e.g., with HIV). However, malaria is an acute disease, and long-term therapeutic regimens are rare; indeed, current standard treatments for uncomplicated falciparum malaria are just 3 d long. Since SUB1 plays an essential role in the development and release of exoerythrocytic (liver-stage) merozoites that initiate blood-stage infection (14, 15), medicines based on SUB1 inhibitors have prophylactic as well as therapeutic potential. Optimized SUB1 inhibitors could
also potentially be combined with inhibitors of other essential enzymes in the erythropathway, including PKG (35–37) and the SUB1 aspartic protease maturase plasminogen X (38–41). Such combinations could yield additive or synergistic enhancement of potency and decrease opportunities to select for drug resistance.

In conclusion, we have produced substrate-based peptidic boronic acids that block asexual blood-stage of *P. falciparum* pro- liferation through direct, effectively irreversible inhibition of intracellular PISUB1. Further investigation of the pharmacoki-netic properties and structure-based improvement of these compounds has the potential to generate compounds suitable for preclinical trials in animal models of Malaria.

**Materials and Methods**

*P. falciparum Maintenance and Manipulation.* Asexual blood stages of *P. falciparum* (clones 3D7 and B11) (42) were routinely maintained at 37 °C in human erythrocytes at 1 to 4% hematocrit in RPMI 1640 containing Albumax II (Thermo Scientific) supplemented with 2 mM L-glutamine in a low oxygen atmosphere using standard procedures (43). Human blood was obtained from anonymized donors through the UK National Blood and Transplant service and was used within 2 wk of receipt. No ethical approval is required for its use. For synchronization, mature schizont-stage parasites were isolated on cushions of 70% (volume/vol/vol) Percoll (GE Healthcare) adjusted to isotonicity as described (43). Routine microscopic examination of parasite growth was performed by fixing air-dried thin blood films with 100% methanol before staining with 10% Giemsa stain (VWR International) in 6.7 mM phosphate buffer, pH 7.1.

Generation of the B11-EXPO2-mNeonGreen line was achieved by fusing mNeonGreen to the endogenous C terminus of EXPO2 using Cas9-mediated gene editing, following the methods of ref. 23. A pair of guide RNAs were designed targeting a region toward the 3′ end of the EXPO locus (oligo 1: ATTGATATT- ATGATATAGCTCA, oligo 2: AAATTTAGATCTTAGCTATAAT (Sigma). This oligonucleotide pair was annealed with T4 PNK ligase (New England Biolabs) and ligated with T4 ligase (New England Biolabs) into the U6 cassette of the Cas9 vector (pD2Cas9-U6-HDHFR) previously digested with BsaB1-HF (New England Biolabs). The plasmid was propagated under ampicillin selection in *Escherichia coli* (England Biolabs). The plasmid was propagated under ampicillin selection in *Escherichia coli* (England Biolabs). The plasmid was propagated under ampicillin selection in *Escherichia coli* (England Biolabs) and was used within 2 wk of receipt. No ethical approval is required for its use. For synchronization, mature schizont-stage parasites were isolated on cushions of 70% (volume/vol/vol) Percoll (GE Healthcare) adjusted to isotonicity as described (43). Routine microscopic examination of parasite growth was performed by fixing air-dried thin blood films with 100% methanol before staining with 10% Giemsa stain (WXR International) in 6.7 mM phosphate buffer, pH 7.1.

Parasite Growth, Egress, and Invasion Assays. The impact of the peptidic boronic acids on replication of asexual blood-stage of *P. falciparum* (clone 3D7) was assessed using a SYBR Green I assay, essentially as described by Smilkstein et al. (22). Synchronization was performed by dissolving in dimethyl sulfoxide (DMSO) at concentrations ranging from 4 mM to 5 μM) were added in triplicate to wells of flat bottomed, 96-well microtitre plates (1 μM per well). Wells were then supplemented with 100 μL per well of a *P. falciparum* parasite culture at 0.1% parasitaemia, 1% hematocrit. Each assay plate also included DMSO-only control wells (1% vol/vol), as well as additional control wells containing uninfected RBCS only. Plates were incubated in sealed, humidified gassed chambers at 37 °C for 96 h to allow the parasites to undergo two entire cycles of erythrocytic growth. Wells were then supplemented with 100 μL per well of a *P. falciparum* parasite culture at 0.1% parasitaemia, 1% hematocrit. Each assay plate also included DMSO-only control wells (1% vol/vol), as well as additional control wells containing uninfected RBCs only. Plates were incubated in sealed, humidified gassed chambers at 37 °C for 96 h to allow the parasites to undergo two entire cycles of erythrocytic growth. Wells were then supplemented with 100 μL 1:5,000 dilution of stock SYBR Green I (Life Technologies, catalog no. S7563) diluted in 20 mM Tris HCl pH 7.5, 5 mM EDTA, 0.008% (weight/vol) saponin, 0.08% (vol/vol) Triton x100. Plates were agitated to mix, incubated for a further 1 h in the dark at room temperature, then 150 μL samples from each well transferred to a fresh white microcell plate and fluorescence quantified using a SpectraMax M5e plate reader and SoftMax Pro 6.3 software (Ex 485 nm/Em 530 nm). IC50 values were determined from dose–response curves obtained after subtracting background fluorescence values (obtained from the RBC-only wells) for all experimental readings.

