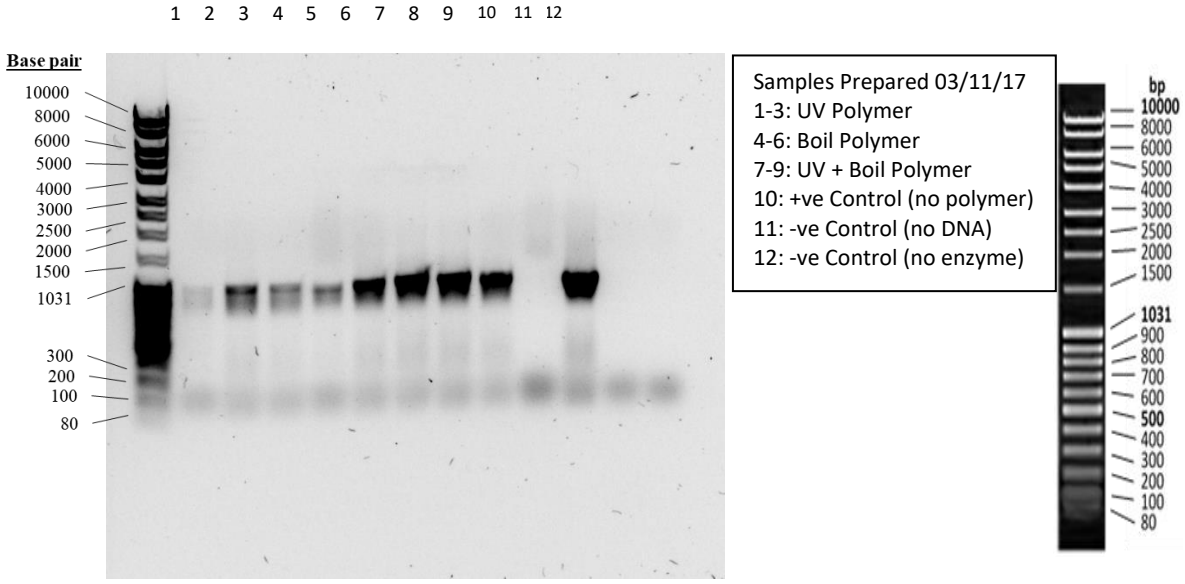
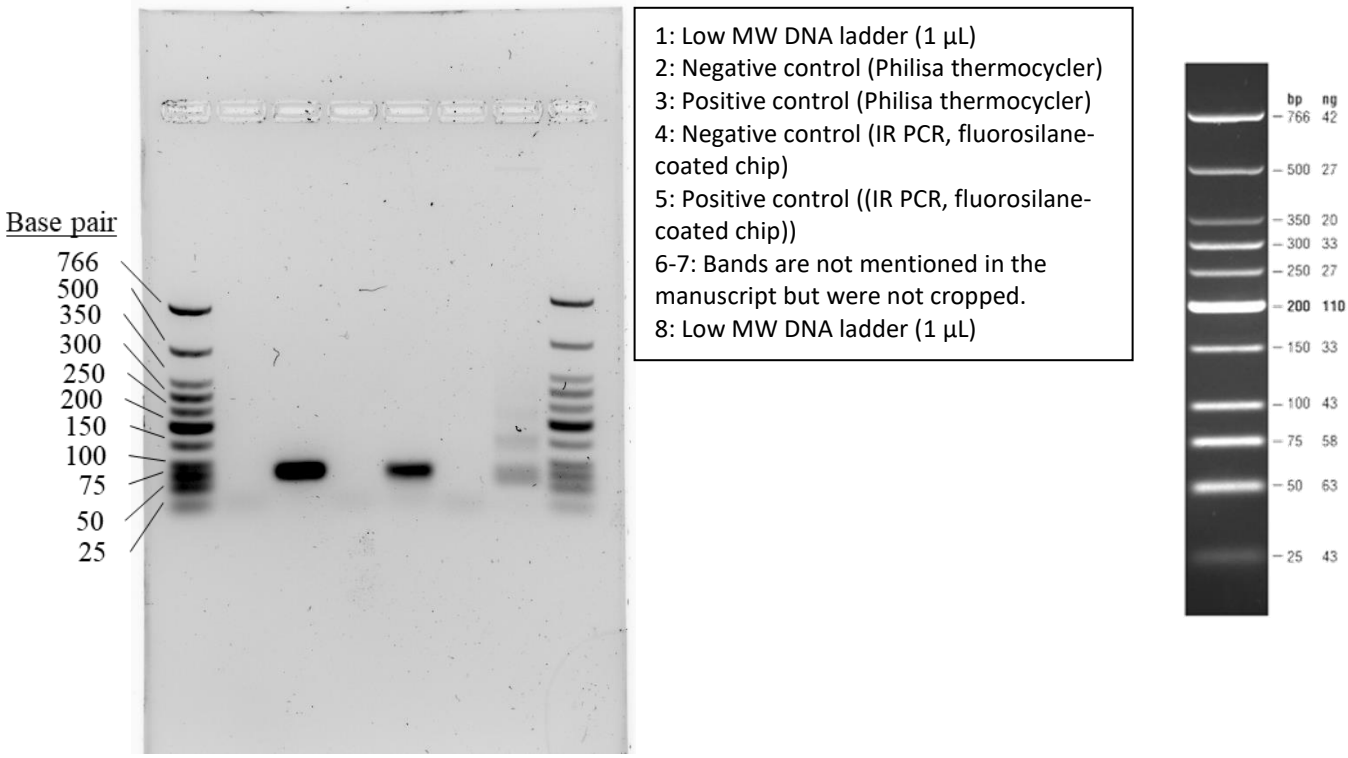
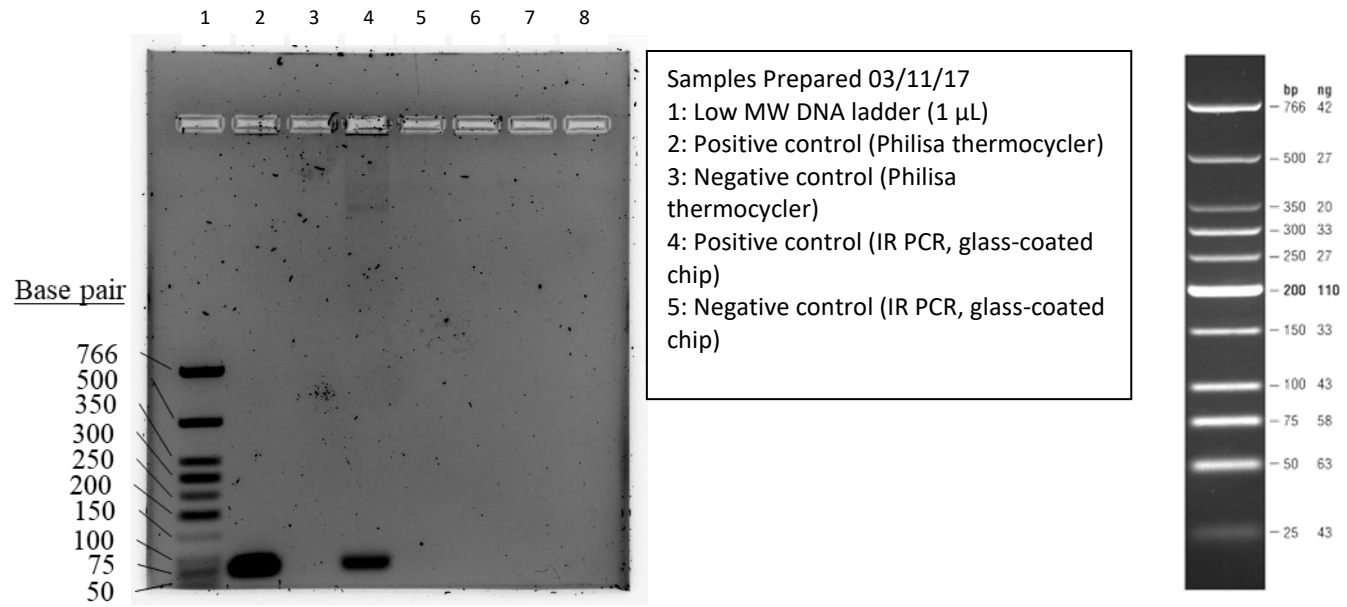
Raw agarose gel images presented in Figure 5 and 8 of the revised manuscript: **Fabrication routes via projection stereolithography for 3D-printing of microfluidic geometries for nucleic acid amplification,** submitted in PLOS one. The brightness of the images is not adjusted.



