



# Airway eosinophils in older teenagers with outgrown preschool wheeze: a pilot study

*To the Editor,*

Preschool wheezing affects one-third of all children growing up in the UK [1]. It varies in clinical presentation and severity, and there is evidence to suggest the co-existence of different wheeze phenotypes [2], some of which have been associated with adult asthma and chronic obstructive pulmonary disease [3]. Using data from the Leicester Respiratory Cohort studies [4], we have previously applied objective data-driven methods to distinguish three distinct preschool wheeze phenotypes: “atopic” and “non-atopic” persistent wheeze (PW) and transient viral wheeze (TVW) [5]. In children with PW, attacks of wheeze with and without colds were observed both at preschool age (0–5 years) and when followed-up at early school-age (4–8 years). Children with TVW, triggered predominantly by colds, had symptoms at age 0–5 years but not at age 4–8 years. Prognosis 5 years later (aged 8–13 years) was markedly better in children with TVW compared to the two PW phenotypes [6].

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Persistent wheeze in older children and adults, diagnosed as asthma, is known to be associated with eosinophilic airway inflammation [7, 8]. Whether eosinophilic inflammation plays a pathophysiological role in TVW remains unclear, due to difficulties in obtaining sputum from preschool children [9]. Little is known about the natural course of such inflammatory changes after the symptoms have resolved. A study of young schoolchildren (aged <10 years) with outgrown “viral” and “atopic” wheeze found low-grade eosinophilic airway inflammation in children with previous atopic wheeze only [10]. Although many children become asymptomatic at school-age, asthma frequently recurs in adulthood, often after long intervals [11]. It is unclear whether this represents a flaring up of a persisting subclinical inflammation, or a *de novo* development of disease.

The aim of our pilot study was to test whether airway eosinophils are present into adulthood in individuals with outgrown childhood wheezing. We also wanted to investigate if it was only present in individuals with previous PW, or also in adolescents with former TVW. Based on previous findings [10], we expected to find airway eosinophils in teenagers with previous PW only.

We used a nested case-control study design. Subjects were sampled from the second Leicester respiratory cohort study, where parents had completed respiratory questionnaires at six time points since 1998. In 2012, when the children were aged 15–16 years, we approached randomly sampled subjects from three groups: individuals with outgrown PW, individuals with outgrown TVW, and individuals who had never reported wheeze. For this pilot study we combined children with atopic and non-atopic PW.

Groups were defined based on symptoms reported in 1998, 2001 and 2003–2010. Selection criteria were: 1) PW group, current wheeze and shortness of breath OR >3 attacks in the past 12 months OR hospitalisation/emergency healthcare visit for wheeze over several years from 1998 to 2001 with no reported wheeze at the latest survey (2003, 2006, 2008 or 2010); and 2) TVW group, wheezing reported with colds in 1998 but no current wheeze in 2001 or at the latest survey. Healthy controls (HC) had no wheezing reported at any survey.

163 subjects, selected at random, received an invitation letter, which was followed by one reminder in cases of nonresponse. This constituted a small subset of the 1739 cohort members (63 PW, 101 TVW and 1575 HC) who fitted the eligibility criteria. 35 teenagers agreed to participate, but three later withdrew.

32 teenagers attended our lung function laboratory between September 2012 and April 2013, of whom three were excluded because of a reported wheezing episode since the last questionnaire survey. All subjects were free from respiratory infections. In total, data from PW n=7, TVW n=12 and HC n=10 were obtained. The study participants were representative for all eligible subjects for cohort for: age, sex, gestational age, birth weight, ethnicity, preschool respiratory symptoms, personal and family history of atopy, pre- and postnatal parental smoking, and socio-economic data (tables S1–S3).

The East Midlands Research Ethics Committee approved all aspects of this study (Ref 09/H0403/92) and informed written consent was obtained from all participants.

We measured forced expiratory volume in 1 s (FEV<sub>1</sub>) and forced vital capacity (FVC) in accordance with America Thoracic Society and British Thoracic Society/Association for Respiratory Technology and Physiology guidelines, using a PC-based spirometer (Vitalograph Pneumotra with Spirotrac IV software; Vitalograph Ltd., UK). Exhaled nitric oxide fraction (F<sub>e</sub>NO) was measured using a portable electrochemical analyser (NIOX MINO, Aerocrine AB, Solna, Sweden). Sputum was induced by hypertonic nebulised saline delivered by high flow nebuliser (Omron UltraAir U17, Omron, the Netherlands) using our standard protocol and processed as previously described [12]. The sputum differential cell count was obtained by counting 400 sequential inflammatory cells on a cytospin slide by an experienced technician blinded to the clinical status. Data were analysed using Prism 6.0 (GraphPad Software; La Jolla, CA, USA). Demographic, sputum inflammatory cell, lung function and F<sub>e</sub>NO data are presented in the table as medians and interquartile ranges (IQR). For the principal outcome measures comparisons between the three groups were made using Kruskal–Wallis testing. Where this test was significant further group differences were explored using the Mann-Whitney U test and 95% confidence intervals between medians were calculated (table 1).