Short-term egress, invasion, and washout assays were performed essentially as described previously (27, 28, 42). Briefly, highly synchronous mature Percoll-enriched schizonts with or without added fresh RBCs (~5% parasitaemia final) were incubated with compounds under test or vehicle only (DMSO, 1% vol/vol). For washout assays, schizonts were treated with C2 or various concentrations of inhibitor 3 j for 1 to 4 h, then washed extensively (at least four times) prior to addition to fresh RBCs where required. After incubation at 37 °C for just 1 to 4 h (or overnight for invasion assays) to allow schizont rupture, cells were pelleted. Clarified culture supernatants were assessed for the absorbance of hemoglobin release (a measure of schizont rupture) by absorption spectroscopy at 415 nm as described previously (28) or analyzed by Western blot using antibodies against SERAS (7, 11). To quantify generation of new rings, samples of the cultures were fixed with 4% paraformaldehyde/0.02% glutaraldehyde and stained with SYBR Green I (Life Technologies) and then analyzed by flow cytometry on a BD FACSVerse using BD FACSuite software. Data were analyzed using FlowJo software. All cultures were also routinely analyzed by microscopic examination of Giemsa-stained thin films to visually assess parasite morphology.

**Protease Inhibition Assays:** IC50 Calculations and Progress Curve Kinetics. Proteolytic activity of PISUB1 was quantified at room temperature by monitoring cleavage of the peptidic fluorescent substrate SERA411F-6R12 (Ac-CXITAQGDFEDE-OC possessing tetramethylrhodamine labeling of both cysteine residues) (13). Fluorescence of this peptide is quenched by rhodamine dimerization in the intact substrate but increases upon cleavage at the internal Q-D bond. Chymotrypsin-treated PISUB1 (expressed and purified as described previously (16)) was stored at −80 °C as a 228 U/mL stock in 20 mM Tris-HCl pH 8.2, 150 mM NaCl, 10% glycerol, and diluted for use (1:500 or 1:600) in reaction buffer (20 mM Tris–HCl pH 8.2, 150 mM NaCl, 12 mM CaCl2, 25 mM CHAPS). Peptidic boronic acid inhibitors were dissolved in 100% DMSO at 10 or 20 μM, then further diluted in DMSO to generate stock solutions ranging from 500 to 0.01 μM and then used diluted 1:100 in the enzyme reactions. All reactions were performed in wells of white 96-well microcell plates (Nunc); 50 μL diluted PISUB1 was preincubated for 5 min with 1 μL each of the serially diluted boronic acid inhibitors, followed by addition of 50 μL substrate solution (0.1 μM final). Subsequent fluorescence increase was continuously monitored using a SpectraMax M5e plate reader and SoftMax Pro 6.3 software, with readings taken every 5 min for 60 min using excitation and emission values of 552 and 580 nm, respectively. Initial rates were calculated over the first 25 min of the assay, during which progress curves were linear, and IC50 values were calculated with GraphPad Prism 8.0 using the nonlinear regression, [inhibitor] versus response, variable slope (four parameters). All experiments were performed in duplicate.

Details of the methodology used to evaluate the effects of compound 3 j on the proteolytic activity of the mammalian serine proteases trypsin, chymotrypsin, and elastase are provided in SI Appendix.

**Progress Curve Kinetic Analysis of Compound 3 j.** Progress curves of SERA411F-6R12 cleavage by PISUB1 were acquired at seven concentrations of inhibitor compound 3 j over a period of 35 min, during which fluorescence increases in the absence of inhibitor were linear, and IC50 values were calculated with GraphPad Prism 8.0 using the time-dependent inhibition equation:

\[
P(t) = Vs \ast t + (Vi - Vs) \ast k_{on} \ast (1 - \exp\left(-k_{off} \ast t\right))
\]

In the equation, *Vi* is the initial velocity, *Vs* is the final steady-state velocity, and *koff* reflects the observed pseudo-first-order rate of inactivation. The obtained *koff* values were plotted against compound concentration using a linear least squares fit. All statistical analysis was carried out using GraphPad Prism 8.

**Covalent Docking.** Flexible covalent docking of peptidyl boronic acid compounds into the active site of PISUB1 (Protein Data Bank: 4LVN) was performed using the Internal Coordinate Mechanics software (ICM-Pro) package version
3.9-10cMacOSX (Molsoft LLC). The inhibitors were drawn using the ICM chemistry molecular editor and compiled into an sdf docking table. After adding hydrogen atoms to the structure, the C-terminal region of the SUB1 propeptide (P4 to P1 positions that occupy the SUB1 active site) was used to define boundaries within the enzyme active site for the docking procedure and then removed from the active site along with all water molecules prior to docking. The catalytic histidine (His428, \(\text{Ni}^3\)) was protonated as part of the catalytic process, resulting from covalent binding of the boron atom ligand to the active Ser (Ser606 O\(_\text{e}\)). The boronic acid covalent mechanism was selected from the ICM program reactions list. Potential energy maps of the SUB1 receptor pocket and docking preferences were set up using the program default parameters. Energy terms were based on the all-atom vacuum force field ECEPP/2, and conformational sampling was based on the biased Monte Carlo procedure (45). Four independent docking runs were performed per compound, with a length of simulation (throughness) varied from three to four and the selection of two docking poses. Ligands were ranked according to their ICM energetics (ICM score, unitless), which weights the internal force-field energy of the ligand combined with other ligand-receptor energy parameters.


28. C. R. Collins et al., The malaria parasite sheddase SUB2 governs host red blood cell membrane sealing at Invasion. eLife 9, e61121 (2020).