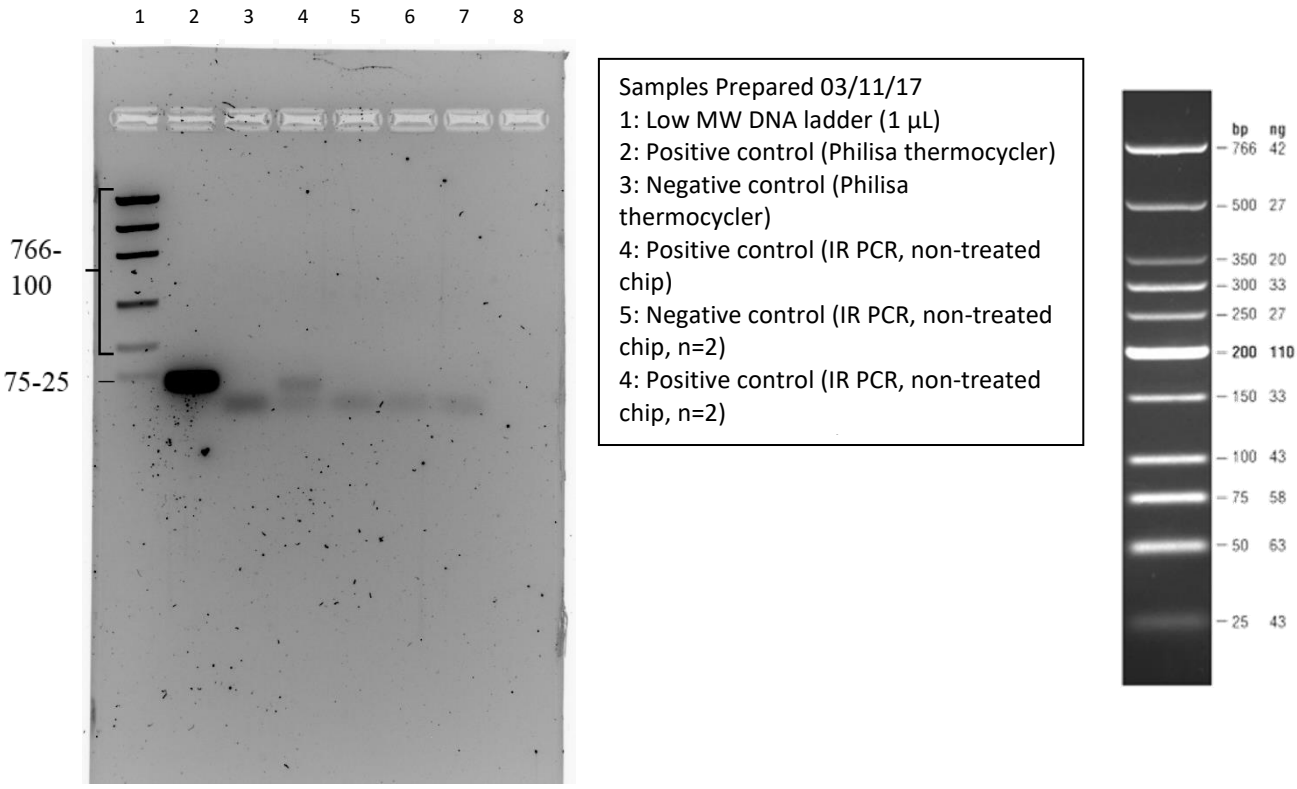
S1 Figure 5-b Twelve PCR reactions to amplify 1,000 base pair long sequence from plasmid template performed in the presence of differently post-processed resin printed specimens (9 reactions plus three controls). The mixtures were stained with Sybr Safe and loaded in numerical order as labelled on a 1% agarose gel prepared in TAE buffer and ran for 35 minutes, 90V. The gel was imaged in a BioRad UV transilluminator.