The three groups (PW, TVW and HC) did not differ with respect to age, sex or atopic status. The median percentage of squamous cells in sputum was low in all three groups suggesting good sample quality. In all children, with the exception of one teenager with outgrown TVW, airway eosinophils were below the cut-off point of 2.5% reported in subjects with symptomatic asthma [7]. Nonetheless, sputum eosinophils were present in significantly higher numbers in outgrown PW (0.3%) and TVW (0.8%) compared to controls (0.0%) (figure 1).

Subjects with previous PW and TVW both had significantly lower FEV<sub>1</sub> compared to healthy controls (figure 1). None of the participants had a significant improvement in FEV<sub>1</sub> (>12%) after the use of bronchodilators in keeping with asthma remission. This is consistent with previous reports and longitudinal lung function measurements. Children with transient early wheeze (similar to our TVW group) in the Tucson [13] and Perth [14] birth cohorts had reduced lung function shortly after birth, and continued to have reduced lung function compared to children who never wheezed at school age [13]. Moreover, the Oslo Environment and Childhood Asthma prospective birth cohort study also reported reduced lung function and increased bronchial responsiveness in teenagers aged 16 years with previous doctor diagnosed preschool wheeze. Unfortunately, the study did not distinguish PW and TVW [15].

TABLE 1 Subject demographics, baseline lung function and sputum analysis

	Persistent wheeze	Transient viral wheeze	Healthy
<b>Subjects</b>	7	12	10
<b>Male</b>	3 (43)	4 (33)	4 (40)
<b>Age years</b>	15 (15–16)	16 (16–16)	16 (15–16)
<b>Age at first questionnaire (1998) years</b>	1.3±0.1	1.5±0.3	1.4±0.2
<b>Age at second questionnaire (2001) years</b>	4.6±0.1	4.8±0.3	4.7±0.3
<b>Atopy<sup>#</sup></b>	3 (43)	6 (50)	5 (50)
<b>FEV<sub>1</sub> z-score</b>	-0.95 <sup>¶</sup> [-1.35– -0.08] (0.76–2.3)	-0.39 <sup>¶</sup> [-1.13–0.29] (0.42–2.08)	0.69 (0.25–1.29)
<b>Subjects</b>			9
<b>p value</b>	0.001	0.005	
<b>FVC z-score</b>	-0.8 <sup>¶</sup> [-1.3–0.1] (1.7–36.7)	-0.5 <sup>#</sup> [-1.1–0.0] (4.9–32.7)	1.0 (0.2–1.4)
<b>Subjects</b>		10	9
<b>p value</b>	0.03	0.013	
<b>FEV<sub>1</sub>/FVC %</b>	85.0 (83.0–89.0)	89.5 (88.0–92.3)	88.0 (80.0–91.3)
<b>Subjects</b>		10	9
<b>F<sub>e</sub>NO ppb</b>	18 (17–22)	13 (9–26)	17.5 (15.75–23.25)
<b>Total sputum leukocyte count ×10<sup>6</sup> cells·g<sup>-1</sup></b>	4.2 <sup>¶</sup> [1.6–4.9] (-4.2– -0.59)	1.0* (0.5; 2.0) [-3.9– -0.09]	0.8 (0.5–1.1)
<b>p value</b>	0.009	0.035	
<b>Sputum eosinophils %</b>	0.25 <sup>¶</sup> (0.25–0.5) [-0.5–0.0]	0.75 <sup>¶</sup> (0.21–1.23) [-1.2– -0.2]	0.0 (0.0–0.06)
<b>p value</b>	0.011	0.0025	
<b>Sputum neutrophils %</b>	23.5 (14–44.5)	27.1 (22–83.9)	55.8 (8.7–93.6)
<b>Sputum macrophages %</b>	74.0 (49.0–84.0)	54.0 (14.8–73.4)	33.0 (4.6–77.1)
<b>Sputum squamous cells %</b>	7.0 (4.1–11.4)	17.2 (5.3–26.1)	15.2 (13.7–26.3)

Data are presented as n, n(%), median (interquartile range (IQR)), mean±sd or median (IQR) (95% CI). FEV<sub>1</sub>: forced expiratory volume in 1 s; FVC: forced vital capacity; F<sub>e</sub>NO: exhaled nitric oxide fraction; #: atopic status was defined as having reported hay fever in the last 12 months and/or any history of eczema; ¶: significant p<0.05 compared to “healthy” using Mann-Whitney U test; \*: significant p<0.05 compared to “persistent wheeze” using Mann-Whitney U test.



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