

Background

Zevatars

- Zevatars involve implanting fresh or cryopreserved patient tumor pieces from biopsy samples into zebrafish and screening for best drug response before treating patients in the clinic
- The main advantage of using zevatars is rapid drug response screening and ability to assess tumor proliferation, angiogenesis, and metastasis by imaging and finally by implanting tumor pieces takes into account the tumor microenvironment as compared to current methods involving injecting dissociated tumor cells
- In this study we describe the application of Zevatars for personalized medicine in pancreatic adenocarcinoma and colorectal cancer.

Humanizing zebrafish

- Currently we cannot use Zevatars to test immunotherapy so therefore we aimed to generate humanized fish
- *cmyb* mutant fish fail to develop definitive hematopoietic cells and therefore provides us a way to completely replace fish system with a human hematopoietic system

Refer to poster number : 2145C



Zevatars determine patient tumor drug response



Figure 1. Differential drug response in cryopreserved liver metastasis biopsy (primary pancreatic cancer): A (a,c,e): Images of implanted tumor (patient ID : 1-01302019-A) on Day 1 of either FOLFIRINOX (4.2mM Fluorouracil, 1mM folinic acid, 0.08mM irinotecan, 0.08mM oxaliplatin) or Gemcitabine (0.1mM) + abraxane (0.005mM) treatment. A(b,d,f): Images of tumors 3 days post treatment. (B) Percentage of fish that survived after drug treatment (C) Change in area of the tumor between Day 1 and Day 3 in each embryo in each group (D) Average change in area between treatment groups. Error bars represent standard deviation (E) Percentage of fish with metastasis after drug treatment.

Zevatars; the future of personalized cancer medicine

Shaila Mudambi Ph.D; Anna Zdunek; Mary Pasquale B.S; Kamden Gray; Seray Er; Ceylan Metin; Paige Burkhardt, B.S; Saumya Kasliwal B.S; Russell Sillmon M.S; Stephen Byers Ph.D; Michael Pishvaian M.D; Ph.D Eric Glasgow Ph.D

Department of Oncology, Lombardi Comprehensive Cancer Centre, Georgetown University, Washington D.C.

Recapitulating patient drug responses in the clinic using Zevatars Hematopoietic chimeras and immune tolerance in cmyb^{+/-} zebrafish embryos (*Refer to oral presentation 104*)



folfirinox. Bars represent maximum-minimum interval range with mean. Interestingly, response varies greatly from

- Zebrafish embryos exhibit immune tolerance when whole kidney marrow is transplanted prior to development of adaptive immune response
- Human peripheral blood was collected and red blood cells (RBCs) were lysed and removed. RBC free blood was then labelled with Dil.
- Stage of development is important for immune tolerance so Dil labelled blood was injected into circulation of wild type zebrafish embryos at different stages of development (2dpf-7dpf)
- Human blood engrafted and fish survived at least until 14 days post injection when injected into 2dpf embryos

Kidney	ubi:zebrabow	Α	B	C	D	
Myelomonocytes	9.36	0.84	6.72	0.41	7.7	0.6
Precursors	2.04	0.37	1.69	0.07	1.99	0.0
Lymphocytes	8.07	0.33	2.73	0.54	7.54	0.0
Erythrocytes	4.73	1.84	4.86	1.84	4.88	0.6
Blood	ubi:zebrabow	Α	B	С	D	E
Myelomonocytes	0.71	0.03	1.75	0.05	0.55	1
Precursors	0.29	0	0.2	0	0.24	0.72
Lymphocytes	2.12	0.05	3.21	0.09	0.55	0.24
Erythrocytes	68.46	1.46	64.37	3.64	6.67	32.4
Spleen	ubi:zebrabow	Α	B	С	D	E
Myelomonocytes	0.52	0.64	0.4	0.31	1.14	0.32
Precursors	0.1	0.05	0.26	0.08	0.15	0.0
Lymphocytes	1.54	0.04	7.7	0.48	2.56	0.0
Erythrocytes	2.21	1.79	9.16	1.43	4.65	0.7

Humanizing zebrafish embryos

Human blood cells persisted in injected fish for at least 15 days. Two larvae injected at 2dpf were dissociated after 12 days and Dye I positive blood cells were observed by flow cytometry



MedStar Georgetown University Hospital