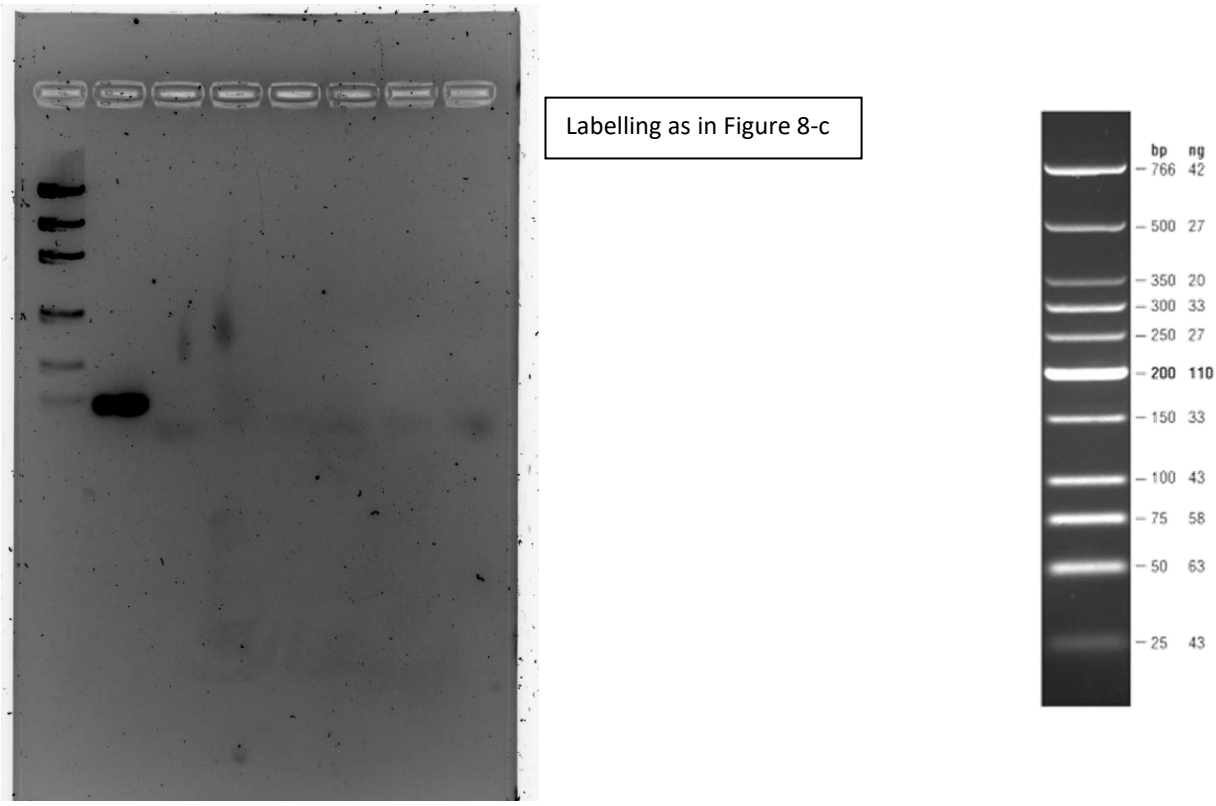
S1 Figure 8-a A negative and positive control 2-step PCR to amplify a 75 bp sequence from subgenomic DNA ran on Philisa commercial thermocycler and on a fluorosilane-coated 3D printed chip on the IR PCR thermocycler. The amplicons were stained with Sybr Safe, loaded in numerical order as labelled on a 2% agarose gel and run for 45 minutes at 90V. The gel was imaged in a BioRad UV transilluminator.



S1 Figure 8-b A positive and negative control 2-step PCR to amplify a 75 bp sequence from subgenomic DNA ran on Philisa commercial thermocycler and on a glass-coated 3D printed chip on the IR PCR thermocycler. The amplicons were stained with Sybr Safe DNA stain and loaded in numerical order as labelled on a 2% agarose gel and run for 45 minutes at 90V. The gel was imaged in a BioRad UV transilluminator.



S1 Figure 8-c A positive and negative control 2-step PCR to amplify a 75 bp sequence from subgenomic DNA ran on Philisa commercial thermocycler and on a non-treated fully post-processed 3D printed chip on the IR PCR thermocycler. The amplicons were stained with Sybr Safe and loaded in numerical order as labelled on a 2% agarose gel and run for 45 minutes at 90V. The gel was imaged in a BioRad UV transilluminator.



S1 Figure 8-c A The gel presented in Figure 8-c ran under the same conditions for an additional 5 minutes. The low MW DNA ladder bands start to separate from each other, however the amplicon of the reaction performed on the non-treated chip has been already fully absorbed. The run was not complete as the result conveys the message of intense reactivity of the SLA polymer and the necessity of surface optimization for printing of nucleic acid testing devices.